

Immunome Research

Open Access

Elevated Levels of Autoantibodies against ATP2B4 and BMP-1 in Sera of Patients with Atherosclerosis-related Diseases

Takaki Hiwasa¹', Toshio Machida², Xiao-Meng Zhang¹, Risa Kimura¹, Hao Wang^{1,3}, Katsuro Iwase¹, Hiromi Ashino¹, Akiko Taira¹, Emiko Arita¹, Seiichiro Mine^{4,5}, Mikiko Ohno⁶, Po-Min Chen⁶, Eiichiro Nishi⁶, Kenichiro Kitamura⁷, Rika Yamazoe⁸, Hirotaka Takizawa⁹, Koichi Kashiwado¹⁰, Ikuo Kamitsukasa¹¹, Takeshi Wada¹², Akiyo Aotsuka¹², Eiichi Kobayashi⁵, Tomoo Matsutani⁵, Yasuo Iwadate⁵, Naokatsu Saeki⁵, Masahiro Mori¹³, Akiyuki Uzawa¹³, Mayumi Muto¹³, Kazuo Sugimoto^{1,13}, Satoshi Kuwabara¹³, Yo Iwata¹⁴, Takashi Nakayama¹⁵, Jun-ya Harada¹⁵, Yoshio Kobayashi¹⁵, Minoru Takemoto¹⁶, Kazuki Kobayashi¹⁶, Harukiyo Kawamura¹⁶, Ryoichi Ishibashi¹⁶, Ken-ichi Sakurai¹⁶, Masaki Fujimoto¹⁶, Koutaro Yokote¹⁶, Ken-ichiro Goto^{1,17}, Ryutaro Matsumura¹⁸, Takao Sugiyama¹⁹, Haruyuki Hayashi²⁰, Ritsuko Hasegawa²¹, Hideaki Shimada²², Masaaki Ito²², Takashi Kudo²³, Hirofumi Doi²³, Rika Nakamura^{1,24}, Go Tomiyoshi^{1,24}, Natsuko Shinmen^{1,24} and Hideyuki Kuroda²⁴

¹Department of Biochemistry and Genetics, Chiba University, Graduate School of Medicine, Chiba, Japan

²Department of Neurosurgery, Chiba Cerebral and Cardiovascular Center, Chiba, Japan

³Department of Anesthesia, The First Affiliated Hospital, Jinan University, Guangdong, P. R. China

⁴Department of Neurological Surgery, Chiba Prefectural Sawara Hospital, Chiba, Japan

⁵Department of Neurological Surgery, Chiba University, Graduate School of Medicine, Chiba, Japan

⁶Department of Cardiovascular Medicine, Graduate School of Medicine, Kyoto University, Kyoto, Japan

⁷Department of Internal Medicine 3, University of Yamanashi School of Medicine, Yamanashi, Japan

⁸Department of Nephrology, Kumamoto University Graduate School of Medical Sciences, Kumamoto, Japan

⁹Port Square Kashiwado Clinic, Kashiwado Memorial Foundation, Chiba, Japan

¹⁰Department of Neurology, Kashiwado Hospital, Chiba, Japan

¹¹Department of Neurology, Chiba Rosai Hospital, Chiba, Japan

¹²Department of Internal Medicine, Chiba Aoba Municipal Hospital, Chiba, Japan

¹³Department of Neurology, Chiba University, Graduate School of Medicine, Chiba, Japan

¹⁴Department of Cardiovascular Medicine, Chiba Cerebral and Cardiovascular Center, Chiba, Japan

¹⁵Department of Cardiovascular Medicine, Chiba University, Graduate School of Medicine, Chiba, Japan

¹⁶Department of Clinical Cell Biology and Medicine, Chiba University, Graduate School of Medicine, Chiba, Japan

¹⁷Department of Orthopedics, National Hospital Organization Chiba-East-Hospital, Chiba, Japan

¹⁸Department of Rheumatology, National Hospital Organization Chiba-East-Hospital, Chiba, Japan

¹⁹Department of Rheumatology, Shimoshizu National Hospital, Chiba, Japan

²⁰Department of Surgery, Sanai Memorial Soga Hospital, Chiba, Japan

²¹Department of Internal Medicine, Sanai Memorial Soga Hospital, Chiba, Japan

²²Department of Surgery, School of Medicine, Toho University, Tokyo, Japan

²³Celish Fd Inc., Chiba Japan

²⁴Medical Project Division, Research Development Center, Fujikura Kasei Co., Saitama, Japan

Abstract

Background: Atherosclerosis-related life-style diseases such as cerebral infarction (CI), cardiovascular disease (CVD), diabetes mellitus (DM), and chronic kidney disease (CKD) are a serious problem in the recently aging society. The development of novel and sensitive diagnostic markers is necessary and expected for the early treatment. **Methods and Results:** Through the first screening by phage expression cloning, we identified ATPase, Ca⁺⁺ transporting, plasma membrane 4 (ATP2B4) and bone morphogenetic protein 1 (BMP-1) as antigens recognized by IgG antibodies in the sera of patients with atherosclerosis. The presence of autoantibodies against these antigens in serum specimens was confirmed by Western blotting. We then compared serum antibody levels against recombinant ATP2B4 and BMP-1 proteins between healthy donors (HD) and patients with atherosclerotic diseases, such as CI, transient ischemic attack (TIA), CVD, DM, or CKD, by the Alpha (amplified luminescent proximity homogeneous assay)-LISA method. The results revealed that both antibody levels were significantly higher in patients with these diseases than in HD and exhibited most prominent differences in CKD vs. HD. Correlation analysis showed that both antibody levels were related to hypertension, whereas anti-BMP-1 antibodies were related to smoking habits.

Conclusions: The serum antibody levels against ATP2B4 and BMP-1 can be useful diagnostic markers for atherosclerosis and its related diseases caused by hypertension and smoking habits, respectively.

Keywords: Atherosclerosis; Diabetes mellitus; Chronic kidney disease; Cerebral infarction; Cardiovascular disease; Antibody biomarker

Abbreviations: ABI: Ankle-Brachial Pressure Index; ATP2B4-Abs: Anti-ATP2B4 Antibodies; ATS: Atherosclerosis; AUC: Areas Under the Curve; BMP-1-Abs: Anti-BMP-1 Antibodies: CAVI: Cardio-Ankle Vascular Index: CI: Cerebral Infarction: CKD: Chronic Kidney Disease; CVD: Cardiovascular Disease; DM: Diabetes Mellitus; *E. coli: Escherichia coli*; GST: Glutathione-S-Transferase; HD: Healthy Donor; HDL: High Density Lipoprotein; IMT: Intima-media Thickness; IPTG: Isopropyl-β-D-Thiogalactoside; LDL: Low Density Lipoprotein; *Corresponding author: Takaki Hiwasa, Department of Biochemistry and Genetics, Chiba University, Graduate School of Medicine, Inohana 1-8-1, Chuo-ku, Chiba 260-8670, Japan, Tel: +81 432262541; E-mail; hiwasa takaki@faculty.chiba-u.jp

Received July 25, 2015; Accepted August 14, 2015; Published August 17, 2015

Citation: Hiwasa T, Machida T, Zhang XM, Kimura R, Wang H, et al. (2015) Elevated Levels of Autoantibodies against ATP2B4 and BMP-1 in Sera of Patients with Atherosclerosis-related Diseases. Immunome Res 11: 097. doi: 10.4172/17457580.1000097

Copyright: © 2015 Hiwasa T, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Page 2 of 9

ROC: Receiver Operating Curve; SEREX: Serological Identification of Antigens by Recombinant cDNA Expression Cloning.

Introduction

The development of atherosclerosis (ATS) leads to the onset of cerebral infarction (CI), cardiovascular disease (CVD), and chronic kidney disease (CKD), which are also caused by diabetes mellitus (DM) [1]. Thus, these ATS-related diseases are regarded as typical life-style diseases and are major causes of mortality worldwide [2,3]. Thus far, many risk factors have been identified for these diseases, including hypertension, hyperlipidemia, body mass index (BMI)/obesity, smoking habits, and family history [4-6]. Blood test parameters, such as highdensity lipoprotein (HDL)-cholesterol, low-density lipoprotein (LDL)cholesterol, total cholesterol (TC), glycohemoglobin (HbA1c) [7], and uric acid [8], were also introduced for evaluating the development of ATS-related diseases. However, these parameters are still insufficient for predicting the onset of CI and CVD. It is possible to prevent the onset of these diseases by early treatment and/or changing the life-style. The development of an early diagnosis and prediction system using novel and sensitive biomarkers is indispensable.

