

Phytosomes: A Modernistic Approach for Novel Herbal Drug Delivery - Enhancing Bioavailability and Revealing Endless Frontier of Phytopharmaceuticals

Sudhir Kumar¹, Ashish Baldi^{2*} and Dinesh Kumar Sharma³

¹Department of Pharmacognosy, ISF College of Pharmacy, Moga, Punjab, India; ²Department of Pharmaceutical Sciences and Technology, Maharaja Ranjit Singh Technical University, Bathinda, Punjab, India; ³Institute of Pharmaceutical Sciences, Sanskriti University, Mathura, Uttar Pradesh, India

ABSTRACT

Phytopharmaceuticals are healing the world from millions and billions of years even though their clinical validation is questioned by virtue of their impediments like low lipid solubility, poor stability, large size moiety and needless metabolism in gut. Phytosome technology has emerged as committed and promising targeting novel drug delivery with improved efficacy, quality and target ability of active plant constituents. Novel herbal formulation techniques have assured the researchers to deliver the plant based secondary metabolites to their systemic targets. This review highlights the unique properties of phytophospholipid complex along with their application in the novel natural drug delivery. Various methods employed in phytosomal preparation and characterization along with the phytosomal advantages over conventional herbal extracts is described in the present review. The prospectus of phytosome technique can suggest new directions and endless frontier as novel drug regimen.

Keywords: NDDS; Phospholipids; Phytosome; Bioavailability

Abbreviations: Polydisparsity index (PDI); Thin layer chromatography (TLC); Novel drug delivery system (NDDS)

INTRODUCTION

Phytomedicine are accepted as natural healers in the whole world and were even used by lords in divine era. Advanced herbal drug delivery system such as phytosomes has demarcated the undefined bioavailability of lipid insoluble secondary metabolites [1]. Lipid insoluble herbal extracts can be redesigned into lipid compatible therapeutic candidate by chemically assimilating herbal extracts into phospholipids in specific ratio [2]. Cellular vesicles produced by phytosome technique prevent destruction of water soluble phytoconstituents such as terpenoids, glycosides, flavonoids and phenolics by gastric secretion and microflora of gut [3]. Numerous advantages of phytosome such as hepatoprotective action, reduced dose to produce desired therapeutic effect [2] improved stability due to chemical linkage, ability to permeate through skin [4,5] systematic targeting to transit from hydrophilic to lipophilic environment has revolutionized the phytomedicine industry. Nano size of Phytosomes has resolved the obstacles originating due to poor solubility and permeability of large size hydrophilic phytoconstituents across biological membranes [6]. The present

study overlooked on phospholipids based drug administration can enlighten modern pathways in the formulation of novel herbal dosage forms.

Phytosomes prospects

Phytosomes represents advanced herbal drug technology that offers defined bioavailability of plant drugs over the herbal extract. Figure 1 represents the phytosome as lipid complex between a natural moieties and soy lecithin compounds such as phosphotidylcholine, phosphotidylethanolamine and phosphotidylserine in a stoichiometric ratio [6]. Reduced particle size, upturned rate of dissolution and absorption claims enhanced *in vivo* performance of the herbal extracts [7]. Structural elucidation reveals that the chemical interaction of phospholipids and herbal substrate involves the formation of hydrogen bonds between the polar side front of phospholipids and the polar functional group of the secondary metabolites generating a specific pattern [8]. Phytosomes show chemical reaction in solvents such as acetone, dioxane, methylene chloride, hexane and ethyl acetate and assumes a micelles shape

Correspondence to: Dr. Ashish Baldi, Professor and Dean, Department of Pharmaceutical Sciences and Technology, Maharaja Ranjit Singh Technical University, Bathinda, Punjab, India, Telephone: 8968423848; E-mail: baldiashish@gmail.com

Received: May 20, 2020; **Accepted:** May 29, 2020; **Published:** June 05, 2020

Citation: Kumar S, Baldi A, Sharma DK (2020) Phytosomes: A Modernistic Approach for Novel Herbal Drug Delivery - Enhancing Bioavailability and Revealing Endless Frontier of Phytopharmaceuticals. J Develop Drugs 9:2. doi: 10.4172/2329-6631. 1000195

Copyright: © 2020 Kumar S, et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

network in water appearing liposomal-like cellular configuration in which the active polar moiety is docked to phospholipids behaving as integral part of the cell membrane [9,10]. There are numerous factors which govern the characteristic originality of phytosomes in physical state such as physical size, membrane permeability, entrapment ratio, chemical constitution as well as the quantity and purity of the precursor starting chemical ingredients [11].

MATERIALS AND METHODS

Different methods of preparation

Non-conventional methods are usually employed in construction of phytosome complexes. Modernistic herbal complexes are formed by reaction between equimolar mixture of natural or synthetic phospholipid and active constituents or herbal extract in aprotic organic solvents [12,13]. Common stages in formulation of Phytosomes are depicted in Figures 2 and 3. Various methods of preparation are as follows:

Anti-solvent precipitation process: Specific amount of herbal extract and phospholipids is refluxed with 20 ml of organic solvents such as acetone at specific experimental conditions below 50°C for 2-3 h. The reaction mixture is concentrated to minimum volume up to 10 ml and then on addition of solvent with low polarity such as n-hexane with stirring, precipitates are obtained. Filtered precipitates are stored in desiccators. The dried precipitates are pulverized and powdered complex are stored in dark amber colored glass bottle at room temperature [14].

Rotary evaporation process: Specific weight of herbal extract and phospholipids were mixed in 30 ml water miscible organic solvent such as acetone in round bottom glass container followed by stirring for 2 hours at a temperature less than 50°C in rota evaporator. Antisolvent such as n-hexane can be added to thin film which is obtained after uninterrupted stirring using a stirrer [15]. Precipitate of phytosomes so obtained can be stored in amber colored glass container at controlled temperature under specified humidity.

