

Improving Activity of Native Arbuscular Mycorrhizal Fungi (AMF) for Mycorrhizal Benefits in Agriculture: Status and Prospect

Dipankar Maiti*

Central Rainfed Upland Rice Research Station (Central Rice Research Institute, Indian Council of Agricultural Research), PB No. 48, Hazaribag, 825 301, Jharkhand, India

Abstract

Agricultural importance and ecological implications of arbuscular mycorrhizal (AM) symbiosis with land plants are well known. AM symbiosis facilitates plant growth through enhancing uptake of several macro- and micro-nutrients of low mobility in soil, like phosphorus, zinc, copper etc. Beside nutritional benefits to plant, AM also contributes to numerous ecological advantages like influencing microbial and chemical environment of the mycorrhizosphere, stabilizing soil aggregates, conferring tolerance (plant) to several abiotic and biotic stresses, bioremediation of soil and supplying protective (antioxidants) nutrient components to human being through agricultural products (food). There are two approaches to exploit arbuscular mycorrhiza for crops: (1) soil introduction of non-native inoculum and (2) exploitation of native AM fungal (AMF) population. The approach of soil introduction of non-native inoculum of selected AMF to field crops suffers from (i) cost intensiveness, (ii) inconsistent competitive performance of introduced inoculum due to lack of adaptability to new ecology and (iii) negative ecological consequences in terms of possible introduction of invasive species as unintended contaminants. Exploitation of native AM fungal (AMF) population of soils, keeping it undisturbed by avoiding faulty agricultural practices, is an alternative approach, now promoted for sound ecological management of crop production, particularly under stressful situations. The approach is based on twin attributes of AM symbiosis – ubiquitous nature and lack of host specificity of AMF. Several prospective avenues of enhancing native AM activities through agronomic manipulations of crop management practices and cropping systems for enhanced response to diverse native AMF population have been discussed in the present article

Keywords: Arbuscular mycorrhiza; AM inoculums; Crop rotation; Phosphorus; Tillage

Introduction

Inefficient acquisition of less mobile nutrients (like phosphorus, zinc, copper etc.) by crops is one major constraint of agriculture, particularly under rainfed ecology, prone to drought, occurring in various crop growth stages [1]. This reduces nutrient uptake, particularly the major nutrient of phosphorus (P) by plants [2,3]. The available 'non-occluded soil P' (inorganic P) remains in continuous equilibrium between 'solution P pool' which is readily available to plant and 'labile P pool' that remains adsorbed on soil surface and needs root interception for its acquisition by plant [4]. This 'labile P pool' is accessed by mycorrhizal plants through interception by extraradical (external) mycelial network extended beyond root zone that may extend up to several centimeters out in the soil [5] and help plant to acquire P beyond P depletion zone around root [6] which is otherwise not available to plant. Evidence of releasing nutrients from insoluble forms of inorganic sources like mineral particles and rock surfaces by AMF is conflicting [7]. Although there are some reports supporting mobilization of insoluble nutrients by AMF these effects could depend upon synergistic interactions with other P-solubilizing micro-organisms growing endosymbiotically with AM plants [8]. Other beneficial soil microbes like N fixing bacteria and P solubilising bacteria, may synergistically interact with AM fungi and thereby benefit plant growth [9].

Beside nutritional benefits to plant, AM also provides numerous ecological advantages like influencing microbial and chemical environment of the mycorrhizosphere, more precisely the hyphosphere, the zone surrounding individual hyphae [10], stabilizing soil aggregates, conferring tolerance (plant) to several abiotic and biotic stresses, bioremediation of soil and supplying protective (antioxidants) nutrient components to human being through agricultural products (food) contributing a key role in the earth's ecosystem services [11].

AM fungal association may influence bacterial communities associated with the roots in both direct and indirect ways [10]. While the fungus provides directly energy-rich carbon compounds derived from host assimilates, which are transported to the mycorrhizosphere via fungal extraradical hyphal network, changes in pH of the mycorrhizosphere induced by the fungus, competition for nutrients, and fungal exudation of other inhibitory or stimulatory compounds induces indirect interactions in the form of mycorrhiza mediated effects on host plant growth, root exudation and soil structure.

The extraradical mycelial network of AMF also imparts binding action on the soil and improves soil structure. In addition, the secretion by AM fungi of hydrophobic, 'sticky' proteinaceous substances, known as 'glomalin' [12], also contributes to soil stability and water retention [13]. The combination of extraradical mycelial network and glomalin secretion is considered to be an important element for stabilization of soil aggregates [14], thereby leading to reduced soil erosion and increased soil structural stability and quality [13].