Till date, only few antibody biomarkers have also been identified, for example, oxidized low density lipoprotein and β 2-glycoprotein I for ATS [9,10], and heat shock proteins (Hsps) for CVD [11], Hsp60 for stroke [12], and GAD for DM [13,14]. The SEREX (serological identification of antigen by recombinant cDNA expression cloning) [15] method is effective and convenient for comprehensive screening of tumor antigens [16-18]. We also introduced SEREX into the screening of autoantigens and autoantibodies responsible for ATS-related diseases and identified RPA2 for stroke [19] and TUBB2C for DM and CI [20]. Here, we report anti-ATP2B4 and anti-BMP-1 antibodies as general ATS markers related to CI, CVD, DM, and CKD.

Materials and Methods

Patients and healthy donor (HD) sera

The Local Ethical Review Board of the Chiba University, Graduate School of Medicine as well as those of co-operating hospitals approved the study. Sera were collected from patients after they had given written informed consent. Each serum sample was centrifuged at $3,000 \times \text{g}$ for 10 min, and then the supernatant was stored at -80° C until use. Repeated thawing and freezing of samples were avoided.

The serum samples of CI were obtained from Chiba Prefectural Sawara Hospital, Chiba Rosai Hospital and Chiba Aoba Municipal Hospital. The samples of CVD and DM were obtained from Kyoto University Hospital and Chiba University Hospital, and those of CKD were from the Kumamoto cohort and Sanai Memorial Soga Hospital. Sera from HDs were obtained from Chiba University, Chiba Prefectural Sawara Hospital, Shimoshizu National Hospital and Port Square Kashiwado Clinic.

Screening by expression cloning

Recombinant DNA studies were performed with the official permission of the Chiba University Graduate School of Medicine and were carried out in accordance with the rules of the Japanese government. We used a commercially available human microvascular endothelial cell cDNA library (Uni-ZAP XR Premade Library, Stratagene, La Jolla, CA) to screen for clones that were immunoreactive against serum IgG from patients with severe carotid stenosis as previously described [19]. *Escherichia coli (E. coli)* XL1-Blue MRF' was

infected with Uni-ZAP XR phage and the expression of resident cDNA clones was induced after blotting the infected bacteria onto NitroBind nitrocellulose membranes (Osmonics, Minnetonka, MN) that had been treated with 10 mM isopropyl-β-D-thiogalactoside (IPTG, Wako Pure Chemicals, Osaka, Japan) for 30 min. The membranes with bacterial proteins were rinsed 3 times with TBS-T [20 mM Tris-HCl (pH 7.5), 0.15M NaCl and 0.05% Tween-20], and non-specific binding was blocked by incubation with 1% protease-free bovine serum albumin (Nacalai Tesque, Inc., Kyoto, Japan) in TBS-T for 1 h. The membranes were exposed to 1:2000-diluted sera of patients for 1 h. After three washes with TBS-T, the membranes were incubated for 1 h with 1:5000-diluted alkaline phosphatase-conjugated goat anti-human IgG (Jackson ImmunResearch Laboratories, West Grove, PA). Positive reactions were developed using 100 mM Tris-HCl (pH 9.5) containing 100 mM NaCl, 5 mM MgCl., 0.15 mg/ml of 5-bromo-4-chloro-3indolylphosphate, and 0.3 mg/ml of nitro blue tetrazolium (Wako Pure Chemicals). Positive clones were re-cloned twice until obtaining monoclonality as previously described [17,19,20].

Monoclonal phage cDNA clones were converted to pBluescript phagemids by excision *in vivo* using the ExAssist helper phage (Stratagene). Plasmid pBluescript containing cDNA was obtained from the *E. coli* SOLR strain after transformation by the phagemid. The sequences of cDNA inserts were evaluated for homology with identified genes or proteins within the public sequence database (http://blast.ncbi. nlm.nih.gov/Blast.cgi).

Expression and purification of antigen proteins

The expression plasmids of glutathione-S-transferase (GST)-fused proteins were constructed by recombining the cDNA sequences into pGEX-4T-3 (GE Healthcare Life Sciences, Pittsburgh, PA). The inserted DNA fragments were ligated in frame to pGEX-4T-3 using the Ligation Convenience Kits (Nippon Gene, Toyama, Japan). Ligation mixtures were used to transform ECOS^{**}-competent *E. coli* BL-21 (Nippon Gene), and appropriate recombinants were confirmed by DNA sequencing as well as protein expressions. Treating the transformed *E. coli* with 0.1 mM IPTG for 3 h induced the expression of the GSTfusion proteins. The GST-fused recombinant proteins were purified by glutathione-Sepharose column chromatography according to the manufacturer's instructions (GE Healthcare Life Sciences) and dialyzed against phosphate-buffered saline as previously described [16,21].

Western blotting

GST, GST-ATP2B4, and GST-BMP-1 proteins $(0.3 \ \mu g)$ were electrophoresed through SDS-polyacrylamide gel followed by Western blotting using anti-GST (Rockland, Gilbertsville, PA) or sera from patients with CI (#350, #462, and #692). After incubation with horseradish peroxidase-conjugated secondary antibody, immunoreactivity was detected with the Immobilon (Merck Millipore, Darmstadt, Germany) as previously described [18,22].

AlphaLISA (Amplified Luminescence Proximity Homogeneous Assay)

AlphaLISA was performed using 384-well microtiter plates (white opaque OptiPlate[™], Perkin Elmer) containing 2.5 μ l of 1/100-diluted sera and 2.5 μ l of GST or GST-fusion proteins (10 μ g/ ml) in AlphaLISA buffer (25 mM HEPES, pH 7.4, 0.1% casein, 0.5% Triton X-100, 1 mg/ml dextran-500, and 0.05% Proclin-300). The reaction mixture was incubated at room temperature for 6-8 h. Next, anti-human IgG-conjugated acceptor beads (2.5 μ l of 40 μ g/ml) and glutathione-conjugated donor beads (2.5 μ l of 40 μ g/ml) were added and incubated further for 7-14 days at room temperature in the dark. The chemical emission was read on an EnSpire Alpha microplate reader (PerkinElmer) as previously described [20,23]. Specific reactions were calculated by subtracting Alpha values of GST control from the values of GST-fusion proteins.

Statistical analyses

Student's *t* test and the Mann-Whitney U test were used to determine the significance of the differences between two groups. Correlation was examined by Spearman's correlation analysis. All statistical analyses were carried out using the GraphPad Prism 5 (GraphPad Software, La Jolla, CA). Multivariate logistic regression analysis was used to find a set of variables classifying the subjects into those with and without a history of stroke. The predictive values of markers for diseases were assessed by receiver operating curve (ROC) analysis and the cut-off values were set at the values that maximize the sums of the sensitivity and specificity. All tests were two-tailed and a P value below 0.05 was considered significant.

Results

Identification of BMP-1 recognized by sera of patients with ATS

Expression cloning using the sera of patients with both ATS has identified two clones, TS22I and A1J3, which showed a sequence homology with ATPase, Ca⁺⁺ transporting, plasma membrane 4 (ATP2B4) (Accession number: NM_001001396.2), and bone morphogenetic protein 1 (BMP-1) (Accession number: NM_006129.4), respectively. Recombinant ATP2B4 and BMP-1 proteins were expressed in *E. coli* as GST-fusion proteins and purified by affinity-chromatography using glutathione-Sepharose.