Solvent ether-injection process: This technique involves reaction of lipids dissolved in organic solvent with herbal extracts in aqueous phase. Phospholipids solubilised in diethyl ether are slowly injected drop wise in an aqueous solution of the phytoconstituents which is to be encapsulated. It results in the formation of cellular vesicles on subsequent solvent removal, leading to complex formation [16]. Structure of phytosomes depends upon concentration, amphiphiles in mono state are produced when the concentration is less, but variety of structures with different shapes *viz.* round, cylindrical, disc and cubic or hexagonal vesicles may be formed on increasing the concentration.

Novel methods: Novel methods for the phospholipid complexation include supercritical fluids which include gas solvent technique, compressed solvent process and supercritical solvent method [15].

Superiority of phytosomes over herbal extracts

Phytosomal technique was proposed as a drug carrier since 1989. Numerous phytosomal advantages as presented in fig 2, explains the increased validity and relevance of herbal extracts in bioevaluation and *in-vitro* studies [17,18]. They possess better metabolic profile than old conventional herbal extracts [19]. Various advantages of phytosomal technology are:-

Enhanced bioavailability: Phytospholipid complex allows penetration of hydrophilic herbal extract from intestinal lumen for better absorption. There is remarkable improvement in the bioavailability of secondary metabolites on complexation with lipophilic head of phospholipids [20].

Safe and synergistic: Additives used in the phytosome formulation are approved ensuring it as safe and secure concept as phosphatidylcholine used in complexation is essential part of cell membrane. Synergistic effect has been observed on complexation with hepatoprotective drugs as phosphatidylcholine itself possess hepatoprotective action. Synergistic advantages are vividly seen in protecting the skin against exogenous or endogenous toxin in stressful environmental conditions. Phytosome concept assures increased duration of action at low dose with low risk profile due to upgraded absorption of the active constituent [21].

Low hazard profile: Toxicological outcome are negligible as seen in reported data moreover there is only small scale production.

Cost effectiveness: This technology provides economical delivery of phytoconstituents. Cellular vesicular system is submissive and is accessible for further instant development. It is comparatively easy to produce as no complicated technical investment is required and no complex practical speculation is essential for the manufacture of phytosomes [22].

Transdermal drug delivery: Herbal phytosomes can be also utilized to improve the diffusion of drug through the skin in transdermal drug delivery as they act as foretop for the delivery of huge assorted group of drugs such as peptides and protein [23].

Biodegradable: Phosphatidylcholine utilized in phytosome formulation act as a carrier transporter and are integral portion of cell membrane so there is no obstacle with drug frame-up during formulation manufacturing [24].

High entrapment effectiveness: Drug entrapment effectiveness is very high furthermore no toxic metabolites are produced. Moreover the biomarker itself forms nano cellular vesicles on bonding with soya lipids and drug release can be predetermined [25].

Optimization and characterization techniques

Optimization and characterization of phytosomes can be carried out by estimating drug release, membrane permeability, vesicle shape and size distribution, percentage drug capture, entrapped concentration, chemical composition, quantity of material [18]. Stability of phospholipids complex depends upon various factors like drug to phospholipids ratio, experimental duration of time, temperature, solvent evaporation and type of drying method employed [26]. Physical parameters can be optimized statistically or by multiple evaluation techniques which can authenticate and validate its characteristics properties.

Visualization: Morphological studies are mostly used for observation of the particle size, entrapment behavior, surface attributes, and identifying the probable proportion of impurities on the particle surfaces [27]. High resolution images of phospholipid complexes can be resolved and visualized by scanning electron microscopy techniques. In SEM, posterior scattered electrons describe the atomic state and secondary electrons provide surface topography. However internal structure, crystallographic features, elemental composition of the phytosome complex can be evaluated through X-rays and TEM analysis [1].

Entrapment efficiency: The entrapment efficiency of herbal extract can be estimated by performing centrifugation. Centrifugation of solution containing weighed amount of phytophospholipid complex equivalent to quantity of encapsulated herbal extract in phosphate buffer of pH 6.8 can be carried out for 30 minutes at 5000 rpm. Stirred contents are allowed to remain undisturbed for one to two hours and finally absorbance of supernatant liquid collected by decantation is estimated by UV or HPLC [28]. The drug entrapment percentage (%) is calculated as:

Drug entrapment (%) = Actual amount determined/Theoretical amount present.

Crystallinity and polymorphism: X-ray diffraction studies are majorly accepted for characterization of crystallinity and polymorphism in phospholipid complex. DSC report explain the endothermic peaks, determination of transition temperature of the

vesicular lipid complex, new peaks, peak shape, peak temperature, melting points and relative peak area [29]. The complete absence or reduction in the intensity of large diffraction peaks corresponding to its crystalline drug in phospholipid complex and polymorphism is characterized by XRD.

Vesicle stability: Particle size, polydispersity index (PDI) and zeta potential describes the vesicle stability. PDI determines width of a particle size distribution while zeta potential quantify the surface potential of cellular vesicles. Size of complex may vary from 50 nm to a few hundred μm but phytosomes with PDI value <0.5 are stable and indicating that the sample does have narrow size distribution and small particles in complex do not sediment in contrast to aggregates particles that may slowly settle down and sediment whereas samples with zeta potential $> \pm 30$ mV are evaluated as stable complex [2,8].

