

# HPLC Method Validation for the Quantitative Fast Determination of 8 Organic UV-Filters in Products with a Level of Sun Protection Factor Emulsion-Type

Juan David Marín Arevalo, Elizabeth Claros Ceballos, Alejandra Patricia Guzmán Carantón, Johnbrynnner García Galvis\*

Belcorp Research and Development, Tocancipá-Colombia

## ABSTRACT

A fast High Performance Liquid Chromatography (HPLC) method was developed and validated for the simultaneous quantitative determination of phenylbenzimidazole sulfonic acid (PBMSA), benzophenone-3 (BZ3), diethylamino hydroxybenzoyl hexyl benzoate (DHHB), octocrylene (OC), butyl methoxydibenzoylmethane (BMBM), ethylhexyl methoxycinnamate (EMC), ethylhexyl salicylate (ES), and homosalate (HS) which are eight organic UV-filters of the most commonly used in cosmetics and sunscreens. A solution pH 2.0 of phosphate buffer, methanol, and acetonitrile in proportions 15:3:82 respectively as mobile phase was used in isocratic conditions, combined with a Purospher Star® Performance RP-18e column and eight minutes of analysis time. The proposed method was validated following the recommendations of International Council on Harmonization (ICH) and the Food and Drug Administration (FDA) guidelines. All parameters examined were found to be well within the stated guidelines. From the robustness test, it was possible to identify that pH of the buffer and the vial material are the critical methodological parameters for the analysis of butyl methoxydibenzoylmethane (BMBM), whereas for phenylbenzimidazole sulfonic acid (PBMSA) they were, the vial material, volume of NaOH solution and sonication time and temperature.

**Keywords:** Chemical analysis; HPLC; Sunscreens; Organic UV-filters; Validation

## INTRODUCTION

Exposure to UV radiation in controlled doses has been employed with therapeutic benefits for the psoriasis treatment [1], and even aesthetic as nail drying [2]. Currently, people are exposed to sunlight radiation for different reasons, such as the progressive damage to the ozone layer, geographic location, social customs, professional practices among others [3,4].

UV radiation is characterized by having a wavelength between (10-400) nm, which allows it to be classified as ionizing radiation that can be divided into three categories. The first, known as UV-A which comprises a wavelength between (320-400) nm, penetrates the skin with relative ease and great depth. The UV-B includes a wavelength between (290-320) nm of higher energy and can cross external layers of the skin causing damage [5]. Finally, UV-C radiation spanning a wavelength between (190-290) nm is the radiation emitted by the sun with a wavelength below 290 nm and it is absorbed by the ozone layer (which acts as a blocking screen) before reaching the Earth's surface.

Exposure to UV radiation promotes changes in the energy levels of molecules, especially chromophore molecules, which are part of cellular tissues and triggers a cascade of radical-mediated reactions, and for that reason, the well-known oxygen-reactive species associated with premature aging, non-melanoma skin cancer (NMSC), DNA or RNA chain damage, among others occur [3]. A source of UV radiation is solar radiation (SUVR), which has been classified by the International Agency for Research on Cancer (IARC) in group I as a carcinogen. Therefore, it is considered a public health problem [6].

In this situation, one of the products currently offered and whose objective is providing protection against the solar radiation that reaches the earth's surface are the sunscreens [7], which work similarly to how the ozone layer does. Molecules of organic and inorganic nature, also known as chemical and physical filters, respectively, are used as filters. Those molecules prevent human health damage through different mechanisms. Both prevent damage to the skin by incident radiation. However, this classification gives rise to the possibility of misinterpreting

**Correspondence to:** Johnbrynnner G Galvis, Belcorp Research and Development, Tocancipá-Colombia, Tel: +573188118280; E-mail: johngarcia@belcorp.biz

**Received:** February 02, 2021; **Accepted:** February 16, 2021; **Published:** February 23, 2021

**Citation:** Arevalo JDM, Ceballos EC, Carantón APG, Galvis JG (2021) HPLC Method Validation for the Quantitative Fast Determination of 8 Organic UV-Filters in Products with a Level of Sun Protection Factor Emulsion-Type. J Chromatogr Sep Tech 12: 440

**Copyright:** © 2021 Arevalo JDM, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

concepts, although both filters are chemical substances, they differ in their composition and in their mode of action as a sunscreen.

Physical filters are named due to their characteristic interaction with incident radiation. Absorption, dispersion, and reflection phenomena take place in this type of filter and in turn, are determined by aspects of the system such as film thickness, particle size, among others [8]. Chemical filters work by absorbing energy radiation and promoting molecular structural arrangements [9].