Confirmation of the presence of serum antibodies by Western blotting

The presence of anti-ATP2B4 antibodies (ATP2B4-Abs) and anti-BMP-1 antibodies (BMP-1-Abs) in the sera of patients was confirmed by Western blotting analysis. GST-ATP2B4 and GST-BMP-1 as well as GST proteins were recognized by anti-GST antibody as reactions of 55-



Figure 1: Serum antibodies recognized GST-ATP2B4 or GST-BMP-1. Representative results of Western blotting are shown. The original SDSpolyacrylamide gel was stained with Coomassie Brilliant Blue (CBB), or the membranes blotted with antigen proteins were probed with anti-GST (αGST), or patient sera, numbers #350, #462 and #692. Arrows indicate specific reactions, and the asterisk represents a reaction between a bacterial protein and serum antibodies, which does not affect the AlphaLISA results because the reactivity to only GST-fusion proteins is evaluated by trapping by glutathioneconjugated donor beads. Molecular weights are shown to the left.

kDa, 85-kDa and 28-kDa proteins, respectively (Figure 1). On the other hand, only GST-ATP2B4 or GST-BMP-1 was reacted with the serum antibodies of patients # 350 and #462, respectively. Serum #692 showed the reactivity to GST, which eventually enabled the recognition of both GST-ATP2B4 and GST-BMP-1. Therefore, in the following study, specific reactions against ATP2B4 or BMP-1 proteins were estimated by antibody levels toward GST-tagged antigen proteins subtracted by the levels toward GST.

Levels of ATP2B4-Abs and BMP-1-Abs are increased in patients with CI

We examined the levels of serum ATP2B4-Abs and BMP-1-Abs using the sera of HD and patients with TIA or in the acute phase of CI (aCI) obtained from Chiba Prefectural Sawara Hospital, Chiba Rosai Hospital and Chiba Aoba Municipal Hospital. HD subjects from Kashiwado Clinic and Chiba Prefectural Sawara Hospital were selected as those exhibiting no abnormalities on MRI examination. The results of AlphaLISA showed that both levels of ATP2B4-Abs and BMP-1-Abs were significantly higher in patients with TIA or aCI than in HD (Figures 2a and 2b). When the cut-off value was determined as the average + 2SD of HD, the positive rates of ATP2B4-Abs in HD and patients with TIA and aCI patients were 3.9%, 13.8%, and 17.3%,



Figure 2: Comparison of serum ATP2B4-Ab and BMP-1-Ab levels between HD and patients with TIA or aCI. Antigens used were GST-ATP2B4 (a) and GST-BMP-1 proteins (b). Serum antibody levels subtracted by the levels against control GST examined by AlphalLISA are shown by a box-whisker plot. The box plots display the 10th, 20th, 50th, 80th, and 90th percentiles. P values vs. HD specimens are shown. In Table 1, averages, SDs, cut-off values, total numbers, positive numbers, positive rates (%), and P values are shown. Receiver operating curve (ROC) analysis was carried out for assessing the ability of ATP2B4-Abs (c and e) and BMP-1-Abs (d and f) to detect TIA (c and d) or aCl (e and f). Areas under the curve (AUC) were 0.690 (95% CI: 0.614-0.766) (c), 0.615 (95% CI: 0.537-0.693) (d), 0.619 (95% CI: 0.553-0.684) (e), and 0.577 (95% CI: 0.511-0.642) (f). Numbers in the curves indicate cut-off values of marker levels and those in the parentheses indicate sensitivity (left) and specificity (right). P values are also shown.

respectively, whereas the positive rates of BMP-1-Abs were 0.8%, 3.9%, and 3.9%, respectively (Table 1). Thus, positivity of BMP-1-Abs in TIA and aCI was less prominent as compared with that of ATP2B4-Abs. Receiver operating curve (ROC) analysis was carried out to evaluate the ability of these markers to detect TIA and aCI. The areas under the curve (AUC) of ATP2B4-Abs and BMP-1-Abs for TIA were 0.690 (95% CI: 0.614-0.766) (Figure 2c) and 0.615 (95% CI: 0.537-0.693) (Figure 2d), respectively, whereas those for aCI were 0.619 (95% CI: 0.553-0.684) (Figure 2e) and 0.577 (95% CI: 0.511-0.642) (Figure 2f), respectively. When the cut-off value of the ATP2B4-Ab level was determined to be 40,076, the sensitivity and specificity of the antibody level for the diagnosis of TIA were 40.3% and 90.2%, respectively (Figure 2c).

Levels of BMP-1-Abs are associated with CVD

We next examined the antibody levels of CVD, including acute myocardial infarction and unstable angina, in samples obtained from Chiba University Hospital and Kyoto University Hospital. The levels of both ATP2B4-Abs and BMP-1-Abs were significantly higher in patients with CVD than those in HD; yet, BMP-1-Ab levels differed more than ATP2B4-Ab levels (Figures 3a and 3b), which was reflected in the positive rates. The positive rates of ATP2B4-Abs in HD and patients with CVD were 4.4% and 17.2%, respectively, whereas the positive rates of BMP-1-Abs were 5.7% and 23.4%, respectively (Table 2). ROC analysis revealed that AUCs of ATP2B4-Abs and BMP-1-Abs for CVD were 0.653 (95% CI: 0.567-0.738) (Figure 3c) and 0.680 (95% CI: 0.636-0.725) (Figure 3d), respectively.

Levels of ATP2B4-Abs and BMP-1-Abs are related to DM

Because ATS is closely related to DM, we then compared the specimens of HD and DM obtained from Shimoshizu National Hospital and Chiba University Hospital, respectively. Both ATP2B4-Abs and BMP-1-Abs markedly increased in patients with DM as compared with

		COT ATDODA	COT DMD 4
		GST-ATP2B4	GST-DIVIP-T
	Average	51,248	16,161
	SD	30,365	6,858
	Cut-off value	111,979	29,878
пр	Total No.	128	123
	Positive No.	5	1
	Positive rate (%)	3.9%	0.8%
	Average	72,672	18,806
	SD	36,128	5,871
T I A	Total No.	29	77
IIA	Positive No.	4	3
	Positive rate (%)	13.8%	3.9%
	P (vs HD)	0.0052	0.0042
	Average	74,380	18,806
-	SD	35,931	5,871
-01	Total No.	127	77
aur	Positive No.	22	3
	Positive rate (%)	17.3%	3.9%
	P (vs HD)	7.38E-08	0.0042

Table 1: Comparison of serum antibody levels between HD and patients with TIA or acute CI (aCI) examined by AlphaLISA. Shown are average, SD, cut-off values (average+2SD), total sample number, the number of positive sera of which antibody levels were higher than the cut-off value, and the positive rate (%) of HD; and average, SD, total sample number, number of positive sera of which the antibody levels were higher than the cut-off value, and the positive rate (%) of patients; and P values of statistical comparison between HD and patients. Antigens used were purified GST-ATP2B4 and GST-BMP-1 proteins. P values lower than 0.05 and positive rates higher than 10% are marked in bold.



Figure 3: Comparison of serum ATP2B4-Ab and BMP-1-Ab levels between HD and patients with CVD. Antigens used were GST-ATP2B4 (a) and GST-BMP-1 proteins (b). Serum antibody levels examined by AlphalLISA are shown by a box-whisker plot as described in the legends of Figure 2. In Table 2, averages, SDs, cut-off values, total numbers, positive numbers, positive rates (%), and P values are shown. The results were also evaluated by ROC analysis (c and d). AUCs of ATP2B4-Abs and BMP-1-Abs were 0.653 (95% CI: 0.567-0.738) (c) and 0.680 (95% CI: 0.636-0.725) (d), respectively.

		ATP2B4	BMP-1
ЦП	Average	21,325	4,859
пр	SD	12,132	3,386
	Cut-off value	45,588	11,630
	Total No.	113	192
	Positive No.	5	11
	Positive rate (%)	4.4%	5.7%
	Average	29,923	8,667
	SD	16,926	7,258
CVD	Total No.	64	380
CVD	Positive No.	11	89
	Positive rate (%)	17.2%	23.4%
	P (vs HD)	0.00054	1.14E-16

 Table 2: Comparison of serum antibody levels between HD and patients with CVD examined by AlphaLISA. Antigens used were purified GST-ATP2B4 and GST-BMP-1 proteins. See Table 1 for further details.

those in HD (Figure 4). The positive rates of ATP2B4-Abs in HD and patients with CVD were 4.9% and 28.0%, respectively, whereas the positive rates of BMP-1-Abs were 2.5% and 17.8%, respectively (Table 3). ROC analysis revealed that AUCs of ATP2B4-Abs and BMP-1-Abs for DM were 0.714 (95% CI: 0.655-0.772) (Figure 4c) and 0.670 (95% CI: 0.606-0.733) (Figure 4d), respectively.

