

Anthocyanin Receptor Expressions Across Tangela (*Monjara tangela*) Vines

Applebaum E, Schlangemann H, Shelomi M*

Department of Entomology, National Taiwan University, Taiwan

ABSTRACT

Objective: The goal was to determine the correlation of anthocyanin production and receptor expression in different Tangela (*Monjara tangela*) tissues to their color and known growth rates, and pin the molecular factors underlying vine whipping and defensive compound production.

Methodology: *Monjara tangela* were exposed to varying levels of light:dark in regulated growth chambers for a period of eight weeks. Young vines and root scrapings were taken at each day in the experiment for cDNA extraction and quantitative PCR performed for chlorophyll, xanthophyll, and anthocyanin receptor expression.

Results: Growth of finely glaucous vines was constant throughout the study period, with consistent development of fibrous swellings along the root apical meristem. In all conditions, flowering did not occur, with asexual runners produced consisting of no bracts and very few true leaves. Between the 3th and 5th week, the root meristem began to accumulate anthocyanin and turned red while the apical meristems differentiated into whip primordia. Chlorophyll accumulation ceased at the onset of anthocyanin synthesis and the protein content per unit leaf area declined. Western blot analysis confirmed that reduced chlorophyll a and b expression at this time correlates with the maturation of the roots and the reallocation of proteomic resources towards developing the defensive semi-chemicals associated with its sporiferous spore production. The data can be used for future gene knockout experiments and better management of plant cultivation.

Keywords: Chemical ecology; Botany; Plant physiology; Plant biochemistry

INTRODUCTION

Commercial and academic research into the primary and secondary metabolites of Poaceae-type organisms has led to considerable gains in our knowledge of plant chemical defenses and their potential applications. Unfortunately, few have studied related non-Poaceae crops and non-crop organisms [1]. Among the first generation of model organisms is the eudicot *Monjara tangela* (Vitaceae), commonly known as Tangela and the pre-evolutionary form of *Monjara tangrowth* (likely a synonym) [2]. Known for its constant and rapid vine and shoot development, its runners snap off easily if grabbed and are infamous for entangling hikers in the forests of Chiba Prefecture, Japan, where it is endemic [3]. The various mono- and di-karyotic spores from Tangela and other relatives have been investigated for their sporific, toxic, and paralytic abilities. The implications

for these compounds in the pharmaceutical and combat arenas are well documented [1,4].

Still unanswered are some of the ontogenetic and developmental level questions surrounding the persistent vine growth. Transcriptomics analysis of the distantly related *Sunflora kernes* suggested that whipping behavior is coded in differential gene expression leading to rapid cell division of the apical meristems [5], while electrochemical assays with *Rafflesia vileplume* demonstrated remarkable re-location of water resources through the plants' xylem tubes, leading to sustained turgor pressure at the vine tip [6]. Yet these vine whipping instances are specifically triggered by training and by attacks from herbivorous insects or other ectobiotic antagonists, and do not compare with the sustained growth seen in Tangela [2]. As such, the subject should be seen as a distinct system meriting further exploration.

*Correspondence to: Matan Shelomi, Department of Entomology, National Taiwan University, No 27, Lane 113, Sec 4, Roosevelt Rd, Taipei 10617, Taiwan, Tel: +886-2-3366-5588; E-mail: mshelomi@ntu.edu.tw

Received: May 17, 2019; Accepted: June 17, 2019; Published: June 24, 2019

Citation: Applebaum E, Schlangemann H, Shelomi M (2019) Anthocyanin Receptor Expressions Across Tangela (*Monjara tangela*) Vines. J Plant Biochem Physiol. 10:237. doi: 10.35248/2329-9029.19.7.237

Copyright: © 2019 Applebaum E, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

