

Contribution of Plant Growth Regulators in Mitigation of Herbicidal Stress

Sugandha Varshney, M Iqbal R Khan, Asim Masood, Tasir S. Per, Faisal Rasheed and Nafees A Khan*

Department of Botany, Aligarh Muslim University, Aligarh, Uttar Pradesh, India

Abstract

Weeds contribute to severe loss of agricultural crop productivity. Herbicides are efficient, cheap and effective method for their control, but their multiple uses at higher concentration have resulted in induced toxicity and stress in non-target crops. The herbicide-induced toxicity affects growth, photosynthesis, physiological and biochemical characteristics of plants directly impacting yield of plants. It is therefore, necessary to find ways to mitigate the herbicidal effects by increasing the tolerance of crops to their application. Plant growth regulators are known to promote growth and development of plants under optimal and stressful environment. They induce various physiological and biochemical responses and also confer tolerance of plants to abiotic stresses. The present review covers the aspects of herbicidal response of plants and evaluates the contribution of plant growth regulators in mitigating herbicidal stress effects and increasing the tolerance of plants. Further, future research in this direction to enhance our understanding on the relation between plant growth regulators and herbicides to improve tolerance of crop plants is discussed. The study suggests the use of plant growth regulators as a tool in mitigating effects of herbicidal stress together with improved growth and development.

Keywords: Agriculture; Crop loss; Herbicides; Stress; Plant growth regulators; Tolerance

Introduction

Weeds are undesirable plants that grow along with the cultivated crops that compete with the crop for water, nutrients and light, make harvesting difficult and cause a considerable yield loss [1,2]. The yield losses of field crops (cereals, legumes, Brassica and vegetables) due to weeds are considerable. In Indian context, it has been reported that the loss is 10-60% in *Triticum aestivum*, 10-100% in *Oryza sativa*, 30-40% in *Zea mays*, 16-65% in *Pennisetum glaucum* and 50% in *Eleusine coracana*, 10-45% in *Vigna radiata*, 10-50% in *Cicer arietinum*, 30-35% in *Lens culinaris*, 10-100% in *Glycine max.*, 10-50% in *Pisum sativum* and 20-30% in *Cajanus cajan*, 20-30% in *Brassica* spp and 20-30% in vegetables [3]. The measures for management of weeds and to overcome the reduction in yield are necessary for sustainable agricultural production. There are different methods available for the control of weeds which include traditional method and advanced technological method, both of which have their own limitation. For example, hand weeding is effective but is cost expensive and difficult owing to labour scarcity and often results in high yield loss due to late weeding. Harrowing is another method for controlling intra-row weeds but is also less effective. Hoeing-ridging method effectively controls both inter and intra row weeds and reduces competitive ability and seed production of uncontrollable weeds [4]. The adoption of intercropping agriculture, a practice to grow two or more crops simultaneously in the same field, reduces the chances of growth of weeds. Another option available is the cover crops which includes suppression of weeds grown at same time of plant growth [5]. These available options require knowledge of soil crop management system, interaction between plants and weeds, and environmental influences associated with the weed control operations [6]. Moreover, there is also growing concern on the involvement of heavy cost in the use of advanced technology methods, such as sensing/guidance technology and robotic method for the control of weeds. Presently, herbicides are commonly used for the control of weeds in the agricultural practices. The herbicides are considered as chemical agents that provide an efficient and cost effective method for the control of weeds. The efficiency of herbicides in the control of weeds has increased their use to protect yield loss of crop plants and sustainability of agriculture, but their prolonged use has prompted the shift in weed population behavior [7] by which they become distinct in morphological, physiological and biochemical nature and may develop resistance to a number of herbicides [8]. Moreover, as the concentration of herbicides is increased it causes inhibition in growth and physiological functions of plants [9]. It is therefore, essential to evaluate the effects of herbicides on non-target

plants and impact on soil health, and develop methods that can have positive effects on development of crop plants in the presence of herbicides. Plant growth regulators have been recognized to stimulate growth and development of plants under optimal conditions. They are also involved in tolerance of plants to biotic and abiotic stresses [10-12]. They induce expression of stress related genes for maintaining photosynthetic activity, antioxidant enzymes activation and osmoprotectants accumulation against oxidative stress [13-15]. They can also prove beneficial for reducing the risks of herbicides exposure through modulating plant response to oxidative stress induced by herbicides. However, the importance of plant growth regulators in the control of herbicides needs more attention and collected information is necessary for understanding their role in herbicidal resistance. The present review evaluates the physiological, growth and yield responses of plants to herbicides treatment, and the impact of plant growth regulators in counteracting the adverse effects of herbicides. It also identifies the gap and delineates future prospects of research.

Responses of plants to herbicides

Herbicides control weed plants by disrupting or altering their metabolic processes but also induce stressful conditions and cause damage to the non-target plants. Moreover, they leave residues in soil that are harmful ecologically [16,17]. The plants may or may not recover from the damage caused by herbicide depending on its dose and the severity of the damage. The chances that both crop plants and weeds may develop resistance to the herbicides are also prominent [18]. The interaction between herbicides, weeds, crops and soil explaining the effects of herbicides on weeds, crop plants and soil, and chances of development of resistance in weeds and toxicity in cultivated plants is presented in Figure 1. The nature of herbicides and the dose of application are the important factors determining damage to crop plants as the herbicides may have different capacity to remove weeds

*Corresponding author: Nafees A Khan, Department of Botany, Aligarh Muslim University, Aligarh-202 002, Uttar Pradesh, India, E-mail: naf9@lycos.com

Received August 24, 2015; Accepted October 18, 2015; Published October 25, 2015

Citation: Varshney S, Khan MIR, Masood A, Per TS, Rasheed F, Khan NA, (2015) Contribution of Plant Growth Regulators in Mitigation of Herbicidal Stress. J Plant Biochem Physiol 3: 160. doi:10.4172/2329-9029.1000160

Copyright: © 2015 Varshney S, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

without affecting crop. The herbicides with common chemistry are grouped in families and shown in Figure 2. Two or more families may have the same mode of action and thus express same herbicide activity and injury symptoms. Herbicide family and mode of action of some gradient herbicides are presented in Table 1. The commonly growing weeds along with the crop plants and herbicides used are shown in Table 2. The effects of herbicides on various characteristics of plants have been evaluated in the following pages.

Photosynthesis

Herbicides negatively impact photosynthetic characteristics of plants. Their use has shown to decrease photosynthetic pigments, carbon assimilation and efficiency of pigment system II (PSII). Application of pendimethalin on *Foeniculum vulgare* leaves [19]; chevalier in *Triticum aestivum* cultivars [20] and metosulam in *Vicia faba* plants [21] has shown to decrease photosynthetic pigments. In a study on *Zea mays*, application of glyphosate at 10 mM resulted in decreased photosynthesis-related proteins [22]. The inhibition in carbon assimilation with 0.20 g L⁻¹ flauzifop and 0.14 g L⁻¹ haloxyfop in *Cucumis sativus* has been found [23]. Studies have shown that chlorophyll synthesis and content were decreased in plants with herbicides application. The decreased content of chlorophyll was found in *Triticum aestivum* with chlorimuron-ethyl at 300 µgkg⁻¹, isoproturon at 2 mg kg⁻¹, prometryne at 4 mg kg⁻¹, sekator at 150 mL ha⁻¹ and zoom at 120 gha⁻¹ [24-27]; in *Vicia faba* with fluometuron at 2.98 kg ha⁻¹ [28], in *Glycine max.* cultivars with diuron at 1, 2 and 5 ppm [29]. The decrease in photosynthetic characteristics with herbicides might have involved its effects on chlorophyll biosynthetic pathway and enzymes of carbon assimilation. The details on the key points in the biosynthetic pathway of photosynthetic pigments and activity of carbon assimilation enzymes are still lacking and needs attention. Chlorophyll fluorescence is a simple and widely used method to study the mechanism of photosynthesis and is used as marker to detect stress in plants. The herbicides application may block photosynthesis related intermediate metabolites and affect fluorescence emission. The herbicides that inhibit glutamine synthase, protoporphyrinogen oxidase and carotenoid biosynthesis affect chlorophyll fluorescence [30], and even lipid synthesis inhibitor herbicides induce chlorophyll fluorescence. Application of 1.96 kg ha⁻¹ Terbutryn to *Vicia faba* plants strongly decreased F_v/F_m ratio and CO₂ assimilation capacity [31]. Recently, Sousa et al. [32] have shown that application of imidazolinone group of herbicides to *Oryza sativa* changed the photosynthetic metabolism of plants and chlorophyll a fluorescence.

