

Contaminants of Emerging Concern Detected By Comprehensive Target Analysis in a Sediment Core Collected From Osaka Bay, Japan

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ABSTRACT

The contaminants of emerging concern (CECs) have been one of the major concerns in recent. The objective of this study was to find potential CECs using an analytical screening method “comprehensive target analysis with an automated identification and quantification system (CTA-AIQS)” which uses gas and liquid chromatography combined with mass spectrometry (GC-MS and LC-MS). CTA-AIQS was applied to analyze sediment core samples collected from Osaka Bay, Japan, in 2019. Among the 74 chemicals were detected in sediment core samples, six sterols, PPCPs (fexofenadine, diphenhydramine, and clarithromycin), and tris(2-isopropylphenyl)phosphate isomers (TIPPPs) showed increasing profile from bottom to surface layer of the sediment core. Through the screening with chemical properties (persistence, bioaccumulation potential, and toxicity) of those chemicals, TIPPPs were categorized as potential CECs in the marine environment in Japan.

Keywords: Persistent Organic Pollutants (Pops), Contaminants Of Emerging Concern (Cecs), Automated Identification And Quantification System (Aiqs), Marine Sediment Core; Phosphate Flame Retardants (Pfrs), Tris(2-Isopropylphenyl)Phosphate Isomers (Tippps)

INTRODUCTION

The contaminants of emerging concern (CECs) have been one of the major concerns in recent. CECs have been defined as naturally occurring or anthropogenic chemicals that “have now been discovered or are suspected to be present in various environmental compartments and whose toxicity or persistence is likely to significantly alter the metabolism of a living being” [1]. The screening surveys of potential CECs have been conducted using the model-based technique among more than 100,000 chemicals in commercial use [2-6]. Those potential CECs were classified as various categories as; alkyl phenols; brominated flame retardants (BFRs); perfluorinated compounds; pharmaceuticals and personal care products (PPCPs); current-use pesticides including insecticides, fungicides, and herbicides; single-walled carbon nanotubes; and organophosphate flame retardants (PFRs) [7,8]. Most of these CECs are currently unregulated because their environmental levels, fates, and effects on organisms are poorly understood.

Although numerous numbers of potential CECs have been reported, few compounds have been analyzed as monitoring surveys. Because considerable numbers of chemicals of potential CECs are present in environmental media, and it is not realistic to develop the method of “target analysis” for individual potential CECs. For this reason, a number of research groups have attempted to detect CECs in the environment using the “comprehensive analysis” as screening method with gas and liquid chromatography coupled with high-resolution mass spectrometry (GC-HRMS and LC-HRMS) and the potential CECs have been detected by suspect screening [9-14] and nontarget screening [15-18].

In this study, Comprehensive Target Analysis with an Automated Identification and Quantification System (CTA-AIQS) was used as a tool to detect potential CECs in environment. CTA-AIQS is one of suspect screening technique that can be used to identify and semiquantify approximately 1500 chemicals in environmental media [19].

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19]. CTA-AIQS has been applied to assess water and sediment quality in various countries [20-23]. In addition, CTA-AIQS has been used as the first pollution screening scheme on the scene of natural disasters such as earthquakes in Japan [24,25].

In this study, CTA-AIQS was used to analyze sediment core samples for screening the potential CECs in Japanese coastal environment. The vertical profile of chemical concentrations in sediment core samples is expected to be useful for estimating the historical trends of chemical loads in marine environments. The chemical concentrations in sediment core sample showed increasing trend from bottom to surface may indicate the increasing trend of environmental loads of those chemicals. Our research group conducted to find potential CECs by analyzing sediment core samples collected in Beppu Bay, Japan using CTA-AIQS. The results indicated that three polycyclic aromatic hydrocarbons (PAHs; anthracene, chrysene, and fluoranthene) and tris(2-isopropylphenyl)phosphate isomers (TIPPPs) could be categorized as potential CECs in marine environments in Japan [26]. To verify this result, this study planned to analyze sediment core samples collected in Osaka Bay, which receives large volumes of sewage and industrial wastewater flows, because coastal area of Osaka Bay is one of the biggest economic and industrial regions in Japan. Environmental surveys of dioxin-related chemicals and heavy metals using sediment core samples in Osaka Bay have been conducted [27,28]; however, sediment core samples have not previously been analyzed by comprehensive screening to find potential CECs.

The objective of this study was to find chemicals that might be classified as potential CECs by assessing the historical trends of chemical concentrations in sediment core samples collected from Osaka Bay, Japan, by means of CTA-AIQS.

MATERIALS AND METHODS

Sample Collection and Sediment Core Dating

Osaka Bay is located on the eastern part of the Seto Inland Sea, central Japan. The sea has a seawater volume of 41 km³, an area of 1,450 km², an average depth of 28 m, and a maximum depth of 197 m. The Yodo, Yamato, and Muko rivers are the main inflowing rivers, with a total estimated flow of 13.1 km³ yr⁻¹. Osaka Bay is bounded by Osaka and Hyogo prefectures, which contain the largest metropolitan areas in Japan, with populations of 9 million and 5 million people, respectively. Osaka and Kobe are some of the largest international ports in Japan, and a major industrial area is located along the coast. The bay is surrounded by the islands of Honshu and Awaji and is classified as an enclosed coastal sea because it has only two narrow mouths. Water exchange in the bay is therefore limited, and there is potential for accumulation of toxic pollutants associated with human activity. Two sediment cores (OS2-1, core length 84 cm; OS2-2, core length 78 cm) were obtained in Osaka Bay (OS2; 34°35'30.96" N, 135°16'7.8" E; Figure S1) from the research vessel *Kaikomaru* by an underwater diver using an acrylic pipe (inner diameter = 10 cm) in December 2019. The cores were collected without disturbing the sediment-water interface, held horizontally during transport to a laboratory, and cut into 2-cm layers. The layers were stored at -20 °C until analysis. To obtain samples that were large enough for both comprehensive and target analysis, we combined the sectioned layers to yield eight samples, from depths of 0-10, 10-20, 20-30, 30-40, 40-50, 50-60, 60-70, and 70-78 cm. These samples were

dried under flowing nitrogen for several days at room temperature and then stored at -20 °C prior to chemical analysis.

