

Raphides in Food – An Unsafe Menu

Naveen Tripathi¹, Chandra Bose¹, Srijoni Basu¹, Nabajit Das¹, Susmit Maitra², Arindam Sikdar¹ and Sukant Khurana^{1*}

¹Indian Institute of Science Education and Research Kolkata (IISER-K), Mohanpur, West Bengal-741246, India

²Kalinga Institute of Industrial Technology (KIIT) University, Bhubaneswar, Odisha-751024, India

Abstract

Calcium oxalate in plant bodies lead to stones in kidney, upon consumption. Calcium oxalate is frequently found in plants in the form of tiny needle like raphides. Out of the 5 types of calcium oxalate crystals, raphides are the predominant ones. Calcium oxalate gets incorporated in our body through plant derived food that contains them; a little amount of them is also synthesized in humans endogenously. Both the sources contribute to kidney problems. Occurrence of calcium oxalate is not limited to higher plants only but also extends to algae, fungi and lichens. Out of all the 3 forms of calcium oxalate, monohydrate form is the one widely reported to cause kidney problems. In this study, we review raphides and explore their possible remediation in order to utilize plants of food and medicinal importance the better way. We also review traditional knowledge of raphide neutralization and point to the methods of removal of calcium oxalate and raphides.

Introduction

Plants are the key sources of food and medicine. Calcium oxalate, a potential causative agent of human kidney stones, can range from 3 to 80% of the dry weight of various plants [1-3]. Calcium oxalate exists in varying crystal shapes and sizes in plants, with raphides being the predominant crystal form [2,4-8]. Deleterious influence of raphides, in addition to promoting kidney stone formation, include irritation to throat, mouth and skin [7,9-18]. Excess presence of raphides, in conjugation with cytotoxic compounds [5,19], can render the food poisonous and is responsible for mentionable fatalities every year [20-22]. Calcium oxalate can contribute up to 70% or 75% of the composition of kidney stones [23,24]. It is present in mono, di and trihydrate forms [25-28]. Monohydrate form is the least soluble and the main constituent of nephroliths [23,24,29]. Monohydrate form readily attaches to the cell surface of the renal tubules [24,30]. In addition to major absorption of calcium oxalate from plant food sources, smaller amount of calcium oxalate can be synthesized endogenously from free oxalic acid or directly from several other biosynthetic precursors [23,31-33]. Liver is the main site of endogenous synthesis of oxalic acid [34]. Inter-conversion rates between monohydrate and dihydrate state governs the attachment of calcium oxalate to renal membrane [24]. Oxalate formation can deplete the human body of divalents, including calcium [35,36]. This depletion of calcium due to calcium oxalate formation aids in osteoporosis [37-39].

In addition to medicines [40,41], dietary control is frequently recommended in the treatment of kidney stones [33,37]. Some proteins that can regulate kidney stone formation [24,42,43] have been identified, which raises a hope for novel drug development. Preclinical studies indicate that few plant extracts can inhibit the growth of calcium oxalate crystals [44], as well as block their adhesion to renal epithelial cells [45,46]. Given that the removal of calcium oxalate from diet can have such a large impact on human health, we review crystals of calcium oxalate in plants, calcium oxalate biosynthesis, and possible ways of calcium oxalate neutralization.

Crystalline Calcium Oxalate; Importance of Raphides

Calcium oxalate is found in several crystal shapes in plants. In some plants presence or absence of a certain kind of crystals can help in taxonomic identification [9] and evolutionary studies [47]. The major kinds of crystals include: needle shaped “raphides” [5-7,48,49], cuboidal or pencil shaped “styloids” [6,7], block shaped aggregates called “crystal sand” [2,11], prism shaped structures [8], and mace-

head or rosette shaped aggregates called “druses” [4]. Raphides are found to have subdivisions in their shapes. According to their shapes, they are classified from type 1 to 4 [9] and a recent study has reported 2 more additional kinds [50]. The calcium oxalate crystals are formed in specialized vacuoles of idioblast cells. The shape of idioblasts, which is under genetic influence, governs the shape of the crystals [51] and is species specific [52]. There can be several kinds of crystals present in the same plant [4,9]. Figure 1 shows the common kinds of Calcium Oxalate crystals [4,17,22,53-59].