The quantity and chemical nature of molecules in personal care products with the properties previously mentioned are regulated by various entities, depending on the geographical area where it intends to market, one of them being the Food and Drug Administration (FDA) in the United States.

Butyl Methoxydibenzoylmethane, benzophenone-3, octocrylene, ethylhexyl salicylate, ethylhexyl methoxycinnamate, homosalate [10] are part of the main organic UV-filters used at industrial level and approved by the FDA; however, other molecules that meet the same objective and are used industrially are diethylamino hydroxybenzoyl hexyl benzoate and phenylbenzimidazole sulfonic acid. These strict regulations respond to recent research on the negative impact that organic filters can have on the environment and human health [11-14].

In order to satisfy the requirements and thus commercialize safe products for the users and the environment, various methods of identification and quantification of sunscreens in emulsion-type matrices have been developed [15,16]. This work shows the development and validation of a fast method for the quantification of eight organic filters simultaneously (phenylbenzimidazole sulfonic acid, benzophenone-3, diethylamino hydroxybenzoyl hexyl benzoate, octocrylene, butyl methoxydibenzoylmethane, ethylhexyl methoxycinnamate, ethylhexyl salicylate, homosalate) in sunscreen products, using Liquid Chromatography of High Efficiency (HPLC).

## MATERIALS AND METHODS

### Standards

The following standards were purchased from USP: phenylbenzimidazole sulfonic acid (p.a. 99.5%), benzophenone-3 (p.a. 99.9%), octocrylene (p.a. 98.9%), Butyl methoxydibenzoylmethane (p.a. 99.5%), ethylhexyl methoxycinnamate (p.a. 99.2%), ethylhexyl salicylate (p.a. 99.7%), homosalate (p.a. 99.5%). Diethylamino hydroxybenzoyl hexyl benzoate (p.a.99.4%) was purchased from Dr. Ehrenstorfer.

### Solvents

Acetonitrile ( $\text{CH}_3\text{CN}$ , p.a.  $\geq 99.9\%$ , Merck) and methanol ( $\text{CH}_3\text{OH}$ , p.a.  $\geq 99.9\%$ , Merck) were HPLC grade. Monopotassium phosphate ( $\text{KH}_2\text{PO}_4$ , Merck), phosphoric acid ( $\text{H}_3\text{PO}_4$ , p.a. 85%, Merck), sodium hydroxide solution ( $\text{NaOH}$ , 0.1N, Merck) were reagent grade. HPLC water was prepared from milli-Q purification system (Burnstead, EasyPure II D7401,  $\geq 18 \text{ M}\Omega\text{cm}$ ).

### Blank matrix

The placebo used was an oil-in-water emulsion formulated and prepared in the laboratory (Chemical Analysis; Belstar S.A), in accordance with the manufacturing protocol for products that declare sun protection factor. This emulsion was used to determine the specificity of the method.

### Equipment

Continued A VWR-HITACHI Elite Chrom L-2450 HPLC system equipped with modules automatic injection, oven, diode array detection (DAD) was used. Data acquisition and processing were performed using Open lab EZChrome software.

### HPLC operating conditions

A Purospher Star® Performance RP18e ( $5 \mu\text{m}$ ) 150-4, 6 mm, C18 was used. The mobile phases used were filtered and degassed. Chromatography was developed under isothermal conditions at  $40^\circ\text{C}$  and isocratic elution at a flow rate of  $1.2 \text{ mLmin}^{-1}$ . The detection wavelengths were: 308 nm for PBMSA, OC, EMC, ES, and HS, and 354 nm for BMBM, BZ3, and DHHB.

### Mobile phase and diluent solution

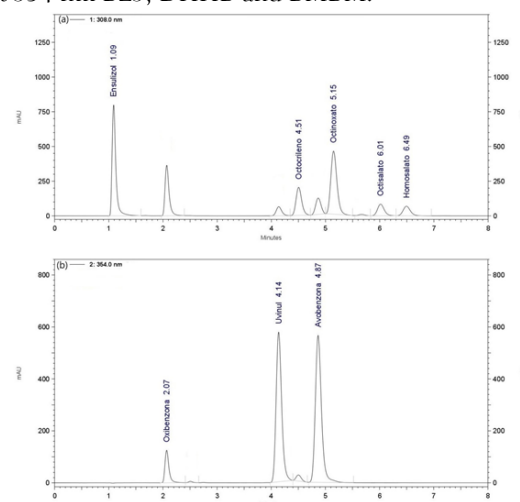
A homogenous mixture of a phosphate buffer solution pH 2.0, methanol, and acetonitrile in proportions 15:3:82 respectively, was employed.