Sediment chronology analysis was performed for the OS2-1 core based on the constant rate of supply (CRS) method of ²¹⁰Pb dating [29] and verified using the ¹³⁷Cs peak for the year 1963 [30]. Details of the CRS-based chronological method are provided in the Supplementary Information (Figure S2). Dried samples were sealed in holders for a month to allow ²²²Rn and its short-lived decay product (²¹⁴Pb) to equilibrate. The activity of supported ²¹⁰Pb was estimated by measuring the activity of ²¹⁴Pb, whereas that of excess ²¹⁰Pb was determined from the difference between the total and the supported ²¹⁰Pb (²¹⁰Pb excess = ²¹⁰Pb total - ²¹⁴Pb). The ¹³⁷Cs, ²¹⁰Pb, and ²¹⁴Pb activities were determined by gamma counting using a germanium detector (GCW-3523 CANBERRA, Mirion Technologies, Inc., USA) equipped with a multi-channel analyzer at Idea Co. Ltd., Osaka. The age of a given sample mass depth was calculated using the ²¹⁰Pb excess inventory, which was obtained by numerical integration of the radioactivity of ²¹⁰Pb excess versus the mass depth profile [30]. The dating of core OS2-2 was estimated indirectly based on stratigraphic correlations of the depth profiles of magnetic susceptibility between cores OS2-1 and OS2-2. For the correlation, the marked peaks in the profiles were used as time-reference horizons. Samples from core OS2-2 were dated by linear interpolation from the depths and ages of two time-reference horizons. Magnetic susceptibility was measured using an SM-30 meter (ZH instruments, Brno, Czech Republic).

Sample Preparation for CTA-AIQS

For CTA-AIQS (GC), samples were prepared according to the method previously reported with slight modifications [31,32,24,26]. A mixture of surrogate standard added prior to extraction and mixture of internal standard added before injection were in Table S1. For CTA-AIQS (LC), samples were prepared according to the previously reported method [33-35]. A mixture of surrogate standard added prior to extraction and mixture of internal standard added before injection were in Table S2. The moisture content of sediment samples was calculated by oven-drying an aliquot of the sample at 130 °C overnight.

Instrument for CTA-AIQS

The organic micropollutants in the samples were identified and semiquantified by means of the AIQS scheme using a private database composed of retention times, mass spectra, and calibration curves. The database for AIQS contained 970 chemicals for GC (Table S3) and 501 organic micropollutants for LC-MS (Table S4). The GC-MS conditions (Table S5) have been reported previously [36]. The LC-MS measurement conditions (Table S6) have previously been reported [34,26].

Targeted Analysis of PFRs

Targeted analysis of PFRs was conducted by the method previously reported with slight modifications [37]. Sixteen monomeric PFRs were selected as target compounds (abbreviations are explained in Table S7). Detailed information about the multiple-reaction-monitoring (MRM) transitions is given in Table S7.

Quality Control

For CTA-AIQS, recovery tests for the entire procedure (both GC and LC) have previously been carried out for chemicals with a wide range of structures, functional groups, boiling points, and

other physicochemical properties. For CTA-AIQS (GC), model compounds spiked prior to extraction showed recoveries of 69%–110% in water samples [38,39] and sediment samples [31,32]. When samples were spiked with surrogate standards prior to extraction, recoveries ranged from 69% to 115%, and the relative standard deviation (RSD) was less than 20% [40]. Recovery tests for CTA-AIQS (LC) have also been conducted for water samples and for soil samples [33–35]. When soil samples were spiked with model compounds prior to extraction, recoveries ranged from 76% to 125%. In another study, mean recoveries of surrogate standards in influent and effluent water samples collected from a sewage treatment plant were 77% and 85% (with RSDs < 23%) [34]. Method detection limits (MDLs) for CTA-AIQS (GC and LC) were estimated from instrument detection limits in the AIQS database on the basis of a 2.5-g dry sediment sample and an injection volume of 1 μL (Tables S3 and S4). Because the standard deviations of the procedural blanks for several of the chemicals (e.g., phthalate-related chemicals) were high, these chemicals were not quantified in this study. CTA-AIQS (GC) was validated by analysis of a certified reference material (NIST 1941b), and the values obtained were 50%–150% of the reference values (average 78%, RSD < 20% for each chemical: Table S8).

For target analysis, samples were analyzed in accordance with the established laboratory quality assurance and quality control procedures. Calibration curves obtained from the LC-ESI-MS/MS data showed good linearity ($R > 0.99$). Average recoveries of internal standards for all samples ranged from 61% to 75%. The instrumental detection limits of the analytes were calculated from signal-to-noise ratios determined from the LC-ESI-MS/MS chromatographs. The MDLs for the analytes in all the samples are summarized in Table S8. The concentrations of analytes in the procedural blank were below the MDL values.

Statistical Analysis

The concentrations of chemicals in soil samples are expressed in nanograms per gram in dry weight. For calculation of total concentrations, values less than the MDL were considered to be zero. The Wilcoxon rank sum test was applied to evaluate group differences with the JMP software suite (ver. 12, SAS Institute Inc.). In this study, a p value of less than 0.05 was considered to indicate statistical significance.

RESULTS & DISCUSSION

Chronology in Osaka Bay

The results of ^{210}Pb radioactivity for the chronological OS2-1 core (Figure S3) demonstrated that ^{210}Pb radioactivity in the core declined non-exponentially with depth, implying that sediment accumulation rates are not constant and that the CRS model was the most appropriate for determining sediment age [29]. The CRS-based ^{210}Pb chronologies indicated that the calendar year corresponding to a core depth of 53 cm (layer at 52–54 cm depth) was 1927 ± 17 AD (Figure S4). The calendar age errors in core OS2-1 were estimated to be <2 yr after 1974 and <9 yr after 1944. The slope of the excess ^{210}Pb plot was steeper in the levels below 17 cm than in the higher levels (Figure S5). There was a minor ^{137}Cs pulse at 37–43 cm and large fluctuations in ^{137}Cs between the detection limit and 0.002 Bq g^{-1} below 47 cm depth (Figure S3). These variations likely resulted from heterogeneous vertical mixing of sediments by macrobenthos. A minor peak of ^{137}Cs was