Osmolytes

The production of osmolytes is an adaptation against stress in plants [33]. These osmolytes include polyols (sugars, fructans, mannitol, pinitol and inositol), amino acids and their derivatives (proline, isoleucine, valine, betaine, glycinebetaine). Proline is widely studied osmolyte that acts as a cellular protector in several plant species by scavenging ROS produced in response to abiotic stress [34-36]. It provides resistance against stress either by uniting oxygen and free radicals generated in stress [37] or by efficiently scavenging OH⁻ and O₂⁻ and inhibit lipid peroxidation, and acts as a source of carbon and nitrogen during stress. Proline may be accumulated in plants in large amounts during stress either due to enhanced synthesis or reduced degradation. Herbicide treatment results in accumulation of proline as a measure of resistance to stress. Application of 5-25 mg kg⁻¹ chlorotoluron to *Triticum aestivum* [38] and 10⁻⁵, 10⁻⁶, 10⁻⁷% metosulam to *Vicia faba* plants [21] resulted in accumulation of proline. Soluble carbohydrates increased in *Glycine max.* by increasing diuron application from 1<2<5 ppm [29]. The increase in soluble sugars content in plants is an

adaptive response to support basal metabolism under stress. Soluble amino acids have been found increased more in leaves of susceptible cultivars of *Glycine max.* plants than resistant cultivars in response to glyphosate application at 2:100 ratio with water [39], but the content of soluble amino acids increased in roots in all the cultivars. The content of myo-inositol increased in roots of *Brassica napus* seedlings after glyphosate application in a concentration dependent manner from 40-2000 µM [40]. In contrast, application of chlorimuron-ethyl at 20 g ha⁻¹ decreased the amino acids; valine, leucine and isoleucine in *Triticum aestivum* and *Zea mays* plants [41]. These studies are however, limited to the quantification of osmolytes, and the key points influenced by the herbicides are not known. Also, studies are limited to few osmolytes and leaves scope for determination of array of osmolytes that plants require for stress tolerance.

Nitrogen and sulfur assimilation

Nitrogen is primary nutrient element and forms backbone for proteins and nitrogenous compounds required for growth and development of plants. The energy needed for N assimilation comes from the photosynthetic process. There exists a strong correlation between N uptake and chlorophyll fluorescence parameters as shown in *Triticum aestivum* [42]. Herbicides may influence N assimilation directly or through its effects on photosynthesis. The activity of nitrate reductase (NR) and nitrite reductase (NiR) was found inhibited on application of isoproturon at 1 kg ha⁻¹, accompanied with the inhibition of electron transfer [43]. The reduction in carbon dioxide fixation also reduced the activity of NR, as energy and carbon skeleton required for N assimilation are provided by photosynthesis. Zabalza et al. [44] reported that the decrease in nitrate uptake due to 52 µM imazethapyr was partly related to a reduced stomatal conductance in *Glycine max.* The decrease in NR activity with 3 L ha⁻¹ butachlor and 20 g ha⁻¹ chlorimuron-ethyl was found in *Triticum aestivum* and *Zea mays* [41]. In contrast, recently Amare et al. [45] found increased N uptake with 1.50 kg ha⁻¹ isoproturon application to *Triticum aestivum*. Sulfur makes a part of vitamins, co-enzymes, phytohormones and reduced sulfur compounds involved in the regulation of metabolism of plants under optimal and stressed conditions [46]. Reduced glutathione (GSH) is formed from its assimilation and reduction [47] and may have role in herbicide detoxification [48] since GSH has well accepted role in protection of plants from oxidative damage by keeping ROS under control [49]. In spite of the recognized role of S and S-compounds in abiotic stress tolerance, only few studies have been conducted to show the influence of herbicides on S assimilation. Application of 10

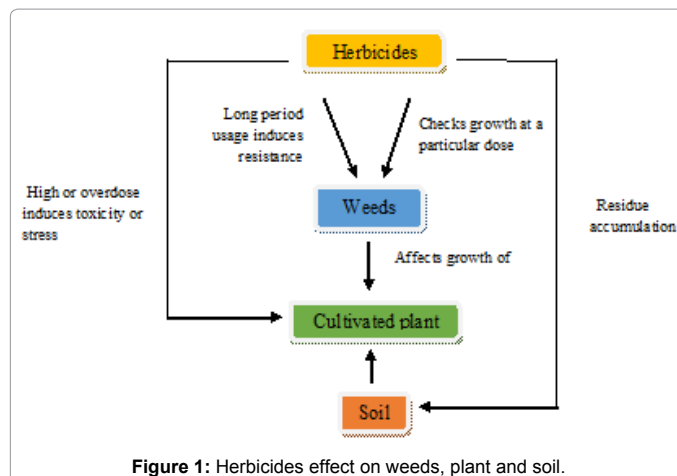


Figure 1: Herbicides effect on weeds, plant and soil.

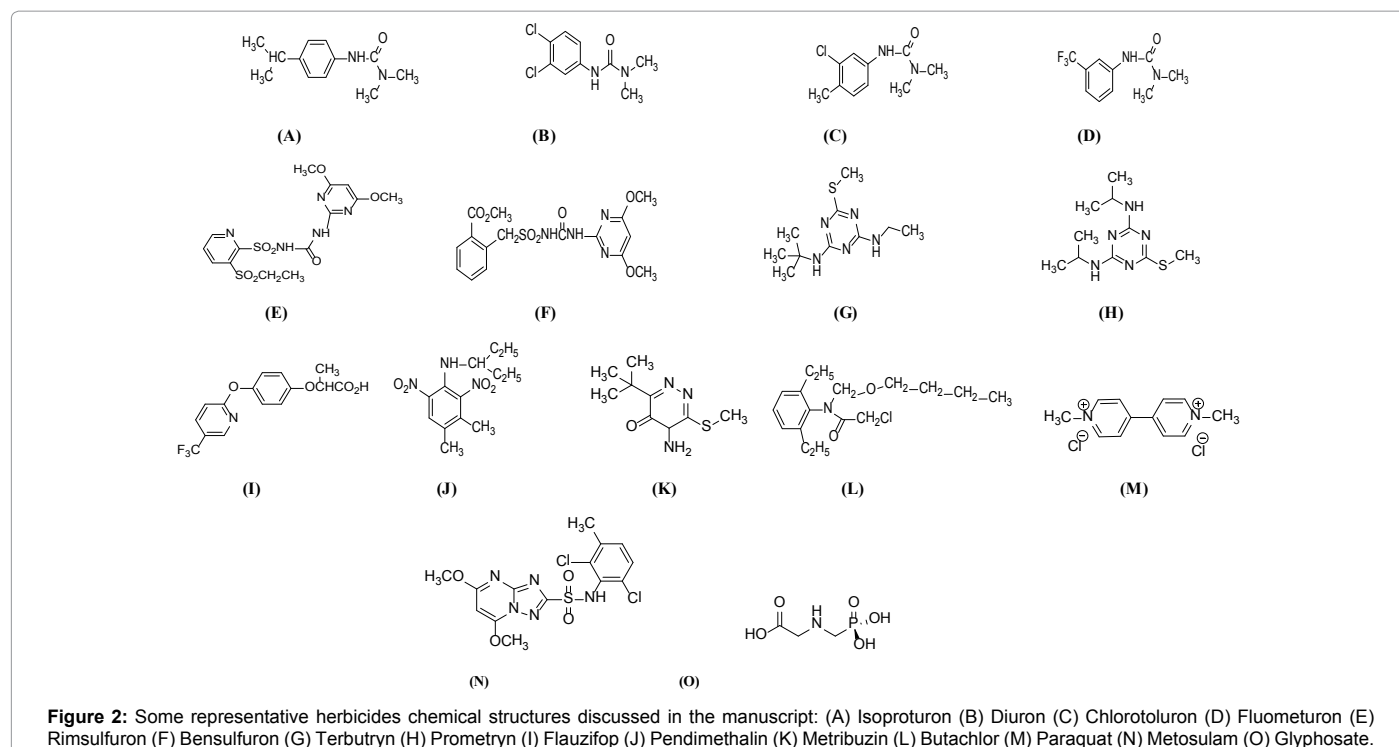


Table 1: Family, mode of action and site of action of gradient herbicides used for crops discussed in the review.