AM association has been proved to be beneficial to agriculture under abiotic and biotic stressed environment including drought

***Corresponding author:** Dipankar Maiti, Central Rainfed Upland Rice Research Station (Central Rice Research Institute, Indian Council of Agricultural Research), PB No. 48, Hazaribag, 825 301, Jharkhand, India, Tel: +91 6546 222263; Fax: +91 6546 223697; E-mail: dipankar_maiti@live.in

Received November 01, 2011; **Accepted** December 15, 2011; **Published** December 25, 2011

Citation: Maiti D (2011) Improving Activity of Native Arbuscular Mycorrhizal Fungi (AMF) for Mycorrhizal Benefits in Agriculture: Status and Prospect. J Biofertil Biopestici S1:001. doi:10.4172/2155-6202.S1-001

Copyright: © 2011 Maiti D. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

(abiotic) and pathogen infection (biotic). AM, under moisture stress benefits in two ways by increasing (i) moisture retention property of soil [15] and (ii) greater exploration of soil moisture [16]. Subsequently, Ruiz-Lazano et al. [17] suggested involvement of modulating drought-induced genes of AM plant in imparting drought tolerance.

Despite several reports of pathogen (soil borne) suppression by AM fungi [18], the underlying mechanism is still not well understood. Some possible mechanisms could be (i) improvement of plant nutrition and competition for photosynthates [19], (ii) AM induced stimulation of saprotrophs and plant growth promoting microbes [20], (iii) AM induced anatomical and morphological changes in root system and (iv) AM induced local (at infection site) elicitation of plant defence mechanism [21]. However, additive pathogen suppressing effect of combined application of other antagonistic microbes with AM indicates possible function of AMF as vectors for the antagonists [10].

Improvement of nutritional quality in plant products as a result of mycorrhizal association was demonstrated in lettuce [22]. Such AM induced improvement was found to be dependent on phosphorus sources also. While copper, iron, starch and protein concentrations were increased in lettuce leaves under application of water insoluble P source, water soluble source increased mostly nonstructural sugars. The role of AMF in soil bioremediation, however, is mostly through encouraging associated microflora directly involved in bioremediation [23] as compared to direct role of ecto-mycorrhizas.

The well-drained, aerobic soil conditions of upland ecology supporting native AM activities [24] indicated that AM has greater potential in this ecology. Benefits from AM fungi (AMF) can be accrued by exploiting native AM flora or by application of external exotic inoculum. The former approach is considered to be more effective [25] owing to its stronger ecological adaptation and suitability due to less negative ecological consequences [26]. Thus, AMF inoculum developed from native sources is considered to be more efficient [27] and also cost effective. So, attempts have been made in the present article to discuss possible options of exploiting native AMF by enhancing its activity through (i) manipulations in agro-practices, (ii) adoption of AM supportive cropping systems/rotations and (iii) development and application of AMF inoculum of native origin.

Options for Exploiting Native AM Fungi

Manipulation in agro-practices

Optimizing tillage schedule: Tillage is one important agricultural operation that influences activities of native AMF in soil. Off-season tillage is an agronomic recommendation for management of weed and soil borne plant pathogens. On the other hand, this operation results in soil disturbance induced (SDI) deleterious effects on natural AMF by disrupting established mycelial network [28] leading to delayed colonization in subsequent crops and less P acquisition. Disruption AMF mycelia network in soil causes a delay in the colonizing roots of the next crop, because more time is needed for the inoculum around the roots to accumulate. Tillage induced reductions in mycorrhizal activity and phosphate nutrition is well known in corn [29], and the effect could be replicated in laboratory systems [30] for confirmation. The timing of the reduction in colonization is important, because the crop demands adequate phosphate early in the season for yield potential to be reached [31]. Hence, a compromise between no-tillage (most suitable for native AMF) and optimum tillage for accruing both mycorrhizal and agronomic benefits was felt to be worked out for recommendation. Magnitude of SDI effects depends upon length of undisturbed period

(no-till period) [32,33] and degree of soil disturbance in terms of extent of soil pulverization [34]. Under temperate ecology of Canada, 'reduced tillage' (only spring disking) was observed to have less severe negative impact on the abundance of soil hyphae and mycorrhizal colonization in corn by native AMF than 'conventional tillage' (fall mould-board ploughing + spring disking) [35]. A threshold of undisturbed period in terms of maintaining gap between two consecutive tillage operations (using bullock drawn country plough, tilling up to a depth of 10-15cm) of 13 weeks has been worked out for rainfed uplands of eastern India (tropical ecology) [36]. Having maintained this gap, two options of off-season tillage schedules (summer tillage alone and initial tillage after harvest + summer tillage) have been recommended for rainfed, mono-cropped (rice), upland ecosystem under study for maintaining optimum activities of native arbuscular mycorrhizal fungi. Heavy tillage both in terms of frequency and use of fine pulverizing machine reduced size of post tillage soil blocks. Resultant (post tillage) soil cutting blocks less than 4 cubic cm reduced AM induced P uptake in maize [34]. Thus, maintaining both (i) threshold of undisturbed period and (ii) use of coarser pulverizing machines are important for sustenance of native AMF activities in soil.