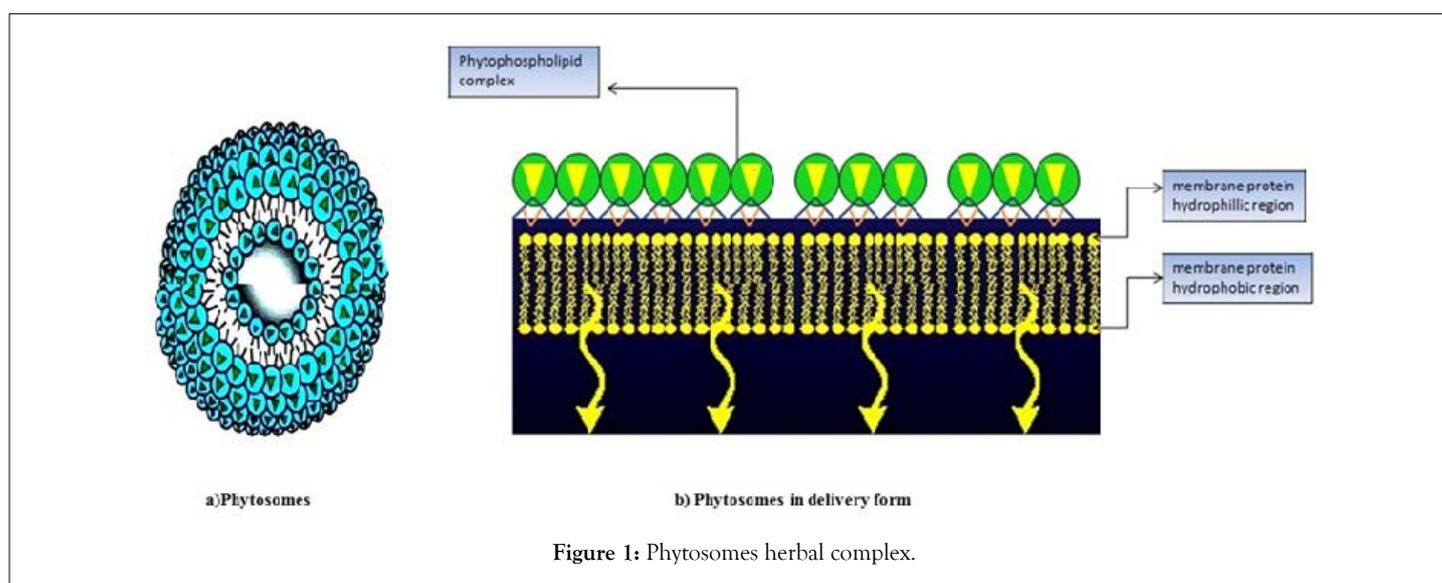


Figure 1: Phytosomes herbal complex.

Table 1: Recipients in phytosomes development.

Chemical	Examples	Uses	References
Phospholipid	Soyaphosphatidylcholine, egg phosphatidylcholine, dipalmitoylphosphatidylcholine, distearylphosphatidylcholine	Cellular vesicles generating component	[39,40]
Solvent	Dioxane, acetone, methylene chloride	Aprotic solvent	[41]
Non-solvent	Aliphatic hydrocarbons or n-hexane	Complex precipitation	[41]
Alcohols	Ethanol, methanol	As a solvent	[42,43, 44]
Color and Dyes	Rhodamine 6G, DHPE-rhodamine, fluorescein, 6 carboxy fluorescence	Cono focal scanning laser microscopy study	[45,46]
Buffering agent	Saline phosphate buffer (pH 6.5) 7 % v/v ethanol tris buffer (pH 6.5)	Hydrating medium	[44,43,47]

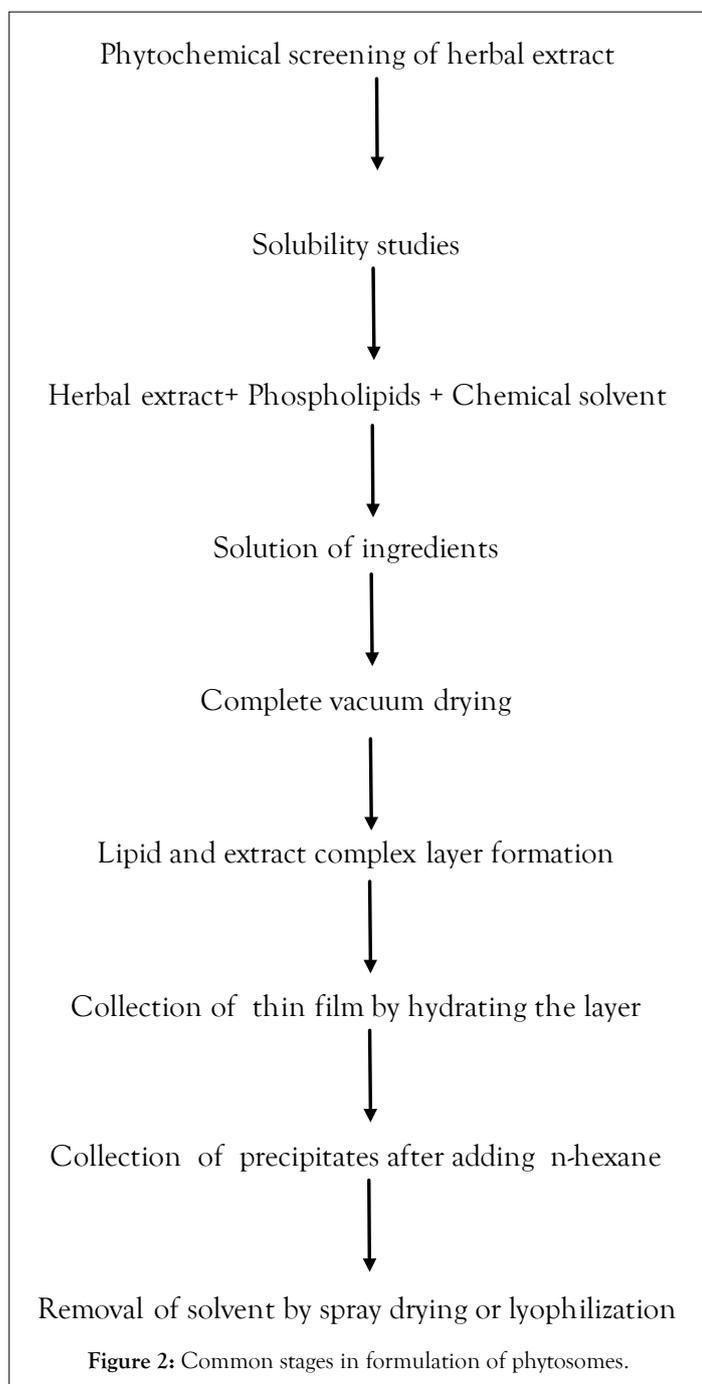
Table 2: Patents of phytosomal dosage form.