| S. No | Herbicide | IUPAC name | Herbicide family | Mode or site of action | Reference |
|-------|--------------------|--|----------------------------|---|-----------|
| 1. | Quizalofop-p-ethyl | ethyl (2R)-2-[4-(6-chloroquinoxalin-2-yl)oxyphenoxy] propanoate | Aryloxy phenoxy propionate | Lipid synthesis inhibitors at acetyl CoA carboxylase enzyme (ACCase) | [67] |
| | Flauzifop | (RS)-2-[4-[5-(trifluoromethyl)-2-pyridyloxy]phenoxy] propionic acid | | | [23] |
| | Haloxifop | (RS)-2-[4-[(3-chloro-5-trifluoromethyl-2-pyridyloxy) phenoxy]propionic acid | | | [56] |
| 2. | Imazethapyr | 5-ethyl-2-[(RS)-4-isopropyl-4-methyl-5-oxo-2-imidazolin-2-yl]nicotinic acid | Imidazolinones | Branched chain amino acid synthesis inhibitors at acetolactate synthase (ALS) | [44] |
| 3. | Rimsulfuron | 1-(4,6-dimethoxypyrimidin-2-yl)-3-(3-ethylsulfonyl-2-pyridylsulfonyl) urea | Sulfonylurea | Branched chain amino acid synthesis inhibitors at acetolactate synthase (ALS) | [28] |
| | Bensulfuron-methyl | methyl 2-[(4,6-dimethoxypyrimidin-2-yl) carbamoylsulfamoylmethyl]benzoate | | | [66] |
| 4. | Terbutryn | N ² -tert-butyl-N ⁴ -ethyl-6-methylthio-1,3,5-triazine-2,4-diamine | Triazine | Photosynthesis inhibitors at PS II | [31] |
| | Prometryn | N ² ,N ⁴ -diisopropyl-6-methylthio-1,3,5-triazine-2,4-diamine | | | [26] |
| 5. | Metribuzin | 4-amino-6-tert-butyl-3-methylsulfanyl-1,2,4-triazin-5-one | Triazinones | Photosynthesis inhibitor at PS II | [41] |
| 6. | Diuron | 3-(3,4-Dichlorophenyl)-1,1-dimethylurea | Phenylurea | Photosynthesis inhibitor at PS II | [72] |
| | Isoproturon | 3-(4-isopropylphenyl)-1,1-dimethylurea | | | [25] |

| | | | | | |
|-----|---------------|---|---------------------------------------|---|------|
| | Chlorotoluron | 3-(3-chloro- <i>p</i> -tolyl)-1,1-dimethylurea | | | [38] |
| | Fluometuron | 1,1-Dimethyl-3-[3-(trifluoromethyl)phenyl]urea | | | [28] |
| 7. | Paraquat | 1,1'-Dimethyl-4,4'-bipyridinium dichloride | Bipyriduliums | Photosynthesis inhibition at PS I | [55] |
| 8. | Glyphosate | N-(phosphonomethyl)glycine | Glycine | Aromatic amino acid inhibitor at EPSP synthase | [62] |
| 9. | Pendimethalin | 3,4-Dimethyl-2,6-dinitro- <i>N</i> -pentan-3-yl-aniline | Dinitroanilines | Mitotic disruptors or microtubule assembly inhibitors | [19] |
| 10. | Butachlor | N-(Butoxymethyl)-2-chloro-N-(2,6-diethylphenyl)acetamide | Acetanilide | Long chain fatty acid inhibitor | [41] |
| | Acetachlor | 2-Chloro- <i>N</i> -(ethoxymethyl)- <i>N</i> -(2-ethyl-6-methylphenyl)acetamide | | | [66] |
| 11. | Metosulam | 2',6'-dichloro-5,7-dimethoxy-3'-methyl[1,2,4]triazolo[1,5- <i>a</i>]pyrimidine-2 sulfonanilide | Triazolopyrimidine (TP) sulfonanilide | Acetolactate synthase ALS or (acetohydroxyacid synthase AHAS) inhibitor | [21] |

Table 2: Some common weeds present in crop plants and herbicides used to control the weeds.

| S No | Crop | Weeds | Herbicides | References |
|------|-------------------------------------|--|--|--------------|
| 1. | Wheat (<i>Triticum aestivum</i>) | <i>Phalaris minor</i> <i>Avena fatua</i> <i>Argemone Mexicana</i> <i>Rumex dentatus</i> | Clodinafop Fenoxaprop Sulfosulfuron Isoproturon | [2] |
| 2. | Maize (<i>Zea mays</i>) | <i>Amaranthus spinosus</i> <i>Cynodon dactylon</i> <i>Commelina benghalensis</i> <i>Bidens pilosa</i> | Metribuzin Butachlor Chlorimuron ethyl | [86] [41] |
| 3. | Rice (<i>Oryza sativa</i>) | <i>Fimbristylis miliacea</i> <i>Cyperus iria</i> <i>Cyperus difformis</i> | Acetachlor Bensulfuron-methyl | [87] [66] |
| 4. | Black gram (<i>Vigna mungo</i>) | <i>Parthenium</i> <i>Tridax</i> <i>Hyptis</i> <i>Cortaderia selloana</i> | Quizalofop-p-ethyl | [88] [67] |
| 5. | Mung bean (<i>Vigna radiata</i>) | <i>Cynodon dactylon</i> <i>Echinochloa colona</i> <i>Chenopodium album</i> <i>Amaranthus viridis</i> | Pendimethalin Oxyfluorfen Quizalofop-p-ethyl | [89] |
| 6. | Soyabean (<i>Glycine max</i>) | <i>Trianthema portulacastrum</i> <i>Celosia</i> <i>Echinochloa colona</i> | Flauzifop Pendimethalin Trifluralin | [90] |
| 7. | Potato (<i>Solanum tuberosum</i>) | <i>Amaranthus retroflexus</i> <i>Sinapsis arvensis</i> | Metribuzin | [91] |

| | | | | |
|-----|-------------------------------------|--|---|--------------|
| 8. | Onion (<i>Allium cepa</i>) | <i>Anthemis palastina</i> <i>Amaranthus spp.</i> <i>Sinapsis arvensis</i> <i>Trigonella arabica</i> | Oxyfluorfen Trifluralin Pendimethalin Metribuzin | [92] |
| 9. | Pea (<i>Pisum sativum</i>) | <i>Chenopodium album</i> <i>Echinocloa crus-galli</i> <i>Sonchus arvensis</i> <i>Fallopia convolvulus</i> | Imazethapyr Metribuzin Pendimethalin | [93] [94] |
| 10. | Cucumber (<i>Cucumis sativus</i>) | <i>Capsella bursa-pastoris</i> <i>Stellaria media</i> <i>Chenopodium album</i> <i>Brassica kaber</i> | Trifluralin Sethoxydim Clomazone | [95] |

mM glyphosate increased glutathione-S-transferase (GST) activity in *Pisum sativum* [50] and *Zea mays* [22]. Yin et al. [25] and Jiang and Yang [26] also found increase in GST activity in *Triticum aestivum* on treatment with 5 and 10 mg kg⁻¹ isoproturon and low concentration of prometryne at 4, 8 and 12 mg kg⁻¹. The detailed studies on S assimilation and S-compounds could provide information on better tolerance of plants to herbicide treatment.