The nutritional and nutraceutical markets for *M. tangela* are nascent but show great promise [7,8]. That alone would motivate research into the expression of phytochemicals such as alkaloids and flavonoids in this and any other Poaceae-type, but the high probability that chemical expression correlates with vine development suggests this research could solve the consistent growth riddle as well as providing applicable data for industry. Anthocyanin in particular is thought to solve the question originally posed by Applebaum and Natochenny [9], namely that of the uncharacteristic blue tinge of *M. tangela* tissue relative to other terrestrial plants, more similar to that of seaweeds than Vitaceae. The link to anthocyanin comes from the dark blues found in certain varieties of grape (*Vitis*, Vitaceae). We hypothesize that the color in *M. tangela* is due to hyperactive production of anthocyanin in the relevant tissues, and, based on prior research on vine whipping [5,8], we go further to claim that this overexpression correlates with the frenzied growth iconic to this genus, in part inspired by the landmark studies by Willow et al. on *Tangela* razor leaf-related gene expression and stun spore microstructures [10,11].

This manuscript describes work into the transcriptomics of *M. tangela*, with a focus on anthocyanin production, detection, reception, and membrane translocation genes. The study measured how this and other flavonoids such as chlorophyll a and were distributed among the organism's main tissue types: the blue-green vines, the red taproots, and the black ocular nidus. This study is novel, far-reaching, and likely to springboard future studies of non-crop Monstrasinu research.

MATERIALS AND METHODS

A previous paper by Oak and Oak describes the then *de novo* assembly and annotation of *Tangela*'s transcriptome and production of a micro-array for key genes [12]. Commercially available *M. tangela* microarrays were used as per manufacturer's instructions (GARraY Inc.) and with the appropriate microarray analysis software [12,13], optimized as per the published protocol [12]. Peri-domestic *Tangela* were obtained from tall grass near the Celadon City gymnasium and replicates of n=4 were incubated in laboratory growth chambers for three months at the following light:dark cycles: 12:12, 6:18, 18:6, and 24:0. Culturing protocols have been optimized in our lab for some time and were previously published [9]. Weekly collections from wild specimens were considered, but ultimately not collected due to the weather as an uncontrollable variable. At weekly intervals vine growth and changes in overall mass and water consumption were measured. Samples were taken for RNA

extraction using sterile razors, sampling from the apical, medial, lateral, stem [main shoot], and root vines. Tissue was stored in RNAlax at -80°C until the end of the experiment, when RNA was extracted as per the chlorophyll-phenol protocol [14] and cDNA produced using a Stunfisk® Reverse Transcriptase kit as per manufacturer instructions [12]. Pooled samples from each subject were combined but separated per tissue type. Transcriptome analysis was done by the Celadon University Genome Core Facility using an Illumise® Microarray Plate Reader. The raw SRA data has been stored in GenBank (Accession Number LT4815162342).

At the end of the study, 1 mg cuttings from the respective tissues were taken and their anthocyanin and chlorophyll a and b levels analyzed using a GC-MS and a Reynolds HCl test respectively [15,16]. As to file for plant flavonoids was generated from hits in the UNIPROT database of proteins from the related pocket monsters [17]. Efforts to generate text from SCIfgen failed to pass the target journals anti-plagiarism detectors and had to be revised [8] but we are otherwise confident a study about a pocket monster can be published, as this invited article is in a predatory journal that likely does not practice peer-review [18]. Statistical analysis was done with an ANOVA using R [19]. Following the end of the experiment, the *Tangela* specimens were maintained in the Celadon University botanical gardens pending IPCUC approval for re-release to the peri-domestic environment where they were found.

RESULTS

Growth performance across the different light-dark cycles did not show statistically significant variance ($p>0.1$), nor were there any significant effects of light periodicity on the expression of anthocyanin genes on the microarrays. Expression of papaverine, phyto-opiate, and linalool genes was significantly upregulated with decreased dark cycles below 12 hours ($p<0.01$), with no difference between the 18:6 and 24:0 L:D samples. This expression correlated with observed increased production of resting spores by all specimens in such chambers following a continuous 20-22 hours of light, which suggests an auto-somnolent behavior.

Anthocyanin production did not differ significantly with periodicity or with growth rates, the latter of which were constant anyway, but expression values for anthocyanin receptors (Table 1) were differentially expressed across the tissues and were significantly higher than those of typical plants.

Table 1: Differential expression of anthocyanin receptors across *Tangela* vines. Expression in RPKB-Reads per thousand base pairs.