Optimizing P amendment: Higher soil P concentration was observed to reduce AMF activity [37] probably by reducing root colonization due to reduced root membrane permeability resulting in decreased loss of metabolites [38]. At the same time, plant rich in P are poor in carbohydrate content which reduces AMF colonization [39]. Effects of plant tissue P concentration on AMF colonization, however, is dependent on nitrogen (N) concentration. Root colonization remained unaffected with increasing P concentration when plants were N deficient, but increasing P inhibited mycorrhizal formation when plants were N sufficient [40,41]. Corroborating these findings, P fertilization was also observed to reduce mycorrhizal development in several crops [42,43]. P amendment through organic sources, however, did not adversely affect AMF activities possibly by improving soil biological properties favoring AMF [35]. Since organic manure is not available in required quantity everywhere, optimum dose of inorganic P, supporting maximum native AMF activities, without sacrificing crop yield, for each crop-ecosystem (at micro-level) combination is needed to be worked out. In a similar attempt, P optimum of 20 kg P₂O₅/ha was worked out for upland rice under AM supportive, two years crop rotation of maize (*Zea mays* L.) relay cropped by horse gram (*Dolichos biflorus* L.) in the first year followed by upland rice in the second year' [44] as compared to the recommended dose of 30 kg P₂O₅/ha. This P optima, however, was not effective under rice mono-cropping because soil P concentration threshold for maximum benefits from the AM symbiosis were observed to be lower than P concentration threshold for maximum plant benefit without enhanced AMF activities (inoculation) in crops like *Phaseolus mungo* and *Triticum aestivum* [45].

Adoption of AM supportive cropping systems/rotations

Reduction in AMF population under fallow (no crop) due to 'fallow disorder' was confirmed by several workers [46,47,48]. Even, contrary to the earlier speculations [49] and subsequent report [50], growing non-host (to AMF) crops, as compared to fallow, not only maintained better AMF activities in terms of colonizing the succeeding host crops [51] and spore population of native AMF [52], in certain cases, pre-cropping of non-host crop like oilseed rape (*Brassica napus* L.) significantly increased colonization in succeeding host crop like barley (*Hordeum vulgare* L.). This can be attributed to the previous findings that AM fungal hyphae can make some hyphal growth around the roots of non-host plants without colonizing the roots due to absence

of signals from non-host roots required by AM fungi for successful colonization [53]. Such roots surrounded by AM hyphal growth are more efficient in colonizing host plants than chlamydo spores or other inoculum source [51]. Such phenomenon is also influenced by soil P level. In non-host Swedes, although not infected by native AMF, magnitude of spore associations was found to be similar as that of host crops like barley and potato [49] grown under both low and high doses of P. The intermediate P level, however, increased spore population only with host crops (barley and potato) indicating the intermediate dose to be P optimum for maximum native AMF activity under AM favorable/supporting environment (barley and potato rhizosphere).

Mono-cropping of a particular crop leads to narrowing of AMF species diversity index [54] and distribution of AM fungi [52] due to obvious reason of encouraging a particular species favored by the particular crop in continuity without any break [55]. Apart from adverse ecological consequences of reducing AMF diversity, mono-cropping also was reported to reduce proliferation of AMF population than mixed cropping [56,57] due to three main reasons: (1) higher root density under mixed cropping favors AMF multiplication [58], (2) higher plant density exhausting soil nutrients faster stimulates AMF activities in soil [59] and (3) under mixed cropping with legumes, the companion crops get additional nitrogen from the legumes through AM fungi and thereby favoring AMF colonization [56].

Having information on 'fallow disorder' and adverse effects of mono-cropping on AMF, improving native AMF efficiency for enhancing P nutrition of crops through removing or reducing the factors (fallowing and mono-cropping) favoring these disorders/adverse effects were thought of. The factors could be removed/reduced by increasing cropping intensity through introduction of AM supportive crops, suitable to specific ecology, in the cropping rotation or cropping system. Several crop combinations suitable to various ecologies were evaluated by different researchers under diverse ecologies worldwide for the purpose. Native AMF population build up varied with different cropping regimes in the same soil under temperate [49,60] and tropical climates [58,61].

While enhancement of mycorrhizal colonization resulting in improved nutrient (P) uptake by cereals under mixed cropping of 'cereals - non-legumes' or 'cereal - cereal' combinations like rice (*Oryza sativa* L.) - finger millet (*Eleusine coracana* L. Gaertn) [62] was attributed to higher root density per volume of soil favoring the spread of the symbiotic fungi [60], higher root volume coupled with N backup to the symbiotic system in 'legume - legume' combination, like of berseem clover (*Trifolium alexandrinum* L.) - Persian clover (*T. resupinatum* L.) [63], and 'cereals - legumes' combination like maize (*Zea mays* L.) - berseem (*T. alexandrinum* L.) [64], rice (*Oryza sativa* L.) - pigeon pea (*Cajanus cajan* L.) [62] and rice - peanut (*Arachis hypogea* L.) [57] led to additive mycorrhizal benefits in terms of nutrient uptake and growth promotion. Ability of AM fungi to interconnect crop species grown together might allow translocation of N from legumes to cereals in mixed cropping.

