

Current Insights in Mammalian Glycosylation: Implication of Glycoproteomics as Next-Generation Biomarkers in Diabetes Mellitus

Alok Raghav^{1*}, Jamal Ahmad¹, Zeeshan Ahmad Khan², Brijesh Kumar Mishra³ and Rai Pawankumar Anirudh⁴

¹Rajiv Gandhi Centre for Diabetes and Endocrinology, J.N Medical College, Aligarh Muslim University. Aligarh-U.P, 202002, India

²Biological Rhythm Lab, Institute of Bioresourses and Sustainable Development, Imphal, Manipur-795001, India

³Department of Endocrinology, Guru Teg Bahadur Hospital, Dilshad Garden, Delhi-110095, India

⁴ICAR-Department of Medicinal and Aromatic Plant Research, Ananad, Gujarat, India

*Corresponding author: Alok Raghav, Rajiv Gandhi Centre for Diabetes and Endocrinology, J.N Medical College, Aligarh Muslim University, Aligarh-202002, UP, India, Tel: +91-571-2721173; E-mail: alokalig@gmail.com

Received date: February 19, 2017; Accepted date: March 14, 2017; Published date: March 21, 2017

Copyright: © 2017 Raghav A, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Glycosylation, an enzyme-directed, site-specific process is one of the major co-translational and post-translational modifications in proteins. So formed product, glycoproteins, mediate multiple functions of signal transductions, cell association, and inter and intra cell signalling. Glycoproteomics exploits all these biological functions and serves as the next generation diagnostic marker for analyzing proteins that are associated with micro- and macro-vascular complication of diabetes mellitus. This tool can unreveal the secrets of the patterned expression of glycoproteins in healthy and patients with diabetes with secondary complications. The expression profile of several O-linked and N-linked mammalian glycoproteins delivers the knowledge and helps to create a database for expression in diabetes mellitus, for proposing a mechanistic approach to understanding the cellular pathway severely involved in the regulation of all glycoproteins. This future next generation glycopeptides tool will describe a novel strategy for the quantification of the several glycoproteins based on a specific chemical ligation described as reverse glycoblotting techniques, two-dimensional gel electrophoresis, western blotting, multiple reaction monitoring, LC-MS/MS analysis of glycosylated peptides along with lectin chromatography. These recent tools will reveal the expression pattern of several glycoproteins that will serve as a power tool precise diagnosis of diabetes associated complications.

Keywords: Glycoproteomics; Glycoproteins; Glycoproteins; Diabetes; Biomarkers; Post translational modifications

Introduction

Proteins are believed to be the building blocks of the body and have a keen involvement in the biological and physiological homeostasis of the organism [1]. Advancement in the proteomics in the past decades has made it an intensive and powerful diagnostic tool. One of the major areas of proteomics is glycoproteomics. Glycoproteomics relies on the study of glycoproteins that derived from glycosylation. The glycoproteomics identifies, characterizes and index proteins and peptides with carbohydrate moieties, including the post-translational modifications (PTMs) [2]. There are numerous PTMs that include ubiquitination, glycosylation, acetylation and phosphorylation [3]. It is more complicated and diverse class of protein modifications that plays a vital role in biological functions as well as disease progression and prognosis. It has been found recently that 80% mammalian proteins undergo PTMs [4] amongst them glycosylation constitute about 50% [5].

Glycosylation of proteins involves the addition of carbohydrate or glycans covalently to deliver protein function and activity. Glycans are most abundant natural biopolymer and constitute the major component of the cells. The diverse function of glycans creates a new field of glycobiology to understand the structure, synthesis, regulation and metabolism of diverse glycans. Human blood groups, discovery gave birth to the importance and clinical applications of glycans function [6]. With numerous developments of the past decades in glycobiology, several parallel approaches evolve that includes genomics, proteomics, lipidomics, glycoproteomics, glycomics that interestingly imparts role of biomolecules in the disease process.

Glycoproteomics and glycomics are interrelated disciplines of glycobiology that dealt with knowing mechanism of glycoproteins and glycans respectively. Carbohydrate or glycans derived modifications significantly involved in the host-pathogen association, cell signalling, inflammation, growth, and development Expression profiling of cellular glycoproteins, glycopeptides library, with comparative between a normal homeostatic and diabetes homeostatic, may reveal the fundamentals and underpinnings of the disease mechanism and prognosis. It also facilitates the determination of structurally and functionally modified proteins. Scheduled glycosylation maintains the homoeostatic environment in body, however if turn aberrant may result in abnormal and impaired changes in biological and physiological activity of protein that may, in turn, be the major culprit in molecular recognition of disease.