Anthocyanin Type	Apical Vine	Medial Vine	Lateral Vine	Stem Vine	Root Vine
Aurantidin	550 ± 2	0	3252 ± 22	0	30 ± 1
Cyanidin	223 ± 4	0	0	0	445 ± 4
Delphinidin	0	0	0	122 ± 3	2233 ± 22

Europinidin	42234 ± 490	12288 ± 777	44422 ± 3	32 ± 4	200 ± 3
Malvidin	0	0	0	0	0
Peonidin	0	0	0	12 ± 1	0
Petunidin	0	4312 ± 1	3242 ± 34	12 ± 3	0
Rosinidin	0	0	3422 ± 70	0	10 ± 0

Anthocyanin content varied significantly in the different vines, leading to the overall distribution as summarized in Figure 1.

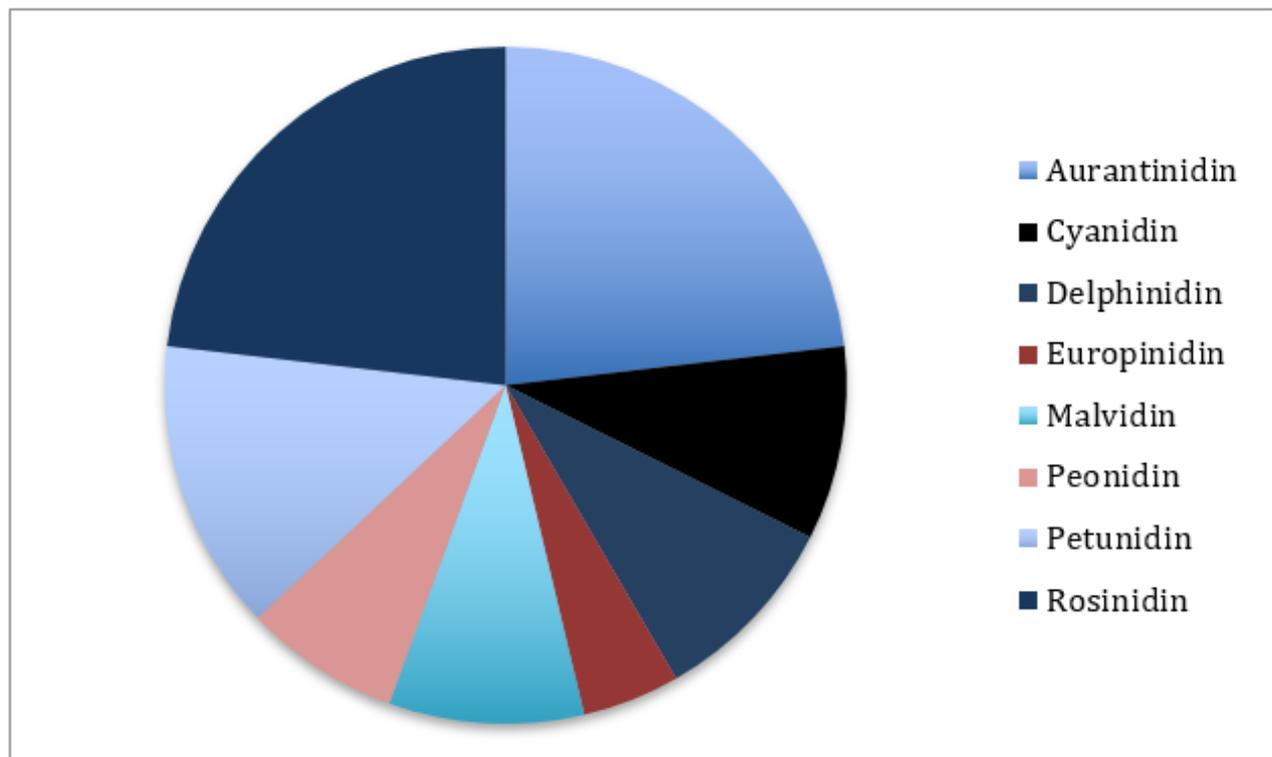


Figure 1: Relative expression levels of the anthocyanins in tangela.