Continuous mixed cropping of 'cereal - legume' and 'legume - legume' combinations, however, enhances chances of developing sick-plot of soil-borne plant pathogens, mostly for legumes, particularly under rainfed agro-ecosystem having mono-modal rainfall pattern with single annual crop season. For this ecology, crop rotations are safer options. Advantages of crop rotations in terms of mycorrhizal benefits can be attributed to (1) soil mycorrhizal potential left by both non-host and host pre-crops [65,66,67,61] and (2) reducing 'fallow disorder' [61]. The residual soil mycorrhizal potential of the pre-crop is also

long-lasting as evident from the resulting mycorrhizal benefits drawn by mycorrhiza responsive crop like sugarcane even after 2-4 years cycle [68]. Such residual effects can also be exploited under comparatively less AMF-favorable ecology like wetland rice with anaerobic soil condition. Pre-cropping rice seed bed (dry seedbed) with several host crops like fodder varieties of maize, sorghum (*S. bicolor*), Dinanath grass (*Pennisetum pedicellatum*), finger millet (*Elusin indica*) and little millet (*Panicum miliare*) enhanced AMF colonization of rice seedlings and P uptake [69] after transplanting at maturity under rainfed conditions. In pre-colonized roots, AMF remained dormant during inundated period and became active during intermittent drought periods [70] leading to moisture stress which is very common under rainfed ecology.

Development and application of AMF inoculum of native origin

Practical and ecological advantages of native AMF inoculum over that of non-native one have been discussed in the previous chapter (Introduction). AMF inoculum developed from native source are more effective [71] mainly due to its ecological adaptation beside other advantages of less negative ecological consequences in terms of possible invasive species introduction as unintended contaminants [26] and cost effectiveness. Having known these advantages, developing AMF inoculum of native origin has been thought of. Several small scale inoculum production techniques developed by different researchers, time to time, have been reviewed by Marleen et al. (2011) [72]. In the present review we have emphasized on various on-farm protocols for mass-production of native AMF inoculum suitable to various agro-eco systems. Soil-root based AMF inoculum produced by (i) growing pre-colonized (by native AMF) Bahia grass (*Paspalum notatum* Flugge) on fumigated plots [73] or raised beds amended with vermiculite and compost [74], (ii) multiplying native AMF fungal consortium on Sorghum roots (*Sorghum bicolor*) grown in partially sterilized (by soil solarization) [75] have been effectively used respectively as amendment to horticultural potting media for production of vegetable seedlings and as band placement in field for growing direct sown upland rice. A more recent approach of multifunctional microbial consortium inoculum facilitates integrated crop production system. The microbial consortium may include combination of compatible beneficial microorganisms having various plant growth promoting and pest controlling functions catering to the diversified crop cultivation need by one inoculum. Co-inoculation of multifunctional microorganisms combinations like (i) AMF + PGPR + PSB in lettuce [76], (ii) AMF + PGPR (*Azospirillum*, *Azotobacter*, *Pseudomonas* etc.) in Rhodes grass (*Chloris gayana* Kunth) [77], (iii) AMF + PSB in clover [78] and in English mint (*Mentha piperita* L.) [79] not only expressed individual beneficial effects, also resulted in additive or synergistic effects on plant growth promotion. Such results prompted the researchers to develop microbial consortium inoculum. In this effort many microbial consortium inoculum have been developed, tested [80] and commercialized. Integration of native AM supportive components is likely to produce additive or synergistic effects on plant growth promotion. In such effort, integration of AM supportive components of (i) crop rotation and (ii) application of on-farm produced native AMF inoculum under blanket practice of optimum tillage schedule and P amendment enhanced native AMF activity, P uptake and grain yield of upland rice under rainfed ecology [62]. Further validation, however, are required for their efficiency under farmers field condition for assessing technical feasibility and necessary fine tuning. While producing AMF inoculums of native origin, however, precautions should be taken to check the efficacy of

the mixed fungal composition in terms nutrient acquisition and growth improvement of the target crops.

Conclusion

Possibilities of exploiting native AMF for mycorrhizal benefits in agriculture through several eco-friendly avenues including manipulation of agro-practices, crop rotations and application of native AMF inoculum was well documented and their practical feasibility has been validated for adoption and recommendation as integrated crop production component. The following suggestions have been made to further strengthen the research for harnessing additional benefits from native AMF and other beneficial microorganisms for developing more ecologically sound integrated crop management strategy.