### Standard solution

50 mL of a stock solution containing  $1.5 \text{ mgmL}^{-1}$  approximately, of each standard was prepared separately, each solution was made up to volume with diluent solution, with exception of PBMSA due to its high polarity and low solubility in ethanol, which was prepared using 5 mL of  $\text{NaOH}$  (0.1 N) and was diluted to volume with deionized water.

A 500  $\mu\text{L}$  aliquot of each standard solution was transferred to the same 25 mL volumetric flask and was made up to volume with diluent solution.

The order elution of each compound analyzed is show in Figure 1. It was the following: at 308 PBMSA, OC, EMC, ES, and HS, and, at 354 nm BZ3, DHHB and BMBM.



**Figure 1:** Chromatographic profile of a mix of standard at 308nm (a) and 354 nm (b).

### Sample preparation

50 mg of sample was weighed and transferred to a 25 mL volumetric flask, 1 mL of NaOH 0.1 N was added, followed by vortex treatment until complete dispersion, later 18 mL of diluent solution was added followed by an ultrasonic treatment of 15 min at 50°C. After the treatment, the sample reached room temperature and was made up to volume with diluent solution. Subsequently, the sample was filtered through with Millipore filter membrane pre size 0.45 µm. 10 µL of each sample were injected into the chromatograph.

### Protocol and process of validation

The validation process was based on the guidelines The International Council for Harmonization of Technical Requirements for Pharmaceuticals for Human Use (ICH), Validation of analytical procedures Q2 (R1) [17,18] The validation parameters evaluated were: specificity, linearity, repeatability, accuracy, intermediate precision, reproducibility, robustness, limit of detection and limit of quantification.

**Specificity:** It was determined from the comparison of chromatographic profiles of the mobile or diluent phase, of the mixture of all standards, the placebo, and the placebo enriched also the chromatographic profiles of the combination of standards, placebo, and enriched placebo after 4 hours exposed to daylight (photolysis) and 4 hours exposed to 60°C (thermolysis) independently. This comparison included observation of additional or missing signals.

**Linearity:** The linearity of the system and the method were evaluated, both separately by constructing five-level calibration curves, each level was prepared three times and injected in duplicate. The linearity of the system was evaluated using the mixture of all standards; the linearity of the method was evaluated with the placebo enriched with all the standards.

This parameter was evaluated by means of linear least squares regression analysis, relating the area of each signal and the concentration of the injected standard.

**Repeatability:** The repeatability of the system and method was evaluated at three different levels of concentrations, six preparations per level and two injections per preparation, calculated by the relative standard deviation (RSD). For the repeatability of the system, the mixture of all standards was used.

**Accuracy:** The percent recovery of each standard was evaluated at three different concentration levels in the placebo with three preparations per level and two injections per preparation.

**Intermediate precision:** Using commercial samples, intermediate precision was calculated by the relative standard deviation (RSD) of three sample preparations on two different days by two distinct analysts.

**Reproducibility:** The same commercial products were analyzed in two different laboratories, three preparations per product were performed and each preparation was injected in duplicate, calculated by the relative standard deviation (RSD).

**Robustness:** The ability of the method to remain unchanged by parameter variations was evaluated applying the Youden-Steiner model for 5 variables (pH of the buffer, ultrasound temperature, ultrasound time, vial material, volume of NaOH) using the placebo enriched. The method conditions were changed for the alternative conditions of each of the variables.

**Limit of Detection (Lod) and Limit of Quantification (Loq):** The LOD and LOQ for the system and the method were theoretically determined according to the definition of the ICH Q2 (R1) [17,18], Based on the Standard Deviation of the Response and the Slope and verified experimentally, using the following mathematical expressions, Equation 1 and 2:

$$\text{LOD}: 3.3(\text{sB}/\text{mB}) \quad (1)$$

$$\text{LOQ}: 10(\text{sB}/\text{mB}) \quad (2)$$

Where sB the standard deviation of the response and mB the slope of the calibration curve.

Measurements of these limits were made using the mixture of all standards for the method and the enriched placebo for the system. For the LOD, two preparations and two injections per preparation were performed; for the LOQ, precision and accuracy were evaluated from six preparations, two injections per preparation.