Patent no.	Patent detail	References
EP/1844785	Phytosomes of olive fruits	[51]
EP1813280	Phytosomes containing <i>Ginkgo biloba</i> derivatives	[52,53]
EP1640041	Phytosome containing cosmetic and dermatological preparation	[54]
US 7691422	Oral phytosome for the treatment of cellulite	[55]
US/2007/001 5698	Thymosin $\beta 4$ phytosome for skin and wound repair	[21]
WO/2004/045 541	Soluble isoflavone phytosomal compositions	[56]
EP1214084	Antioxidant phytosome formulation	[38]
EP2228062 A1	Phospholipid-curcumin complex and piperine formulation	[57,58]
EPO283713	Saponins with phospholipid phytosome	[59,60]

Table 3: Commercial registered phytosome products.

Biological Source	Synonym	Phytoconstituents Complexes	Commercial product	Therapeutic indications	References
<i>Aesculus hippocastanum</i>	Horse Chestnut	Saponins	Escin β sitosterol Phytosome™	Anti-oedema and vasoactive properties	[61]
<i>Ammi visnaga</i>	Khella	Visnadine	Visnadex™	Improve microcirculation	[61]
<i>Centella asiatica</i>	Brahmi	Asiatic acid, madecassic acid	Centella triterpenoid Phytosome™	Skin disorders, antiulcer, wound healing, anti-hair loss agent	[61]
<i>Citrus aurantium</i>	Bitter orange	Naringenin	Naringenin Phytosome™	Antioxidant	[30,61]
<i>Crataegus oxyacanthoides</i>	Hawthorn	Hyperin, quercitin	Hawthorn Phytosome™	Nutraceutical, cardioprotective and antihypertensive	[61]
<i>Cucurbita pepo</i>	Pumpkin	Tocopherols, steroids, carotenoids	Cucurbita Phytosome™	Anti-inflammatory, Benign prostatic hyperplasia	[61,62]
<i>Fraxinus ornus</i>	Flowering ash	Esculoside (Esculin)	Esculoside Phytosome™	Vasoactive, anticellulite	[61,62]
<i>Ginkgo biloba</i>	Maiden hair Tree	Ginkgo flavonoids, Gingoic acids, ginkgo flavon glucosides, ginkgolides, bilobalide	Gingkoselect Phytosome™	Cognition enhancer	[22,61]
			Ginkgo bilobaterpene	Raynaud's disease,	
			Phytosome™ Ginkgo	Antiageing, anti-asthmatic	
<i>Glycine max</i>	Soya	Genistein and daidzein	Soyselec Phytosome™	Anti-amnesic, antidepressant,	[61]
				Cardioprotective, dermatitis, Anti-Inflammatory	
<i>Glycyrrhiza glabra</i>	Mulethi	Glycyrrhetic acid	Glycyrrhetic acid Phytosome™	Anti-inflammatory, used in dermatitis	
<i>Melilotus officinalis</i>	Sweet clover	Melilotoside, flavanoids and terpenoids	Lymphaselect™	Anti-inflammatory, in oedema, thrombophlebitis	[22]
<i>Olea europaea</i>	Olive tree	Verbascoside, tyrosol, hydroxytyrosol	Oleaselect Phytosome™	Antioxidant, antihyperlipidemic, anticancer and anti-inflammatory.	[22]
<i>Panax ginseng</i>	Ginseng	Ginsenosides	Ginseng Phytosome™	Nutraceutical, immunomodulatory	[61]
<i>Panicum miliaceum</i>	Millet	Mineral salts, vitamins unsaturated fatty acids, aminoacids	Millet Phytosome™	Antistress, beauty food for skin, nails and hairs	[61]
<i>Curcuma longa</i>	Turmeric	Curcumin	Curcumin Phytosome™ Curcuvet®(Meriva®)	Anti-inflammatory, osteoarthritis, anticancer	[63,64]
<i>Camellia sinensis</i>	Tea	Epigallocatechin, epicatechin-3-O-gallate, epigallo catechin-3-O-gallate, catechin	Green tea Phytosome™	Nutraceutical, anticancer, Antioxidant, atherosclerosis, hepatoprotective, antidiabetic, anti-inflammatory	[61,65]
<i>Echniacea angustifolia</i>	Cone flower	Echinacosides and Inulin	Echniacea Phytosome™	Nutraceutical, immunomodulatory	[66]
<i>Pinus maritime</i>	Pine	Procyanidins	Pycnogenol Phytosome™	Anti-inflammatory, antiwrinkle, Antiallergic	[67]
<i>Radix puerariae</i>	Kudzu root	Puerarin	Puerarin and phospholipid complex	Anti-inflammatory, cardiovascular diseases	[68,69]
<i>Ruscusa culeatus</i>	Butchers broom	Ruscogenin, neoruscogenin,	Ruscogenin Phytosome™	Anti-inflammatory, anti-ageing, Sunscreen agent	[61]
<i>Santalum album</i>	Sandal wood	Ximenynic acid, ethyl ximenynate	Ximilene and Ximenoil Phytosome™	Improve microcirculation	[61]
<i>Serenoa repens</i>	Saw palmetto	Phytosterols	Phytosterols	Noncancerous prostate Enlargement	[22]
<i>Silybium maranium</i>	Milk Thistle	Silybin, silycristin, isosilbin	Silybin Phytosome™ (Siliphos®)	Hepatoprotective, hepatitis, cirrhosis and inflammation	[61,63,70]
<i>Swertia alternifolia</i>	Swertia	Xanthones 26	Swertia Phytosome™	Antidiabetic	[71]
<i>Syzygium cumini</i>	Jamun	Tannins	Madeglucyl Phytosome™	Antihyperglycemic, anti-inflammatory, antioxidant	[61]