1. Location, soil type and agro-ecosystem specific optimum tillage schedule and type in terms of tillage depth and pulverization (favoring native AMF/supporting minimum damage to native AMF) need to be worked out at micro level for location specific recommendation.
2. Development of soil fertility and agro-system based prediction model of mycorrhizal activity would provide basis for regular updating of location specific fertilizer and microbial inoculums amendment schedule as recommendation which would reduce cost of cultivation beside mycorrhizal advantages.
3. Farmers' wisdom need to be considered for fine tuning suitable AM supportive crop rotation recommendations with the help of extensive farmers' participatory on-farm trials.
4. Further refinement of native AMF based inoculum production protocol in terms of quality, longevity and broader ecological adaptability would make the technology more suitable.
5. Inclusion of multifunctional microorganisms like bio-controlling (pests) agents beside biofertilizer agents in the microbial consortium inoculum would strengthen ecologically sound crop management strategy.
6. The research on native AMF aided agricultural benefits needs to be integrated with exploiting other beneficial microorganisms.
7. Strengthening research on integration of beneficial microorganisms' supportive agricultural components followed by proper validation under farmers' participatory mode would result in tangible recommendation as component of integrated crop management strategy.

Acknowledgement

Financial support from the Indian Council of Agricultural Research is gratefully acknowledged.

References

1. Tuong TP, Kam SP, Wade L, Pandey S, Bounman B et al. (2000) Characterizing and understanding rainfed environments. Proc. of the International Workshop on Characterizing and understanding Rainfed Environments. International Rice Research Institute, Bali, Indonesia.
2. Fageri NK, Barbosa Filho MP, Catvalho JRP (1980) Response of upland rice to phosphorus fertilization on an Oxisol of central Brazil. *Agronomy J* 74: 51-56.
3. Jupp AP, Newman EI, Ritz K (1987) Phosphorus uptake from soils by *Lolium perenne* during and after severe drought. *J Appl Ecology* 24: 969-978.
4. Tiessen H, Stewart JWB, Cole CV (1984) Pathways of phosphorus transformation in soils of differing pedogenesis. *Soil Sci Soc of America J* 48: 853-858.
5. Rhodes LH, Gerdemann JW (1975) Phosphate uptake zones of mycorrhizal and non-mycorrhizal onions. *New Phytologist* 75: 555-561.
6. Smith SE, Read DJ (1997) *Mycorrhizal Symbiosis*. (2nd edn), Academic Press, London.
7. Finley RD (2008) Ecological aspects of mycorrhizal symbiosis: with special emphasis on the functional diversity of interactions involving the extraradical mycelium. *Journal of Experimental Botany* 59: 1115-1126.
8. Jargeat P, Cosseau C, Ola'h B, Jauneau A, Bonfante P, et al. (2004) Isolation, free-living capacities, and genome structure of *Candidatus Glomeribacter gigasporarum*, the endocellular bacterium of the mycorrhizal fungus *Gigaspora margarita*. *Journal of Bacteriology* 186: 6876-6884.
9. Puppi G, Azcon R, Höflich G (1994) Management of positive interactions of arbuscular mycorrhizal fungi with essential groups of soil microorganisms. Birkhäuser Verlag, Basel, Switzerland.
10. Johansson JF, Paul LR, Finlay RD (2004) Microbial interactions in the mycorrhizosphere and their significance for sustainable agriculture. *FEMS Microbiology Ecology* 48: 1-13.
11. Gianinazzi S, Gollotte A, Binet Marie-Noëlle, Tuinen D van, Redecker D, et al. (2010) Agroecology: the key role of arbuscular mycorrhizas in ecosystem services. *Mycorrhiza* 20: 519-530.
12. Rillig MC, Wright SF, Nichols KA, Schmid WF, Torn MS (2002) The role of arbuscular mycorrhizal fungi and glomalin in soil aggregation: Comparing effects of five plant species. *Plant and Soil* 238: 325-333.
13. Bedini S, Pellegrino E, Avio L, Pellegrini S, Bazzoffi P, et al. (2009) Changes in soil aggregation and glomalin-related soil protein content as affected by the arbuscular mycorrhizal fungal species *Glomus mosseae* and *Glomus intradices*. *Soil Biol Biochem* 41: 1491-1496.
14. Rillig MC, Mummey D (2006) Mycorrhizas and soil structure. *New Phytol* 171: 41-53.
15. Augé RM, Stodola AJW, Tims JE, Saxton AM (2001) Moisture retention properties of a mycorrhizal soil. *Plant and Soil* 230: 87-97.
16. Augé RM (2001) Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. *Mycorrhiza* 11: 3-42.
17. Ruiz-Lozano JM, Porcel R, Aroca R (2006) Does the enhanced tolerance of arbuscular mycorrhizal plants to water deficit involve modulation of drought-induced plant genes? *New Phytologist* 171: 693-698.
18. Caron M (1989) Potential use of mycorrhizae in control of soilborne diseases. *Can J Plant Pathol* 11: 177-179.
19. Azcon-Aguilar C, Barea JM (1996) Arbuscular mycorrhizas and biological control of soil-borne plant pathogens – An overview of the mechanisms involved. *Mycorrhiza* 6: 457-464.
20. Kapoor R, Mukherji KG (1998) Microbial interactions in mycorrhizosphere of *Anethum graveolens* L. *Phytomorphology* 48: 383-389.
21. Gianinazzi-Pearson V, Gollotte A, Dumas-Gaudot E, Franken P, Gianinazzi S (1994) Gene expression and molecular modifications associated with plant responses to infection by arbuscular mycorrhizal fungi. Kluwer, Dordrecht.
22. Baslam M, Pascual I, Sanchez-Díaz M, Erro J, García-Mina J, et al. (2011) Improvement of nutritional quality of greenhouse-grown lettuce by arbuscular mycorrhizal fungi is conditioned by the source of phosphorus nutrition. *J Agric Food Chem* 59: 11129-11140.
23. Jøner EJ, Johansen A, Loibner AP, Cruz MAD, Szolar OHJ, et al. (2001) Rhizosphere effects on microbial community structure and dissipation and toxicity of polycyclic aromatic hydrocarbons (PAHs) in spiked soil. *Environmental Science and Technology* 35: 2773-2777.
24. Wangiyana W, Cornish PS, Morris EC (2006) Arbuscular mycorrhizal fungi dynamics in contrasting cropping systems on vertisol and regosol soils of Lombok, Indonesia. *Experimental Agriculture* 42: 427-439.
25. Caravaca F, Alguacil MM, Barea JM, Roland A (2005) Survival of inocula and native AM fungi species associated with shrubs in a degraded Mediterranean ecosystem. *Soil Biol Biochem* 37: 227-233.
26. Schwartz MW, Hoeksema JD, Gehring CA, Johnson NC, Klironomos JN, et al. (2006) The promise and the potential consequences of the global transport of mycorrhizal fungal inoculum. *Ecology Letters* 9: 501-515.