## RESULTS AND DISCUSSION

### Specificity

There was no evidence of signal interference between the placebo blank and the mobile phase at the two wavelengths proposed in the method. There is no difference between the retention times of the same standard when evaluated individually, in the mixture or in the placebo. The chromatographic profiles of the samples subjected to heat stress and exposure to sunlight showed no interference in their response due to the addition or loss of signals compared to the chromatographic profiles before stress. These results indicated that the extraction method together with the wavelengths used in the data acquisition were the appropriate conditions to specifically analyze the target analytes.

### Linearity

The most representative linearity parameters of the system and method for each of the analytes evaluated are shown in Table 1. The five concentration levels for the construction of each calibration curves were (0.01, 0.03, 0.06, 0.09 and 0.12) mgmL<sup>-1</sup>.

The linearity of the curve for each filter was constructed by relating the concentration (x) and the signal area (y). The trend line that best adjust the experimental data was obtained using the least-squares linear regression method with determination coefficients (r<sup>2</sup>) >0.99. The linear relationship between the two variables studied (concentration and response) was statistically verified by variance analysis using a Fisher test ( $F_t \rightarrow 4.196$ ,  $\alpha \rightarrow 0.05$ ). These results suggest that there is a significant linear adjustment of each one of the studied systems in the evaluated concentration range.

**Table 1:** Range of concentrations, linear equation.

Sunscreen (INCI name)	Range of concentrations ( $\mu\text{g mL}^{-1}$ )		Linear equation (standard form)		$r^2 \geq 0.99$	
	System	Method	System	Method		
Phenylbenzimidazole Sulfonic Acid	6.0-122.5	2.8-120.0	$y=119162127x + 251336$	$y=127849571x + 162183$	0.9982	0.9996
Benzophenone-3	6.7-124.4	3.0-120.4	$y=21493108x + 2657$	$y=21768838x - 22364$	0.9979	0.9995
Diethylamino Hydroxybenzoyl Hexyl Benzoate	5.3-120.1	2.7-121.0	$y=140851470x - 342$	$y=137755166x - 47889$	0.9986	0.9996
Octocrylene	6.8-120.0	3.7-121.6	$y=50278794x + 12439$	$y=49780444x - 75114$	0.9976	0.9993
Butyl Methoxydibenzoylmethane	7.6-123.0	5.0-119.8	$y=144340254x + 135751$	$y=152580950x + 179551$	0.9972	0.9987
Ethylhexyl Methoxycinnamate	7.0-124.1	3.1-122.4	$y=118972297x + 4034$	$y=119293588x - 194150$	0.9976	0.9995
Ethylhexyl Salicylate	6.7-122.3	3.6-120.5	$y=24726096x + 2809$	$y=24327255x - 12250$	0.9978	0.9993
Homosalate	6.5-119.6	2.6-118.7	$y=21898827x + 3146$	$y=22025616x - 28121$	0.9978	0.9997

In addition, using t student ( $t_{ab} \rightarrow 2.0484$ ,  $\alpha \rightarrow 0.05$ ), it was confirmed that the slopes are significantly different from zero and that the intercepts are not statistically different from zero.

The range of working concentrations was redefined from the data obtained from LOQ and for the system and method (Table 2).

**Table 2:** Limit of detection (LOD) and limit of quantification (LOQ).

Sunscreen (INCI name)	LOD ( $\mu\text{g mL}^{-1}$ )		LOQ ( $\mu\text{g mL}^{-1}$ )	
	System	Method	System	Method
Phenylbenzimidazole Sulfonic Acid	1.984	0.928	6.012	2.812
Benzophenone-3	2.197	0.979	6.657	2.966
Diethylamino Hydroxybenzoyl Hexyl Benzoate	1.742	0.906	5.28	2.744
Octocrylene	2.249	1.226	6.814	3.716
Butyl Methoxydibenzoylmethane	2.494	1.663	7.557	5.04
Ethylhexyl Methoxycinnamate	2.319	1.034	7.028	3.133
Ethylhexyl Salicylate	2.194	1.183	6.648	3.586
Homosalate	2.158	0.843	6.54	2.556

### Precision

The repeatability results of the system and method for each analyte expressed as coefficient of variation are shown in Table 3. In neither of the two systems evaluated this value exceeded 2.00%, which shows an acceptable degree of closeness between the data obtained in a relatively short period of time and under the same instrumental conditions.