measured at 55 cm (Figure S3), which probably formed as a result of downward transport of sediments with high ^{137}Cs values by macrobenthos. The minor pulse in ^{137}Cs at 37–43 cm is most likely associated with the bomb-derived fall-out maximum in Japan in 1963 [30]. However, the depth corresponding to 1964 based on the CRS age–depth model was 34 cm (Figure S4), 3–9 cm higher than the ^{137}Cs minor pulse. The occurrence of moderate values above the pulse suggests steady-state rapid mixing in the surface layer [41]. Theoretical mixing modeling demonstrated that steady-state rapid mixing in the surface layer redistributes bomb-derived ^{137}Cs both upward and downward, resulting in an alteration of the original vertical profile with a sharp ^{137}Cs pulse toward 3 cm depth and a minor ^{137}Cs pulse together with moderate values in the upper layer [41]. A strong steady-state mixing experiment (surface 6-cm layer) did not yield a sharp ^{137}Cs pulse; instead, it exhibited constant ^{137}Cs values in the upper 7 cm and a rapid decrease in values below that depth. In contrast, in a moderate mixing experiment (surface 3 cm layer) the sharp ^{137}Cs pulse broadened and shifted 2.5 cm downward, and the surface values increased relative to the original vertical profile. Our observed depth profile of ^{137}Cs was consistent with that of a mixing experiment for the surface 3 cm of sediment. Therefore, steady-state mixing of the 3-cm surface layer may produce moderate values of ^{137}Cs in the upper layer together with a 3-cm downward shift of the ^{137}Cs pulse and yield constant excess ^{210}Pb values in the upper 6 or 7 cm of sediment. This finding indicates that the approximately-half thickness of an upper layer with a constant excess ^{210}Pb value corresponds to the thickness of the steady-state mixing and the depth of the downward shift of the ^{137}Cs pulse. Our data show a vertical profile pattern similar to the latter, suggesting moderate surface steady-state mixing at this site. Given the observed constant excess ^{210}Pb values in the upper 12 cm (Figure S5) and the minor ^{137}Cs pulse at 37–43 cm (Figure S3), steady-state mixing probably occurred in the uppermost 6 cm, and the original ^{137}Cs pulse might have been located at 31–37 cm, consistent with the CRS age–depth model (Figure S4).

Profiles of magnetic susceptibilities and pigments (Figure S6) contained four time-reference horizons (Table S9), which were considered to represent contemporary environmental or biological events in the study site of Osaka Bay. We estimated the ages of core OS2-2 based on these four reference layers and the core top (0 cm, 2019.9 AD). The estimated ages for core OS2-2 were 1908.9, 1935.8, 1953.5, and 1993.1 AD at core depths of 63, 47, 37, and 17 cm, respectively.

Comprehensive Analysis with CTA-AIQS

In this study, CTA-AIQS was used for comprehensive screening to detect CECs in samples from a sediment core (OS2-1) collected in 2019 from Osaka Bay, Japan. Among 74 compounds detected, 70 compounds were found with CTA-AIQS (GC), and 4 compounds were detected by CTA-AIQS (LC) (Figure 1 and Table S10). The median total concentration (range, minimum–maximum) of chemicals detected in sediment core samples was $8100 (1000\text{--}24000) \text{ ng g}^{-1}$ dry weight. The detected chemicals in sediment core samples were classified into four categories (Figure 1 and Table S10): sterols, alkanes, PAHs, and miscellaneous chemicals (including POPs, PPCPs, and industry).

Overview of Vertical Profiles

The total concentration of chemicals detected in sediment samples by CTA-AIQS was lowest in the bottom layers and gradually

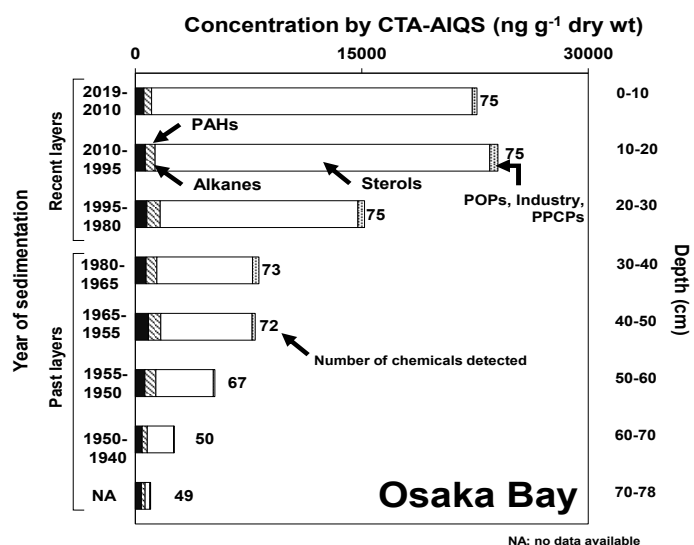


Figure 1: Vertical profiles of chemical concentrations detected by CTA-AIQS (GC and LC) in samples from a sediment core collected from Osaka Bay, Japan in 2019

increased toward the surface layers (Figure 1). In the surface layer, sterols were the predominant chemicals (94%), followed by alkanes (2.5%), PAHs (2.3%), and miscellaneous chemicals (1.4%).

As the first step to discuss the vertical profile of chemical concentrations, POPs must be the important chemical marker to understand appropriateness. The concentrations of POPs, which are known to be CECs, e.g., PCBs, DDTs, and related chemicals, were relatively low compared to those of other types of chemical in this study (Table S10). The vertical profile of POPs in Osaka Bay showed lower concentrations in bottom and surface layers and the highest concentrations in the layers representing the 1930s–1950s (Figure S7). Previous studies of sediment core samples collected from various places in Japan, such as Tokyo Bay, Osaka Bay, Lake Biwa, and Beppu Bay, reported that the highest concentrations of POPs were detected in the layers deposited around the 1970s, because large quantities of these chemicals were used in Japan during the 1960s and 1970s [42-46]. Although concentration profile of POPs in this study seems reasonable, discordance of the sedimentation year on the highest concentration was observed. As discussed above, sediment disturbance by biological and anthropogenic activity might be the reason for this discordance in this study.

Sterols were the chemical category with the highest concentration in this study (Figure 1). Six sterols (cholesterol, cholestanol, campesterol, stigmasterol, beta-sitosterol, and stigmastanol) were detected in all layers (Table S10). Almost the same sterols were detected in marine surface sediments collected from Dokai Bay, Tokyo Bay, and Beppu Bay, Japan [31,47]. The sterol concentrations in this study are comparable to those in Dokai Bay and Tokyo Bay, but 10 times higher than those in Beppu Bay. Sterols and related chemicals in marine and lake sediments can be derived from sewage effluent, but also from terrestrial plants, phytoplankton, and animal feces [48-50]. The higher concentrations of sterols in Osaka Bay than in Beppu Bay are consistent with the large amount of sewage effluent loading from urban areas. The vertical profile of sterol concentrations in this study showed gradually increasing from bottom to surface (Figure S8). A similar increasing trend of sterol concentrations was reported in sediment core samples collected in Beppu Bay. These results suggest a historical increase in sewage effluent loading from urban areas surrounding Osaka Bay. Additionally, degradation of sterols in the deeper layers due

to their short half-lives (e.g., cholesterol < 40 days in sediment) might be another reason for the decline in sterol concentrations with depth [51].