Diabetes has several biomarker but glycoprotein open new vistas in its diagnostics. It is a novel biomarker for an early detection of diabetes, its associated complications and also provides detailed information about risk factors and associated prognosis factors. Analysis of diabetes related glycoproteins' expression (DRGE) may prove a novel next generation future biomarker in prognosis of diabetes and its associated micro and macro vascular complications. Diverse type of PTMs found in living organisms, including (1) Nlinked glycosylation (2) O-linked glycosylation (3) C-linked glycosylation [7] and (4) S-linked glycosylation (occurs in bacteria) [8].

Furthermore, components of glycoproteins i.e., glycans plays important role in vast biological processes, including immune response, cell attachment and interaction, embryonic development that majorly involves sugar-sugar or sugar-protein interaction and recognition [9]. Mammalian glycans are resultant of an endogenous portfolio of several cellular metabolic enzymes and their substrate that have been conserved during the whole evolution spanning 1%-2% genome. Slight variation in the patterned expression of glycomes of organisms may be the molecular basis for interspecies recognition elements. Mammalian glycans are highly conserved along with species specific variations that may lead to the development of distinct trait [10]. Consequently, aberrant pattern of glycosylation of mammalian glycoproteins results in non-homeostatic expression of glycopeptides library in diseased individual compared to healthy. This has been implicated in several diseases, including immune deficiencies, genetic disorders, cardiovascular diseases, neurodegenerative disorders and cancer [11].

Several published literature reported implications of glycoproteins as targeted clinical biomarkers in complications, including Her2/neu (breast cancer), PSA (prostate specific antigen, marker for prostate cancer) and CA-125 (ovarian cancer) [12]. Indeed, impaired glycosylation in disease and associated complications showed a significant amount of biological information about mammalian glycome to be the clinical diagnostic marker [12]. In order to diagnose abrupt glycosylated protein expression in disease situations, responsive, specific, swift and strong methods are needed. Although, massive work is routinely going in various laboratories for identification of these glycopeptides markers, but the study of glycoproteins remains challenging. Frequent, comprehensive reviews related to this approach have been published in recent years covering identifications of glycoproteins [13]. This review focuses on the involvement of mammalian glycoproteome in diagnosis of diabetes and its related disorders, along with an emphasis on precise method development to create expression library of these glycopeptides in order to improve the glycoproteomics approach of human health.

Construction and Topology of Mammalian Glycosylation Mechanism

Monosaccharides play major role in mammalian glycosylation. Overall, nine monosaccharides participate in this enzyme mediate process of glycosylation. Ubiquitously precursor presents in the dietary components supply these nine monosaccharides to undergo processes of conserved biological synthesis.

Mammalian glycosylation involves primarily three diverse forms of modifications, including N-linked, O-linked and C-linked. Among these N-linked additions of sugars being the prime one. Nglycosylation in mammals occurs on asparagine's amino residue at specific sequences of Asn-X-Ser/Thr occasionally on Cys residues with X being any of the amino acids except proline. The initiation of these modifications occurs on the cytoplasmic side of rough endoplasmic reticulum (RER), with binding of oligosaccharides Man5GlcNAc2 considered to be the precursor of dolichol pyrophosphate. Glc3Man9GlcNAc2-PP-dol is considered to be the appropriate core glycan precursor [14]. Depending upon the types and position of the monosaccharides encompass within chain, the N-glycans auxiliary grouped into high-mannose type, hybrid type and antennary type [14]. Components of high-mannose type of N-glycans are large mannose in the core structure. The Antennal region contains complex type of Nglycans Galß1-3/4GlcNAc in N-terminal regime.

Furthermore, O-linked mammalian glycosylation occurs on serine or threonine residues with addition of N-acetyl glucosamine, N-acetylgalactosamine, fucose, mannose or xylose [14]. The most preferred O-linked glycosylation occurs with N-acetylgalactosamine linked with α -glycosidic bond to Ser/Thr.

A recent literature demonstrated that O-linked glycosylation in mucin may involve in immune response, cell adhesion and invasion that may be served as biomarker for diagnosis of cancer [12]. The nascent protein undergoes this post-translational modification after secreted from the classical secretory pathway from Golgi body. Hydroxyl (OH) group present on the Ser/Thr actively participates in O-linked glycosylation by addition of galactose/N-acetylglucosamine. Post addition of GalNAc followed by further elongation and termination (sialylation) confirm a large number of O-linked glycopeptides synthesized by glycosyltransferases [15]. Compared to N-linked glycosylation, the quantification of O-linked modifications has proven to be tough. The probable hurdles may due to the presence of readily available predicted sequences (-Asn-Xaa-Ser/thr- or rarely -Asn-Xaa-Cys-;Xaa is nay amino acid except Proline in N-linked glycosylation. In disparity, Ser/Thr in O-linked glycosylation can be predicted only in the beginning. Secondly, N-linked glycopeptides are well separated, while the O-linked glycopeptides are located in clusters thereby making it more difficult to identify.