Levels of ATP2B4-Abs and BMP-1-Abs are elevated in patients with CKD

CKD is also closely related to ATS and was divided into three groups as follows: type 1, diabetic kidney disease; type 2, nephrosclerosis; and type 3, glomerulonephritis; samples were obtained from the Kumamoto cohort. ATP2B4-Ab and BMP-1-Ab levels in patients with CKD were higher in all three types than those in HD (Figures 5a and

Page 5 of 9



Figure 4: Comparison of serum ATP2B4-Ab and BMP-1-Ab levels between HD and patients with DM. Antigens used were GST-ATP2B4 (a) and GST-BMP-1 proteins (b). Serum antibody levels examined by AlphalLISA are shown by a box-whisker plot as described in the legends of Figure 2. In Table 3, averages, SDs, cut-off values, total numbers, positive numbers, positive rates (%), and P values are shown. The results were also evaluated by ROC analysis (c and d). AUCs of ATP2B4-Abs and BMP-1-Abs were 0.714 (95% CI: 0.655-0.772) (c), and 0.670 (95% CI: 0.606-0.733) (d), respectively.

		GST-ATP2B4	GST-BMP-1
	Average	6,865	2,150
	SD	3,507	928
ЦБ	Cut-off value	13,880	4,006
пО	Total No.	81	81
	Positive No.	4	2
	Positive rate (%)	4.9%	2.5%
	Average	11,833	3,013
	SD	8,388	1,859
DM	Total No.	275	275
DM	Positive No.	77	49
	Positive rate (%)	28.0%	17.8%
	P (vs HD)	1.05E-13	3.71E-08

 Table 3: Comparison of serum antibody levels between HD and patients with DM

 examined by AlphaLISA. Antigens used were purified GST-ATP2B4 and GST-BMP-1 proteins. See Table 1 for further details.

5b). The positive rates of ATP2B4-Abs in HD, CKD type 1, type 2 and type 3 were 5.0%, 37.2%, 43.8%, and 19.5%, respectively, whereas the positive rates of BMP-1-Abs were 3.8%, 35.9%, 37.5%, and 20.3% respectively (Table 4). Little difference was found among CDK types. The total positive rates of ATP2B4-Abs and BMP-1-Abs were 30.7% and 29.7%, respectively. BMP-1-Ab levels were also examined in serum samples obtained from patients on dialysis in Sanai Memorial Soga Hospital, and similar differences between HD and patients with CKD were observed (Figure 5c and Table 5). ROC analysis revealed that AUCs of ATP2B4-Abs and BMP-1-Abs for CKD type 1 were 0.789 (95% CI: 0.730-0.848) (Figure 5d) and 0.770 (95% CI: 0.708-0.831) (Figure 5e), respectively. Thus, CKD was most associated with both ATP2B4-Abs and BMP-1-Abs with such high sensitivity and specificity.

Levels of BMP-1-Abs are related to cancer

Autologous antibodies frequently developed in patients with cancer [24]. We therefore examined the samples from patients with benign glioma, malignant glioma or esophageal squamous cell carcinoma (SCC) obtained from Chiba University Hospital and Toho University Hospital. The levels of BMP-1-Abs were significantly elevated in esophageal SCC as compared with HD (Table 6). ATP2B4-Ab levels were also elevated in esophageal SCC but less prominently. No apparent difference in both ATP2B4-Abs and BMP-1-Abs was observed between HD and patients with benign or malignant gliomas.

Correlation analysis between ATP2B4-Abs and BMP-1-Abs and atherosclerosis indices

We then carried out Spearman correlation analysis and multivariate logistic regression analysis between antibody marker levels and data on study individuals, including gender, age, height, weight, BMI, blood pressure, smoking habit, and the degree of artery stenosis such as maximum intima-media thickness (max IMT), as well as complication of DM, hypertension, CVD, lipidemia, and CI. Blood test data, such as data on total protein, albumin/globulin ratios, aspartate aminotransferase, alanine aminotransferase, alkaline phosphatase, lactate dehydrogenase, total bilirubin, γ -glutamyl transpeptidase, albumin, blood urea nitrogen, creatinin, estimated glomerular filtration rates, uric acid, amylase, total cholesterol, HDL-cholesterol, LDL-cholesterol, triglyceride, Na, K, Cl, Ca, P, Fe, C-reactive protein, blood



Figure 5: Comparison of serum ATP2B4-Ab and BMP-1-Ab levels between HD and patients with CKD. Serum antibody levels against GST-ATP2B4 (a) and GST-BMP-1 proteins (b) were compared between HD and patients with CKD types 1, 2, and 3. (c) Serum antibody levels against GST-BMP-1 proteins were also compared between HD and dialysis patients in the Sanai Memorial Soga Hospital. Results are shown as described in the legends of Figure 2. In Tables 4 and 5, averages, SDs, cut-off values, total numbers, positive numbers, positive rates (%), and P values are shown. The results were also evaluated by ROC analysis to detect CKD type 1 (d and e). AUCs of ATP2B4-Abs and BMP-1-Abs were 0.789 (95% CI: 0.730-0.848) (d), and 0.770 (95% CI: 0.708-0.831) (e), respectively.