Alkanes exhibited the second highest concentration in the sediment core samples in this study (Figure 1). The 25 alkanes (C9–C33) were detected in all layers (Figure S9). The essentially uniform alkane concentrations in all layers yielded a consistent vertical profile. Because alkanes can be derived from various sources, several molecular markers are often used to estimate their origins. The carbon preference index (CPI), which is the ratio of odd-to-even carbon number *n*-alkanes, was applied to differentiate petrogenic and biogenic alkanes. A CPI value of less than 2 indicates a petrogenic (oil) source; a value greater than 2 indicates a biogenic source [52-55]. The CPI range in Osaka Bay was 1.9–2.5, with a value of less than 2 for most levels (Figure S10). This result suggests that the alkanes detected in Osaka Bay could have been derived from petrogenic sources, which, because Osaka Bay is surrounded by urban and industrialized areas, seems reasonable. The uniform vertical profiles in Osaka Bay indicate continuous input of alkanes from petrogenic sources.

PAHs were detected in all layers of the sediment core samples, and 22 species were identified (16 native PAHs and 6 methylated PAHs; Table S10). The vertical profiles of PAHs concentrations were relatively uniform in all layers (Figure S11). For estimation of the major sources of PAHs, the ratio of the concentration of methylated phenanthrenes to those of phenanthrene (Σ MPhen/Phen) was applied. A Σ MPhen/Phen value of more than 1 indicates a petrogenic (oil) origin; a value less than 1 suggests a pyrogenic (combustion) source [56]. The average Σ MPhen/Phen ratios in the sediment core sample in this study were approximately 1, with a value of more than 1 for several layers (Figure S12). This result suggests that the PAHs in Osaka Bay are derived mainly from pyrogenic sources but partly from petrogenic sources. Half lives. This conclusion is plausible because Osaka Bay is a major industrial and urbanized area in Japan, and the result for PAHs is consistent with that for alkanes.

The concentrations of Miscellaneous (PPCP, and industry) were relatively low compared to those of other types of chemicals in Osaka Bay (Figure 1). Concentrations of PPCPs (including clarithromycin, fexofenadine, and diphenhydramine) and industrial chemicals (including TIPPPs and 4-methyl-2,6-di-*t*-butylphenol) were gradually increased from the bottom to the surface of the sediment profile (Figures S13 and S14). Although no research has been conducted on these chemicals in sediment core samples, this result might suggest a recent increase in the input these substances into the marine environment from sewage effluent. TIPPPs were also reported to exhibit an increasing trend in sediment core samples collected from Beppu Bay [26]. Detailed profiles of individual chemicals are discussed in the next section.

Screening of Potential CECs

In this study, 74 compounds were identified and semiquantified in sediment core samples using CTA-AIQS (GC and LC). Among these chemicals, screening was conducted to select potential CECs following a three-step protocol.

In step 1, sediment core layers were divided into two groups, “past layers” and “recent layers”, to compare the chemical concentrations of the two groups. The deeper five layers (<1920–1970) were defined as “past layers”, which included the high consumption of POPs around the 1970s in Japan. The shallower three layers

(1970–2019) were defined as “recent layers”, which included the generation of post-POPs (Figure 1). In step 2, chemicals exhibiting significantly higher concentrations in recent layers were statistically selected as candidates ($p < 0.05$, Wilcoxon’s rank sum test). In step 3, among the candidate chemicals, the properties of Persistence, Bioaccumulation, and Toxicity (PBT) criteria were used to select potential CESs. The PBT criteria has been applied to identify chemicals with a high impact on the environment, with high persistence, bioaccumulation, and toxicity: half-life in soil/sediment > 180 days, bioconcentration factor (BCF) > 2000 , and toxicity (NOEC) < 0.01 mg L⁻¹ [58], respectively.

Through the steps 1 and 2, 11 compounds were selected (Table 1): 2,6-dimethylnaphthalene, six sterols (cholesterols, beta-sitosterol, stigmasterol, campesterol, cholestanol, and stigmastanol), TIPPPs, and three PPCPs (fexofenadine, diphenhydramine, and clarithromycin). The vertical profiles of one representative chemical for each category are shown in Figure 2. For the selected chemicals, screening by using the PBT criteria (step 3) was conducted (Table 1). For sterols, no reliable data on PBT criteria were found, although their half-lives and toxicity for laboratory animals were shorter and lower than the criteria [59]. Assessment surveys of dimethylnaphthalene reported that its half-life in sediment/water, NOEC, and BCF on aquatic ecosystems were < 14 days, 0.19 mg L⁻¹, and 700 – 1200 , respectively [60,61]. According to our assessment, cholesterols and PAHs did not meet the PBT criteria (Table 1). Assessment surveys of clarithromycin reported that the BCF calculated by the EPA/BCFWIN system and the NOEC for *Daphnia magna* were 56 [62] and > 2100 µg L⁻¹ [63], respectively, but no report was found for environmental half-life. For fexofenadine, a BCF of 43.9 for *Daphnia pulex* [64] and an NOEC on aquatic ecosystems of > 330 – 570 mg L⁻¹ [65] have been reported, but we could find no information on the environmental half-life. A BCF of 4.2 – 53.3 for fathead minnows [66] and a 10-day NOEC for *Daphnia magna* of 0.00294 mg L⁻¹ [67] have been measured for diphenhydramine, but there is no information on its environmental half-life. According to these assessments, these PPCPs do not satisfy the PBT criteria (Table 1); however, because information for these chemicals is limited, further investigations of their environmental behavior and PBT properties are required. The remaining compound in Table 1 was TIPPPs. The reported

half-life in sediment, BCF, and chronic NOEC for *Daphnia magna* of TIPPPs are > 180 days [68], > 2000 [69], and < 0.01 mg L⁻¹ [70], respectively. These results indicate that TIPPPs meet the PBT criteria [68]; thus, on the basis of our three-step screening protocol, TIPPPs can be identified as potential CECs (Table 1).

TIPPPs as Potential CECs

By applying the protocol described above, TIPPPs can be categorized as potential CECs; however, there are limitations to the sensitivity and accuracy of CTA-AIQS as an analytical method. To confirm the existence of TIPPPs with higher sensitivity and accuracy, target analysis was conducted for PFRs by using LC-MS/MS on the same sediment core samples in this study. From this analysis, several PFRs (EHDPP, TDMPP, TIPPPs, and TEHP) were detected. The concentration of TIPPPs ranged from $< \text{MDL}$ to 2.2 ng g⁻¹ dry (Table S11). Although the range of TIPPPs concentrations in Osaka Bay was lower than those in Beppu Bay [26], the vertical profile of TIPPPs in Osaka Bay exhibited increasing trend from bottom to surface which was similar to Beppu Bay (Table S11). These results indicate a ubiquitous distribution and increasing load of TIPPPs in coastal environments of Japan. However, halogenated PFRs (e.g., TCEP and TDCPP) were not detected in sediment core samples from Osaka Bay in this study and also from Beppu Bay. Halogenated PFRs can become concentrated as CECs in samples of water and organisms, and they were one of the most abundant CECs analyzed in previous studies [71,72]. Because halogenated PFRs have relatively higher water solubility than other PFRs, these chemicals do not accumulate in sediment.