A final and unexpected example of mammalian glycosylation is Clinked. This modification involves a-mannose group of certain tryptophan residues in proteins which have been previously described by Hofsteenge and their colleagues. C-mannosylation is known to occur on the Dol-P-Man consensus sequence along with Trp-Xaa-Xaa-Trp sequence. However, present knowledge for C-linked glycosylation is limited. The degree of complexity, diversity, structural changes is subject to further research and investigations.

Regulation of Mammalian Glycosylation

Regulation of mammalian glycosylation is also complex as their structure and conformations. Glycans are specific targets of glycosyltransferase and glycosidase enzymes. Glycans chains are synthesized by glycosyltransferase while hydrolyzed by glycosidases enzyme class. Being anabolic components of glycosylation, glycosyltransferase in collaboration with glycosidases determine the structural and functional outcome of the biosynthesis of glycans [16]. Glycosaminoglycans and O-glycans biosynthetic pathways involve single enzyme reaction that completely depends on the topology of glycans, create by earlier enzymes for being the substrate for next [17].

Mammalian glycosylation mechanism of secretory proteins is a dynamic mechanism that involves alterations in the activity and accessibility of glycosyltransferase and glycosidase towards its substrate. Structural, topological variety in the glycan repertoire in the biosynthetic pathways generates numerous clinical biomarkers; some of these may correlate with the disease and its associated complications in every species to distinguish it from normal homeostasis. A previously supported literature proves elevated truncated O-glycans served as cancer biomarkers [18]. Transcription process modulates RNA expression of biosynthetic pathways of glycans formation governed by glycosyltransferases and glycosidases. The microarray analysis of genes involved in the biosynthetic pathways of glycans plays a key role in the cellular expression pattern of glycans that may perhaps be predictive in providing vast information as biomarkers [19]. Similarly, gene transcription shows that core 2 GlcNAcT-I is induced by transcription factor T-bet in T helper type 1 lymphocytes [20]. Although, numerous transcription factors regulate the glycosylation pathways gene expression, the concise effect is still unclear.

The existent evidence reflects that regulation of mammalian glycans occurs on post-transcriptional and post-translational stages too. Foremost alterations in glycoproteins are induced by loss of multiprotein complexes along with few chaperones that impair the glycosyltransferase mediated trafficking via endoplasmic reticulum and Golgi machinery [21]. A potential regulatory approach of glycoyltransferases and glycosidases is through phosphorylation event of their cytoplasmic tails, which might contribute to the intracellular trafficking and intermolecular rearrangements. Another potential regulatory factor in glycan formation is the disintegration of glycosyltransferases and glycosidases from their locations via proteolysis. The mammalian biosynthetic pathway enzymes are type 2 transmembrane glycoproteins containing luminal catalytic domain along with luminal proximal stem domain. Specific cleavage at stem domain may cleave catalytic domain separately, thereby may be the probable cause of inflammation as reported earlier [10]. This cleavage mechanism is still unclear and their ability to catalyze glycan formation among extracellular compartments has vanished. Lysosome mediated degradation of mammalian cell surface glycans is a key feature of infection causing pathogens like influenza virus [22]. Mammalian glycan formation is broadly governed by biosynthesis and availability of nucleotide sugar donors. Obstruction in the donor may lead to functional loss of transporters residing in endoplasmic reticulum and Golgi membrane and may abolish cellular glycans productions [23]. The Previously published study showed that extra supplements of glucosamine to hexosamine pathway elevate the glycans in mammalian cells [24]. The controlled regulation of this mammalian glycosylation is not yet known and careful investigations are required to study modulating mammalian glycan expression.

Glycoproteomic Development of Biomarker

The discovery of novel glycoproteins as novel biomarker has proven its clinical application in diabetes screening, diagnosis, and prognosis. The construction of glycoproteins expression library focuses on translational research in diabetes to fight against its associated complications. Mass spectroscopic (MS) glycoproteomics approach may be used for early diagnostic biomarkers to differentiate diseased from healthy individuals. Although the conventional proteomics approach in biomarker discovery, some recent advance MS technologies have been introduced for identifications, characterization and quantifications of glycoproteins expression (Figure 1).