<i>Terminalia serica</i>	Silver cluster	Sericoside	Sericoside	Anti-aging, skin restructuring	[61,72]
<i>Vaccinium angustifolium</i>	Blue berry	Anthocyanosidestocotrienol complex,	VitaBlue Phytosome™	Anti-oxidant, improves vision, memory enhancer	[61]
<i>Vaccinium myrtillus</i>	Bilberry	Anthocyanosides	Mirtoselect Phytosome™	Antioxidants, antiinflammatory, vasoprotective	[22,73]
<i>Vitis vinifera</i>	Grapes	Resveratrol, catechin, quercetin, epicatechin,	Biovin and leucoselect	Cardioprotective, systemic	[22,61]
			Masquiliers Phytosome™	antioxidant, nutraceutical	
<i>Zanthoxylum bungeanum</i>	Tumburu	Hydroxy-a-sanshool	Zanthalene Phytosome™	Soothing and Anti-reddening	[61]



Spectroscopic evaluation

The spectroscopic analysis of the designed phytosome can be validated by comparing spectrum of complex formed with soya lipid with the herbal extract. Spectroscopic analysis confirms the formation of lipid compatible complex [30,31]. Phytosphospholipid complexation and molecular bonding interactions can be analyzed

by employing different spectroscopic techniques like ¹HNMR, ¹³CNMR, ³¹PNMR, and IR spectroscopy as follows:

¹HNMR: Complex formation between the active phytoconstituents and phospholipid can be confirmed by NMR spectra. Marked alteration in signals emerging in ¹HNMR evolving from atoms involved in the formation of complex describes the chemical bonding. The broad signals belonging to phytoconstituents and phospholipids and chemical shift corresponding to the N-methyl of choline confirm the formation of phytosomes [1,32].

¹³CNMR: The shift in signals corresponding to the fatty acids chains can be interpreted in ¹³CNMR of the phytoconstituents and the stoichiometric complex of phosphatidylcholine and herbal extract [1,5].

FTIR: Comparative spectrum study of phytosomes complex in solid form after lyophilization with that of micro dispersion in water at different times as seen in FTIR [6].

Retention Time: Retention factor in thin layer chromatography (TLC) is a simple method for characterization of phytosome. Retention time or retention factor varies for the phytoconstituents and phospholipids. The difference in retention time or retention factor indicates the generation of a new complex termed as phytosome [33-40].

Design of phytosomal formulation

Design of phytosomal formulation depends upon mode of delivery of the product either by topical or oral route [41-45]. With the advancement in pharmaceutical sciences, phytosomes finds applications in formulating various dosage forms as pharmaceuticals, nutraceuticals and cosmeceuticals. Several pharmaceutical companies involved in production of phytosome products are Indena, Jamieson natural resources, Thorne research, Natural factors and Nature herb [46-50]. Significant aspects as considered are:

Screening of herbal extractive: Phytosomes involve chemical interaction between hydrophilic herbals and lipophilic phospholipids moieties so aqueous extractive can be utilized for optimum bonding. Herbal extracts can be phytochemically screened for terpenoids, tannins, flavonoids as they show optimum bonding with phospholipids. Moreover drug release from phytosomal complex can be defined by optimizing various inherent properties of phytoconstituents. Fundamental inherent characteristics such as hydrophilicity, lipophilicity, cellular permeability, biodegradability, release characteristics and size of phytosome complex should be considered while selecting the phytoconstituents [51-55].

Selection of additives: Additives used in phytosomal formulation can be selected on the basis of dosage form and its mode of drug delivery. Phospholipids, solvent, dyes, buffering agents used in formulation of phytosomes are mentioned in Table 1.

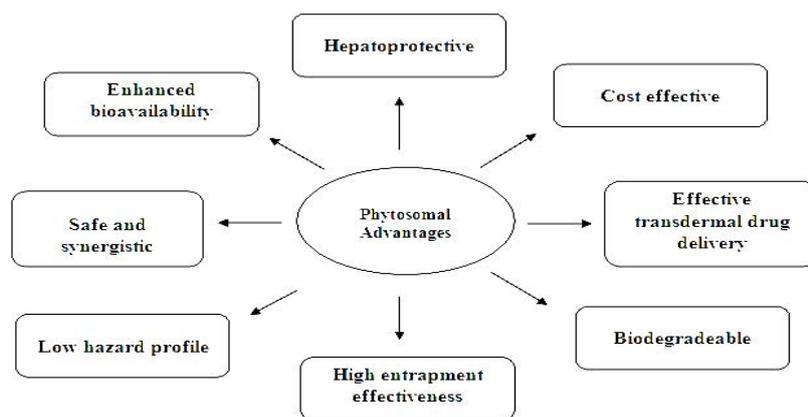


Figure 3: Advantage of phytosomes.