Antioxidant system

The enhanced production of reactive oxygen species (ROS; superoxide radicals, O²⁻; hydroxyl radicals, OH; perhydroxyl radicals, HO²; alkoxy radicals, RO; hydrogen peroxide, H₂O₂; singlet oxygen, ¹O₂) takes place under abiotic stress conditions which damage plants primary metabolites; protein, lipid, carbohydrate, DNA and halt metabolic activities of cell [51]. Plants require activation of antioxidative system, especially ROS-scavenging systems within the cells that help in stress tolerance and contribute to continued plant growth under stress conditions [52]. The effects of herbicides on ROS and antioxidant system have been shown in plants. Application of isoproturon increased malondialdehyde production, while decreased the content of GSH and ascorbic acid and reduced the activity of SOD, CAT and APX in *Triticum aestivum* seedlings [53]. In another study, application of isoproturon resulted in increased CAT activity and decreased peroxidase (POD) activity [54]. Lipid peroxidation increased in *Triticum aestivum* cultivars by application of chevalier at 0.9, 1.2, 1.5 mg pot⁻¹ [20]. The activity of antioxidant enzymes; SOD and APX increased by application of 50 μM haloxyfop-ethoxyethyl and 15 μM paraquat to *Triticum aestivum* [55,56]. A concentration dependent change in antioxidant activity has been reported with prometryne in *Triticum aestivum*; low concentration of prometryne at 4, 8, 12 mg kg⁻¹ increased the activity of SOD, POD, CAT, APX activity in leaves, while higher concentration 16, 20, 24 mg kg⁻¹ decreased the activity [26]. The induced accumulation of superoxide anion (O₂⁻) and H₂O₂ in leaves and peroxidation of plasma membrane lipids were found with 5-25 mg kg⁻¹ chlorotoluron to *Triticum aestivum* in a concentration dependent manner. Moreover, the total activity of POD in roots was enhanced by 5 mg kg⁻¹, but CAT activity generally suppressed under the chlorotoluron exposure at 20-25 mg kg⁻¹ [38]. In contrast, glyphosate application at 10 μM increased CAT activity in *Zea mays* [22]. The treatment of *Zea mays* with herbicides 2.98 kg ha⁻¹ fluometuron and 0.015 kg ha⁻¹ rimsulfuron increased POD activity [28]. However, the application of increasing concentration of metosulam herbicide from 10⁻⁷ to 10⁻⁵% in plants resulted in decrease of POD activity and increase of CAT and

APX in *Vicia faba* plants [21]. The activity of SOD and CAT declined in proportion to concentration of paraquat (0.1, 1, 2 μM) applied to *Lactuca sativa*, *Phaseolus coccineus* and *Pisum sativum* [57].

Growth and yield

The use of herbicides negatively influences growth and yield of crop plants. The application of 8.5 mL L⁻¹ pendimethalin reduced growth characteristics; shoot length, fresh and dry weight of *Foeniculum vulgare* plants [19]. Application of phenoxy herbicide, MCPA (4 -chloro-2-methyl-phenoxy acid) at 1.5 L ha⁻¹ has been reported to reduce shoot length in *Triticum aestivum* [58], while the use of metosulam at 10⁻⁷ to 10⁻⁵% induced cytotoxic effects in *Vicia faba* and decreased fresh and dry weight of plants [21]. Glyphosate treatment at 2 to 10 mM concentration in two varieties of *Vigna radiata* produced differential effects. The germination percentage was similar to control at 6 mM, but was increased at 8 mM in one variety. On the other hand, fresh weight declined by two-fold on increasing concentration to 10 mM in both the varieties [59]. In another study, glyphosate application at 0.62 and 0.84 kg ha⁻¹ to *Triticum aestivum* [60], 100, 250 and 500 μg L⁻¹ to *Allium cepa* [61] and 1, 2, 4, 6, 8, and 10 mM to six *Vigna mungo* genotypes [62] was found to inhibit germination in a concentration dependent manner, and decreased leaf area and shoot dry biomass of *Glycine max.* at 800, 1200, and 2400 g ha⁻¹ [63]. Higher concentration of glyphosate at 500 μg L⁻¹ was found inhibitory on root growth in *Allium cepa* [61] and at 10 mM for growth of six *Vigna mungo* genotypes [62]. Haloxyfop-ethoxyethyl at 50 μM inhibited growth of *Triticum* leaves [56], while increasing dose of chevalier at 0.9, 1.2 and 1.5 mg pot⁻¹ resulted in a greater decrease of roots than shoots of *Triticum aestivum* [20]. Root length and shoot length [25], shoot fresh and dry weights of *Triticum aestivum* were decreased after treatment with isoproturon [53] at 2-20 mg kg⁻¹ and 2.5 L ha⁻¹, respectively. Application of 1 kg ha⁻¹ metribuzin, 3 L ha⁻¹ butachlor and 20 g ha⁻¹ chlorimuron-ethyl to *Triticum aestivum* and *Zea mays* seedlings resulted in decrease of shoot fresh weight [41]. Pendimethalin at 0.5-10 ppm reduced the germination percentage of *Zea mays* [64], while isoproturon at 2.5 kg ha⁻¹ significantly reduced the fresh and dry weight of shoot and root of *Zea mays* seedlings [65]. Fresh weight of *Oryza sativa* shoot and root was reduced by the application of acetachlor at 3.2 μmol L⁻¹ and bensulfuron-methyl at 0.96 μmol L⁻¹ [66]. Higher concentration of quizalafop-p-ethyl at 1, 1.5 and 2% decreased the yield of *Vigna mungo* [67]. Reduction in number of tillers per plant and grain yield in *Triticum aestivum* was recorded with 1 kg ha⁻¹ isoproturon [43], while bromoxynil decreased the number of spikes, main spike length, grains weight and straw per plant [68].

Pendimethalin decreased yield components of *Foeniculum vulgare* [19]. Table 3 presents the summary of studies on the effects of different doses of herbicides on various characteristics of plants.

Role of plant growth regulators in herbicidal stress tolerance

Among the several strategies adopted by plants to counteract the adverse effects of abiotic stress, plant growth regulators provide signals to allow plants to survive under stress conditions. The modulation of their levels in plants in environmental stressed conditions modifies the signaling pathways and induces resistance in plants [69,70]. They are one of the key systems integrating metabolic and developmental events in the whole plant, and are essential for many processes throughout the life of a plant, influencing the yield and quality of crops. The compounds

that have been considered as plant hormones are described as classical hormones (auxin, cytokinin, gibberellins, ethylene, abscisic acid) and newly added compounds (brassinosteroids, salicylic acid, nitric oxide, jasmonic acid).