In general, increasing trend of PFR consumption has been reported since the use of BFRs (tetra to heptabromodiphenyl ether formulations in 2009, hexabromocyclododecane in 2013, and decabromodiphenyl ether in 2017) was restricted by the Stockholm Convention [73-75]. Since penta-BDE formulations were phased out, TIPPPs contained in Firemaster 550 and ITP organophosphate aryl ester technical mixtures have increasingly been used to treat polyurethane foam in residential upholstered furniture [76]. The environmental monitoring data for TIPPPs are limited. However, TIPPPs were detected in air [77], house dust [78,79], sediment, and bivalves.

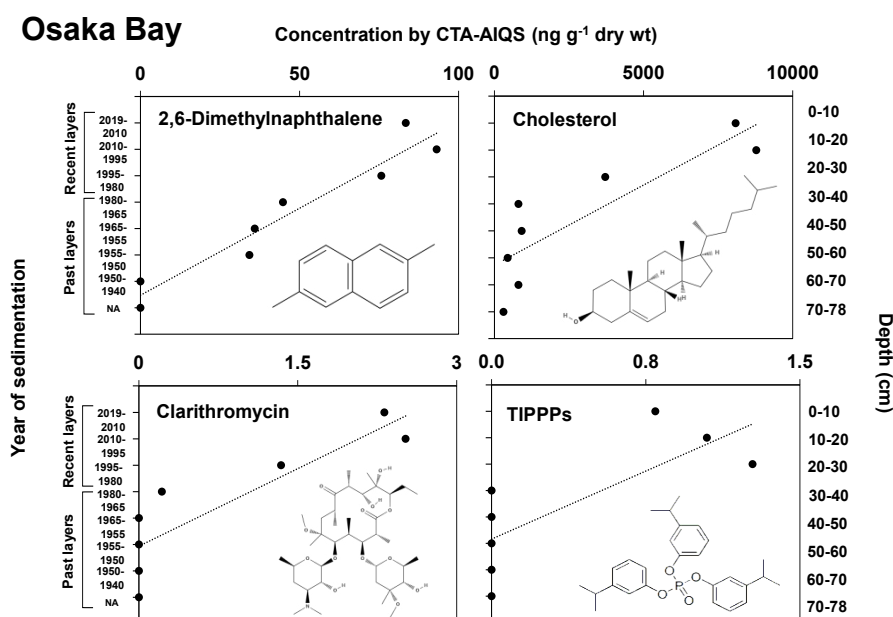


Figure 2: Vertical profiles of representative micropollutants concentrations of which increased with decreasing sediment depth, as detected by CTA-AIQS (GC and LC), in samples from a sediment core collected from Osaka Bay, Japan, in 2019.

Table 1: Screening of potential CECs among the chemicals detected by CTA-AIQS in sediment core sample collected in Osaka Bay, Japan in 2019

Compounds	Increasing trend	Persistence*	Bioaccumulation	Toxicity	Potential of CECs	Reference
PAHs						
2,6-Dimethylnaphthalene	✓	x	x	x		a), b)
Sterol						
Cholesterols	✓	x	x	x		j), k)
Beta-sitosterol	✓	na	na	na		
Stigmasterol	✓	na	na	na		
Campesterol	✓	na	na	na		
Cholestanol	✓	na	na	na		
Stigmastanol	✓	na	na	na		
Industry						
Tris(2-isopropylphenyl)phosphate (TIPPPs)	✓	✓	✓	✓	✓	c), l), m)
PPCPs						
Fexofenadine	✓	na	x	x		d), e)
Diphenhydramine	✓	na	x	✓		f), g)
Clarithromycin	✓	na	x	x		h), i)

*: Significant higher concentrations found in recent layers (1970-2019) (Wilcoxon rank sum test)

✓: Meet the criteria

x: Not meet the PBT criteria

na: No data available

a) RIVM, 2018

b) Neff, 2002

c) Environment Agency UK, 2009

d) FDA, 2010

e) Sundelin, 2015

f) Kristofco et al., 2015

g) Nichols et al., 2015.

h) Baumann et al., 2015.

i) Oguz and Mihciokur, 2014.

j) Christian 1986

k) Sun and Wakeham 1998

l) Cleveland et al. 1986

m) Sanders et al. 1985

In addition, the POP-like characteristics of TIPPPs (overall persistence and [80,81] long-range transport potential) were evaluated by using the OECD Pov and LRTP Screening Tool [82,83]. respectively). Vertical and horizontal lines denote POP criteria for P_{ov} and CTD or TE, respectively. (P_{ov} (days) is a measure of the time scale of degradation of the chemical in the whole environment. CTD (km) and TE (%) represents a transport-oriented metric and a target-oriented indicator, respectively. Details of these parameters are described in an earlier paper. The tool requires the three degradation half-lives in air, water, and soil, and also two partition coefficients such as 1-octanol/water partition coefficient (K_{ow}) and air/water partition coefficient (K_{aw}) for the chemical of interest. The half-lives were estimated using EPI-Suite ver. 4.11 (<https://www.epa.gov/tsca-screening-tools/epi-suitetm-estimation-program-interface>). The log K_{ow} and log K_{aw} values were estimated using EPI-suite ver. 4.11 and UFZ-LSER Database [84]. Since TIPPPs might have POPs potential (persistence and long-range transport), those should be categorized as potential CECs [85,86].

CONCLUSION

Comprehensive analytical method of CTA-AIQS (which combines

GC and LC with MS) was applied to detect CECs from a sediment core collected in Osaka Bay, Japan, in 2019. Among the chemicals detected, TIPPPs concentrations showed increasing trend from bottom to top layer among the core samples. Through the screening processes, TIPPPs were categorized as potential CECs in marine environments in Japan.