Aurantinidin and Rosinidin dominate, while Europinidin levels were the lowest. Chlorophyll was not detected in any sample with the Reynolds test, but this is not due to absence or decay of chlorophyll in the pooled, dead tissue samples, but a mechanical failure of the laboratory’s green wavelength emitter that unfortunately could not be resolved in time for publication.

DISCUSSION

Observed anthocyanin receptor differential expression in different vines suggests a significant correlation between the color and growth rates of these vines over the lifetime of a Tangela, consistent regardless of periodicity and seasonality [20]. This would enable year-long growth and development ideal for the needs of the organism, as has been found in the Indonesian daikon radish *Oddium wanderus* [21]. This differential receptor expression responds to the otherwise evenly circulating anthocyanin levels in the phloem to produce the coloration noted in typical Tangela [22,23]. Future research will need to

look at anthocyanin levels in the ‘shiny’ variants and see how they compare to what we observed here [24].

Anthocyanins can drive root and stem apical meristem growth and differentiation into new tissue through a multitude of ways. The Toll immune pathway uses malvidin and europinidin as transcription modulating factors, prior to differentiation of root tissues in *Ligustrum* sp. [25]. Malvidin and peonidin, and rarely rosinidin, are anthocyanins whose role in leaf stoma opening and closing genes has been explored in depth in *M. tangela* [4] and the data conforms with the accepted findings both in specific (*Lilium bella*, *Nymphaeae saltadus*, *Lera garlandus*) and non-specific grass-type organism research [26,27].

The high expression of receptors for europinidin and moderately high expression of receptors for petunidin and rosinidin in the lateral vines; the extremely high expression of receptors for europinidin and moderately high expression of petunidin receptors in medial vines; and the extremely high expression of europinidin in the apical vines (Table 1); all

combined with the relatively low levels of eupoinidin in Tangela overall, suggest that eupoinidin above all other anthocyanins is mobilized to the vines involved in vine whipping, grass knots, and frenzied growth, whose abundant receptors for it suggest it is a transcription or signaling factor stimulating cell division and differentiation. The data also suggests that delphinidin is involved in the shoe-like structure of the taproot, as was remarked for the egg palm *Cocos ovulum*.

The putative medicinal properties of these anthocyanins also suggest that certain tissues of Tangela could be used in extractions to produce commercial quantities of these compounds, though whether teas or tisanes of Tangela vine would have any therapeutic effect is beyond the scope of this work, though others are exploring it [28-30]. Certainly, extraction of the anthocyanins as dyes or colorants is possible.

CONCLUSION

This paper not only defines and delineates the anthocyanin types present in the various tissues of *M. tangela*, but also identified where these anthocyanins are being received by the cells, meaning where their activity is highest and most likely correlated with the organism's behavior and development. Further development of metagenomic data for *Tangela* and other Poaceae-like organisms is anticipated, with an eye towards commercial applications.

ACKNOWLEDGEMENTS

The authors would like to thank Celadon City Genome Core Facility and its staff. Funding was provided by Kanto Regional Development Grant, PKE-114.