Matrix assisted laser desorption/ionization-time of flight (MALDI-TOF), electron spray ionization-mass spectroscopy (ESI-MS) and surface enhanced laser desorption/ionization (SELDI) in combination with liquid chromatography mass spectroscopy (LC-MS) may contribute in the validation and identification of novel glycopeptides biomarkers in diabetes and its associated complications. A recent study demonstrated the application of the MS approach to identify Sp1 as novel biomarker for hyperglycemia development due to activation of hexosamine pathway that has been easily implicated in diabetes and its associated pathogenesis [25].

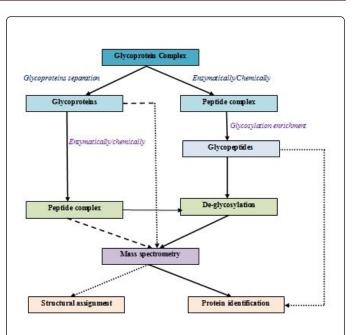


Figure 1: Showing workflow of glycoproteomics identification and characterization of all glycosylated proteins.

Recently, due to generation of innovative novel MS-based glycoproteomics technologies the diabetes glycoproteomics area is explored too much for the human welfare and day by day novel biomarkers are evolving. Additionally, high throughput capacity of MS-based approach coupled with LC-MS for identification of biomarker in diabetes [26] is a new hope for development of glycoproteomics expression library. Indeed, large efficient efforts were made to sincerely investigate the diabetes glycoproteomics with various human specimens including adipose tissues, cell lines, serum, plasma, urine and saliva for creating a database of an expression library of glycosylated proteins in diabetes associated complications. The MSbased glycoproteomics research incorporates two principle approaches (1) glycoprotein identifications (2) pattern expression and recognition. Both approaches require high throughput automated computing and bioinformatics systems to create database for generation of expression library of glycoproteome associated with diabetes and its complications.

Specifically, glycoproteomics identifications involve glycoprotein isolation, enrichment, proteolytic digestion and identifications with advance MS based approaches. Table 1 showed various enrichment techniques of glycoproteins. Since, various biological samples of serum, plasma constitute mixtures of glycoproteins, therefore, extensive chromatographic separations is required that uses ionexchange, affinity column separation, size exclusion and hydrophobic separation to minimize the complexity of proteins and enhance accuracy [27]. Recently develops a bottom-up approach has been developed for analysis of glycoprotein [28]. In this approach, glycoproteins are digested and undergo deglycosylation event with peptide N-glycosidase F (PNGase F) that cleaves glycosydic bonds between Asn and innermost Glc-NAc of N-linked glycoproteins with few exceptions [29]. Digested and deglycosylated peptides were subjected to MS analysis coupled with MS tool, bioinformatics with numerous algorithm and glycoproteomic database analyzing software that identifies the specific expression of glycans and glycopeptides

Page 3 of 7

Page 4 of 7

separately [30]. Database libraries of Uniprot and Peptide Atlas also have information mass spectra information of glycoproteins and glycopeptides [31]. Integration of database acquired with recent tools is necessary to create an expression library of glycans and glycoproteins [32]. A study differentiates between healthy and diseased subject based on co-immune purification [33] on hypothesis based manner. Thus, various approaches make fields of glycoproteomics feasible for study and biomarker development.

Enrichment techniques	Advantages	Disadvantages	References
Hydrazide Chemistry	 High coupling efficiency for all types of glycans. Reducing chances of non- specific binding of complex proteins. 	 Oxidative coupling renders loss of structural database of glycans. Only applicable for N-linked glycosylation using PNGase F. 	[44]
Lectins affinity chromatography	 Nourishes and enriches glycan structure. Coupled to several isomers. 	 Enriches only a particular class of glycoconjugates. Deliver weak binding of glycans. Nonspecific binding. 	[45]
Activated graphitized carbon	(1) Retains glycoconjugates and resolve structural isomers.	(1) Non-specific separation of hydrophilic glycans.	[46]
Boronic acid based solid phase extraction chromatography	(1) High coupled throughput efficiency of all glycans.	 Non-specific for glycans that terminate in N-acetylgalactosamine. 	[47]
Size exclusion chromatography	(1) Simple, non- selective and efficient.	 Non-specific for glycans. Only for N-linked glycans. 	[48]
Hydrophilic interaction liquid chromatography	(1) Efficient and specific separation of glycopeptides.	(1) Not specific for glycans.	[49]

Table 1: Glycoproteome enrichment approach coupled with mass spectrometry tool for construction, identification and characterization of expression library of glycoproteins and glycans.