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Conflicts of Interest/Competing Interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

REFERENCES

- Sauvé S, Desrosiers M. A review of what is an emerging contaminant. *Chemi Central J.* 2014;1:7.
- Muir DC, Howard PH. Are there other persistent organic pollutants? A challenge for environmental chemists. *Environmental science & technology.* 2006;40(23):7157-7166.
- Brown TN, Wania F. Screening chemicals for the potential to be persistent organic pollutants: A case study of Arctic contaminants. *Environ Sci Technol.* 2008;42(14):5202-5209.
- Howard PH, Muir DCG. Identifying new persistent and bioaccumulative organics among chemicals in commerce. *Environ Sci Technol.* 2010;44:2277-2285.
- Howard PH, Muir DC. Identifying new persistent and bioaccumulative organics among chemicals in commerce II: pharmaceuticals. *Environ Sci Technol.* 2011;45(16):6938-6946.
- Öberg T, Iqbal MS. The chemical and environmental property space of REACH chemicals. *Chemosphere.* 2012;87(8):975-981.
- Anderson PD, Denslow ND, Drewes JE, Olivieri AW, Schlenk D, Scott GI, et al. Monitoring strategies for chemicals of emerging concern (CECs) in California's aquatic ecosystems—recommendations of a science advisory panel. Costa Mesa, CA: Southern California Coastal Water Research Project. 2012:219.
- Dodder NG, Maruya KA, Ferguson PL, Grace R, Klosterhaus S, La Guardia MJ, et al. Occurrence of contaminants of emerging concern in mussels (*Mytilus* spp.) along the California coast and the influence of land use, storm water discharge, and treated wastewater effluent. *Marine pollution bulletin.* 2014 Apr 30;81(2):340-346.
- Jobst KJ, Shen L, Reiner EJ, Taguchi VY, Helm PA, McCrindle R, et al. The use of mass defect plots for the identification of (novel) halogenated contaminants in the environment. *Anal Bioanal Chem.* 2013;405(10):3289-3297.
- Cariou R, Omer E, Léon A, Dervilly-Pinel G, Le Bizet B. Screening halogenated environmental contaminants in biota based on isotopic pattern and mass defect provided by high resolution mass spectrometry profiling. *Anal chimica acta.* 2016;936:130-138.
- Yau A, Favela KA, Isaacs KK, McEachran A, Grulke C, Richard AM, et al. Suspect screening analysis of chemicals in consumer products. *Environmental science & technology.* 2018 Feb 6;52(5):3125-3135.
- Léon A, Cariou R, Hutinet S, Hurel J, Guitton Y, Tixier C, et al. HaloSeeker 1.0: a user-friendly software to highlight halogenated chemicals in nontargeted high-resolution mass spectrometry data sets. *Analytical chemistry.* 2019;91(5):3500-3507.
- Rostkowski P, Haglund P, Aalizadeh R, Alygizakis N, Thomaidis N, Arandes JB, et al. The strength in numbers: comprehensive characterization of house dust using complementary mass spectrometric techniques. *Anal bioanal chem.* 2019;411(10):1957-1977.
- Creusot N, Casado-Martinez C, Chiaia-Hernandez A, Kiefer K, Ferrari BJ, Fu Q, et al. Retrospective screening of high-resolution mass spectrometry archived digital samples can improve environmental risk assessment of emerging contaminants: A case study on antifungal azoles. *Environ Int.* 2020;139:10570-10578.
- Hoh E, Dodder NG, Lehotay SJ, Pangallo KC, Reddy CM, Maruya KA. Nontargeted comprehensive two-dimensional gas chromatography/time-of-flight mass spectrometry method and software for inventorying persistent and bioaccumulative contaminants in marine environments. *Environ Sci Technol.* 2012;46(15):8001-8008.
- Ionas AC, Ballesteros Gómez A, Leonards PE, Covaci A. Identification strategies for flame retardants employing time-of-flight mass spectrometric detectors along with spectral and spectra-less databases. *J Mass Spectro.* 2015 Aug;50(8):1031-1038.
- Schymanski EL, Singer HP, Slobodnik J, Ipolyi IM, Oswald P, Krauss M, et al. Non-target screening with high-resolution mass spectrometry: critical review using a collaborative trial on water analysis. *Anal Bioanal Chem.* 2015;407(21):6237-6255.
- Hollender J, Schymanski EL, Singer HP, Ferguson PL. Nontarget screening with high resolution mass spectrometry in the environment: ready to go? *Environ Sci Technol.* 2017;51:11505-11512.
- Kadokami K, Tanada K, Taneda K, Nakagawa K. Novel gas chromatography-mass spectrometry database for automatic identification and quantification of micropollutants. *J Chromatography A.* 2005;1089(1-2):219-226.
- Allinson G, Allinson M, Kadokami K. Combining passive sampling with a gc-MS-database screening tool to assess trace organic contamination of rivers: a pilot study in Melbourne, Australia. *Water, Air, & Soil Pollution.* 2015;226(7):1-1.
- Li X, Shang X, Luo T, Du X, Wang Y, Xie Q, et al. Screening and health risk of organic micropollutants in rural groundwater of Liaodong Peninsula, China. *Environmental Pollution.* 2016;218:739-748.
- Chau HT, Kadokami K, Ifuku T, Yoshida Y. Development of a comprehensive screening method for more than 300 organic chemicals in water samples using a combination of solid-phase extraction and liquid chromatography-time-of-flight-mass spectrometry. *Environmental Science and Pollution Research.* 2017;24(34):26396-26409.
- Hai DM, Qiu X, Xu H, Honda M, Yabe M, Kadokami K, et al. Contaminants in Liquid Organic Fertilizers Used for Agriculture in Japan. *Bulletin of environmental contamination and toxicology.* 2017;99(1):131-137.
- Matsuo Y, Miyawaki T, Kadokami K, Nakai K, Tatsuta N, Nakata H, et al. Development of a novel scheme for rapid screening for environmental micropollutants in emergency situations (REPE) and its application for comprehensive analysis of tsunami sediments deposited by the great east Japan earthquake. *Chemosphere.* 2019;224:39-47.
- Matsuo Y, Nakata H, Agusa T, Miyawaki T, Kadokami K, Sato K, et al. Comprehensive target analysis of micropollutants in soil at debris storage sites of the Kumamoto earthquake. *Soil and Sediment Contamination: An International Journal.* 2020;29(4):452-463.
- Nishimuta K, Ueno D, Takahashi S, Kuwae M, Kadokami K, Miyawaki T, et al. Use of comprehensive target analysis for determination of contaminants of emerging concern in a sediment core collected from Beppu Bay, Japan. *Environmental Pollution.* 2021;272:115-587.