GST-ATP2B4GST-BMP-1Average12,8255,023SD8,0743,163Cut-off value28,97311,349Total No.8080Positive No.43Positive rate (%)5.0%3.8%Average29,20410,881SD21,8358,530Total No.145145Positive rate (%)37.2%35.9%Positive rate (%)37.2%35.9%Positive rate (%)31,99110,642Positive rate (%)31,99110,642SD17,8665,083Type 2 CKDSD1412Positive rate (%)43.8%37.5%Positive rate (%)143.8%37.5%Fype 3 CKDSD13,5375,521Positive rate (%)13,5375,521Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)300300CKD totalPositive rate (%)30.7%Positive rate (%)30.7%29.7%				
HDAverage12,8255,023SD8,0743,163Cut-off value28,97311,349Total No.8080Positive No.43Positive rate (%)5.0%3.8%Average29,20410,881SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%Positive rate (%)37.2%35.9%Positive rate (%)31,99110,642SD17,8665,083SD17,8665,083Type 2 CKDNo.1412Positive rate (%)43.8%37.5%P (vs HD)1.14E-067.85E-07Positive rate (%)13,5375,521Total No.123123Positive rate (%)13,5375,521Total No.123123Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)300300CKD totalPositive rate (%)30.7%Positive rate (%)30.7%29.7%			GST-ATP2B4	GST-BMP-1
HDSD8,0743,163Cut-off value28,97311,349Total No.8080Positive No.43Positive rate (%)5.0%3.8%Average29,20410,881SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%Positive rate (%)37.2%35.9%Positive rate (%)37.2%36.38-12Positive rate (%)31,99110,642SD17,8665,083Type 2 CKDTotal No.32Positive rate (%)43.8%37.5%Positive rate (%)14412Positive rate (%)43.8%37.5%Positive rate (%)11.4E-067.85E-07SD13,5375,521Total No.123123Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)300300CKD totalPositive rate (%)30.7%Positive rate (%)30.7%29.7%		Average	12,825	5,023
HDCut-off value28,97311,349Total No.8080Positive No.43Positive rate (%)5.0%3.8%Average29,20410,881SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%Positive rate (%)37.2%36.95Positive rate (%)31,99110,642Positive rate (%)31,99110,642SD17,8665,083Total No.3232Positive rate (%)1412Positive rate (%)1412Positive rate (%)14.623.056SD13,5375,521Positive rate (%)13,5375,521Positive rate (%)123123Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)300300Positive rate (%)300300Positive rate (%)300300Positive rate (%)30.7%29.7%		SD	8,074	3,163
HDTotal No.8080Positive No.43Positive rate (%)5.0%3.8%Positive rate (%)5.0%3.8%Average29,20410,881SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%Positive rate (%)37.2%35.9%Positive rate (%)31,99110,642SD17,8665,083Total No.3232Positive rate (%)1412Positive rate (%)43.8%37.5%Positive rate (%)43.8%37.5%Positive rate (%)11,4E-067.85E-07Positive rate (%)11,5375,521Total No.123123Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)300300CKD totalPositive rate (%)30.7%Positive rate (%)30.7%29.7%	ЧР	Cut-off value	28,973	11,349
Positive No.43Positive rate (%)5.0%3.8%Positive rate (%)5.0%3.8%SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%Positive rate (%)37.2%35.9%Positive rate (%)37.2%36.38Positive rate (%)31,99110,642SD17,8665,083Total No.3232Positive rate (%)43.8%37.5%Positive rate (%)11412Positive rate (%)114E-067.85E-07Positive rate (%)13,5375,521Total No.123123Positive rate (%)113,5375,521Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)19.5%30.0Positive rate (%)300300Positive rate (%)30.7%29.7%	пр	Total No.	80	80
Positive rate (%)5.0%3.8%Average29,20410,881SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%P (vs HD)5.62E-143.63E-12Average31,99110,642SD17,8665,083Total No.3232Positive rate (%)43.8%37.5%Positive no.1412Positive rate (%)43.8%37.5%Positive rate (%)11.4E-067.85E-07Positive rate (%)13,5375,521Total No.123123Positive no.123123Positive no.2425Positive rate (%)19.5%20.3%P (vs HD)2.30E-081.46E-06Positive rate (%)300300CKD totalPositive no.9289Positive rate (%)30.7%29.7%		Positive No.	4	3
Average29,20410,881SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%P (vs HD)5.62E-143.63E-12Average31,99110,642SD17,8665,083Total No.3232Positive rate (%)43.8%37.5%Positive Ro.1412Positive rate (%)43.8%37.5%Positive rate (%)43.8%37.5%Positive Rote (%)11.4E-067.85E-07P (vs HD)1.14E-067.85E-07SD13,5375,521Total No.2425Positive Rote (%)19.5%20.3%P (vs HD)2.30E-081.46E-06Positive rate (%)300300CKD totalPositive No.9289Positive rate (%)30.7%29.7%		Positive rate (%)	5.0%	3.8%
SD21,8358,530Total No.145145Positive No.5452Positive rate (%)37.2%35.9%P (vs HD)5.62E-143.63E-12Average31,99110,642SD17,8665,083Total No.3232Positive rate (%)43.8%37.5%Positive rate (%)43.8%37.5%Positive rate (%)43.8%37.5%Positive rate (%)1.14E-067.85E-07P (vs HD)1.14E-067.85E-07SD13,5375,521Total No.123123Positive No.2425Positive rate (%)19.5%20.3%P (vs HD)2.30E-081.46E-06Positive rate (%)300300CKD totalPositive No.9289Positive rate (%)30.7%29.7%		Average	29,204	10,881
Type 1 CKDTotal No.145145Positive No.5452Positive rate (%)37.2%35.9%P (vs HD)5.62E-143.63E-12P (vs HD)5.62E-143.63E-12Average31,99110.642SD17.8665.083Total No.3232Positive No.1412Positive rate (%)43.8%37.5%P (vs HD)1.14E-067.85E-07P (vs HD)1.14E-067.85E-07SD13,5375,521Total No.123123Positive No.2425Positive rate (%)19.5%20.3%P (vs HD)2.30E-081.46E-06Positive No.300300CKD totalPositive No.9289Positive rate (%)30.7%29.7%		SD	21,835	8,530
Positive No. 54 52 Positive rate (%) 37.2% 35.9% P (vs HD) 5.62E-14 3.63E-12 Average 31,991 10,642 SD 17,866 5,083 Total No. 32 32 Positive rate (%) 43.8% 37.5% Positive rate (%) 43.8% 37.5% P (vs HD) 1.14E-06 7.85E-07 P (vs HD) 1.14E-06 8,056 SD 13,537 5,521 Total No. 123 123 Positive rate (%) 19.5% 20.3% Positive rate (%) 19.5% 20.3% Positive rate (%) 300 300 Positive rate (%) 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		Total No.	145	145
Positive rate (%)37.2%335.9%P (vs HD)5.62E-143.63E-12Average31,99110,642SD17,8665,083Total No.3232Positive No.1412Positive rate (%)43.8%37.5%P (vs HD)1.14E-067.85E-07P (vs HD)1.14E-068,056SD13,5375,521Total No.123123Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)300300CKD totalPositive No.9289Positive rate (%)30.7%29.7%	туретско	Positive No.	54	52
P (vs HD)5.62E-143.63E-12Average31,99110,642SD17,8665,083Total No.3232Positive No.1412Positive rate (%)43.8%37.5%P (vs HD)1.14E-067.85E-07Average21,6628,056SD13,5375,521Total No.123123Positive rate (%)19.5%20.3%Positive rate (%)19.5%20.3%Positive rate (%)300300CKD totalPositive No.9289Positive rate (%)30.7%29.7%		Positive rate (%)	37.2%	35.9%
Average 31,991 10,642 SD 17,866 5,083 Total No. 32 32 Positive No. 14 12 Positive rate (%) 43.8% 37.5% P(vs HD) 1.14E-06 7.85E-07 Average 21,662 8,056 SD 13,537 5,521 Total No. 123 123 Positive rate (%) 19.5% 20.3% Positive rate (%) 19.5% 20.3% Positive rate (%) 300 300 Positive No. 92 89 Positive rate (%) 30.7% 29.7%		P (vs HD)	5.62E-14	3.63E-12
SD 17,866 5,083 Total No. 32 32 Positive No. 14 12 Positive rate (%) 43.8% 37.5% Positive rate (%) 43.8% 37.5% Positive rate (%) 1.14E-06 7.85E-07 Positive SD 11,537 5,521 Total No. 123 123 Positive No. 24 25 Positive rate (%) 19.5% 20.3% P(vs HD) 2.30E-08 1.46E-06 Positive No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		Average	31,991	10,642
Type 2 CKD Total No. 32 32 Positive No. 14 12 Positive rate (%) 43.8% 37.5% Positive rate (%) 43.8% 37.5% P (vs HD) 1.14E-06 7.85E-07 Average 21,662 8,056 SD 13,537 5,521 Total No. 123 123 Positive No. 24 25 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Positive No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		SD	17,866	5,083
Positive No. 14 12 Positive rate (%) 43.8% 37.5% P (vs HD) 1.14E-06 7.85E-07 P (vs HD) 1.14E-06 7.85E-07 Average 21,662 8,056 SD 13,537 5,521 Total No. 123 123 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Positive No. 300 300 CKD total Positive nate (%) 30.7% 29.7%		Total No.	32	32
Positive rate (%) 43.8% 37.5% P (vs HD) 1.14E-06 7.85E-07 Average 21,662 8,056 SD 13,537 5,521 Total No. 123 123 Positive No. 24 25 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 KD total Positive No. 300 300 Positive rate (%) 92 89 89 Positive rate (%) 30.7% 29.7%	Type 2 CKD	Positive No.	14	12
P (vs HD) 1.14E-06 7.85E-07 Average 21,662 8,056 SD 13,537 5,521 Total No. 123 123 Positive No. 24 25 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Positive No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		Positive rate (%)	43.8%	37.5%
Average 21,662 8,056 SD 13,537 5,521 Total No. 123 123 Positive No. 24 25 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Total No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		P (vs HD)	1.14E-06	7.85E-07
SD 13,537 5,521 Total No. 123 123 Positive No. 24 25 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Total No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		Average	21,662	8,056
Type 3 CKD Total No. 123 123 Positive No. 24 25 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Total No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		SD	13,537	5,521
Positive No. 24 25 Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Total No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		Total No.	123	123
Positive rate (%) 19.5% 20.3% P (vs HD) 2.30E-08 1.46E-06 Total No. 300 300 Positive No. 92 89 Positive rate (%) 30.7% 29.7%	Type 5 CKD	Positive No.	24	25
P (vs HD) 2.30E-08 1.46E-06 Total No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		Positive rate (%)	19.5%	20.3%
Total No. 300 300 CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		P (vs HD)	2.30E-08	1.46E-06
CKD total Positive No. 92 89 Positive rate (%) 30.7% 29.7%		Total No.	300	300
Positive rate (%) 30.7% 29.7%	CKD total	Positive No.	92	89
		Positive rate (%)	30.7%	29.7%

 Table 4: Comparison of serum antibody levels between HD and patients with CKD examined by AlphaLISA. CKD types 1, 2 and 3 were diabetic kidney disease, nephrosclerosis and glomerulonephritis, respectively. Antigens used were purified GST-ATP2B4 and GST-BMP-1 proteins. See Table 1 for further details.