Raphides provide plant defense against herbivore [60,61]. Herbivory has been demonstrated to increase the amount of raphides in plants [60]. Frequently raphides are co-present with cysteine proteases and other chemical defense [5]. The needle shaped raphides bruise the lining of the throat, gut, and intestine, while cysteine proteases add to the irritation [15]. Studies using larvae and caterpillars have shown additive effect of the irritants such as proteases and raphides [62]. Raphides can act as needles or syringes to deposit cysteine proteases in herbivore cells [4]. More acidity measurement studies are needed to evaluate the synergistic action of raphides and cysteine proteases on human taste perception [5].

Raphide crystals also play a role in reducing metal toxicity. This suggestion has largely been based on the observation that such crystals can have many other divalents [63-65]. Raphides have also been implicated in light scattering and increasing efficiency of photosynthesis [12,18] but work is required on other crystal types to see if it is exclusively a phenomenon limited to raphide containing plants. Some studies have also suggested a clear role of raphides in

***Corresponding author:** Sukant Khurana, Indian Institute of Science Education and Research Kolkata (IISER-K), Mohanpur, West Bengal-741246, India, Tel: +91-33-2587-3017; Fax: +91-33-2587-3028; E-mail: sukant.khurana@iiserkol.ac.in sukantkhurana@gmail.com

Received February 04, 2015; **Accepted** March 03, 2015; **Published** March 10, 2015

Citation: Tripathi N, Bose C, Basu S, Das N, Maitra S, et al. (2015) Raphides in Food – An Unsafe Menu. J Plant Biochem Physiol 2: 143. doi:10.4172/2329-9029.1000143

Copyright: © 2015 Tripathi N, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

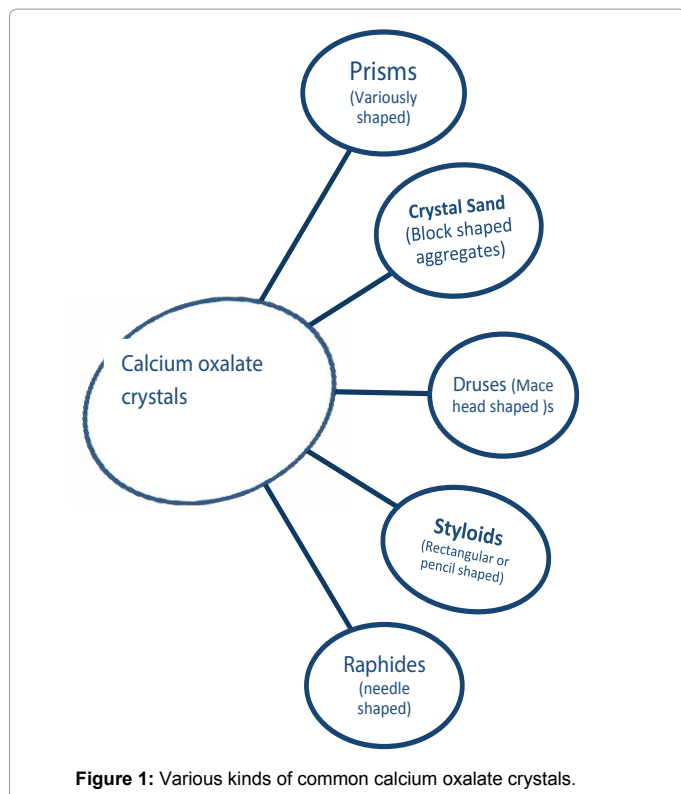


Figure 1: Various kinds of common calcium oxalate crystals.

structural support to the plant [7,66]. So far, other crystal shapes have not been explored much in defense. Several crystal shapes have been implicated in calcium regulation [67-70] and play an important role in ion regulation.