27. Sakai S, Deguchi S, Takatsuki H. Comparative study on time trends and sedimentation flux of PCDDs/DFs and PCBs in sediment cores from Japan. *Environ Sci.* 2002;15:361-376.
28. Hosono T, Su CC, Okamura K, Taniguchi M. Historical record of heavy metal pollution deduced by lead isotope ratios in core sediments from the Osaka Bay, Japan. *J Geochemical Exploration.* 2010;107(1):1-8.
29. Appleby PG, Oldfield F. The calculation of lead-210 dates assuming a constant rate of supply of unsupported 210Pb to the sediment. *Catena.* 1978;5(1):1-8.
30. Appleby PG. Chronostratigraphic techniques in recent sediments. In *Tracking environmental change using lake sediments.* Springer. 2002:171-203.
31. Kadokami K, Li X, Pan S, Ueda N, Hamada K, Jinya D, Iwamura T. Screening analysis of hundreds of sediment pollutants and evaluation of their effects on benthic organisms in Dokai Bay, Japan. *Chemosphere.* 2013;90(2):721-728.
32. Miyawaki T, Tobiishi K, Takenaka S, Kadokami K. A rapid method, combining microwave-assisted extraction and gas chromatography-mass spectrometry with a database, for determining organochlorine pesticides and polycyclic aromatic hydrocarbons in soils and sediments. *Soil and Sediment Contamination: An Int J.* 2018;27(1):31-45.
33. Miyawaki T, Tobiishi K, Takenaka S, Kadokami K. Development of rapid screening method of organic pollutants in soil by LC/MS: study of extraction method and the optimization of the conditions. *Bunseki Kagaku= J Japanese Society for Anal Chem.* 2015;64(7):533-541.
34. Kadokami K, Ueno D. Comprehensive target analysis for 484 organic micropollutants in environmental waters by the combination of tandem solid-phase extraction and quadrupole time-of-flight mass spectrometry with sequential window acquisition of all theoretical fragment-ion spectra acquisition. *Anal Chem.* 2019;91(12):7749-7755.
35. Miyawaki T, Tobiishi K, Takenaka S, Kadokami K. Development of rapid screening method of organic pollutants in soil by LC/MS: study of extraction method and the optimization of the conditions. *Bunseki Kagaku= J Japanese Soci Anal Chem.* 2015;64(7):533-541.
36. Duong HT, Kadokami K, Trinh HT, Phan TQ, Le GT, Nguyen DT, et al. Target screening analysis of 970 semi-volatile organic compounds adsorbed on atmospheric particulate matter in Hanoi, Vietnam. *Chemosphere.* 2019;219:784-795.
37. Matsukami H, Suzuki G, Someya M, Uchida N, Tue NM, Viet PH, et al. Concentrations of polybrominated diphenyl ethers and alternative flame retardants in surface soils and river sediments from an electronic waste-processing area in northern Vietnam, 2012–2014. *Chemosphere.* 2017;167:291-299.
38. Jinya D, Iwamura T, Kadokami K, Miyagawa H, Nakagawa K, Kondo T, et al. System Performance Check Standards for Gas Chromatography Quadrupole Mass Spectrometry in Simultaneous Analysis of Semi-Volatile Compounds. *Bunseki Kagaku/Japan Analyst.* 2011;60(12).
39. Jinya D, Iwamura T, Kadokami K. Comprehensive analytical method for semi-volatile organic compounds in water samples by combination of disk-type solid-phase extraction and gas chromatography-mass spectrometry database system. *Anal Sci.* 2013 Apr 10;29(4):483-486.
40. Duong HT, Kadokami K, Chau HT, Nguyen TQ, Nguyen TT, Kong L. Groundwater screening for 940 organic micro-pollutants in Hanoi and Ho Chi Minh City, Vietnam. *Environ Sci Poll Res.* 2015;22(24):19835-19847.
41. Robbins JA, Krezoski JR, Mozley SC. Radioactivity in sediments of the Great Lakes: post-depositional redistribution by deposit-feeding organisms. *Earth and Planetary Science Letters.* 1977;36(2):325-333.
42. Sugiura K, Kitamura M, Matsumoto E, Goto M. Polychlorobiphenyls (PCBs) in sediments of Tokyo bay, Japan. *Arch Environ Cont Toxicol.* 1986;15(1):69-76.
43. Yamashita N, Kannan K, Imagawa T, Villeneuve DL, Hashimoto S, Miyazaki A, et al. Vertical profile of polychlorinated dibenzo-p-dioxins, dibenzofurans, naphthalenes, biphenyls, polycyclic aromatic hydrocarbons, and alkylphenols in a sediment core from Tokyo Bay, Japan. *Environmental Science & Technology.* 2000;34(17):3560-3567.
44. Yao Y, Takada H, Masunaga S, Nakanishi J. PCDDs, PCDFs and co-PCBs in Tokyo Bay: sources and contribution. *Organohalogen Compounds.* 2000;46:491-494.
45. Hosomi M, Matsuo T, Dobashi S, Katou S, Abe H. Survey of dioxins in Tokyo Bay bottom sediment. *Marine pollut bulletin.* 2003;47(1-6):68-73.
46. Hosomi M, Matsuo T, Dobashi S, Katou S, Abe H. Survey of dioxins in Tokyo Bay bottom sediment. *Marine pollution bulletin.* 2003;47(1-6):68-73..
47. Pan S, Kadokami K, Li X, Duong HT, Horiguchi T. Target and screening analysis of 940 micro-pollutants in sediments in Tokyo Bay, Japan. *Chemosphere.* 2014;99:109-116.
48. Nishimuta K, Ueno D, Takahashi S, Kuwae M, Kadokami K, Miyawaki T, et al. Use of comprehensive target analysis for determination of contaminants of emerging concern in a sediment core collected from Beppu Bay, Japan. *Environ Pollut.* 2021;272:115-587.
49. Bujagić IM, Grujić S, Jauković Z, Laušević M. Sterol ratios as a tool for sewage pollution assessment of river sediments in Serbia. *Environ Pollut.* 2016;213:76-83.
50. Nakakuni M, Takehara K, Nakatomi N, Higuchi J, Yamane M, Yamamoto S. Sterol and stanol compositions in sediments from the Bungaku-no-ike pond, Tokyo, Japan: Examination of stanol sources. *Res Organic Geochemistry.* 2017;33(1):7-16.
51. Sun MY, Wakeham SG. A study of oxic/anoxic effects on degradation of sterols at the simulated sediment-water interface of coastal sediments. *Organic Geochemistry.* 1998;28(12):773-784.
52. Guo JX, Fang J. The distribution of n-alkanes and polycyclic aromatic hydrocarbons in water of Taihu Lake. *Procedia Environ Sci.* 2012;12:258-264.
53. Xu HM, Tao J, Ho SS, Ho KF, Cao JJ, Li N, et al. Characteristics of fine particulate non-polar organic compounds in Guangzhou during the 16th Asian Games: Effectiveness of air pollution controls. *Atmos Environ.* 2013;76:94-101.
54. Ladjji R, Yassaa N, Balducci C, Cecinato A. Particle size distribution of n-alkanes and polycyclic aromatic hydrocarbons (PAHS) in urban and industrial aerosol of Algiers, Algeria. *Environ Sci Pollut Res.* 2014;21(3):1819-1832.
55. Gupta S, Gadi R, Mandal TK, Sharma SK. Seasonal variations and source profile of n-alkanes in particulate matter (PM 10) at a heavy traffic site, Delhi. *Environ Monit Asses.* 2017;189(1):43.
56. Lima AL, Farrington JW, Reddy CM. Combustion-derived polycyclic aromatic hydrocarbons in the environment—a review. *Environ Forensic.* 2005;6(2):109-131.
57. Prince RC, Garrett RM, Bare RE, Grossman MJ, Townsend T, Suflita JM, et al. The roles of photooxidation and biodegradation in long-term weathering of crude and heavy fuel oils. *Spill Sci Technol Bulletin.* 2003;8(2):145-156.
58. Rorije E, Verbruggen EM, Hollander A, Traas TP, Janssen MP. Identifying potential POP and PBT substances: Development of a new Persistence/Bioaccumulation-score.
59. Christian M. Final report on the safety assessment of cholesterol. *J Am Coll Toxicol.* 1986;5:491-516.
60. Neff JM. Bioaccumulation in marine organisms: effect of contaminants from oil well produced water. Elsevier. 2002:16.