The experimental data of intermediate precision and reproducibility expressed as coefficient of variation did not exceed 4% as shown in Table 3. This allows concluding that there is no high dispersion between experimental data from the same samples obtained in two different days by different analysts, and additionally there is no high dispersion between data from the same samples obtained by two different laboratories under the proposed method.

**Table 3:** Repeatability, intermediate precision, Reproducibility, and repeatability at stability conditions.

Sunscreen (INCI name)	Repeatability (RSD ≤ 2%)		Intermediate precision (RSD global ≤ 4 %)	Reproducibility (RSD global ≤ 4 %)	Repeatability (Stability Conditions) (RSD ≤ 4 % global)
	System	Method			
Phenylbenzimidazole Sulfonic Acid	1.97	1.89	2.9	1.87	1.3
Benzophenone-3	1.99	1.98	3.63	3.64	1.83
Diethylamino Hydroxybenzoyl Hexyl Benzoate	1.96	1.71	1.36	1.94	2.31
Octocrylene	1.74	1.99	1.49	3.05	1.14
Butyl Methoxydibenzoylmethane	1.97	1.87	2.36	3.52	1.97
Ethylhexyl Methoxycinnamate	1.92	1.99	2.63	1.41	2.08
Ethylhexyl Salicylate	1.74	1.96	2.25	1.55	3.49
Homosalate	1.96	1.99	2.54	1.38	2.13

The averages of the recovery percentage can be seen in Table 4. All the experimental values obtained in this evaluation are within the allowed range (98-102) % for an analytical method of quantification of chromatographic active ingredients. Using t student ( $t_{ab} \rightarrow 2.306$ ,  $\alpha \rightarrow 0.025$ ) statistically significant difference was not find between the mean recovery value and 100% theoretical. Therefore, it concludes the extraction step of the analysis method is acceptably accurate.

**Table 4:** Percent recovery.

Sunscreen (INCI name)	Recovery % (98-102%)
Phenylbenzimidazole Sulfonic Acid	99.97
Benzophenone-3	99.50
Diethylamino Hydroxybenzoyl Hexyl Benzoate	100.34
Octocrylene	99.81
Butyl Methoxydibenzoylmethane	100.25
Ethylhexyl Methoxycinnamate	100.51
Ethylhexyl Salicylate	100.08
Homosalate	100.42

### Robustness

The combinations carried out experimentally with the variations of the five parameters already mentioned can be seen in Table 5. The difference caused by each evaluated parameter (D) was quantified as indicated in the Equations 3-7.

$$D_{pH}: |(Y1+Y4+Y5+Y7)-(Y2+Y3+Y6+Y8)| \cdot 4^{-1} \quad (3)$$

$$D_T: |(Y1+Y2+Y5+Y8)-(Y3+Y4+Y6+Y7)| \cdot 4^{-1} \quad (4)$$

$$D_i: |(Y1+Y2+Y3+Y7)-(Y4+Y5+Y6+Y8)| \cdot 4^{-1} \quad (5)$$

$$D_m: |(Y2+Y3+Y4+Y8)-(Y1+Y5+Y6+Y7)| \cdot 4^{-1} \quad (6)$$

$$D_{NaOH}: |(Y3+Y4+Y5+Y7)-(Y1+Y2+Y6+Y8)| \cdot 4^{-1} \quad (7)$$

Where  $Y_i$  is the signal area of I-experiment in the Table 5.

**Table 5:** Range of variation of the parameters in the robustness test. Buffer pH + (2.1), - (1.9); Ultrasound temperature +(60°C), - (40°C); Ultrasound time+(20 min), - (10 min); Vial material+(plastic), - (glass); Volume of NaOH + (1.5 mL), - (0.5ml)

Experiment No.	Factor Buffer pH	Ultra-sound temperature	Ultra-sound time	Vial material	Volume of NaOH
1	+	+	+	-	-
2	-	+	+	+	-
3	-	-	+	+	+
4	+	-	-	+	+
5	+	+	-	-	+
6	-	-	-	-	-
7	+	-	+	-	+
8	-	+	-	+	-

The effect of each variable was compared with the critical value of each analyte, it was found that for two analytes there are significant effects due to the slight variation in some variables. Regarding phenylbenzimidazole sulfonic acid all the slight changes in the evaluated variables except the pH of the buffer, critically affect the result, and with respect to butyl methoxydibenzoylmethane, the critical variables were the pH of the buffer and the material of the vial.