Biosynthesis of Calcium Oxalate

Many recent studies have established that several precursors can be used to synthesize calcium oxalate. The major precursors are ascorbate, glycolate, glyoxylate, oxaloacetate and isocitrate [53,71-75]. Different precursor use by the different parts of a plant remains to be explored. Biosynthetic pathways have been very well reviewed at length in several recent works [53,71-76] so we are merely summarizing the key findings and focusing on amelioration in the next section. The synthesis of ascorbate, a precursor of calcium oxalate takes place in the vacuoles of idioblast cells itself. Concentration of oxalate is higher in idioblasts and calcium is incorporated to make calcium oxalate [18,25]. This higher oxalate in idioblasts results in predominantly the monohydrate form and not the di or trihydrate form [2]. The process of calcium oxalate crystal formation takes place in a matter of hours [77], raising hopes of its possible quick removal. Figure 2 summarizes the common biosynthetic pathways and possible amelioration approaches.

Traditional Remediation of Raphides

Not much modern analysis has been conducted of the traditional approaches, creating a problem of lack of peer-reviewed scientific material on the particular topic of traditional methods. We are bringing forward such information with the hope that modern analysis of these approaches follow soon. Traditional approaches work largely by neutralizing the cysteine proteases. Milk and coconut milk have been used in India to ameliorate the acidity of raphides and they likely work by providing protein substrates to interact with the active poisons present along with raphides. Tamarind [78], lime [79] and various other acidic treatments have also been used because acid can

neutralize toxins. Heating, boiling, frying [80], baking [81], battering, mashing, fermentation and sun drying [82] have also been used for the neutralization of cysteine proteases and release of raphides from idioblasts. Changes in crystal structures of calcium oxalate itself due to these traditional approaches should be explored systematically but this topic has not received attention yet.

Removal of Calcium Oxalate

While several methods neutralize raphides, very few have addressed the neutralization of the long-term damage of kidney stones that increases by eating foods rich in calcium oxalate. There are few traditional approaches that offer hope and few established methods that are yet to gain popularity. Peeling of foods where plant pericarp has high calcium oxalate [83] seems to be an effective and economical way of reducing calcium oxalate. In addition, there are emerging pieces of evidence that raise the hope that several plants with huge drug and food potential can be better utilized by reduction or removal of calcium oxalate. Few approaches, such as treatment with strong acid, sodium bicarbonate and tetracycline treatment have been shown to work but it is not clear as of now that any of these approaches can be applied at mass scale due to unacceptable alteration of food and the associated costs. Some fungi have been shown to be able to degrade calcium oxalate [84] and they offer a hope for future biotech applications to remove calcium oxalate. The *Flammulina velutipes* study shows formation of formic acid but formic acid is extremely unpleasant and additional enzymatic steps might be required to use the enzymes from this fungus [85]. Transgenic tomatoes have been made using OXDC gene from *Flammulina velutipes* like fungi, are port from bacteria also suggests break down of calcium oxalate [86-91]. CoD genes, as observed in *Medicago truncatula*, regulate the formation of calcium oxalate crystals and this gene might also have good biotechnology potential [14]. Heat at the levels used in traditional cooking removes only a small fraction of calcium oxalate, if any, [92] but we suspect that its potential might lie in combination with other approaches. Fast synthesis of calcium oxalate in idioblasts [77] raises hope that an equally fast removal of calcium oxalate might be a possibility. We would expect next few years of research to provide us with signaling and extrinsic factors.