61. RIVM 1,4-dimethylnaphthalene. National Institute for Public Health and the Environment. <https://echa.europa.eu/registry-of-clh-intentions-until-outcome/-/dislist/details/0b0236e181f0654a>. 2018.
62. Oğuz M, Mihçioğulları H. Environmental risk assessment of selected pharmaceuticals in Turkey. *Environ Toxicol Pharmacol*. 2014;38(1):79-83.
63. Baumann M, Weiss K, Maletzki D, Schüssler W, Schudoma D, Kopf W, et al. Aquatic toxicity of the macrolide antibiotic clarithromycin and its metabolites. *Chemosphere*. 2015;120:192-198.
64. Sundelin A. <https://www.diva-portal.org/smash/get/diva2:827050/FULLTEXT01.pdf>. 2015.
65. https://www.accessdata.fda.gov/drugsatfda_docs/nda/2011/201373Orig1s000EA.pdf. 2010.
66. Nichols JW, Du B, Berninger JP, Connors KA, Chambliss CK, Erickson RJ, et al. Observed and modeled effects of pH on bioconcentration of diphenhydramine, a weakly basic pharmaceutical, in fathead minnows. *Environ Toxicol Chem*. 2015;34(6):1425-1435.
67. Kristofco LA, Du B, Chambliss CK, Berninger JP, Brooks BW. Comparative pharmacology and toxicology of pharmaceuticals in the environment: diphenhydramine protection of diazinon toxicity in *Danio rerio* but not *Daphnia magna*. *The AAPS J*. 2015;17(1):175-183.
68. https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/290854/scho0809bqug-e-e.pdf. 2020.
69. Cleveland L, Mayer FL, Buckler DR, Palawski DU. Toxicity of five alkyl-aryl phosphate ester chemicals to four species of freshwater fish. *Environmental Toxicology and Chemistry: An Int J* 1986;5(3):273-282.
70. Sanders HO, Hunn JB, Robinson-Wilson E, Mayer Jr FL. Toxicity of seven potential polychlorinated biphenyl substitutes to algae and aquatic invertebrates. *Environ Toxicology and Chem: An Int J* 1985;4(2):149-154.
71. Sengupta A, Lyons JM, Smith DJ, Drewes JE, Snyder SA, Heil A, et al. The occurrence and fate of chemicals of Emerging concern in coastal urban rivers receiving discharge of treated municipal wastewater effluent. *Environmental Toxicology and Chemistry*. 2014;33(2):350-358.
72. Maruya KA, Dodder NG, Sengupta A, Smith DJ, Lyons JM, Heil AT, et al. Multimedia screening of contaminants of emerging concern (CECS) in coastal urban watersheds in southern California (USA). *Environ Toxicol Chem*. 2016;35(8):1986-1994.
73. Watanabe I, Sakai SI. Environmental release and behavior of brominated flame retardants. *Environ Int*. 2003;29(6):665-682.
74. Van der Veen I, de Boer J. Phosphorus flame retardants: properties, production, environmental occurrence, toxicity and analysis. *Chemosphere*. 2012;88(10):1119-1153.
75. <http://chm.pops.int/Portals/0/download.aspx?d=UNEP-POPS-PUB-Brochure-16NewPOPs-201706.English.pdf>. 2017.
76. Phillips AL, Hammel SC, Konstantinov A, Stapleton HM. Characterization of individual isopropylated and tert-butylated triarylphosphate (ITP and TBPP) isomers in several commercial flame retardant mixtures and house dust standard reference material SRM 2585. *Environ Sci Technol*. 2017;51(22):13443-13449.
77. Kurt-Karakus P, Alegria H, Birgul A, Gungormus E, Jantunen L. Organophosphate ester (OPEs) flame retardants and plasticizers in air and soil from a highly industrialized city in Turkey. *Sci Total Environ*. 2018;625:555-565.
78. Vykoukalová M, Venier M, Vojta Š, Melymuk L, Bečanová J, Romanak K, et al. Organophosphate esters flame retardants in the indoor environment. *Environ Int*. 2017;106:97-104.
79. Brits M, Brandsma SH, Rohwer ER, De Vos J, Weiss JM, de Boer J. Brominated and organophosphorus flame retardants in South African indoor dust and cat hair. *Environ Pollut*. 2019;253:120-129.
80. Peverly AA, O'Sullivan C, Liu LY, Venier M, Martinez A, Hornbuckle KC, et al. Chicago's Sanitary and Ship Canal sediment: Polycyclic aromatic hydrocarbons, polychlorinated biphenyls, brominated flame retardants, and organophosphate esters. *Chemosphere*. 2015;134:380-386.
81. Choi W, Lee S, Lee HK, Moon HB. Organophosphate flame retardants and plasticizers in sediment and bivalves along the Korean coast: Occurrence, geographical distribution, and a potential for bioaccumulation. *Marine Pollut Bulletin*. 2020;156:111275.
82. <http://www.oecd.org/env/ehs/risk-assessment/oecd-pov-and-lrtp-screening-tool.htm>. 2020.
83. Wegmann F, Cavin L, MacLeod M, Scheringer M, Hungerbühler K. The OECD software tool for screening chemicals for persistence and long-range transport potential. *Environ Modelling Software*. 2009;24(2):228-237.
84. Klasmeier J, Matthies M, Macleod M, Fenner K, Scheringer M. Application of multimedia models for screening assessment of long-range transport potential and overall persistence. *Environ. Sci. Technol*. 2006;40:53-60.
85. Klasmeier J, Matthies M, MacLeod M, Fenner K, Scheringer M, Stroebe M, et al. Application of multimedia models for screening assessment of long-range transport potential and overall persistence. *Environ Sci Technol*. 2006;40:53-60.
86. Wegmann F, Cavin L, MacLeod M, Scheringer M, Hungerbühler K. The OECD software tool for screening chemicals for persistence and long-range transport potential. *Environ Modelling Software*. 2009;24(2):228-237.