Role of Glycopeptides in Diabetes and Its Associated Complications as Biomarkers

The beta cells are unique to contains large amount of OGT that enables them to respond to physiological increase in the glucose concentration by transforming the glucose to the OGT substrate UDP-GlCNAc thereby disruption in the O-linked glycosylation pathway [34]. A recent research in relation to diabetes associated hyperglycemia revels that increased sugar level in subjects with diabetes induce mitochondrial superoxide overproduction that in turn activate the hexosamine pathway and induces the plasminogen activator inhibitor-1 expression due to increased production of Sp1 glycosylation [32]. This study reveals the alterations in the pattern of glycoprotein expression in diabetes complications. A specific study on type 2 diabetes mellitus patients states that hyperglycemia induced endothelial glycoproteome changes lead to alterations in homeostatic endothelial signalling pathways [35]. In another recent study, glycoproteins are found to be the major components of the cardiac extracellular matrix that plays important role in cardiovascular disease progression [36]. Thrombospondin, tenascin-C and periostin are some of the extracellular matrix glycoproteins that modulate cardiac remodelling post myocardial infractions [37]. Hexosamine biosynthetic pathway flux and OGlcNAc level increased in hyperglycaemic conditions. A recent study examined the modification of OGlcNAc alters Ca^{+2} binding proteins that may lead to cardiac dysfunction in patients with diabetes [38]. Subsequently Hu et al. reported increased OGT expression and OGlcNAc in heart from STZ induced diabetic mice that may lead to cardiomyocyte dysfunction [38]. Table 2 clearly demonstrates glycomarkers proteins and transcription factors express during diabetes and its associated complications.

Proteins	Sources	O-GIcNAc effect on activity		
Transcription factors				
Sp1	NRVM,Mesangial cells	↑Expression		
Mef-2	NRVM	↓Expression		
P53	ARVM	↑Expression		
Nf-κβ	Mesangial Cells	↑Expression		
Cytokines				
PAI-1	Mesangial Cells, BAEC	↑Expression		
TGF-β	BAEC	↑Expression		
Insulin pathway and glucose biochemical metabolism				
IRS 1, IRS2	Skeletal muscles, HCAEC	Impaired insulin signalling		
IR-β	RIN-β	↓Phosphorylation		
РІЗК	HCAEC	↓Expression		
GLUT4	Skeletal muscles, primary adipocytes	↓Translocation		
Akt, Akt1, Akt2	Nueroblastoma, primary adipocytes	↓Insulin mediated response		
GSK3β	RIN-β, 3T3-L1	↓Response to insulin		
Glycogen synthase	Adipocytes, 3T3-L1	↓Activation		
Calcium signalling				
SERCA	NRVM	↓Expression		
Related signalling pathways				
PLC-β1	C1C12 myotubules ↓Expression			
ΡΚCβ,ε/ΡΚC α,ε	Mesangial cells/SVG cells	Altered translocation		
eNOS	BAEC	↓Expression		

Table 2: List of proteins and transcription factors contributed in diabetic associated complication that are modified and impaired directly or indirectly by O-GlcNAc. These proteins/transcription factors may contribute to the diagnostic biomarkers in the patients with altered glycosylation having diabetes or related complications.

[Sp1: Specificity Protein 1; Mef-2: Myocyte Enhancer Factor-2, Nf-κß: Nuclear Factor κβ; IRS1/2: Insulin Receptor Substrate ½; IR-β: Insulin Receptor: PI3K: Phosphatidylinositol-3-Kinases; GLUT4: Glucose Transporter Type 4; Akt: Protein Kinase B (PKB); GSK3ß: Glycogen Synthase Kinase 3 Beta; SERCA: Sarco/endoplasmic Reticulum Ca²⁺-ATPase; PLC-ß1: Phosphoinositide-Phospholipase C ß1; PKC: Protein Kinase C, eNOS: Endothelial Nitric Oxide Synthase; NRVM: Neonatal Rat Ventricular Myocytes; ARVM: Adult Rat Ventricular Myocytes; BAEC: Bovine Aortic Endothelial Cells; HCAEC: Primary Human Coronary Artery Endothelial Cells; RIN-β: Cell Line for *Rattus norvegicus*, 3T3-L1: Cell Line; SVG: Human Fetal Glial Cell Line; Increased expression: ; decreased expression:].