Selection of dosage form: Phytosome complex can be formulated into oral and topical dosage form however relevant dosage form required for drug release can be selected on the basis on effectiveness and efficiency of biomarker compounds. Solubility is an important criterion to determine the stability of complex and selection of solvent can be done according to phytoconstituents whether hydrophilic or lipophilic [56-60]. Suspension formulated by dispersing phytosome in biocompatible edible or semi-synthetic oily vehicles [61-67]. Phytosomal capsule of herbal extract and lipid complex can be formulated manually or by auto filling method without compressing the complex. Phytosome complex can be chemically incorporated into prepared emulsion or ointment base. Different topical phytosomal formulations such as solution, emulsion, lotion are proposed after optimizing the solubility of ingredients and other pharmacokinetic parameters [68-73].

RESULTS AND DISCUSSION

Phytosomes and liposomes both involve chemical interaction of active compounds and phospholipids but there is difference in chemical bonding, bioavailability and molecular arrangement. Phytosome are vesicular system in which each phytoconstituent molecule is surrounded and bounded by phospholipid molecule through chemical hydrogen bonds assuring enhanced bioavailability and permeability through cellular membranes, whereas liposomes are aggregate of many phospholipid molecules that encloses each chemical entity without specifically bonding to them possessing lesser bioavailability as compared to phytosome. In liposome, hundreds and thousands of phosphatidylcholine molecules surround the water soluble molecule but in phytosomes phospholipid and phytoconstituent interact in 1:1 or 2:1 ratio depending on chemical entity. Numerous patents of phytosomes have been granted in past 10 years mentioned in Table 2 explaining the future of phytopharmaceuticals. Phytosomes as novel herbal vesicular drug delivery systems assure to deliver the drug through the pathway channelizing the active phytoentity to the desired site of action. Cellular shaped nano herbal particles assure the mode of delivering the therapeutic agent to the tissues of need improving the therapeutic efficacy and reducing the other allied effects. Phytospholipid complexation technique has evolved as an important revolution for herbal medicines which were not able to demonstrate a remarkable effect at *in vivo* and *in vitro* level. Commercially available registered phytosomal products by various manufactures as summarized in Table 3 claims safe and synergistic therapeutic benefits in various pathological conditions. Hence

phytosome technology has emerged as safe and promising future of phytomedicine.

CONCLUSION

Phytospholipid complex technique has evolved as advanced frontier aspect in defining systemic absorption of herbal extracts. This technique has effectively resolved the irrational queries of plant based drugs. Aimed with predetermined lipid penetration at higher concentration with sustained and constant therapeutic levels in plasma, allows more quantity of active biomarkers to reach at desired site of action. However, it needs more emphasis including complete characterization with optimization, quantitative and qualitative exploration the lipid based system and its impact in different pathological states. However these novel complexes can act as reliable candidates for improved drug dosage therapy. As seen phytomedicines have been healing the world long back time and presently have major acceptance. Initially the phytosome complexes as used in cosmetics, but they are now widely utilized in therapies such as antioxidants, cardioprotective, antiinflammatory, liver protective, antitumor and anti-cancer. With this emerging formulation tool phytosomes has re-explained the relevance of herbals in modern drug targeting approaches.

INTEREST CONFLICT

No conflict of interest by authors.

ACKNOWLEDGEMENT

We are thankful to Chairman, Sh. Parveen Garg; Director, Prof. GD Gupta and Pharmacognosy Department, ISF College of Pharmacy for providing support for this study.

REFERENCES

1. Kumari P, Singh N, Cheriyan P, Neelam. Phytosomes: A novel approach for phytomedicine. *Int J Inst Pharm Life Sci.* 2011;1:89-100.
2. Saha S, Sharma A, Saikia P, Chakrabarty T. Phytosomes: A brief overview. *Sch Acad J Pharm.* 2013;2:12-20.
3. Srikanth V, Laxmaiah CH, Gopi SB, Naveen P, Chiranjeeb B. Phytosome: A novel drug delivery system for improving bioavailability of herbal medicine. *Int J Pharm Res Dev.* 2011;3(6):175-84.
4. Saraf S, Kaur CD. Phytoconstituents as photoprotective novel cosmetic formulations. *Pharmaco Rev.* 2010;4:1-11.
5. Amin T, Bhat S. A review on phytosomes technology as a novel