Classical hormones

Auxins have been reported to impart resistance to herbicide toxicity. Application of 2, 4-dichlorophenoxyacetic acid at 60-120 g ha⁻¹ was found suitable for protecting cultivated plants against the phytotoxic action of 20-60 g ha⁻¹ clodinafop-propargyl [71]. The protective effect of benzyladenine at 1, 10, 100 μM reduced the decrease in chlorophyll, carotenoid and ascorbic acid contents of *Zea mays* leaves caused by paraquat. Pretreatment with benzyladenine

Table 3: Some representatives herbicides used in agricultural practices and their effect on various parameters studied. Abbreviations: -: Decrease; +: Increase; APX: Ascorbate peroxidase; CAT: Catalase; GOGAT: Glutamine oxoglutarate aminotransferase; GR: Glutathione reductase; GST: Glutathione-S-transferase; LPO: Lipid peroxidation; MDA: Malondialdehyde; NR: Nitrate reductase; POX: Peroxidase; SOD: Superoxide dismutase.

| S No | Herbicides | Plants name | Concentration of herbicides | Response | Affected parameters | References |
|------|--|--|--|----------|--|------------|
| 1. | Isoproturon | <i>Triticum aestivum</i> | 2, 3.5, 5, 10, 20 mg kg ⁻¹ | - | Growth, photosynthesis, chlorophyll content | [25] |
| 2. | Isoproturon | <i>Triticum aestivum</i> | 2, 3.5, 5, 10 and 20 mg kg ⁻¹ | + | SOD, APX, POD and GST | [25] |
| 3. | Pendimethalin | <i>Zea mays</i> | 0.5, 1.0, 2.5, 5.0 and 10.0 ppm | - | Seed germination | [64] |
| 4. | Chlorotoluron | <i>Triticum aestivum</i> | 5, 10, 15, 20 and 25 mg kg ⁻¹ | - | Biomass | [38] |
| 5. | Chlorotoluron | <i>Triticum aestivum</i> | 5, 10, 15, 20 and 25 mg kg ⁻¹ | + | Enhanced total activity of POD and APX, and proline content | [38] |
| 6. | Chlorotoluron | <i>Triticum aestivum</i> | 5, 10, 15, 20 and 25 mg kg ⁻¹ | - | CAT activity | [38] |
| 7. | Metribuzin | <i>Triticum aestivum</i> , <i>Zea mays</i> | 1 kg ha ⁻¹ | - | Shoot fresh and dry mass, GS and GOGAT activity | [41] |
| 8. | Butachlor | <i>Triticum aestivum</i> , <i>Zea mays</i> | 3 L ha ⁻¹ | - | Shoot fresh and dry mass, GS and GOGAT activity | [41] |
| 9. | Chlorimuron-ethyl | <i>Triticum aestivum</i> , <i>Zea mays</i> | 20 g ha ⁻¹ | - | Shoot fresh and dry mass, GS and GOGAT activity | [41] |
| 10. | Acetachlor | <i>Oryza sativa</i> | 3.2 μmol L ⁻¹ | - | Root and shoot biomass, NR activity | [66] |
| 11. | Bensulfuron-methyl | <i>Oryza sativa</i> | 0.96 μmol L ⁻¹ | - | Root and shoot biomass, NR activity | [66] |
| 12. | Glyphosate | <i>Vigna radiata</i> | 2, 4, 6, 8, and 10mM | - | Germination percentage, Root length, Fresh weight | [59] |
| 13. | Glyphosate | <i>Vigna radiata</i> | 2, 4, 6, 8, and 10mM | + | CAT, POD, GST | [59] |
| 14. | Paraquat | <i>Lactuca sativa</i> , <i>Phaseolus coccineus</i> , <i>Pisum sativum</i> | 0.1, 1, and 2 μM | - | Chlorophyll and carotenoid content, SOD, CAT | [57] |
| 15. | Alachlor | <i>Lactuca sativa</i> , <i>Phaseolus coccineus</i> , <i>Pisum sativum</i> | 0.2, 1, 2, 20, 100, and 200 μM | - | Chlorophyll and carotenoid content, SOD, CAT | [57] |
| 16. | Metolachlor | <i>Lactuca sativa</i> , <i>Phaseolus coccineus</i> , <i>Pisum sativum</i> | 0.2, 1, 2, 20, 100, and 200 μM | - | Chlorophyll and carotenoid content, SOD, CAT | [57] |
| 17. | Paraquat | <i>Triticum aestivum</i> | 15, 30 and 60 μM | - | Chlorophyll fluorescence, chlorophyll and carotenoid content | [55] |
| 18. | Paraquat | <i>Triticum aestivum</i> | 15, 30 μM | + | SOD and POD | [55] |
| 19. | TOPIK (active ingredient-clodinafop-propargyl) | <i>Secale cereale</i> , <i>Zea mays</i> | 8, 80 and 800 μg L ⁻¹ | + | LPO Intensity; CAT and APX | [9] |
| 20. | Metosulam | <i>Vicia faba</i> | 10 ⁻⁵ , 10 ⁻⁶ and 10 ⁻⁷ % | - | Photosynthetic pigment, Fresh and dry weight, Mitotic index; Cell division | [21] |
| 21. | Metosulam | <i>Vicia faba</i> | 10 ⁻⁵ , 10 ⁻⁶ and 10 ⁻⁷ % | + | Chromosomal aberrations, Proline content | [21] |
| 22. | Prometryn | <i>Triticum aestivum</i> | 4, 8, 12, 16, 20 and 24 mg kg ⁻¹ | - | Growth and biomass, Chlorophyll content | [26] |

significantly increased SOD activity after paraquat treatment [72]. Phenylurea cytokinin at 0.1 mM reduced the deleterious effects of 10 mM glyphosate in *Zea mays* plants. It induced its protective action by hardiness in the antioxidant defense systems [22]. Treatment of *Zea mays* leaves with exogenous abscisic acid led to significant decrease in the content of malondialdehyde, the percentage of ion leakage and the level of protein oxidation under 50 μM paraquat stress. Abscisic acid induced protection against oxidative damage generated by paraquat through mitogen activated protein kinase cascade in *Zea mays* leaves [73]. Pretreatment with 5 μM abscisic acid enhanced tolerance of *Oryza sativa* plants to 10 μM phosphinothricin and reduced ammonium ion accumulation in leaves [74]. The increase in endogenous abscisic acid content in leaves was closely related to phosphinothricin tolerance.

Newly added plant hormones

Besides these classical hormones, salicylic acid is an important signaling molecule that plays a vital role in protecting plants against herbicide toxicity [75]. It regulates plant response to organic contaminants and provides a basis to control herbicide contamination in crop production. Salicylic acid at 500 $\mu\text{mol L}^{-1}$ mediates 10 $\mu\text{mol L}^{-1}$ paraquat tolerance in *Hordeum vulgare* [76] and in *Zea mays* seedling by enhancing chlorophyll and carotenoid content and leaf total protein. In addition, salicylic acid at 0.5 and 1.0 mM significantly reduced free proline, lipid peroxidation and CAT activity in 10 mM paraquat stressed *Zea mays* seedlings [77]. It improved growth of napropamide treated *Brassica napus* and reduced napropamide levels in plants. Its application at 0.1 mM also decreased the abundance of O_2^- and H_2O_2 as well as activity of SOD, CAT, APX and increased activities of POD, guaiacol peroxidase (GPX) and GST in 8 mg L^{-1} napropamide-exposed plants [78]. Nitric oxide has been shown to reduce herbicide toxicity by its protective effects on plants. The reduction in protein content in *Oryza sativa* leaves by paraquat was prevented by nitric oxide by reducing the level of malondialdehyde and improving the activity of antioxidant enzymes, such as SOD, POD and GR [79]. Nitric oxide given as sodium nitroprusside at 50, 100 and 200 μM protected *Glycine max* plants by scavenging the ROS generated by 168 g ha^{-1} lactofen [80]. The protective role of brassinosteroids against biotic and abiotic stress has been documented in literature, but the action of brassinosteroids against damage by herbicides is not well validated. There are few studies that demonstrate that brassinosteroids function to protect metabolism against effects of herbicides. Study of Pinol and Simon [31] has shown protective action of brassinosteroids against photosynthesis-inhibiting herbicides. The study has shown that epibrassinolide at 2×10^{-6} or 2×10^{-5} mM induced the recovery from damage caused by 1.96 kg ha^{-1} terbutryn in *Vicia faba* plants, as detected by chlorophyll fluorescence and photosynthetic CO_2 assimilation. Polyamines (putrescine, spermidine and spermine) are also known to participate in plants for protection against abiotic stress conditions [81,82]. Spermidine offered protection against paraquat toxicity in *Raphanus sativus* seedlings [83]. Diethylenetriamine polyamine increased the protection against 2, 4-D herbicide in the triticale plants [84]. Application of polyamines significantly prevented the loss of chlorophyll content, reduced the lipid peroxidation caused by diuron herbicide in leaves of *Zea mays* and also increased the activity of SOD, GPX and GR [85]. The use of plant growth regulators results in the development of resistance against herbicides through activation of defense mechanisms, such as control of ROS production, enhanced osmolytes biosynthesis, nutrients homeostasis and activation of antioxidant system resulting in protection of cells from damage. These effects together ensured higher photosynthesis, growth and yield. The possible mechanisms involved in the herbicidal stress tolerance through the use of plant growth

regulators are shown in Figure 3. However, there is need to study the extent of mechanisms induced by individual plant growth regulators in mitigation of herbicidal stress.