27. Oliveira RS, Vosátka M, Dodd JC, Castro PML (2005) Studies on the diversity of arbuscular mycorrhizal fungi and the efficacy of two native isolates in a highly alkaline anthropogenic sediment. *Mycorrhiza* 16: 23-31.
28. Jasper DA, Abbot LK, Robson AD (1991) The effect of soil disturbance on vesicular arbuscular mycorrhizal fungi in soils from different vegetation type. *New Phytol* 118: 471-476.
29. McGonigle TP, Millar MH (1993) Response of mycorrhizae and shoot phosphorus of maize to the frequency and timing of soil disturbance. *Mycorrhiza* 4: 63-68.
30. McGonigle TP, Millar MH (1991) Mycorrhizal development and phosphorus absorption in maize under conventional and reduced tillage. *Soil Sci Soc of Amer J* 57: 1002-1006.
31. Grant CA, Flaten DN, Tomaszewicz DJ, Sheppard SC (2001) The importance of early season phosphorus nutrition. *Can J Plant Pathol* 81: 211-224.
32. Fairchild G L, Miller MH (1988) Vesicular-arbuscular mycorrhizas and the soil disturbance induced reduction of nutrient absorption in maize. II. Development of the effect. *New Phytol* 110: 75-84.
33. Fairchild GL, Miller MH (1990) Vesicular arbuscular mycorrhizae and the soil disturbance induced reduction of nutrient absorption in maize. III. Influence of P amendments to soil. *New Phytol* 114: 641-650.
34. McGonigles TP, Evans DG, Miller MH (1990) Effect of degree of soil disturbance on mycorrhizal colonization and phosphorus absorption in maize in growth chamber and field experiment. *New Phytol* 116: 629-636.
35. Kabir Z, O'Halloran IP, Fyles JW, Hamel C (1997) Seasonal changes of arbuscular mycorrhizal fungi as affected by tillage practices and fertilization: Hyphal density and mycorrhizal root colonization. *Plant and Soil* 192: 285-293.
36. Maiti D, Variar M, Singh RK (2010) Optimizing tillage schedule for maintaining activity of the arbuscular-mycorrhizal fungal population in a rainfed upland rice (*Oryza sativa* L.) agro-ecosystem. *Mycorrhiza* 21: 167-171.
37. Habte Mitiku, Manjunath Aswathanarayan (1987) Soil solution phosphorus status and mycorrhizal dependency in *Leucaena leucocephala*. *Applied and Environmental Microbiology* 53: 797-801.
38. Graham JH, Leonard RT, Menge JA (1981) Membrane mediated decrease in root exudation responsible for phosphorus inhibition of vesicular-arbuscular mycorrhiza formation. *Plant Physiol* 68: 548-552.
39. Jasper DA, Robson AD, Abbott LK (1979) Phosphorus and the formation of vesicular arbuscular mycorrhizas. *Soil Biol. Biochem* 11: 501-505.
40. Sylvia DM, Neal LH (1990) Nitrogen affects the phosphorus response of VA mycorrhiza. *New Phytol* 115: 303-310.
41. Smith SE, St Jojn BJ, Smith FA, Bromly JL (1986) Effects of mycorrhizal infection on plant growth, nitrogen and phosphorus nutrition in glasshouse-grown *Allium cepa* L. *New Phytol* 103: 359-373.
42. Abbott LK, Robson AD (1991) Factors influencing the occurrence of vesicular-arbuscular mycorrhizas. *Agric. Ecosyst Environ* 35: 121-150.
43. Bethlenfalvay GJ (1992) *Mycorrhizae and crop production*. ASA, Madison, WI.
44. Maiti D, Barnwal MK, Singh RK (2011) Optimum phosphorus dose favoring efficient native arbuscular mycorrhizal (AM) activity for enhanced phosphorus nutrition of upland rice under AM supportive rice (*Oryza sativa* L.) based crop rotations in rainfed ecology, communicated to *Mycorrhiza*.
45. Shukla A, Kumar A, Jha A, Ajit, Nageawar Rao DVK (2011) Phosphorus threshold for arbuscular mycorrhizal colonization of crops and tree seedlings. *Biol and Fertl of Soils*.
46. Black RLB, Tinker PB (1977) Interaction between effects of vesicular-arbuscular mycorrhizas and fertilizer phosphorus on yields of potatoes in the field. *Nature* 267: 51-511.
47. Harinikumar KM, Bagyaraj DJ (2005) Effect of crop rotation on the vesicular arbuscular mycorrhizal propagules in soil. *Plant and Soil* 110: 77-80.
48. Thompson JP (1987) Decline of vesicular-arbuscular mycorrhizae in long fallow disorder of field crops and its expression in phosphorus deficiency of sunflower. *Aust J Agric Res* 38: 847-867.
49. Hayman S, Johnson AM, Ruddlesdiin M (1975) The influence of phosphate and crop species on *Endogone* spores and vesicular-arbuscular mycorrhiza under field conditions. *Plan and Soil* 43: 489-495.