		GST-BMP-1
	Average	2,021
	SD	1,542
ЧР	Cut-off value	5,104
	Total No.	78
	Positive No.	5
	Positive rate (%)	6.4%
	Average	3,292
	SD	3,221
CKD	Total No.	111
CND	Positive No.	17
	Positive rate (%)	15.3%
	P (vs HD)	0.00040

 Table 5: Comparison of serum antibody levels between HD and patients with CKD

 examined by AlphaLISA. Antigens used were purified GST-ATP2B4 and GST-BMP-1 proteins. See Table 1 for further details.

sugar, and HbA1c were also included as described [20,23]. A total of 741 specimens from the Sawara Hospital including 139 specimens from HD, 79 from patients with deep and subcortical white matter hyperintensity, 15 patients with asymptomatic CI, 29 patients with TIA, 227 with aCI, 58 with chronic CI, and 194 from disease controls. Both Spearman and multivariate analyses showed similar results. The degree of artery stenosis, such as max IMT and IMT values, smoking habit and smoking period were well correlated with both ATP2B4-Ab and BMP-1-Ab levels (Table 7). In addition, ATP2B4-Abs were associated well with blood pressure and complication of hypertension.

A similar correlation analysis was also performed for 384 specimens obtained from patients with CKD of the Kumamoto cohort were analyzed. The levels of ATP2B4-Abs showed a positive correlation with max IMT and cardio-ankle vascular index (CAVI) (Table 8), suggesting a close relation between ATP2B4-Ab levels and ATS. ATP2B4-Ab levels also correlated with ferritin, Fe and K levels. On the other hand, the levels of BMP-1-Abs were correlated with ferritin, cigarette smoking habit, AST and triglyceride but showed inverse correlation with HDLcholesterol. This implicates that the high BMP-1-Abs levels may reflect liver malfunction caused by smoking habit. No other patient data significantly correlated with the levels of ATP2B4-Abs or BMP-1-Abs.

Discussion

Phage expression cloning, also called SEREX, is effective for the screening of tumor antigens [15-18] and biomarkers of stroke [19] and DM [20]. Through the expression cloning method, we have identified two antigens that were recognized by serum IgG antibodies in patients with ATS. The presence of serum antibodies against ATP2B4 and BMP-1 in patient sera was confirmed by Western blotting (Figure 1). Furthermore, AlphaLISA enabled us to evaluate those antibody levels, and thereby, to compare the levels between HD and patients.

ATP2B4 is a member of the plasma membrane Ca^{2+} -ATPase family and involved in calcium homeostasis [25]. Several mutations in the ATP2B4 gene have been found in familial spastic paraplegia [26]. The positive rates of ATP2B4-Abs were higher in patients with DM (28.7%) and CKD (30.7%) than those in patients with aCI (17.3%), CVD (17.2%), and TIA (13.8%) (Tables 1-4). Similar high positive rates of ATP2B4-Abs in patients with DM and CKD may suggest that these

		GST-ATP2B4	GST-BMP-1
	HD Av	14,221	4,645
	HD SD	13,869	4,367
ЦБ	HD Av+2SD	41,959	13,380
пр	HD Total No.	111	111
	HD Positive No.	4	5
	HD Positive (%)	3.6%	4.5%
	P Av	14,882	5,078
	P SD	9,280	4,078
Cliama B	P Total No.	83	83
Giloina-D	P Positive No.	1	6
	P Positive (%)	1.2%	7.2%
	P (P vs HD)	0.692	0.479
	P Av	14,469	5,125
	P SD	10,366	4,151
Gliama M	P Total No.	90	90
Giloffia-Ivi	P Positive No.	2	3
	P Positive (%)	2.2%	3.3%
	P (P vs HD)	0.885	0.427
	P Av	20,144	6,895
	P SD	12,377	4,918
Esophag-	P Total No.	100	100
SCC	P Positive No.	5	11
	P Positive (%)	5.0%	11.0%
	P (P vs HD)	0.0012	0.00058

 Table 6: Comparison of serum antibody levels between HD and patients with cancer examined by AlphaLISA. Cancer specimens included benign glioma (Glioma-B), malignant glioma (Glioma-M) and esophageal squamous cell carcinoma (Esophag-SCC). Antigens used were purified GST-ATP2B4 and GST-BMP-1 proteins. See Table 1 for further details.

Page 6 of 9

Page 7 of 9

		ATP2B4			BMP-1	
	Spea	irman	Multivariate	Spe	arman	Multivariate
	r value	P value	r value	r value	P value	r value
Gender	0.012	0.762	0.025	-0.006	0.868	-0.005
Age	0.124	0.001	0.140	0.066	0.091	0.077
Height	-0.072	0.065	-0.100	-0.031	0.425	-0.046
Weight	-0.058	0.136	-0.081	-0.034	0.385	-0.033
BMI	-0.019	0.621	-0.044	-0.016	0.687	-0.018
IMT (right)	0.150	0.001	0.115	0.096	0.040	0.088
IMT (left)	0.104	0.027	0.119	0.120	0.011	0.130
max IMT	0.152	0.001	0.129	0.124	0.008	0.119
Blood pressure	0.128	0.001	0.125	0.070	0.080	0.067
Smoking	0.102	0.009	0.109	0.139	0.000	0.150
Smoking period	0.151	0.000	0.188	0.181	< 0.0001	0.201
DM (Complication)	0.050	0.202	0.016	0.038	0.323	0.030
Hypertension (Complication)	0.097	0.012	0.103	0.030	0.442	0.034
CVD (Complication)	0.033	0.398	0.034	0.010	0.803	0.004
Lipidemia (Complication)	-0.033	0.395	-0.064	-0.023	0.550	-0.030
CI (Complication)	-0.018	0.649	-0.012	0.013	0.732	0.008
Total protein	-0.047	0.239	-0.031	-0.099	0.013	-0.075
Albumin/globulin ratio	-0.062	0.119	-0.087	0.049	0.225	0.046
Aspartate aminotransferase	0.056	0.149	0.004	0.020	0.611	0.004
Alanine aminotransferase	-0.021	0.587	-0.015	0.017	0.666	0.022
Alkaline phosphatase	0.100	0.015	0.069	0.067	0.100	0.060
Lactate dehydrogenase	0.049	0.219	0.098	0.026	0.512	0.069
Total bilirubin	0.023	0.560	0.019	0.014	0.732	0.010
γ-glutamyl transpeptidase	-0.026	0.522	0.023	0.042	0.297	0.062
Albumin	-0.066	0.094	-0.092	-0.021	0.599	-0.015
Blood urea nitrogen	-0.042	0.286	0.037	-0.055	0.159	0.011
Creatinin	-0.018	0.640	0.015	-0.021	0.589	-0.002
Estimated glomerular filtration rate	-0.002	0.965	-0.008	0.039	0.365	0.033
Uric acid	-0.056	0.216	0.037	-0.022	0.626	0.040
Amylase	-0.081	0.100	-0.086	-0.092	0.060	-0.090
Total cholesterol	-0.093	0.028	-0.087	-0.065	0.122	-0.060
HDL-cholesterol	-0.003	0.954	0.022	-0.013	0.792	0.016
LDL-C	-0.097	0.073	-0.111	-0.066	0.220	-0.078
Triglyceride	-0.058	0.212	-0.039	0.007	0.874	0.054
Na	-0.044	0.263	-0.077	-0.030	0.455	-0.028
κ	-0.054	0.174	-0.047	-0.077	0.052	-0.099
CI	-0.028	0.479	-0.065	-0.008	0.838	-0.020
Са	-0.063	0.218	-0.077	-0.088	0.086	-0.109
Inorganic phosphate	-0.019	0.741	-0.016	-0.002	0.973	-0.012
Fe	-0.053	0.352	0.001	-0.015	0.792	0.022
C-reactive protein	0.069	0.133	0.026	0.012	0.800	0.005
Blood sugar	0.076	0.065	0.046	0.036	0.381	0.062
HbA1c	-0.039	0.383	0.047	-0.063	0.161	0.039