Conclusion

The use of herbicides has become a potential tool in modern agriculture to get rid of weeds menace, but unknowingly the indiscriminate application of herbicides has resulted in adverse effects on the morphological, physiological and biochemical characteristics of crop plants. The production of biomass of crop plants is negatively affected due to impaired photosynthetic activity generated by toxicity induced by herbicides. The traditional as well as modern methods for the control of weeds are not cost effective. Phytohormones have potential in protecting and/or alleviating adverse herbicidal effects in plants as they have significant role in antioxidant metabolism and synthesis of metabolites known for providing stress tolerance. However, the studies related to plant growth regulators action in protection against herbicides are few and in depth experimental studies are required to explain the mechanisms responsible for counteracting herbicidal effects. The developmental stage of plants is important in the use of herbicides to evaluate their effects and the dose of applied herbicide needs to be evaluated for crops to reduce their effects. The target points in biosynthetic pathways are to be investigated on which herbicides act, so that these key points can be manipulated for reversing the effects of herbicides. Moreover, the use of molecular tools may help in explaining the proteins involved in providing tolerance to herbicidal effects. In addition, the gap in knowledge and understanding of the effects of herbicides action on various physiological, biochemical and molecular levels are required to explore the regulatory points for combating the effects. The modulation in the endogenous levels of plant growth regulators or signaling for gene expression in plants is required to cope up with the stressful conditions. Further, more studies

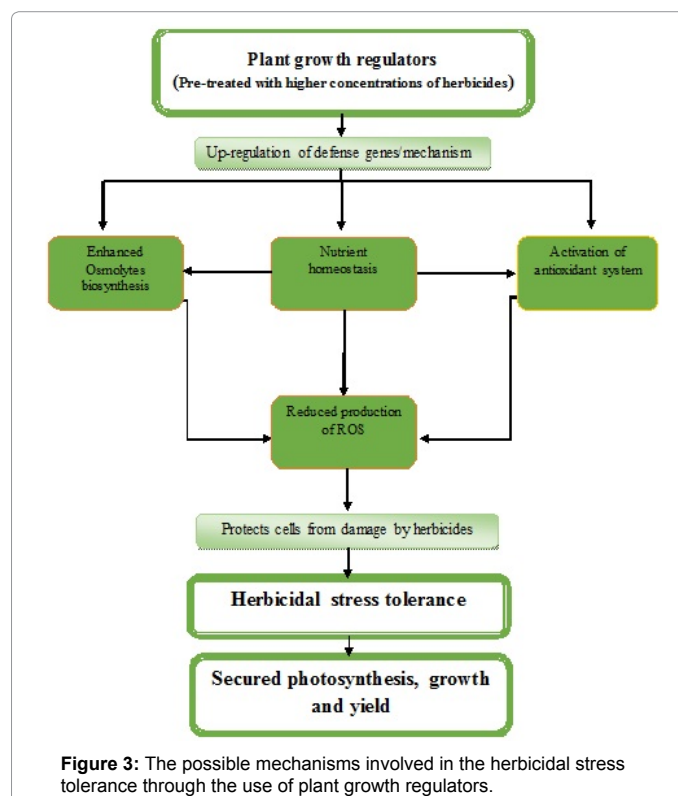


Figure 3: The possible mechanisms involved in the herbicidal stress tolerance through the use of plant growth regulators.

should be focused on unraveling the cross-talk between different plant hormones and other molecules in regulating the toxicity induced by herbicides.

Acknowledgements

Authors (SV and FR) gratefully acknowledge the research fellowship by the University Grants Commission, New Delhi, India.