Glycans Profiling: Future Hope of Glycomics

Concise investigations of transcripts and proteins, the key of genomics and proteomics, have enhanced our knowledge of cell's biochemical machinery. The close relation between cellular metabolism and transcription of genes help to understand the biological complexity of each system. Sequencing and accessibility of whole genome offer blueprint of biochemical makeup of individual. Extensive efforts have been made to relate the genome with its proteome. Murrell et al. further made efforts to relate genome and proteome with the complete repertoire of glycans of an organism to understand glycomic approach [39]. Considering inventory efforts in the field of glycomics and glycoproteomics, new high-throughput techniques have been evolving. Glycans profiling approach can be implicated at several hierarchical levels of organism's structural database content. At first hierarchical level, the aim is to identify the glycoproteins bearing monosaccharide that shows certain modifications with change in physiological homeostasis. A convenient and complete approach to profiling numerous cytosolic and nuclear proteins incorporates the bio orthogonal strategy. A recent work also showed identification of B-O-GlcNAc modification with affinity capture and MS. Another identical profiling study recognized sialylated or mucin type O-linked glycoproteins with the help of azidosugars as substrate. The higher level of profiling includes the identification of specific glycan linkage patterns. Subtle alterations in the monosaccharide units and linkage may have profound metabolic consequences, which further may relate to the development of complications and pathogenesis of diseases. The range of glycans profiling is aided by antibodies and lectins that may use in microarrays along with fluorescently labeled samples to provide detailed information of bound specific class of glycans in the glycoconjugate. Lectin and antibody approach for determination of glycans in the glyco-conjugates. Lectin and antibody approach for determination of glycan profiling, however, limitations of glycan epitopes binding with their complementary antibodies or lectins. Thus, glycan profiling can be effectively being performed with mass spectrometry [40]. Considerable expertise in profiling of glycans is required. Microchips labelled with synthetic glycan are used for high throughput profiling of mammalian cells [41].

Future Challenges and Hurdles in Discovery of Glycol Biomarker

There is a need to construct glycopeptides expression library in diabetes and its associated complications. Several published literature demonstrates the preliminary research in regards to constructing glycoproteome library of few glycoproteins. In a Study by Hiromi Ito available describes the method for constructing O-linked glycopeptides library of humans with glycosyltransferase. Similar studies have been conducted worldwide for determining the glycoproteins expression [42]. Nonetheless, the hypotheses proposed here incorporate the library of glycoproteins whose data are still not available. As well as the implications of them as next generation future diagnostic marker for diabetes related secondary complications are also not so well established. These fields need more concern and research is warranted for the establishment of expression library and diagnostic biomarker. Previously reported literature also showed the there are isoforms of GalNAc-transferase that can be utilized for creation of libraries [43]. Similarly glycomics approach was explained in detail by previously published literature also [44] and was proved that changes in glycoproteome may correlate with the diseased state [45,46]. Previously published literature on CD4⁺ glycoprotein expression profile can be easily correlated with the diseased state [47].

Significance of Glycoproteomics Biomarker Expression Library

Glycoproteomic approach for generating expression functional proteome of all glycoproteins in healthy and subjects with diabetes will deliver the database for further analysis of the glycoproteins. The library database will enhance the ability to understand the mechanistic pathway that primarily involves in the complications arises due to these glycoproteins. These libraries will also help to discover a novel therapeutic approach for prevention of diabetes associated micro- and macro-vascular complications.

Another important significance is the discovery of next generation future diagnostic marker of diabetes. Several glycoproteins alter in its structural and functional proteome whose properties can be exploited for diagnosing chronic disease. Currently, investigations of diabetes diagnosis limit up to HbA1c and fructosamine only, but due to several limitations and shortcomings, these parameters are not fulfilling the scope completely. Apart from these conventional markers, the glycoproteins may be the possible next generation diagnosis marker of diabetes for deciphering the precise and accurate investigations in diabetes.

Conclusion

Glycoproteomics is an important part of proteomics as glycosylation of proteins reveals biological impairments in disease conditions. Extensive research is needed to study glycoproteins and glycans structure and expression altered pattern in diabetes and its associated complication for generation of new diagnostic future marker. Incorporation of various identifying and characterizing approach based on mass spectroscopy coupled with chromatography may enhance the precise of the biomarkers. Still, there is an extensive data processing is needed for studying glycans and its related database with bioinformatics coupled tool with algorithms. These glycoproteins expression pattern in healthy and diabetes patients will add knowledge to the international database so that one can exploit these libraries for proposing a novel therapeutic approach in diabetes mellitus. The research of glycoproteomics is never ending and technically challenged yet very promising to discover novel next generation future diagnostic markers for early diagnosis of diabetes and its associated complications.

Conflict of Interest

The authors declare no conflict of interest.