Table 7: Correlation analysis between ATP2B4 and BMP-1 antibody marker levels and data on study individuals from Sawara Hospital cohort. Correlation coefficient (r) values and P values were calculated by Spearman's correlation analysis. r values obtained by multivariate logistic regression analysis are also shown. Significant correlations are marked in bold.

diseases are interrelated; i.e., one may be the cause of the other and vice versa. However, multivariate logistic regression analysis revealed no significant correlation between ATP2B4-Abs and blood sugar, DM marker HbA1c and complication of DM (Table 7). Thus, the primary cause detected by ATP2B4-Abs may be kidney failure. The close association of ATP2B4-Abs with blood pressure and complication of hypertension (Tables 7 and 8) may be well consistent with the high expression of ATP2B4 in the aorta [27], because the aorta is most susceptible to hypertension produced by heart beating. Therefore, it is

possible that the simple leaking out of ATP2B4 protein by hypertension in the aorta as well as by ATS in the neighboring artery caused the development of the antibodies. Further, continuous high expression of ATP2B4 may disrupt calcium homeostasis leading to tissue calcification, which is the final stage of ATS.

It has been well documented that BMP signals are involved in the development of ATS [28,29]. The expression of BMP-2 and BMP-4 is elevated in atherosclerotic endothelium [30,31]. Plasma BMP-2 levels are elevated in patients with type 2 DM [32]. The chronic infusion

	ATP2B4	BMP-1
Ferritin	<0.0001	0.0035
C-reactive protein	0.0030	0.0578
max IMT	0.0041	0.1836
Fe	0.0042	0.9711
CAVI (right)	0.0111	0.1203
CAVI (left)	0.0167	0.2364
к	0.0215	0.4760
Smoking	0.2445	0.0068
Aspartate aminotransferase	0.0607	0.0114
Triglyceride	0.0563	0.0260

 Table 8: Correlation analysis between ATP2B4 and BMP-1 antibody marker levels

 and data on study individuals from Kumamoto cohort. P values were calculated

 using Spearman's correlation analysis. Significant correlations are marked in bold.

of BMP-4 induced endothelial dysfunction and hypertension [33], and treatment with the BMP antagonist, matrix gla protein, or BMP inhibitors prevent from the development of ATS [34,35]. On the other hand, the knockdown of BMP type II receptor, BMPRRII, accelerates ATS [36]. Therefore, BMP family members may have a subtle regulatory role in the development of ATS.

Unlike other members of the BMP family including BMP-2 - 15, BMP-1 is a metalloproteinase containing an astacin-like domain and is also known as procollagen C-proteinase, [37]. BMP-1 activates BMP signaling by degrading chordin which can inhibit the action of BMPs [38]. The positive rates of BMP-1-Abs were higher in patients with CKD (29.7%) and CVD (23.4%) than those in patients with DM (17.8%) and aCI (3.9%) (Tables 1-4), suggesting that BMP-1-Abs are associated with most, if not all, ATS-related diseases. It should be noted that we have found elevated antibody levels in patients with CI against SOSTDC1 [23], which is an antagonist of BMPs [39,40]. Therefore, these autologous antibodies may not be generated due to the high expression of antigen proteins; however, they may play a role in the development of ATS. This notion further implies that the antigenic cytokines can be therapeutic targets by treating patients with agonists or antagonists as appropriate.

Correlation analysis using specimens from the Kumamoto cohort and Sawara hospital cohort revealed that BMP-1-Abs correlated with smoking habit (Tables 7 and 8). Consequently, this marker can detect CVD accompanied by CKD or DM, of which the primary cause may be the smoking habit. This type of correlation analysis of biomarkers is of utmost importance because the high risk of the disease onset might be reduced by changing only marker-related life-style behaviors of the examinee.

BMP-1-Ab levels, which reflect smoking, were also higher in patients with esophageal SCC than those in HD (Table 6), probably because smoking habit is one of the major causes of esophageal SCC. However, it should be taken into account that DM is also a risk factor for cancers such as colorectal cancer and esophageal carcinoma [41-43]. Consistently, ATP2B4-Ab levels were also higher in patients with esophageal SCC than those in HD but less prominently compared with BMP-Ab levels (Table 6).

Serum samples from patients with CI were collected within two weeks after disease onset. The onset may induce spreading of various antigens, however, the antibodies against these antigens are not immediately produced. Therefore, the antibodies that were specifically detected in patient sera immediately after the onset, such as ATP2B4-Abs and BMP-1-Abs, were probably present prior to disease onset. Consistently, these autoantibody levels were markedly elevated not only in a CI but also in TIA, a harbinger of CI, compared with HD samples (Figure 2). Thus, these antibody markers appear to be prediction markers but not simple risk markers.

Page 8 of 9

ATP2B4-Abs and BMP-1-Abs are promising biomarkers for ATS-related diseases. The positive rates of each marker may not be high enough, this may be because they were associated with different causes, such as hypertension and smoking habit. Further diagnosis using combination of as many markers as possible may improve the sensitivity.

Conclusion

The levels of ATP2B4-Abs and BMP-1-Abs were higher in patients with TIA, aCI, CVD, DM and CKD, and may therefore prove valuable for the early diagnosis of these ATS-related diseases.

Competing interests

This work was performed in collaboration with Fujikura Kasei Co., Ltd. and Celish Fd Inc. RN, GT, NS and HK are employees of Fujikura Kasei Co., Ltd., and TK and HD are employees of Celish Fd Inc.

Acknowledgments

The authors thank Prof. Masaki Takiguchi (Department of Biochemistry and Genetics, Graduate School of Medicine, Chiba University) for valuable discussions.

This work was supported, in part, by Research grant from the Japan Agency for Medical Research and Development (AMED) (Practical Research Project for Life-Style related Diseases including Cardiovascular Diseases and Diabetes Mellitus), Grants-in-Aid of Japan Science and Technology Agency (JST) and Ministry of Education, Culture, Sports, Science and Technology (MEXT) in Japan.

References

- Donnelly R, Emslie-Smith AM, Gardner ID, Morris AD (2000) ABC of arterial and venous disease: vascular complications of diabetes. BMJ 320: 1062-1066.
- Mozaffarian D, Benjamin EJ, Go AS, Arnett DK, Blaha MJ, et al. (2015) Heart disease and stroke statistics--2015 update: a report from the American Heart Association. Circulation 131: e29-322.
- Murray CJ, Vos T, Lozano R, Naghavi M, Flaxman AD, et al. (2012) Disabilityadjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet 380: 2197-2223.
- Carandang R, Seshadri S, Beiser A, Kelly-Hayes M, Kase CS, et al. (2006) Trends in incidence, lifetime risk, severity, and 30-day mortality of stroke over the past 50 years. JAMA 296: 2939-2946.
- Kubo M, Hata J, Doi Y, Tanizaki Y, Iida M, et al. (2008) Secular trends in the incidence of and risk factors for ischemic stroke and its subtypes in Japanese population. Circulation 118: 2672-2678.
- Libby P, Ridker PM, Hansson GK (2011) Progress and challenges in translating the biology of atherosclerosis. Nature 473: 317-325.
- Rollins KE, Varadhan KK, Dhatariya K, Lobo DN (2015) Systematic review of the impact of HbA1c on outcomes following surgery in patients with diabetes mellitus. Clin Nutr.
- Nakanishi N, Okamoto M, Yoshida H, Matsuo Y, Suzuki K, et al. (2003) Serum uric acid and risk for development of hypertension and impaired fasting glucose or Type II diabetes in Japanese male office workers. Eur J Epidemiol 18: 523-530.
- Stemme S, Faber B, Holm J, Wiklund O, Witztum JL, et al. (1995) T lymphocytes from human atherosclerotic plaques recognize oxidized low density lipoprotein. Proc Natl Acad Sci U S A 92: 3893-3897.
- Matsuura E, Lopez LR, Shoenfeld Y, Ames PR (2012) β2-glycoprotein I and oxidative inflammation in early atherogenesis: a progression from innate to adaptive immunity? Autoimmun Rev 12: 241-249.
- Carbone F, Nencioni A, Mach F, Vuilleumier N, Montecucco F (2013) Evidence on the pathogenic role of auto-antibodies in acute cardiovascular diseases. Thromb Haemost 109: 854-868.