Conclusion

To summarize, we have briefly evaluated the kidney stone formation due to calcium oxalate, explored the various crystalline forms of the compound, with a focus on raphides and looked at its biosynthesis, with the end goal of evaluating novel possible calcium oxalate amelioration methods for better utilization of plants by humans. Modern evaluation of traditional methods has been largely lacking but from knowledge of known ingredients and limited published work, one can infer that milk, heating, boiling, frying, baking, battering, mashing, fermentation

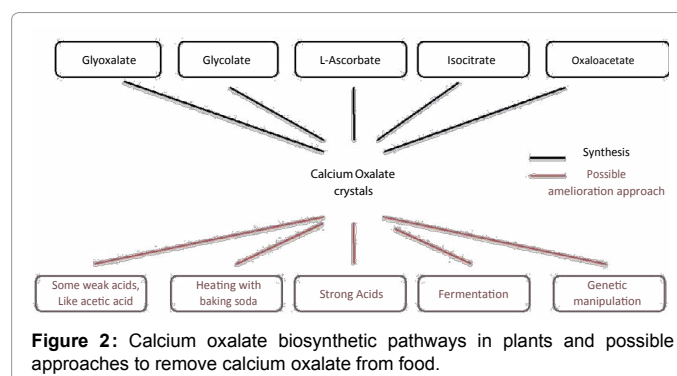


Figure 2: Calcium oxalate biosynthetic pathways in plants and possible approaches to remove calcium oxalate from food.

and sun drying, likely work by neutralization of cysteine proteases or through release of raphides from idioblasts or both. Neutralization of calcium oxalate is a bigger health question than the neutralization of specific crystal form of raphides. A traditional approach of peeling plant pericarp rich in calcium oxalate has beneficial effects. Discovery of fungi and bacteria that can break down calcium oxalate and plant genes that regulate calcium oxalate formation have all offered hope and some initial promising results in genetic engineering to counteract calcium oxalate toxicity. We expect the next few years to witness more chemical, physical and genetic engineering approaches to make food safer for the health of kidneys.