- approaches to improve the bioavailability of nutraceuticals. *Int J Adv Res and Tech.* 2012;1:1-15.
6. Patel A, Tanwar Y, Rakesh S, Patel P. Phytosome: Phytolipid drug delivery system for improving bioavailability of herbal drugs. *J Pharm Sci and Bio Sci Res.* 2013;3:51-57.
 7. Semalty A, Semalty M, Rawat MS, Franceschi F. Supramolecular phospholipids-polyphenolics interactions: The Phytosome® strategy to improve the bioavailability of phytochemicals. *Fitoter.* 2010;81:306-14.
 8. Tripathy S, Patel D, Baro L, Nair S. A review on phytosomes their characterization, advancement and potential for transdermal application. *J Drug Del and Thera.* 2013;3:147-152.
 9. Marena C, Lampertico M, Maria JM. Preliminary clinical development of silybin: A new complex of silybin in toxic liver disorders. *Planta Med.* 1991;57:124-5.
 10. Sharma S, Roy RK. Phytosomes: An emerging technology. *Int J Pharm Res Dev.* 2010;2:1-7.
 11. Patel J, Patel R, Patel N. An overview of phytosomes as an advanced herbal drug delivery system. *Asian J Pharm.* 2009;4 (6):363-371.
 12. Magistretti JM, Bombardelli E. Pharmaceutical compositions containing flavanolignans and phospholipid active principles U.S. 1987;p:EPO209037.
 13. Karimi N, Ghanbarzadeh B, Hamishehkar H, Pezeshki A, Mostafayi H. Phytosomes as novel delivery system for nutraceutical materials. *Int J Curr Microbio App Sci.* 2015;4(6):152-159.
 14. Rathore P, Swami G. Planterosomes: Potential phytophospholipid carriers for the bioavailability enhancement of herbal extracts. *Int J Pharm Sci and Res.* 2015;3(3):737-755.
 15. Singh RP, Parpani S, Narke R, Chavan R. Phytosome: Recent advance research for novel drug delivery system. *Asian J Pharm Res and Dev.* 2014;2(3):15-29.
 16. Khan J, Alexander A, Saraf A, Swarnlata SS. Recent advances and future prospects of phytophospholipid complexation technique for improving pharmacokinetic profile of plant actives. *J Controlled Rel.* 2013;168: 50-60.
 17. Sanghi DK, Tiwle R. Herbal drugs an emerging tool for novel drug delivery systems. *Res J Pharm Techn.* 2013;6:962-66.
 18. Bombardelli E, Curri SB, Del NP, Tubaro A, Gariboldi P. Complexes between phospholipids and vegetal derivatives of biological interest. 1989;60:1-9.
 19. Kulkarni GT. Herbal drug delivery systems: An emerging area in herbal drug research. *J Ch Res Drug Dev.* 2011;2(3):113-19.
 20. Kidd PM. Bioavailability and activity of phytosome complexes from botanical polyphenols: the silymarin, curcumin, green tea, and grape seed extracts. *Altern Med Rev.* 2009;1;14(3):226-46.
 21. Bhattacharya S. Phytosomes emerging strategy in delivery of herbal drugs and nutraceuticals. *Pharmatimes.* 2009;41:3.
 22. Pandey S. Phytosomes: Technical revolution in phytomedicine. *Inter J Pharm Tech Res.* 2010;2:627-31.
 23. Das MK, Kalita B. Design and evaluation of phytophospholipid complexes (Phytosomes) of rutin for transdermal application. *J Appl Pharma Sci.* 2014;4(10):051-57.
 24. Salazar J, Muller RH, Moschwitz JP. Combinative particle size reduction technologies for the production of drug nanocrystals. *J Pharma.* 2014;pp:1-11.
 25. Agarwal A, Kharb V, Saharan VA. Process optimization, characterization and evaluation of resveratrol-phospholipid complexes using box-behnken statistical design. *Int Curr Pharm J.* 2014;3(7):301-8.
 26. Jain NK. Liposomes as drug carriers, controlled and novel drug delivery, 1st edn, CBS publisher, New Delhi, India. 2005.
 27. Babak G, Afshin B, Hamed H. Nanophytosome as a potential food grade delivery system. *Food Biosci.* 2016;15:126-135.
 28. Varde N, Mehta N, Thakor N, Shah V, Upadhyay U. Phytosomes: A potential phospholipid nanoparticulate carrier for the bioavailability enhancement of herbal extracts. *Inter J of Compreh Pharm.* 2012;10:1-7.
 29. Yue PF, Zhang WJ, Yuan HL, Yang M, Zhu WF. Process optimization, characterization and pharmacokinetic evaluation in rats of ursodeoxycholic acid-phospholipid complex. *American Association of Pharmaceutical Scientists; Pharm Sci Tech.* 2008;9(1):322-329.
 30. Semalty A. Cyclodextrin and phospholipid complexation in solubility and dissolution enhancement: A critical and meta-analysis. *Expert Opin Drug Deliv.* 2014;11(8):1255-72.
 31. Sharma S, Roy RK, Shrivastava B. Antiproliferative effect of phytosome complex of methanolic extract of *Terminalia arjuna* bark on human breast cancer cell lines (MCF-7). *Int J Drug Dev & Res.* 2015;7(1):173-182.
 32. Li J, Wang X, Zhang T, Wang C, Huang Z. A review on phospholipids and their main applications in drug delivery systems. *Asian J Pharm Sci.* 2015;10(2):81-98.
 33. Singh R, Parpani S, Narke R, Chavan A. Phytosome: Recent advance research for novel drug delivery system. *Asian J Pharm Res and Develop.* 2014;2(3):15-29.
 34. Gupta NK, Dixit VK. Development and evaluation of a vesicular system for curcumin delivery. *Arch of Derma Res.* 2011;303:89-101.
 35. Mascarella S. Therapeutic and antilipoperoxidant effects of the silybin-phosphatidylcholine complex in chronic liver disease, preliminary results. *Curr Ther Res.* 1993;53:98-102.
 36. Vandana S, Rani B, Nagpal M, Arora S. Phytosomes: Potential carriers for herbal drugs. *Amer J of Pharmatech Res.* 2013;3(1):249-60.
 37. Pawar HA, Bhangale BD. Phytosome as a novel biomedicine: A microencapsulated drug delivery system. *J Bioanal Biomed.* 2015;7(1):006-012.
 38. El-Maghraby GM, Williams AC, Barry BW. Oestradiol skin delivery from ultra-deformable liposomes: Refinement of surfactant concentration. *Inter J Pharm.* 2000;196(1):63-74.
 39. Fuzzati N, Gabetta B, Jayakar K, Pace R, Peterlongo F. Liquid chromatography electrospray mass spectrometric identification of ginsenosides in *Panax ginseng* roots. *J Chromato Ana.* 1999;854(1):69-79.
 40. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L. Positional cloning of the mouse obese gene and its human homologue. *Nature.* 1994;372 (6505):425-32.
 41. Gabetta B, Fuzzati N, Griffini A, Lolla E, Pace R. Characterization of proanthocyanidins from grape seeds. *Fitoter.* 2000;71(2):162-75.
 42. Basnet P, Basnet NS. Curcumin: an anti-inflammatory molecule from a curry spice on the path to cancer treatment. *Molecules.* 2011;16(6):4567-98.
 43. El-Maghraby GM, Barry BW, Williams AC. Liposomes and skin: From drug delivery to model membranes. *Eur J Pharm Sci.* 2008;34(4):203-22.
 44. Chaudhari P, Sharma P, Barhate N, Kulkarni P, Mistry C. Solubility enhancement of hydrophobic drugs using synergistically interacting cyclodextrins and cosolvent. *Current Sci.* 2007;92(11):1586-91.
 45. Aad G, Abajyan T, Abbott B, Abdallah J, Khalek SA. Observation of a new particle in the search for the standard model higgs boson with the ATLAS detector at the LHC. *Phy Lett B.* 2012;716(1):1-29.
 46. Nimbalkar CK, Hatware K. Phytosomes: Novel drug delivery system, *Indian J Drugs.* 2017;5(1):16-36.