Page 9 of 9

- Kramer J, Harcos P, Prohászka Z, Horváth L, Karádi I, et al. (2000) Frequencies of certain complement protein alleles and serum levels of anti-heat-shock protein antibodies in cerebrovascular diseases. Stroke 31: 2648-2652.
- Baekkeskov S, Aanstoot HJ, Christgau S, Reetz A, Solimena M, et al. (1990) Identification of the 64K autoantigen in insulin-dependent diabetes as the GABA-synthesizing enzyme glutamic acid decarboxylase. Nature 347: 151-156.
- 14. Taplin CE, Barker JM (2008) Autoantibodies in type 1 diabetes. Autoimmunity 41: 11-18.
- Sahin U, Türeci O, Schmitt H, Cochlovius B, Johannes T, et al. (1995) Human neoplasms elicit multiple specific immune responses in the autologous host. Proc Natl Acad Sci U S A 92: 11810-11813.
- Nakashima K, Shimada H, Ochiai T, Kuboshima M, Kuroiwa N, et al. (2004) Serological identification of TROP2 by recombinant cDNA expression cloning using sera of patients with esophageal squamous cell carcinoma. Int J Cancer 112: 1029-1035.
- Kagaya A, Shimada H, Shiratori T, Kuboshima M, Nakashima-Fujita K, et al. (2011) Identification of a novel SEREX antigen family, ECSA, in esophageal squamous cell carcinoma. Proteome Sci 9: 31.
- Shimada H, Ito M, Kagaya A, Shiratori T, Kuboshima M, et al. (2015) Elevated serum antibody levels against cyclin L2 in patients with esophageal squamous cell carcinoma. J Cancer Sci Ther 7: 60-66.
- Machida T, Kubota M, Kobayashi E, Iwadate Y, Saeki N, et al. (2015) Identification of stroke-associated-antigens via screening of recombinant proteins from the human expression cDNA library (SEREX). J Transl Med 13: 71.
- Hiwasa T, Zhang XM, Kimura R, Machida T, Kitamura K, et al. (2015) Association of serum antibody levels against TUBB2C with diabetes and cerebral infarction. Gratis J Biomed Sci 1: 49-63.
- Matsutani T, Hiwasa T, Takiguchi M, Oide T, Kunimatsu M, et al. (2012) Autologous antibody to src-homology 3-domain GRB2-like 1 specifically increases in the sera of patients with low-grade gliomas. J Exp Clin Cancer Res 31: 85.
- 22. Hiwasa T, Arase Y, Kikuno K, Hasegawa R, Sugaya S, et al. (2000) Increase in ultraviolet sensitivity by overexpression of calpastatin in ultraviolet-resistant UV/-1 cells derived from ultraviolet-sensitive human RSa cells. Cell Death Differ 7: 531-537.
- 23. Goto K, Sugiyama T, Matsumura R, Zhang XM, Kimura R, et al. (2015) Identification of Cerebral Infarction-Specific Antibody Markers from Autoantibodies Detected in Patients with Systemic Lupus Erythematosus. J Mol Biomark Diagnos 6: 2.
- Shimada H, Yajima S, Oshima Y, Hiwasa T, Tagawa M, et al. (2012) Impact of serum biomarkers on esophageal squamous cell carcinoma. Esophagus 9: 131-140.
- 25. Kunert-Keil CH, Gredes T, Lucke S, Botzenhart U, Dominiak M, et al. (2014) Differential expression of genes involved in the calcium homeostasis in masticatory muscles of MDX mice. J Physiol Pharmacol 65: 317-324.
- 26. Li M, Ho PW, Pang SY, Tse ZH, Kung MH, et al. (2014) PMCA4 (ATP2B4) mutation in familial spastic paraplegia. PLoS One 9: e104790.
- 27. Okunade GW, Miller ML, Pyne GJ, Sutliff RL, O'Connor KT, et al. (2004) Targeted ablation of plasma membrane Ca²⁺-ATPase (PMCA) 1 and 4 indicates a major housekeeping function for PMCA1 and a critical role in hyperactivated sperm motility and male fertility for PMCA4. J Biol Chem 279: 33742-33750.
- Cai J, Pardali E, Sánchez-Duffhues G, ten Dijke P (2012) BMP signaling in vascular diseases. FEBS Lett 586: 1993-2002.
- 29. Dyer LA, Pi X, Patterson C (2014) The role of BMPs in endothelial cell function and dysfunction. Trends Endocrinol Metab 25: 472-480.

- Dhore CR, Cleutjens JP, Lutgens E, Cleutjens KB, Geusens PP, et al. (2001) Differential expression of bone matrix regulatory proteins in human atherosclerotic plaques. Arterioscler Thromb Vasc Biol 21: 1998-2003.
- Sorescu GP, Sykes M, Weiss D, Platt MO, Saha A, et al. (2003) Bone morphogenic protein 4 produced in endothelial cells by oscillatory shear stress stimulates an inflammatory response. J Biol Chem 278: 31128-31135.
- 32. Zhang M, Sara JD, Wang FL, Liu LP, et al. (2015) Increased plasma BMP-2 levels are associated with atherosclerosis burden and coronary calcification in type 2 diabetic patients. Cardiovasc Diabetol 14: 64.
- Miriyala S, Gongora Nieto MC, Mingone C, Smith D, Dikalov S, et al. (2006) Bone morphogenic protein-4 induces hypertension in mice: role of noggin, vascular NADPH oxidases, and impaired vasorelaxation. Circulation 113: 2818-2825.
- Yao Y, Bennett BJ, Wang X, Rosenfeld ME, Giachelli C, et al. (2010) Inhibition of bone morphogenetic proteins protects against atherosclerosis and vascular calcification. Circ Res 107: 485-494.
- 35. Derwall M, Malhotra R, Lai CS, Beppu Y, Aikawa E, et al. (2012) Inhibition of bone morphogenetic protein signaling reduces vascular calcification and atherosclerosis. Arterioscler Thromb Vasc Biol 32: 613-622.
- Kim CW, Song H, Kumar S, Nam D, Kwon HS, et al. (2013) Anti-inflammatory and antiatherogenic role of BMP receptor II in endothelial cells. Arterioscler Thromb Vasc Biol 33: 1350-1359.
- 37. Sarras MP Jr (1996) BMP-1 and the astacin family of metalloproteinases: a potential link between the extracellular matrix, growth factors and pattern formation. Bioessays 18: 439-442.
- Garrigue-Antar L, François V, Kadler KE (2004) Deletion of epidermal growth factor-like domains converts mammalian tolloid into a chordinase and effective procollagen C-proteinase. J Biol Chem 279: 49835-49841.
- Tanaka M, Asada M, Higashi AY, Nakamura J, Oguchi A, et al. (2010) Loss of the BMP antagonist USAG-1 ameliorates disease in a mouse model of the progressive hereditary kidney disease Alport syndrome. J Clin Invest 120: 768-777.
- 40. Rider CC, Mulloy B (2010) Bone morphogenetic protein and growth differentiation factor cytokine families and their protein antagonists. Biochem J 429: 1-12.
- Will JC, Galuska DA, Vinicor F, Calle EE (1998) Colorectal cancer: another complication of diabetes mellitus? Am J Epidemiol 147: 816-825.
- Jarvandi S, Davidson NO, Schootman M (2013) Increased risk of colorectal cancer in type 2 diabetes is independent of diet quality. PLoS One 8: e74616.
- Fujihara S, Kato K, Morishita A, Iwama H, Nishioka T, et al. (2015) Antidiabetic drug metformin inhibits esophageal adenocarcinoma cell proliferation in vitro and in vivo. Int J Oncol 46: 2172-2180.