References

- Zindler E (1976) Oxalate biosynthesis in relation to photosynthetic pathway and plant productivity A survey. *Zeitschrift für Pflanzenphysiologie* 80: 1-3.
- Webb MA (1999) Cell-mediated crystallization of calcium oxalate in plants. *Plant Cell* 11: 751-761.
- Ilarslan H, Palmer R, Imsande J, Horner H (1997) Quantitative determination of calcium oxalate and oxalate in developing seeds of soybean (Leguminosae). *Am J Bot* 84: 1042.
- Coté GG (2009) Diversity and distribution of idioblasts producing calcium oxalate crystals in *Dieffenbachia seguine* (Araceae). *Am J Bot* 96: 1245-1254.
- Konno K, Inoue TA, Nakamura M (2014) Synergistic defensive function of raphides and protease through the needle effect. *PLoS One* 9: e91341.
- Crowther A (2009) Morphometric analysis of calcium oxalate raphides and assessment of their taxonomic value for archaeological microfossil studies. In: *Archaeological science under a microscope: studies*, pp. 102-128.
- Prychid CJ, Rudall PJ (1999) Calcium Oxalate Crystals in Monocotyledons: A Review of their Structure and Systematics. *Annals of botany* 84: 725-739.
- Chairiyah N, Harijati N, Mastuti R (2013) Variation of Calcium Oxalate (CaOx) Crystals in Porang (*Amorphophallus muelleri* Blume). *AJPS* 4: 1765-1773.
- Saadi S, Mondal AK (2011) Studies on the calcium oxalate crystals of some selected aroids (Araceae) in Eastern India. *Advances in BioResearch* 2: 134-143.
- Salinas M L, Ogura T, Soffchi L (2001) Irritant contact dermatitis caused by needle-like calcium oxalate crystals, raphides, in *Agave tequilana* among workers in tequila distilleries and agave plantations. *Contact dermatitis* 44: 94-96.
- Franceschi VR, Nakata PA (2005) Calcium oxalate in plants: formation and function. *Annu Rev Plant Biol* 56: 41-71.
- Franceschi VR, Harry THSJS (1980) Calcium Oxalate Crystals in Plants. *Botanical Review* 46: 361-427.
- Holloway WD, Argall ME, Jealous WT (1989) Organic acids and calcium oxalate in tropical root crops. *J Agric Food Chem* 37: 337-341.
- Korth KL, Doege SJ, Park SH, Goggin FL (2006) *Medicago truncatula* mutants demonstrate the role of plant calcium oxalate crystals as an effective defense against chewing insects. *Plant Physiology* 141: 188-195.
- Perera CO, Hallett I (1990) Calcium oxalate crystals: the irritant factor in kiwifruit. *Journal of Food Science* 55: 1066-1069.
- Coelho EG, Amaral A, Ferreira J, Santos DAG, et al. (2007) Calcium oxalate crystals and methyl salicylate as toxic principles of the fresh leaves from *Palicourea longiflora*, an endemic species in the Amazonas state. *Toxicon* 49: 407-409.
- Sunell LA, Healey PL (1979) Distribution of calcium oxalate crystal idioblasts in corms of taro (*Colocasia esculenta*). *Amer J Bot* 66: 1029-1032.
- Nakata PA (2003) Advances in our understanding of calcium oxalate crystal formation and function in plants. *Plant Science* 164: 901-909.
- Yu HL, Zhu FG, Wu H (2011) Study of toxic proteins on raphides from *Pinellia ternata* and *Pinellia pedatisecta* Schott. *China J Tradit Chin Med Pharm*.
- Chen CL, Fang HC, Chou KJ, Wang JS, Chung HM (2001) Acute oxalate nephropathy after ingestion of star fruit. *Am J Kidney Dis* 37: 418-422.
- Bhandari A, Koul S, Sekhon A, Pramanik SK, Chaturvedi LS, et al. (2002) Effects of oxalate on HK-2 cells, a line of proximal tubular epithelial cells from normal human kidney. *J Urol* 168: 253-259.
- Kausch AP, Horner HT (1983) The development of mucilaginous raphide crystal idioblasts in young leaves of *Typha angustifolia* L. (Typhaceae). *Amer J Bot* 70: 691-705.
