

# Analysis of Chromosomal Aberrations in PBL of Chrome Tanning Industrial Workers

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Research

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## Abstract

In order to monitor the geno-toxic effects of chromium compound, 160 chrome tanning industrial workers (80 each of smoker and non-smoker) were selected from a leather tanning industry to perform the cytogenetic protocol of chromosomal aberrations in the peripheral blood lymphocytes. The workers were further classified into two groups on the basis of their personnel habit (Smoker and Non-Smokers) and on the duration of their employment (1-5, 6-10 and 11-15 years). Even 120 individuals (62 smokers and 58 non-smokers) were also selected as controls for the purpose of comparison. A statistically significant increased frequency of total chromosomal aberrations was observed in the chrome tannery workers ( $6.20 \pm 0.12$ ) as against the control ( $1.61 \pm 0.09$ ). The increased frequencies of aberrations were duration dependent. Even higher incidence of chromosome aberrations was reported among the smoker group of both control and exposed workers. The results clearly establish the mutagenic nature of the chromium compound in human beings.

**Keywords:** Chromium; Chromosomal aberration; Leather tanning; Industrial workers; Peripheral blood lymphocyte; Smokers and non-smokers

## Introduction

Metals are the ubiquitous chemical entities, known to cause mutations in a variety of test systems. Based on the epidemiological studies, some of them are classified as human carcinogen [1] as well as mutagens and the human exposure to these metallic compounds was found among the workers in a large number of professional groups. The occupational exposure to chromium compound was typically found in stainless steel welders, chrome platters [2], leather tanning and chromate production workers [3].

Leather tanning is an age-old practice in India, which is recently recognized as one of the potential polluting industry of considerable importance. The wastes from tannery contain an excess amount of chromium ranging from 100-200 mg/l and the spent chrome liquor contains 2900-4500 mg/l [4]. Chronic exposure, poor working conditions, lack of civic sense and awareness of the potential hazard leads to an occupational health hazardous situations at an industrial set up.

Chromium compounds were known to be potent carcinogens and mutagens [5], which can induce a spectrum of DNA damage [6], gene mutation [7], sister chromatid exchanges [8] and chromosomal aberrations [9]. Several workers already reported the mutagenic and carcinogenic potentialities of chromium compound in bacterial and mammalian cell-based mutagenicity assays [10-13]. The investigations carried out to examine the geno-toxicity in workers occupationally exposed to chromium are meagre [14-18]. However, there are reports on positive geno-toxic effects in populations exposed to chromium also a negative findings [19-24]. The studies on the evaluation of genetic damage in chrome tanning workers exposed to chromium are scanty and rather conflicting. Hence an attempt was made during this study to evaluate the mutagenic potential of chromium compound in occupationally exposed leather tanning industrial workers in a tanning industry situated at Bakaram industrial area, Hyderabad, Andhra Pradesh, India by using the standard cytogenetic protocol of chromosome aberrations.

## **Materials and Methods**

#### Air and blood chromium analysis

Ambient air samples at different locations of the industry were collected to evaluate chromium levels. The samples were collected on membrane filter (37 mm, 0.8  $\mu$ m pore size) and in 1M HNO<sub>3</sub> using KIMOTO air samples at a flow rate of 1-2 LPM for 8hrs in day shift. The collected samples were wet digested with concentrated HNO<sub>3</sub> and analysed for chromium by using atomic absorption spectrophotometer (Double Beam 3100 model, Perkin Elmer, USA) [25].

Blood samples were collected in heparinized vials and were wet digested in microwave digestive system using conc.  $HNO_3$  and were analyzed for the metal by atomic absorption spectrophotometer (Double Beam 3100 model, Perkin Elmer, USA) [26].

#### **Study population**

160 tannery industrial workers (80 each of smoker and non-smoker group) were selected from a leather tanning industry situated at Bakaram industrial area, Hyderabad, Andhra Pradesh, India, to evaluate the genotoxic potential of Chromium compound by using the standard cytogenetic protocol of chromosomal aberrations in peripheral blood lymphocytes. The selected workers belong to the age group of 25-50 years and belong to the same socio-economic status. Simultaneously 120 individuals (62 smokers and 58 non-smoker group) who didn't have any history of exposure were also selected for comparison (control). The data was further analyzed on the basis of their personnel habitat into smokers and non-smokers and also on their duration of service into 1-5, 6-10 and 11-15 years, respectively. All participants were informed about the objectives of the investigation and written consent was obtained from each person. Personnel data and family histories were collected by the interviews and questionnaires. This study was approved by institutional ethics committee.