47. Conti M, Malandrino S, Magistretti MJ. Protective activity of silipide on liver damage in rodents. *Jpn J Pharmacol.* 1992;60:315-21.
48. Gupta A, Ashawat MS, Shailendra S, Swarnlata S. Phytosome: A novel approach toward functional cosmetics. *J Plant Sci.* 2007;2:644-49.
49. Kumar P, Sharma S. Phytosomes: A novel phytophospholipid carriers: An overview. *Inter J Pharma Res and Develop.* 2009;pp:1-7.
50. Kareparamban J, Nikam P, Jadhav A. Phytosome: A novel revolution in herbal drugs. *Inter J Res Pharm and Chem.* 2012;2:299-310.
51. Doering T, Traeger A, Waldmann-Laue M. Cosmetic and dermatological composition for the treatment of aging or photodamaged skin. 2006;p:EP1640041,A2.
52. Comoglio A, Tomasi A, Malandrino S, Poli G, Albano E. Scavenging effect of silipide-a new silybin-phospholipid complex on ethanol derived free radicals. *Biochem Pharmacol.* 1995;50:1313-16.
53. Morazzoni P, Bombardelli E. Phospholipid complexes prepared from extracts of *Vitis vinifera* as anti-atherosclerotic agents. Indena Spa, Milan, Italy. 2001;p:US6297218.
54. Yanyu X, Yunmei S, Zhipeng C, Qineng P. The preparation of silybin-phospholipid complex and the study on its pharmacokinetics in rats. *Inter J Pharma.* 2006;37:77-82.
55. Bombardelli E. Oral compositions for the treatment of cellulite. Indena Spa, Milano, Italy. 2010;p:7691422.
56. Chauhan NS, Gowtham R, Gopalkrishna B. Phytosome: Potential phytophospholipid carriers for herbal drug delivery. *J Pharm Res.* 2009;2:1267-70.
57. Merizzi G. An anti-oxidant preparation based on plant extracts for the treatment of circulation and adiposity problems. 2002;p:EP1214084,A2.
58. Fry DW, White JC, Goldman ID. Rapid secretion of low molecular weight solute from liposomes without dilution. *Anal Biochem.* 1978;90:809-15.
59. Kleinman HK, Goldstein AL. Treatment of skin, and wound repair, with thymosin β 4 US Patent No.20070015698. 2007.
60. Franco PG, Bombardelli E. Complex compounds of bioflavonoids with phospholipids, their preparation and use, and pharmaceutical and cosmetic compositions containing them. Indena, Milan, Italy US Patent No.5043323A. 1991.
61. Singh A, Saharan VA, Singh M, Bhandari A. Phytosomes: Drug delivery system for polyphenolics phytoconstituents. *Iranian J of Pharma Sci.* 2011;7(4):209-19.
62. Esculin (esculoside, asculin). Available at <http://www.bpg.bg./substances/esculin.php>: Last accessed on 29 September 2017.
63. Farinacci M, Gasparido B, Colitti M, Stefanon B. Dietary administration of curcumin modifies transcriptional profile of genes involved in inflammatory cascade in horse leukocytes. *Ita J Ani Sci.* 2009;8:84-86.
64. Kohli K, Ali J, Ansari M, Raheman Z. Curcumin: A natural anti-inflammatory agent. *Ind J Pharmacol.* 2005;37:141-7.
65. Pierro FD, Menghi A, Barreca A, Lucarelli M, Calandrelli A. Greenselect® phytosome as an adjunct to a low-calorie diet for treatment of obesity: A clinical trial. *Alt Med Rev.* 2009;14:154-60.
66. Gandhi A, Dutta A, Pal A. Recent trends of phytosomes for delivering herbal extract with improved bioavailability. *J Pharmacog and Phytochem.* 2012;1(4):6-14.
67. Mullaicharam AR, Deori G, Maheswari RU. Mini review on cosmeceuticals. *Res J Pharm Bio and Chem Sci.* 2013;4(1):1091.
68. Guangxi Z, Hongxiang L, Dianzhou B. Interaction of puerarin with phospholipid in solid dispersion. *Chin Pharm J.* 2003;12:36-40.
69. Li Y, Yang DJ, Chen SL, Chen SB, Chan ASC. Process parameters and morphology in puerarin, phospholipids and their complex microparticles generation by supercritical antisolvent precipitation. *Inter J Pharm.* 2008;359:35-45.
70. Kidd PM. Phosphatidylcholine: A superior protectant against liver disease. *Alter Med Rev.* 1996;1:258-274.
71. Maiti K, Mukherjee K, Gantait A, Saha BP, Mukherjee PK. Enhanced therapeutic potential of naringenin phospholipid complex in rats. *J Pharm Pharmacol.* 2006;58:1227-33.
72. Bombardelli E, Bonati A, Gabetta B, Mustich G. Triterpenoids of *Terminalia sericea*. *Phytochemistry.* 1974;13:2559-62.
73. Naik S, Pilgaonkar V, Panda V. Evaluation of antioxidant activity of *Ginkgo biloba* phytosomes in rat brain. *Phytother Res.* 2006;20:1013-6.