- Williams HE, Wandzilak TR (1989) Oxalate synthesis, transport and the hyperoxaluric syndromes. *J Urol* 141: 742-749.
- Wesson JA, Worcester EM, Wiessner JH, Mandel NS, Kleinman JG (1998) Control of calcium oxalate crystal structure and cell adherence by urinary macromolecules. *Kidney Int* 53: 952-957.
- Frey A (1981) Crystallography of the two hydrates of crystalline calcium oxalate in plants. *Amer J Bot* 68: 130-141.
- Arnott HJ, Pautard F, Steinfink H (1965) Structure of calcium oxalate monohydrate. *Nature* 208: 1197-1198.
- Gardner GL (1975) Nucleation and crystal growth of calcium oxalate trihydrate. *Journal of Crystal Growth* 30: 158-168.
- Grases F, Millan A, Conte A (1990) Production of calcium oxalate monohydrate, dihydrate or trihydrate. *Urological research* 18: 17-20.
- Evan A, Lingeman J, Coe FL, Worcester E (2006) Randall's plaque: pathogenesis and role in calcium oxalate nephrolithiasis. *Kidney Int* 69: 1313-1318.
- Mandel N1 (1994) Crystal-membrane interaction in kidney stone disease. *J Am Soc Nephrol* 5: S37-45.
- Chai W, Liebman M, Kynast-Gales S, Massey L (2004) Oxalate absorption and endogenous oxalate synthesis from ascorbate in calcium oxalate stone formers and non-stone formers. *Am J Kidney Dis* 44: 1060-1069.
- Holmes RP, Assimos DG (1998) Glyoxylate synthesis, and its modulation and influence on oxalate synthesis. *J Urol* 160: 1617-1624.
- Massey LK, Roman-Smith H, Sutton RA (1993) Effect of dietary oxalate and calcium on urinary oxalate and risk of formation of calcium oxalate kidney stones. *J Am Diet Assoc* 93: 901-906.
- Holmes RP, Knight J, Assimos DG (2007) Origin of urinary oxalate. *AIP Conf Proc, Indianapolis, Indiana USA*.
- Weaver CM, Heaney RP, Nickel KP (1997) Calcium bioavailability from high oxalate vegetables: Chinese vegetables, sweet potatoes and rhubarb. *Journal of food Science* 62: 524-525.
- Heaney RP, Weaver CM, Recker RR (1988) Calcium absorbability from spinach. *Am J Clin Nutr* 47: 707-709.
- Curhan GC, Willett WC, Rimm EB, Stampfer MJ (1993) A prospective study of dietary calcium and other nutrients and the risk of symptomatic kidney stones. *N Engl J Med* 328: 833-838.
- Domrongkitchaiporn S, Ongphiphadhanakul B (2002) Risk of calcium oxalate nephrolithiasis in postmenopausal women supplemented with calcium or combined calcium and estrogen. *Maturitas* 41: 149-156.
- Harvey JA, Zobitz MM, Pak CY (1985) Calcium citrate: reduced propensity for the crystallization of calcium oxalate in urine resulting from induced hypercalciuria of calcium supplementation. *J Clin Endocrinol Metab* 61: 1223-1225.
- Finkelstein VA, Goldfarb DS (2006) Strategies for preventing calcium oxalate stones. *CMAJ* 174: 1407-1409.
- Etinger B, Citron JT, Livermore B, Dolman LI (1988) Chlorthalidone reduces calcium oxalate calculous recurrence but magnesium hydroxide does not. *J Urol* 139: 679-684.
- Wesson JA, Johnson RJ, Mazzali M, Beshensky AM, Stietz S, et al. (2003) Osteopontin is a critical inhibitor of calcium oxalate crystal formation and retention in renal tubules. *J Am Soc Nephrol* 14: 139-147.
- Hoyer JR, Asplin JR, Otvos L (2001) Phosphorylated osteopontin peptides suppress crystallization by inhibiting the growth of calcium oxalate crystals. *Kidney Int* 60: 77-82.
- Joshi VS, Parekh BB, Joshi MJ, Vaidya AB (2005) Herbal extracts of *Tribulus terrestris* and *Bergenia ligulata* inhibit growth of calcium oxalate monohydrate crystals in vitro. *Journal of Crystal Growth* 275: e1403-e1408
- Atmani F, Khan SR (2000) Effects of an extract from *Herniaria hirsuta* on calcium oxalate crystallization in vitro. *BJU Int* 85: 621-625.