The above suggests special care during the extraction stage of the analytes from the matrix but also during the chromatographic analysis avoiding pH changes in the buffer, using glassware in each of the stages, therefore it is important to guarantee the conditions proposed in the method.



## CONCLUSION

The experimental results show an HPLC method that complies with the validation parameters indicated by the ICH. Additionally, it is a quick and easy method. Quick because it requires an analysis time of eight minutes, and simple due to the ease in the preparation of the sample and its development under isocratic conditions.

The method can be used in quality control processes for the simultaneous quantification of eight organic UV-filters (phenylbenzimidazole sulfonic acid, benzophenone-3, diethylamino hydroxybenzoyl hexyl benzoate, octocrylene, butyl methoxydibenzoylmethane, ethylhexyl methoxycinnamate, ethylhexyl salicylate, homosalate) in cosmetic emulsion-type products that declare some level of sun protection.

## DATA AVAILABILITY

The data used to support the findings of this study are available from the corresponding author upon request.

## ACKNOWLEDGEMENTS

The authors thank the Innovation and Development department of the company Belstar S.A. for the financial support provided for the development and verification of analysis methodologies. This work was funded by Belcorp. All authors are Belcorp employees at the time of writing.

## REFERENCES

1. Almutawa F, Alnomair N, Wang Y, Hamzavi I, Lim HW. Systematic review of UV-based therapy for psoriasis. *Am J Clin Dermatol*. 2013;14(2):87–109.
2. Bednarczyk P, Pawlikowska M, Czech Z. Primers used in UV-curable nails varnishes. *Int J Adhes Adhes*. 2017;74(January):177–80.
3. World Health Organization. Global Solar UV Index: A Practical Guide. 2002; 28.
4. Moldovan HR, Wittlich M, John SM, Brans R, Tiplica GS, Salavastru C, et al. Exposure to solar UV radiation in outdoor construction workers using personal dosimetry. *Environ Res*. 2020;181:108967.
5. El-Boury S, Couteau C, Boulande L, Papis E, Coiffard LJM. Effect of the combination of organic and inorganic filters on the Sun Protection Factor (SPF) determined by in vitro method. *Int J Pharm*. 2007;340(1–2):1–5.
6. Solar and ultraviolet radiation. Vol. 55, IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Francia; 2011; 35–101.
7. Gasparro FP, Mitchnick M, Nash JF. A Review of sunscreen safety and efficacy. *Photochem Photobiol*. 1998;68(3):243–56.
8. Serpone N, Dondi D, Albini A. Inorganic and organic UV filters: Their role and efficacy in sunscreens and suncare products. *Inorganica Chim Acta*. 2007;360(3):794–802.
9. Claudia M, Issa A. Daily routine in cosmetic dermatology. *Daily routine in cosmetic dermatology*. 2016.
10. Calvo BT. About sunscreens get the best of our health advice what the study found. 2020;1–7.
11. Minu M, Rosca M, Cozma P, Catrinescu CGM. International conference on e-health and bioengineering. In: Ecological and human health risks generated by organic uv filters. Iasi, Romania; 2019; 6–9.
12. Schaap I, Slijkerman DME. An environmental risk assessment of three organic UV-filters at Lac Bay, Bonaire, Southern Caribbean. *Mar Pollut Bull*. 2018;135(July):490–5.
13. Manaia EB, Kaminski RCK, Corrêa MA, Chiavacci LA. Inorganic UV filters. *Brazilian J Pharm Sci*. 2013;49(2):201–9.
14. Schlumpf M, Schmid P, Durrer S, Conscience M, Maerker K, Henseler M, et al. Endocrine activity and developmental toxicity of cosmetic UV filters-An update. *Toxicology*. 2004;205(1-2 SPEC. ISS.):113–22.
15. Kim K, Mueller J, Park YB, Jung HR, Kang SH, Yoon MH, et al. Simultaneous determination of nine UV filters and four preservatives in suncare products by high-performance liquid chromatography. *J Chromatogr Sci*. 2011;49(7):554–9.
16. Nyeberg M, Pissavini M, Lemasson Y, Doucet O. Validation of HPLC method for the simultaneous and quantitative determination of 12 UV-filters in cosmetics. *Int J Cosmet Sci*. 2010;32(1):47–53.
17. US. Food and drug administration. Guideline for Industry: Text on Validation of Analytical Procedures (ICH-Q2A). Food and Drug Administration. 1995; 1–9.
18. US. Food and drug administration. Guidance for Industry Q2B validation of analytical procedures: Methodology, 1996;20857:1–10.