50. Fontenla S, García-Romera JA, Ocampo JA (1999) Negative influence of non-host plants on the colonization of *Pisum sativum* by the arbuscular mycorrhizal fungus *Glomus mosseae*. *Soil Biol Biochem* 31: 1591-1597.
51. Ocampo JA, Hayman DS (1981) Influence of plant interactions on vesicular-arbuscular mycorrhizal infections. *New Phytol* 87: 333-343.
52. Kruckelmann W (1975) Effects of fertilizers, soils, soil tillage and plant species on the frequency of *Endogone* chlamydo spores and mycorrhizal infection in arable soils. Academic Press, London.
53. Ocampo A, Martin J, Hayman D (1980) Influence of plant interaction on vesicular-arbuscular mycorrhizal infections. I. Host and non-host plants grown together. *New Phytologist* 84: 27-35.
54. Oehl F, Sieverding E, Ineichen L, Mander P, Boller T, et al. (2003) Impact of Land Use Intensity on the Species Diversity of Arbuscular Mycorrhizal Fungi in Agroecosystems of Central Europe. *Appl Environ Microbiol* 69: 2816-2824.
55. Schenck NC, Kinloch RA (1980) Incidence of Mycorrhizal fungi on six in monoculture on a newly cleared woodland site. *Mycologia* 72: 445-455.
56. Harinikumar KM, Bagyaraj DJ, Majlesha BC (1990) Effect of intercropping and organic soil amendments on native VA mycorrhizal fungi in an Oxisol. *Arid Soils Res Rehabil* 4: 193-197.
57. Rana SK, Maiti D, Barnwal, M K, Singh RK, Variar M (2002) Effect of rice based intercropping systems on vesicular-arbuscular mycorrhizal colonization, P uptake and yield. *Indian Journal of Agricultural Sciences* 72: 400-403.
58. Harinikumar and Bagyaraj (1988) Effect of crop rotation on native vesicular arbuscular mycorrhizal propagules in soil. *Plant and Soil* 110: 77-80.
59. Bagyaraj DJ, Lakshminpathi R, Balakrishna AN (2005) *Agricultural intensification in tropics - its influence on AM fungi*. International Publ House, New Delhi, India.
60. Strzemska J (1975) Occurrence and intensity of mycorrhizae in cultivated plants. Academic Press, London.
61. Maiti D, Variar, M, Singh RK (2011) Rice based crop rotation for enhancing native arbuscular mycorrhizal (AM) activity to improve phosphorus nutrition of upland rice (*Oryza sativa* L.). *Biol and Fert of Soils*.
62. Maiti D, Neha Nancy Toppo, Variar M (2011) Integration of crop rotation and arbuscular mycorrhizal (AM) fungal inoculum application for enhancing native AM activity to improve phosphorus nutrition of upland rice (*Oryza sativa* L.). *Mycorrhiza* 21: 659-667.
63. Zarea MJ, Ghalavand A, Goltapeh EM, Rejali F, Zamanian M (2009) Effects of mixed cropping, earthworms (*Pheretima* sp.), and arbuscular mycorrhizal fungi (*Glomus mosseae*) on plant yield, mycorrhizal colonization rate, soil microbial biomass and nitrogenase activity of free-living rhizosphere bacteria. *Pedobiologia* 52: 223-235.
64. Frey B, Schüepp H (1992) Transfer of symbiotically fixed nitrogen from berseem (*Trifolium alexandrinum* L.) to maize via vesicular-arbuscular mycorrhizal hyphae. *New Phytol* 122: 447-454.
65. Johansen A, Jakobsen I, Jensen ES (1993) External hyphae of vesicular-arbuscular mycorrhizal fungi associated with *Trifolium subterraneum* L. 3. Hyphal transport of 32 P and 15N. *New Phytol* 120: 61-68.
66. Oliveira AAR, Sanders FE (1999) Effect of management practices on mycorrhizal infection, growth and dry matter production in field-grown bean. *Pesq Agropec Bras* 34: 1247-1254.
67. Harinikumar KM, Bagyaraj DJ (2005) Effect of crop rotation on native arbuscular mycorrhizal propagules in soil. *Plant and Soil* 110: 77-80.
68. Edmilson JA, Rozario A, Heitor C, Gláucia MBA, Eliana A, et al. (2010) Crop rotation biomass and arbuscular mycorrhizal fungi effects on sugarcane yield. *Sc Agric (Piracicaba, Braz.)* 67: 692-701.
69. Maiti D, Barnwal MK, Singh RK (2008) Exploring possibility of utilizing native arbuscular mycorrhizal fungi for improving phosphorus nutrition in transplanted rice of plateau region. *Ind Phytopath* 61: 302-304.
70. Solayman MZ, Hirata H (1997) Effectiveness of arbuscular mycorrhizal colonization at nursery-stage on growth and nutrition in wetland rice after transplanting under different soil fertility and water regimes. *Soil Sci Plant Nutr* 41: 505-514.
71. Oliveria RS, Vosátka M, Dodd JC, Castro PML (2005) Studies on the diversity of arbuscular mycorrhizal fungi and the efficacy of two native isolates in a highly alkaline anthropogenic sediment. *Mycorrhiza* 16: 23-31.