References

1. Khan I, Hassan G, Khan MI, Khan IA (2004) Efficacy of some new herbicidal molecules on grassy and broadleaf weeds in wheat-II. Pak J Weed Sci Res 10: 33-38.
2. Chhokar RS, Sharma RK, Sharma I (2012) Weed management strategies in wheat a review. J Wheat Res 4: 1-21.
3. Rao AN, Wani SP, Ladha JK (2014) Weed management research in India an analysis of past and outlook for future. Monograph, DWR Jabalpur.
4. Pannacci E, Tei F (2014) Effects of mechanical and chemical methods on weed control weed seed rain and crop yield in maize sunflower and soyabean. Crop Prot 64: 51-59.
5. Brennan EB, Smith RF (2005) Winter cover crop growth and weed suppression on the central coast of California. Weed Technol 19: 1017-1024.
6. Kochaki A, Zarifketabi H, Nakhforosh AR (2001) Ecological Approaches on weed management. University of Ferdowsi. p: 457.
7. Owen MDK (2008) Weed species shifts in glyphosate-resistant crops. Pest Manag Sci 64: 377-387.
8. Heap I (2004) The international survey of herbicide resistant weeds.
9. Lukatkin AS, Garkova AN, Bochkarjova AS, Nushtaeva OV, da Silva JAT (2013) Treatment with the herbicide TOPIK induces oxidative stress in cereal leaves. Pestic Biochem Physiol 105: 44-49.
10. Khan NA, Nazar R, Iqbal N, Anjum NA (2012) Phytohormones and abiotic stress tolerance in plants. Springer-Verlag, New York, USA.
11. Ahammed GJ, Zhou YH, Xia XJ, Mao WH, Shi K, et al. (2013) Brassinosteroids regulates secondary metabolism in tomato towards enhanced tolerance to phenanthrene. Biol Plant 57: 154-158.
12. Bhajbhujje MN (2015) *In vitro* fungitoxic effect of some plant growth regulators on spore germination and germ tube emergence of *Alternaria solani*. Int J Life Sci 3: 125-130.
13. Saibo NJ, Lourenço T, Oliveira MM (2009) Transcription factors and regulation of photosynthetic and related metabolism under environmental stresses. Ann Bot 103: 609-623.
14. Fukuda A, Tanaka Y (2006) Effects of ABA, auxin, and gibberellin on the expression of genes for vacuolar H⁺-inorganic pyrophosphatase, H⁺-ATPase subunit A, and Na⁺/H⁺ antiporter in barley. Plant Physiol Biochem 44: 351-358.
15. Tran LSP, Urao T, Qin F, Maruyama K, Kakimoto T, et al. (2007) Functional analysis of AHK1/ATHK1 and cytokinin receptor histidine kinases in response to abscisic acid, drought, and salt stress in Arabidopsis. Proceedings of the National Academy of Sciences 104: 20623-20628.
16. Derksen DA, Anderson RL, Blackshaw RE, Maxwell B (2002) Weed dynamics and management strategies for cropping systems in the Northern Great Plains. Agron J 94: 174-185.
17. Riaz M, Jamil M, Mahmood TZ (2007) Yield and yield components of maize as affected by various weed control methods under rain-fed conditions of Pakistan. Int J Agric Biol 9: 152-155.
18. Szigetzi Z, Lehoczki E (2003) A review of physiological and biochemical aspects of resistance to atrazine and paraquat in Hungarian weeds. Pest Manag Sci 59: 451-458.
19. El-Awadi ME, Hassan EA (2011) Improving growth and productivity of fennel plant exposed to pendimethalin herbicide: stress-recovery treatments. Nat Sci 9: 97-108.
20. Nahiha B, Reda DM, Noureddine Z, Houria B (2014) Differential response to treatment with herbicide chevalier induced oxidative stress in leaves of wheat. Ann Biol Res 5: 1-7.
21. Badr A, Zaki H, Germoush MO, Tawfeek AQ, El-Tayeb MA (2013) Cytophysiological impacts of metosulam herbicide on *Vicia faba* plants. Acta Physiol Plant 35: 1933-1941.
22. Sergiev IG, Alexieva VS, Ivanov SV, Moskova II, Karanov EN (2006) The phenyl urea cytokinin 4PU-30 protects maize plants against glyphosate action. Pestic Biochem Physiol 85: 139-146.
23. Xia XJ, Huang YY, Wang L, Huang LF, Yu YL, et al. (2006) Pesticides-induced depression of photosynthesis was alleviated by 24-epibrassinolide pretreatment in *Cucumis sativus* L. Pestic Biochem Physiol 86: 42-48.
24. Wang ME, Zhou QX (2006) Effects of herbicide chlorimuron-ethyl on physiological mechanisms in wheat *Triticum aestivum*. Ecotoxicol Environ Saf 64: 190-197.
25. Yin XL, Jiang L, Song NH, Yang H (2008) Toxic reactivity of wheat *Triticum aestivum* plants to herbicide isoproturon. J Agric Food Chem 56: 4825-4831.
26. Jiang L, Yang H (2009) Prometryne-induced oxidative stress and impact on antioxidant enzymes in wheat. Ecotoxicol Environ Saf 72: 1687-1693.
27. Hana S, Leila MA, Nedjoud G, Reda DM, Shing WL (2015) Physiology and biochemistry effects of herbicides sekator and zoom on two varieties of wheat (Waha and HD) in Semi-Arid Region. Annu Res Rev Biol 5: 449.
28. Hassan NM, Alla MN (2005) Oxidative stress in herbicide treated broad bean and maize plants. Acta Physiol Plant 27: 429-438.
29. Fayez KA (2000) Action of photosynthetic diuron herbicide on cell organelles and biochemical constituents of the leaves of two soybean cultivars. Pestic Biochem Physiol 66: 105-115.
30. Hess FD (2000) Light-dependent herbicides: an overview. Weed Sci 48: 160-170.
31. Pinol R, Simon E (2009) Effect of 24-epibrassinolide on chlorophyll fluorescence and photosynthetic CO₂ assimilation in *Vicia faba* plants treated with the photosynthesis inhibiting herbicide terbutryn. J Plant Growth Regul 28: 97-105.
32. Sousa CP, Pinto JJO, Martinazzo EG, Perboni AT, Farias ME, et al. (2014) Chlorophyll a fluorescence in rice plants exposed of herbicides of group imidazolinone. Planta 32: 141-150.
33. Parida AK, Das AB (2005) Salt tolerance and salinity effects on plants: a review. Ecotoxicol Environ Saf 60: 324-349.
34. Demiral T, Turkan I (2006) Exogenous glycine betaine affects growth and proline accumulation and retards senescence in two rice cultivars under NaCl stress. Environ Exp Bot 56: 72-79.
35. Ashraf M, Foolad M (2007) Roles of glycine betaine and proline in improving plant abiotic stress resistance. Environ Exp Bot 59: 206-216.
36. Mittal S, Kumari N, Sharma V (2012) Differential response of salt stress on *Brassica juncea*: photosynthetic performance, pigment, proline, D1 and antioxidant enzymes. Plant Physiol Biochem 54: 17-26.
37. Ghorbanli M, Gafarabad M, Amirian T, Allahverdi MB (2013) Investigation of proline, total protein, chlorophyll, ascorbate and dehydro ascorbate changes under drought stress in Akria and Mobil tomato cultivars. Iranian J Plant Physiol 3: 651-658.
38. Song NH, Yin XL, Chen GF, Yang H (2007) Biological responses of wheat *Triticum aestivum* plants to the herbicide chlorotoluron in soils. Chemosphere 68: 1779-1787.
39. Moldes CA, Medici LO, Abrahao OS, Tsai SM, Azevedo RA (2008) Biochemical responses of glyphosate resistant and susceptible soybean plants exposed to glyphosate. Acta Physiol Plant 30: 469-479.
40. Piotrowicz-Cieslak AI, Adomas B, Michalczuk DJ (2010) Different glyphosate phytotoxicity to seeds and seedlings of selected plant species. Pol J Environ Stud 19: 123-129.
41. Alla MN, Badawi AM, Hassan NM, El-Bastawisy ZM, Badran EG (2008) Effect of metribuzin, butachlor and chlorimuron-ethyl on amino acid and protein formation in wheat and maize seedlings. Pestic Biochem Physiol 90: 8-18.
42. Schachtl J, Huber G, Maidl FX, Sticksei E, Schulz J, et al. (2005) Laser-induced chlorophyll fluorescence measurements for detecting the nitrogen status of wheat *Triticum aestivum* L. canopies. Precision Agric 6: 143-156.
43. Singh SP, Pandey P, Kumar M, Singh S, Pandey NS, et al. (2013) Growth and biochemical responses of wheat *Triticum aestivum* L. to different herbicides. Afr J Agric Res 8: 1265-1269.