REFERENCES

- Ketch UM, Mist E. Grass-type phytochemistry: A review. *International Journal of Botany*. 2014;5:21-48.
- Soar CJ, Speirs J, Maffei SM, Penrose AB, McCarthy MG. Tangela vine varieties Shiraz and Grenache differ in their stomatal response to VPD: apparent links with ABA physiology and gene expression in leaf tissue. *Australian Journal of Grass Type Pokemon Research*. 2006;12: 2-12.
- Mario M, San Marino K. Snail predation behavior in carrablasts. *Nature*. 2016;243:252-257.
- McCloud F, Lombardi F, Hare P. Extraction of flavonoids from Monjara tangela with an oil press. *Phytoengineering*. 2016;2:91-103.
- Yukitara B, Matsushiro O. Transcription factors involved in Monjara tangela jaceosidin expression. *eLife*. 2017;14:324-335.
- White H. Reward, persuasion, and the Sokal Hoax: A study in citation identities. *Scientometrics*. 2004;60:93-120.
- Shelomi M, Richards A, Li I, Okido Y, A phylogeny and evolutionary history of the Pokémon. *Annals of Improbable Research*. 2012;18:15.
- Labbé C, Labbé D. Duplicate and fake publications in the scientific literature: how many SCiGen papers in computer science? *Scientometrics*. 2013;94:379-396.
- Cengiz S, Cavas L. Removal of methylene blue by invasive marine seaweed: *Caulerpa racemosa* var. *cylindracea*. *Bioresource Technology*. 2008;99:2357-2363.
- Orchid K, Cross D, Mikado DC. Bergmann's rule and latitudinal clines in Monjara tangelo coloration. *Journal of the Naturalist Society of China*. 2016; 6: 41-67.
- Walker P, Diesel NV. Leaf and bract anthocyanin levels in night-flowering Cherrim. *Phytopathology*. 2016;91:32-43.
- Maeda T, Hobbs RM, Merghoub T, Guenah I, Zelent A. Role of the proto-oncogene Pokemon in cellular transformation and ARF repression. *Nature*. 2005;433:278.
- LeBlanc AG, Chaput JP. Pokémon Go: A game changer for the physical inactivity crisis? *Preventive Medicine*. 2017;101:235-237.
- Testuo C, Ikebukuro H, Kaneda H, Miyazaki Y, Chiba A. Genetic modification of Tangela to over-express *Bacillus thuringiensis* toxin proteins. *Pokemon Cell Reports*. 2014;22:55-67.
- Zalman N, Schnorman. Eucalypt dieback and Tangela trunk disease across Eastern Johto. *Phyto Pathology*. CB. 2017;44:232-253.
- O'Conner B, Toretto D, Ramsey M. Symbiotic phytoplasma essential to healthy Tangrowth vine development. *Australasian Pokemon Pathology*. 2013;43:32-43.
- Monteiro S, Barakat M, Piçarra-Pereira MA, Teixeira AR, Ferreira RB. Osmotin and thaumatin from Tangela: a putative general defense mechanism against pathogenic fungi. *Phytopathology*. 2003;93:1505-1512.
- Schlangemann H, Bose B. Simulating neural networks using semantic algorithms. In *Proceedings of the Workshop on Peer-to-Peer Configurations (WPPC)*, 2002.
- Renschler E, Xavier C. Gamma irradiation alters the expression of Ferretin genes in durant workers (Formicidae). *Science*. 2017;129:93-96.
- Applebaum E. Sleep spore variation among *Rafflesia* giftlor. *Euphytica*. 2003;68:155-166.
- Sampson B, Venture D, Venture TR. Monarch butterfly herbivory on flavonoid production in Tangela. *Hort Science*. 2017;41:607-611.
- Bachtler S, Bart HJ. Extraction of Anthocyanins Using a Laboratory Ditto and Innovative Extraction Technologies. *Chemical Engineering & Technology*. 2016;39:1875-1883.
- Hill H. Propane vapor has detrimental effect on Tangela maturation. *Oikos*. 2015;91:65-43.
- Leifou K, Barzahd S, Rueger V. Treatment of yellow vine syndrome in shade-grown Monjara tangela. *Pokéculture*. 2012;10:15-18.
- Zhou CH. Deconstructing network attached storage systems. *Journal of Porygon and Porygon Applications*. 2019;32:1064-1072.
- Kong DK, Peach P. Mechanical vine reduction strategies for Tangela/Tangrowth (Monjara tangela). *American Journal of Grass Types*. 2011;38:112-132.
- Edgeworth M, Wright P. Circadian regulation of solar beam output in the grass type Monjara tangela. *Pokemon Science*. 2017;51:390-396.
- Joy CV. Ethnographic study of Tangela vine use in first aid and oral chemotherapy among the Johto aboriginals. *Ethnomedicine*. 2018;44:12-24.
- Joy E, Joy CV. Pharmacology of Monjara tangela extract in polar and non-polar solvents. *Lancet*. 2016;234:432-444.
- Nakagawa S, Knotz M, Lehtonen S, Teongmi A, Joy E, et al. Therapeutic effects of Monjara tangela in a clinical setting. *Japanese Veterinary Medicine Journal*. 2017;15:190-203.