Citation: Raghav A, Ahmad J, Khan ZA, Mishra BK, Anirudh RP (2017) Current Insights in Mammalian Glycosylation: Implication of Glycoproteomics as Next-Generation Biomarkers in Diabetes Mellitus. J Glycobiol 6: 125. doi:10.4172/2168-958X. 1000125

References

- Wilkins MR, Sanchez JC, Gooley AA, Appel RD, Humphery-Smith I, et al. (1996) Progress with proteome projects: why all proteins expressed by a genome should be identified and how to do it. Biotechnol Genet Eng Rev 13: 19-50.
- 2. Tissot B, North SJ, Ceroni A, Pang PC, Panico M, et al. (2009) Glycoproteomics: Past, present and future. FEBS Lett 583: 1728-1735.
- Gudepu RG, Wold F (1998) In Proteins: Analysis and Design, ed. Angeletti RH, Academics, San Diego, Calif, USA, pp: 121–207.
- 4. Kornfeld R, Kornfeld S (1985) Assembly of asparagine-linked oligosaccharides. Annu Rev Biochem 54: 631-664.
- Apweiler R, Hermjakob H, Sharon N (1999) On the frequency of protein glycosylation, as deduced from analysis of the SWISS-PROT database. Biochemical et Biophysica Acta 1473: 4-8.
- 6. Landsteiner K (1931) Individual differences in human blood. Science 73: 405-411.
- 7. Wei X, L Li (2009) Comparative glycoproteomics: approaches and applications. Brief Funct Genomics 8: 104-113.
- 8. Floyd N, Vijayakrishnan B, Koeppe JR, Davis BG (2009) Thiyl glycosylate of olefinic proteins: S-linked glycoconjugate synthesis. Angewandte Chemie International Edition 48: 7798-7802.
- 9. Helenius A, Aebi M (2001) Intercellular functions of N-linked glycans. Science 291: 2364-2369.
- Gagneux P, Varki A (1999) Evolutionary considerations in relating oligosaccharide diversity to biological function. Glycobiology 97: 747-755.
- 11. Lowe JB, Marth JD (2003) A genetic approach to mammalian glycan function. Annu Rev Biochem 72: 643-691.
- Diamandis EP (2004) Mass spectrometry as a diagnostic and a cancer biomarker discovery tool: Opportunities and potential limitations. Mol Cell Proteomics 3: 367-378.
- Wuhrer M, Catalina MI, Deelder AM, Hokke CH (2007) Glycoproteomics based on tandem mass spectrometry of glycopeptides. J Chromatogr B Analyt Technol Biomed Life Sci 849: 115-128.
- 14. Rakus JF, Mahal LK (2011) New technologies for glycomic analysis: Toward a systematic understanding of the glycome. Annu Rev Anal Chem (Palo Alto Calif) 4: 367-392.
- 15. Kornfeld R, Kornfeld S (1985) Assembly of asparagine-linked oligosaccharides. Annu Rev Biochem 54: 631-664.
- Schachter H (2000) The joys of HexNAc. The synthesis and function of Nand O-glycan branches. Glycoconj J 17: 465-483.
- Maccioni HJ, Giraudo CG, Daniotti JL (2002) Understanding the stepwise synthesis of glycolipids. Neurochem Res 27: 629-636.
- Kobata A, Amano J (2005) Altered glycosylation of proteins produced by malignant cells and application for the diagnosis and immunotherapy of tumours. Immunol Cell Biol 83: 429-439.
- Underhill GH, Zisoulis DG, Kolli KP, Ellies LG, Marth JD, et al. (2005) A crucial role for T-bet in selection ligand expression in T helper 1 (Th1) cells. Blood 106: 3867-3873.
- Wu X, Steet RA, Bohorov O, Bakker J, Newell J, et al. (2004) Mutation of the COG complex subunit gene COG7 causes a lethal congenital disorder. Nat Med 10: 518-523.
- 21. Ju T, Cummings RD (2005) Protein glycosylation: Chaperone mutation in Tn syndrome. Nature 437: 1252.
- 22. Lubke T, Marquardt T, Etzioni A, Hartmann E, von K Figura, et al. (2001) Complementation cloning identifies CDG-IIc, a new type of congenital disorders of glycosylation, as a GDP-fucose transporter deficiency. Nat. Genet 28: 73-76.
- 23. Smith PL, Myers JT, Rogers CE, Zhou L, Petryniak B, et al. (2002) Conditional control of selection ligand expression and global fucosylation events in mice with a targeted mutation at the FX locus. J Cell Biol 158: 801-815.