-
72. Marleen I, Sylvie C, Stéphane D (2011) Methods for large scale production of AM fungi: past, present and future. *Mycorrhiza* 21: 1-16.
73. Douds DD Jr, Nagahashi G, Pfeffer PE, Reider C, Kayser WM (2005) On-farm production and utilization of arbuscular mycorrhizal fungus inoculum. *Can J Plant Sci* 85: 15–21.
74. Douds DD Jr, Nagahashi G, Pfeffer PE, Reider C, Kayser WM (2006) On-farm production of AM fungus inoculum in mixtures of compost and vermiculite. *Biores Tech* 97: 809-818.
75. Maiti D, Barnwal MK, Singh RK, Variar M (2009) A new protocol for on-farm production method of arbuscular mycorrhizal fungal mass inoculum for rainfed upland rice. *Ind Phytopath* 62: 31-36.
76. Kohler J, Caravaca F, Carrasco L, Roldan A (2007) Interactions between a plant growth-promoting rhizobacterium, an AM fungus and a phosphate-solubilising fungus in the rhizosphere of *Lactuca sativa*. *Applied Soil Ecology* 35: 480-487.
77. Bhowmick SN, Singh CS (2004) Mass multiplication of AM inoculum: Effect of plant growth-promoting rhizobacteria and yeast in rapid culturing of *Glomus mosseae*. *Current Science* 86: 705-709.
78. Souchie EL, Azcon R, Barea JM, Suggin-Junior, da Silva EMR (2006) Phosphate solubilization and synergism between P-solubilizing and arbuscular mycorrhizal fungi. *Pesquisa Agropecuaria Brasileira* 41: 1405-1411.
79. Cabello M, Irrazable G, Bucsinszky AM, Saparrat M, Schalamuk (2005) Effect of an arbuscular mycorrhizal fungus, *Glomus mosseae*, and a rock-phosphate-solubilizing fungus, *Penicillium thomii*, on *Mentha piperita* growth in a soilless medium. *J Basic Microbiology* 45: 182-189.
80. Wu SC, Cao ZH, Li ZG, Cheung KC, Wong MH (2005) Effects of biofertilizer containing N-fixer, P and K solubilizers and AM fungi on maize growth: a greenhouse trial. *Geoderma* 125: 155-166.