46. Atmani F, Farell G, Lieske JC (2004) Extract from *Herniaria hirsuta* coats calcium oxalate monohydrate crystals and blocks their adhesion to renal epithelial cells. J Urol 172: 1510-1514.
47. Arroyo SC, Cutler DF (1984) Evolutionary and taxonomic aspects of the internal morphology in Amaryllidaceae from South America and Southern Africa. Kew Bulletin 39: 467-498.
48. Sakai WS, Hanson M, Jones RC (1972) Raphides with barbs and grooves in *Xanthosoma sagittifolium* (Araceae). Science 178: 314-315.
49. Prychid CJ, Rudall PJ (2000) Distribution of calcium oxalate crystals in monocotyledons. In: Monocots: systematics and evolution, pp. 159-162.
50. Raman V, Horner HT, Khan IA (2014) New and unusual forms of calcium oxalate raphide crystals in the plant kingdom. J Plant Res 127: 721-730.
51. McConn MM, Nakata PA (2002) Calcium oxalate crystal morphology mutants from *Medicago truncatula*. Planta 215: 380-386.
52. Bouropoulos N, Weiner S (2001) Calcium oxalate crystals in tomato and tobacco plants: morphology and in vitro interactions of crystal-associated macromolecules. Chemistry-A European Journal 7: 1881-1888.
53. Kostman TA, Tarlyn NM, Loewus FA, Franceschi VR (2001) Biosynthesis of L-Ascorbic Acid and Conversion of Carbons 1 and 2 of L-Ascorbic Acid to Oxalic Acid Occurs within Individual Calcium Oxalate Crystal Idioblasts. Plant physiology 125: 634-640.
54. Mazen AMA, Zhang D, Franceschi VR (2004) Calcium Oxalate Formation in *Lemna minor*: Physiological and Ultrastructural Aspects of High Capacity Calcium Sequestration. New Phytologist 161: 435-448.
55. Tilton VR (1980) A new type of specialized cell in the gynoecium of *Ornithogalum caudatum* (Liliaceae) with notes on specialized cells in carpels of other taxa. Annals of botany 46: 527-532.
56. Sunell LA, Healey PL (1985) Distribution of calcium oxalate crystal idioblasts in leaves of taro (*Colocasia esculenta*). Amer J Bot 72: 1854-860.
57. Zindler E (1975) On the formation of the pattern of crystal idioblasts in *Canavalia ensiformis* DC. VII. Calcium and oxalate content of the leaves in dependence of calcium nutrition. Zeitschrift für Pflanzenphysiologie 77: 80-85.
58. Ciler M, Dane F (2004) Calcium oxalate crystals in floral organs of *Helianthus annuus* L. and *H. tuberosus* L. (Asteraceae). Acta Biologica Szegediensis 48:19-23.
59. Franceschi VR, Horner HT (1979) Use of *Psychotria punctata* callus in study of calcium oxalate crystal idioblast formation. Zeitschrift für Pflanzenphysiologie 92: 61-75.
60. Molano B (2001) Herbivory and calcium concentrations affect calcium oxalate crystal formation in leaves of *Sida* (Malvaceae). Ann Bot 88: 387-391.
61. Ward D, Spiegel M, Saltz D (1997) Gazelle herbivory and interpopulation differences in calcium oxalate content of leaves of a desert lily. Journal of Chemical Ecology 23: 333-346.
62. Lucas J, Lewis SA (2003) Kiwi fruit allergy: a review. Pediatric allergy and Immunology 14: 420-428.
63. Franceschi VR, Schueren AM (1986) Incorporation of strontium into plant calcium oxalate crystals. Protoplasma 130: 199-205.
64. Yang YY, Jung JY, Song WY, Suh HS, Lee Y (2000) Identification of rice varieties with high tolerance or sensitivity to lead and characterization of the mechanism of tolerance. Plant Physiol 124: 1019-1026.
65. Ma JF, Ryan PR, Delhaize E (2001) Aluminium tolerance in plants and the complexing role of organic acids. Trends Plant Sci 6: 273-278.
66. Lane BG1 (1994) Oxalate, germin, and the extracellular matrix of higher plants. FASEB J 8: 294-301.
67. Franceschi VR, Loewus FA (1995) Oxalate biosynthesis and function in plants and fungi. In: Calcium oxalate in biological, pp. 113-130.
68. Borchert R (1986) Calcium acetate induces calcium uptake and formation of calcium-oxalate crystals in isolated leaflets of *Gleditsia triacanthos* L. Planta 168: 571-578.
69. Borchert R (1985) Calcium-induced patterns of calcium-oxalate crystals in isolated leaflets of *Gleditsia triacanthos* L. and *Albizia julibrissin* Durazz. Planta 165: 301-310.