- 24. Lau K, Partridge EA, Cheung P, Dennis JW, Hexosamine (2005) Nglycans and cytokines signaling a regulatory network. Glycobiology 15: 1196.
- 25. Du XL, Edelstein D, Rossetti L, Fantus IG, Goldberget H, et al. (2000) Hyperglycemia-induced mitochondrial superoxide overproduction activates the hexosamine pathway and induces plasminogen activator inhibitor-1 expression by increasing Sp1glycosylation. PNAS 97: 12222-12226.
- 26. Ottens AK, Kobeissy FH, Wolper RA (2005) A multidemsional differential proteomics platform using dual phase ion-exchange chromatography-polyacrylamide gel electrophoresis/reversed phase liquid chromatography tandem mass spectrometry. Analytical Chemistry 77: 4836-4845.
- 27. Huang HL, Stasky, Morandel S (2005) Enrichment of low abundant serum proteins by albumin/immunoglobulin G immunoaffinity depletion under partly denaturing conditions. Electrophoresis 26: 2843-2849.
- Liu T, Qian WJ, Gritsenko MA (2005) Human plasma N-glycoproteome analysis by immunoaffinity substraction, hydrazide chemistry and mass spectrometry. J Proteome Res 4: 2070-2080.
- 29. Aoki-Kinoshita K (2008) An introduction to bioinformatics for glycomics research. PLoS Comput Biol 4: e1000075.
- Deutsch EW, Lam H, Aebersold R (2008) PeptideAtlas: A resource for target selection for emerging targeted proteomics workflows. EMBO Rep 9: 429-434.
- Uematsu R, Furukawa JI, Nakagawa H (2005) High throughput quantitative glycomics and glycoform-focused proteomics of murine dermis and epidermis. Mol Cell Proteomics 4: 1977-1989.
- 32. Mallick P, Kuster B (2010) Proteomics: A pragmatic perspective. Nat Biotechnol 28: 695-709.
- 33. Konrad RJ, Kudlow JE (2002) The role of O-linked protein glycosylation in beta-cell dysfunction. Int J Mol Med 10: 535-539.
- Hoffmann B, Prisco A, Greene A (2014) Hyperglycemia-driven glycosylation of AT1 Receptor in the vascular endothelium. FASEB J 28: 1086-1088.
- Rienks M, Papageorgiou AP, Frangogiannis NG, Heymans S (2014) Myocardial extracellular matrix: An ever-changing and diverse entity. Circ Res 114: 872-888.
- 36. Xia Y, Dobaczewski M, Gonzalez Quesada C, Chen W, A Biernacka (2011) Endogenous thrombospondin1 protects the pressure overloaded myocardium by modulating fibroblast phenotype and matrix metabolism. Hypertension. 58: 902-911.
- Nishioka T, Onishi K, Shimojo N, Nagano Y, Matsusaka H (2010) Tenascin-C may aggravate left ventricular remodelling and function after myocardial infarction in mice. Am J Physiol Heart Circ Physiol 298: H1072-H1078.
- Hu Y, Belke D, Suarez J, Swanson E, Clark R, Hoshijima M (2005) Adenovirus mediated overexpression of O-GlcNAcase improves contractile function in the diabetic heart. Circ Res 96: 1006-1013.
- 39. Murrell MP, Yarema KJ, Levchenko A (2004) The systems biology of glycosylation. Chembiochem 5: 1334-1347.
- 40. Dell A, Morris HR (2001) Glycoprotein structure determination by mass spectrometry. Science 291: 2351-2356.
- 41. Kiessling LL, Gestwicki JE, Strong LE (2000) Synthetic multivalent ligands in the exploration of cell-surface interactions. Curr Opin Chem Biol 4: 696-703.
- 42. Nakakita S (2008) Chemical liberation of N-linked oligosaccharides from glycoproteins. Experimental Glycosciences pp: 5-6.
- Tarp, Mads Agervig, Clausen Henrik (2008) Mucin-type O-glycosylation and its potential use in drug and vaccine development. Biochimica et Biophysica Acta 1780: 546-563.
- 44. Varki A, Cummings RD, Esko JD, Freeze HH, Stanley P, et al. (2009) Essentials of glycobiology, 2nd Edn. Cold Spring Harbor Laboratory Press: Cold Spring Harbor (NY), USA.

Page 6 of 7

Citation: Raghav A, Ahmad J, Khan ZA, Mishra BK, Anirudh RP (2017) Current Insights in Mammalian Glycosylation: Implication of Glycoproteomics as Next-Generation Biomarkers in Diabetes Mellitus. J Glycobiol 6: 125. doi:10.4172/2168-958X. 1000125

Page 7 of 7

- 45. Dube, Danielle H, Prescher, Jennifer A, Quang, et al. (2006) Probing mucin-type O-linked glycosylation in living animals. Proc Natl Acad Sci U S A 103: 4819-4824.
- 46. Keppler OT, Horstkorte R, Pawlita M, Schmidt C, Reutter W (2001) Biochemical engineering of the N-acyl side chain of sialic acid: Biological implications. Glycobiology 11: 11R-18R.
- Ford, Mandy L, Onami, Thandi M, Sperling, et al. (2003) CD43 modulates severity and onset of experimental immune encephalomyelitis. J Immunol 171: 6527-6533.