## Lymphocyte isolation and cell culture

Blood samples were collected by vein puncture in Heparinized centrifuge test tubes and transported to the laboratory within 2-4 hrs. Lymphocytes were isolated by gradient centrifugation and washed three times in phosphate buffer saline. Lymphocytes from each individual sample were divided into several aliquots. One part was used to prepare 2 ml lymphocyte cultures, with a cell density of 1  $\times$  106/ ml in RPMI – 1640 culture media, supplemented with 15% fetal calf serum , antibiotic and 1% phytohaemagglutinin. The lymphocyte cultures were incubated at 37°C in a humid atmosphere with 5 % CO<sub>2</sub> for 48 hrs. Cell harvesting and slide preparation

Cell harvesting was done followed by colchicine treatment (0.1  $\mu$ g/ml culture medium) for the last 4 hours of the 48 hours incubation, hypotonic treatment in prewarmed 0.075M KCl solution for 15 minutes at 37°C and three fixations in chilled methanol – acetic acid

mixture (v/v, 3:1). The cells were dropped on to cool wet slides. The slides were air dried and stained in 2% Giemsa stain (pH 6.8) mounted with DPX and scored for chromosomal aberrations by adopting the method of [27].

#### Aberration scoring and statistical analysis

Chromosomal aberrations were scored on coded slides by two independent observers using research microscope under  $10 \times \text{ and } 100 \times$ oil immersion. One hundred metaphases per subject were scored in the heavy metal exposure study. Only well spread metaphases with 46 centromeres were selected. Gaps (A chromatic lesions) were counted separately but not included in the frequency of the cells with aberrations. The frequencies of the cells with aberrations were tested statistically by using a Chi-square (2×2 contingency) test to find out the significant levels between the groups tested.

## Results

The result on chromium concentrations in the ambient air and in the blood was depicted in Table 1. The levels of chromium were found in higher concentrations at tanning unit (43.26  $\pm$  20.20 µg/m<sup>3</sup>). Spray dyer (22.14  $\pm$  10.60 µg/m<sup>3</sup>) and in administrative office (3.86  $\pm$  0.61 µg/m<sup>3</sup>). The chromium levels observed are much lower than threshold limit value of 500 µg/m<sup>3</sup>. The mean blood chromium levels in the exposed workers was 6.83  $\pm$  1.32 µg/100 ml which was considered higher when compared to the control value of 2.67  $\pm$  0.34 µg/100 ml.

Chromium	Ambient air µg/m³ Blood(µg/100ml)			Blood(µg/100ml)	
	Tanning unit	Spray dyer	Admn. office	Control	Exposed
	43.26 ± 0.20	22.14 ± 0.60	3.86 ± 0.61	2.67 ± 0.34	6.83 ± 1.32

Values in the ambient air at different work stations were less than the threshold values of 500 µg/m<sup>3</sup>.

#### Table 1: Chromium concentration in different study areas.

The overall result on the incidence of chromosomal aberrations in leather tanning industrial workers is presented in Tables 2 and 3.

Group	No. of Examinees	% of Aberrant cells	P value		
Control	120	1.61 ± 0.09	-		
1-5 yrs	75	4.82 ± 0.33	0.01		
6-10 yrs	50	6.84 ± 0.73	0.05		
11-15 yrs	35	8.22 ± 1.38	0.05		
Total exposure	160	6.20 ± 0.12	0.05		
100 metaphases were scored for each sample. Gaps and polyploids are not included in aberrant cells. Values in parenthesis are percentages ± S.E.					

**Table 2:** Chromosomal aberration frequencies in occupational tanneryworkers (smokers and non-smokers) exposed to chromium.