44. Zabalza A, Gaston S, Ribas-Carbo M, Orcaray L, Igal M, et al. (2006) Nitrogen assimilation studies using ^{15}N in soybean plants treated with imazethapyr, an inhibitor of branched-chain amino acid biosynthesis. J Agric Food Chem 54: 8818-8823.
45. Amare T, Sharma JJ, Zewdie K (2014) Effect of weed control methods on weeds and wheat *Triticum aestivum* L. yield. World J Agr Res 2: 124-128.
46. Nazar R, Iqbal N, Masood A, Syeed S, Khan NA (2011) Understanding the significance of sulfur in improving salinity tolerance in plants. Environ Exp Bot 70: 80-87.
47. Masood A, Iqbal N, Khan NA (2012) Role of ethylene in alleviation of cadmium-induced photosynthetic capacity inhibition by sulphur in mustard. Plant Cell Environ 35: 524-533.
48. Cummins I, Dixon DP, Freitag-Pohl S, Skipsey M, Edwards R (2011) Multiple roles for plant glutathione transferases in xenobiotic detoxification. Drug Metab Rev 43: 266-280.
49. Rausch T, Gromes R, Liedschulle V, Muller I, Bogs J, et al. (2007) Novel insight into the regulation of GSH biosynthesis in higher plants. Plant Biol 9: 565-572.
50. Miteva LE, Ivanov SV, Alexieva VS (2010) Alterations in glutathione pool and some related enzymes in leaves and roots of pea plants treated with the herbicide glyphosate. Russ J Plant Physiol 57: 131-136.
51. Anjum NA, Sofu A, Scopa A, Roychoudhury A, Gill SS, et al. (2014) Lipids and proteins-major targets of oxidative modifications in abiotic stressed plants. Environ Sci Pollut Res 1-23.
52. El-Mashad A, Mohamed H (2012) Brassinolide alleviates salt stress and increases antioxidant activity of cowpea plants *Vigna sinensis*. Protoplasma 249: 625-635.
53. Alla MMN, Hassan NM (2014) Alleviation of isoproturon toxicity to wheat by exogenous application of glutathione. Pestic Biochem Physiol 112: 56-62.
54. Mohammed AMA, El-Mashad AA, Kamel EA (2000) Biochemical changes in germinated wheat grain treated with herbicides. Ann Agric Sci Cairo 2: 457-476.
55. Ekmekci Y, Terzioglu S (2005) Effects of oxidative stress induced by paraquat on wild and cultivated wheat. Pestic Biochem Physiol 83: 69-81.
56. Janicka U, Mioduszewska H, Kielak E, Klocek J, Horbowicz M (2008) The effect of haloxyfop-ethoxyethyl on antioxidant enzyme activities and growth of wheat leaves (*Triticum vulgare* L.). Pol J Environ Stud 17: 485-490.
57. Stajner D, Popovic M, Stajner M (2003) Herbicide induced oxidative stress in lettuce, beans, pea seeds and leaves. Biol Plant 47: 575-579.
58. Zaltauskaite J, Kisonaite G (2014) The effects of phenoxy herbicide MCPA on non-target vegetation in spring wheat (*Triticum aestivum* L.) culture. Biologija 60: 148-154.
59. Basantani M, Srivastava A, Sen S (2011) Elevated antioxidant response and induction of tau-class glutathione S-transferase after glyphosate treatment in *Vigna radiata* (L.) Wilczek. Pestic Biochem Physiol 99: 111-117.
60. Yenish JP, Young FL (2000) Effect of preharvest glyphosate application on seed and seedling quality of spring wheat (*Triticum aestivum*). Weed Technol 14: 212-217.
61. Cavusoglu K, Yalcin E, Turkmen Z, Yapar K, Cavusoglu K, et al. (2011) Investigation of toxic effects of the glyphosate on *Allium cepa*. Tarim Bilimleri Dergisi J Agr Sci 17: 131-142
62. Bajpai J, Srivastava A (2013) Evaluation of germination and seedling tolerance index of black gram genotypes in response to herbicide glyphosate. Ann Plant Sci 2: 351-357.
63. Zobiolo LHS, Kremer RJ, Constantin J (2012) Glyphosate effects on photosynthesis, nutrient accumulation, and nodulation in glyphosate-resistant soybean. J Plant Nutr Soil Sci 175: 319-330.
64. Rajashekhar N, Prakasha, Murthy TCS (2012) Seed germination and physiological behavior of maize cv. Nac-6002 seedlings under abiotic stress pendimethalin condition. Asian J Crop Sci 4: 80-85.
65. Alla MMN, Hassan NM, El-Bastawisy ZM (2008) Changes in antioxidants and kinetics of glutathione-S-transferase of maize in response to isoproturon treatment. Plant Biosyst 142: 5-16.
66. Huang H, Xiong ZT (2009) Toxic effects of cadmium, acetochlor and bensulfuron-methyl on nitrogen metabolism and plant growth in rice seedlings. Pestic Biochem Physiol 94: 64-67.
67. Mahakavi T, Bakiyaraj R, Baskaran L, Rashid N, Sanker GK (2014) Effect of herbicide quizalofop-p-ethyl on growth, photosynthetic pigments, enzymes and yield responses of black gram *Vigna mungo* L. Int Lett Natural Sci 9: 58-65.
68. Shehzad MA, Nadeem MA, Iqbal M (2012) Weed control and yield attributes against post-emergence herbicides application in wheat crop, Punjab, Pakistan. GARJAS 1: 7-16.
69. Ashraf M, Athar HR, Harris PJC, Kwon TR (2008) Some prospective strategies for improving crop salt tolerance. Adv Agron 97: 45-110.
70. Asgher M, Khan MIR, Anjum NA, Khan NA (2015) Minimising toxicity of cadmium in plants role of plant growth regulators. Protoplasma 252: 399-413.
71. Kon KF, Kotzian GR (2004) Herbicidal Composition. WIPO Patent WO/2004/008858A1. January 29, 2004.
72. Durmuş N, Bekircan T (2015) Pretreatment with polyamines alleviate the deleterious effects of diuron in maize leaves. Acta Biol Hung 66: 52-65.
73. Ding HD, Zhang XH, Xu SC, Sun LL, Jiang MY, et al. (2009) Induction of protection against paraquat-induced oxidative damage by abscisic acid in maize leaves is mediated through mitogen-activated protein kinase. J Integr Plant Biol 51: 961-972.
74. Hsu YT, Kao CH (2004) Phosphinothricin tolerance in rice *Oryza sativa* L. seedlings is associated with elevated abscisic acid in the leaves. Bot Bull Acad Sinica 45: 41-48.
75. Sakhautdinova AR, Fatkhutdinova DR, Bezrukova MV, Shakirova FM (2003) Salicylic acid prevents the damaging action of stress factors on wheat plants. Bulg J Plant Physiol 29: 314-319.
76. Ananieva EA, Alexieva VS, Popova LP (2002) Treatment with salicylic acid decreases the effects of paraquat on photosynthesis. J Plant Physiol 159: 685-693.
77. Shahrtash M, Mohsenzadeh S, Mohabatkar H (2011) Salicylic acid alleviates paraquat oxidative damage in maize seedling. Asian J Exp Biol Sci 2: 377-382.
78. Cui J, Zhang R, Wu GL, Zhu HM, Yang H (2010) Salicylic acid reduces napropamide toxicity by preventing its accumulation in rapeseed (*Brassica napus* L.). Arch Environ Contam Toxicol 59: 100-108.
79. Hung KT, Chang CJ, Kao CH (2002) Paraquat toxicity is reduced by nitric oxide in rice leaves. J Plant Physiol 159: 159-166.
80. Ferreira LC, Cataneo AC, Remaeh LMR, Bufalo J, Scavroni J, et al. (2011) Morphological and physiological alterations induced by lactofen in soybean leaves are reduced with nitric oxide. Planta Daninha 29: 837-847.
81. Groppa MD, Benavides MP (2008) Polyamines and abiotic stress: recent advances. Amino acids 34: 35-45.
82. Kasukabe Y, He L, Nada K, Misawa S, Ihara I, et al. (2004) Overexpression of spermidine synthase enhances tolerance to multiple environmental stresses and up-regulates the expression of various stress-regulated genes in transgenic *Arabidopsis thaliana*. Plant Cell Physiol 45: 712-722.
83. Kim HS, Jin CD (2006) Polyamines as antioxidant protectors against paraquat damage in radish *Raphanus sativus* L. cotyledons. J Plant Biol 49: 237-246.
84. Berova M (2001) Protective effect of the polyamine. J Environ Protec Ecol 2: 179-184.
85. Durmuş N, Kadioğlu A (2005) Reduction of paraquat toxicity in maize leaves by benzyladenine. Acta Biol Hung 56: 97-107.
86. Ndam LM, Enang JE, Mih AM, Egbe AE (2014) Weed diversity in maize *Zea mays* L. fields in South Western Cameroon. Int J Curr Microbiol App Sci 3: 173-180.
87. Chauhan BS, Johnson DE (2009) Ecological studies on *Cyperus difformis*, *Cyperus iria* and *Fimbristylis miliacea*: three troublesome annual sedge weeds of rice. Ann Appl Biol 155: 103-112.
88. Babu GP, Hooda V, Audiseshamma K, Paramageetham C (2014) Allelopathic effects of some weeds on germination and growth of *Vigna mungo* (L). Hepper. Int J Curr Microbiol App Sci 3: 122-128.
89. Tamang D, Nath R, Sengupta K (2015) Effect of herbicide application on weed management in green gram *Vigna radiata* L. Wilczek. Adv Crop Sci Tech 3: 2.
90. Chhokar RS, Balyan RS (1999) Competition and control of weeds in soybean. Weed Sci 47: 107-111.

91. Kahramanoglu I, Uygur FN (2010) The effects of reduced doses and application timing of metribuzin on redroot pigweed *Amaranthus retroflexus* L. and wild mustard *Sinapsis arvensis* L. Turk J Agric For 34: 467-474.
92. Ghosheh HZ (2004) Single herbicide treatments for control of broadleaved weeds in onion *Allium cepa*. Crop Prot 23: 539-542.
93. Kostrzevska MK, Jastrzębska M, Treder K, Wanic M (2014) Diversity of segetal weeds in pea (*Pisum sativum* L.) depending on crops chosen for a crop rotation system. Acta Agrobot 67: 55-66.
94. Wagner G, Nadasy E (2005) Effect of pre-emergence herbicides on growth parameters of green pea. Comm Agric Appl Biol Sci 71: 809-813.
95. Al-Khatib K, Kadir S, Libbey C (1995) Broadleaf weed control with clomazone in pickling cucumber (*Cucumis sativus*). Weed Technol 9: 166-172.