70. Pennisi SV, McConnell DB (2001) Inducible calcium sinks and preferential calcium allocation in leaf primordia of *Dracaena sanderiana* Hort. Sander ex MT Mast. (Dracaenaceae). Hortscience 36: 1187-1191.
71. Nuss RF, Loewus FA (1978) Further Studies on Oxalic Acid Biosynthesis in Oxalate-accumulating Plants. Plant Physiol 61: 590-592.
72. Yu L, Jiang J, Zhang C, Jiang L, Ye N, et al. (2010) Glyoxylate rather than ascorbate is an efficient precursor for oxalate biosynthesis in rice. J Exp Bot 61: 1625-1634.
73. Debolt S, Melino V, Ford CM (2007) Ascorbate as a biosynthetic precursor in plants. Ann Bot 99: 3-8.
74. Keates SE, Tarlyn NM, Loewus FA, Franceschi VR (2000) L-Ascorbic acid and L-galactose are sources for oxalic acid and calcium oxalate in *Pistia stratiotes*. Phytochemistry 53: 433-440.
75. Fujii N, Watanabe M, Watanabe Y (1993) Fate of oxalate biosynthesis from glycolate and ascorbic acid in spinach leaves. Soil Science and Plant Nutrition 39: 627-634.
76. Nakata PA (2012) Plant calcium oxalate crystal formation, function, and its impact on human health. Frontiers in biology 7: 254-266.
77. Franceschi VR (1989) Calcium oxalate formation is a rapid and reversible process in *Lemna minor* L. Protoplasma 148: 130-137.
78. Hallson PC, Rose GA (1985) The additive effects of magnesium and tartrate upon inhibition of calcium oxalate crystal formation in whole urine. Urolithiasis and related clinical research 847-850.
79. Penniston KL, Nakada SY, Holmes RP, Assimos DG (2008) Quantitative assessment of citric acid in lemon juice, lime juice, and commercially-available fruit juice products. J Endourol 22: 567-570.
80. Iwuoha CI, Kalu FA (1995) Calcium oxalate and physico-chemical properties of cocoyam (*Colocasia esculenta* and *Xanthosoma sagittifolium*) tuber flours as affected by processing. Food chemistry 54 : 61-66.
81. Oscarsson KV, Savage GP (2007) Composition and availability of soluble and insoluble oxalates in raw and cooked taro (*Colocasia esculenta* var. Schott) leaves. Food chemistry 101: 559-562.
82. Agwunobi LN, Angwukam PO, Cora OO, Isika MA (2002) Studies on the use of *Colocasia esculenta* (taro cocoyam) in the diets of weaned pigs. Trop Anim Health Prod 34: 241-247.
83. Rassam M, Laing W (2005) Variation in ascorbic acid and oxalate levels in the fruit of *Actinidia chinensis* tissues and genotypes. J Agric Food Chem 53: 2322-2326.
84. Mattos J, Santos TB, Alemanno L (2007) Involvement of calcium oxalate degradation during programmed cell death in *Theobroma cacao* tissues triggered by the hemibiotrophic fungus *Moniliophthora perniciosa*. Plant Science 173: 106-117.
85. Chakraborty N, Ghosh R, Ghosh S, Narula K (2013) Reduction of oxalate levels in tomato fruit and consequent metabolic remodeling following overexpression of a fungal oxalate decarboxylase. Plant Physiology 162: 364-378.
86. Allison MJ, Daniel SL, Cornick NA (1995) Oxalate-degrading bacteria. Calcium oxalate in biological.
87. Allison MJ, Cook HM, Milne DB, Gallagher S, Clayman RV (1986) Oxalate degradation by gastrointestinal bacteria from humans. J Nutr 116: 455-460.
88. Allison MJ, Cook HM (1981) Oxalate degradation by microbes of the large bowel of herbivores: the effect of dietary oxalate. Science 212: 675-676.
89. Sidhu H, Allison MJ (2013) Compositions and methods for treating or preventing oxalate-related disease, US Patent Office.
90. Kaur J, Rajkhowa R, Afrin T, Tsuzuki T, Wang X (2014) Facts and myths of antibacterial properties of silk. Biopolymers 101: 237-245.
91. Hoppe B, Beck B, Gatter N, von Unruh G, Tischer A, et al. (2006) *Oxalobacter formigenes*: a potential tool for the treatment of primary hyperoxaluria type 1. Kidney Int 70: 1305-1311.
92. Savage GP, Vanhanen L, Mason SM (2000) Effect of cooking on the soluble and insoluble oxalate content of some New Zealand foods. Journal of Food Composition and Analysis 13: 201-206.