An increased pattern in the frequency of chromosomal aberration was observed in the chromium exposed group when compared to the control. As a result of this the percentage of total chromosomal aberrations got increased from  $6.20 \pm 0.12$  in the exposed workers as against  $1.61 \pm 0.09$  in the control subjects. In order to monitor the longitudinal variations of chromosomal aberrations, further analysis was carried out on the basis of duration of employment and on the basis of their smoking habit. A gradual increase in the frequency of total chromosomal aberrations of  $4.82 \pm 0.33$ ,  $6.84 \pm 0.73$  and  $8.22 \pm$ 1.38 were observed with the increase in the duration of exposure of 1-5, 6-10, 11-15 years respectively (Table 2). Similar trend was observed among the smoker and non-smoker subjects. A  $5.35 \pm 0.36$ ,  $7.76 \pm 0.55$  and  $10.80 \pm 0.84$  increased percentage of chromosomal aberrations were observed among the smoker exposed group as the increase in the duration of exposure at the industry as against the control smoker group of  $3.70 \pm 0.30$ ,  $5.92 \pm 0.48$  and  $8,40 \pm 0.74$ respectively (Table 3).

Group		Smokers		Non-Smokers	
		No. of Examine es	% Aberrant cells ± S.E	No. of Examinee s	% Aberrant cells ± S.E
Control	-	62	134 (2.16 ± 0.18)	58	60 (1.03 ± 0.13)

Duration of	1-5	40	214 (5.35 ± .36)**	40	148(3.70 ± . 30) <sup>**</sup>
exposure	6-10	25	194 (7.76 ± .55) <sup>*</sup>	25	148(5.92 ± .48)*
	11-15	15	162 (10.80 ± . 84) <sup>*</sup>	15	126 (8.40 ± . 74) <sup>*</sup>
Total experien ce	1-15	80	570 (7.12 ± .29)*	80	422(5.27 ± .25)*
100 metaphases were scored for each sample. Values in parenthesis are percentages ± S.E. Gaps and polyploidy are not included in aberrant cells. * P< 0.05 and **P<0.01.					

**Table 3:** Chromosomal aberration frequencies in Smoker and Non-Smoker groups exposed to chromium.

# Discussion

Large number of industries releases the chromium compound into the air, water and soil. In the air this compound is mainly present in the form of fine dust [28]. The permissible exposure limits for chromium in work place during an 8 hr, 40 hrs work weak is 100  $\mu$ g/m3 and the recommended exposure limit is 1  $\mu$ g/m<sup>3</sup>. The investigated result is supported by the observations made by [29] in a chemical based industry.

There is clear evidence that some metals represent a carcinogenic hazard to man and several metallic compounds has been identified as human carcinogens [30]. Evaluation of mutagenic hazards has become an integral part in the toxicological assessment with a number of environmental chemical [31]. The cytogenetic methods were routinely employed in monitoring the populations, exposed to industrial chemicals [32]. A significant increase in the frequency of chromosomal aberrations among the workers of leather tanning industry is once again establishing the mutagenic nature of the chromium compound, as the tannery effluents have the potential to damage the DNA of test organisms [33].

Induction of chromosomal aberrations in human peripheral blood lymphocytes by chromate compounds were reported earlier [34]. However the present results were attributed to the observations made in peripheral blood lymphocytes of chromate workers, exposed to chromium containing fumes [35-39]. Further the results were comparable with that of the observations among the workers exposed to benzene pyrene-epoxides [40-42].

The chromosomal aberrations observed in the study was mainly of chromatid type, which can be capable to induce more number of aberrations in late S1 phase or early G2 phase of the cell cycle [43]. The presence of less iso-chromatid aberrations may reflect a direct effect of the compound on G1 phase. Further, the aberrations recorded even after 11-15 yrs of exposure may be due to the phenomenon of aging of the cells in circulating blood lymphocytes.

A significant increase in chromosomal aberrations in smoker and non-smoker exposed group to chromium compound than their respective controls was due to the effect of cigarette smoke on the genetic material. Even the synergistic interaction would also be possible, but the actual mechanism is not yet to be documented. Similar findings were reported by several workers among the smoker groups, occupationally exposed to rubber, heavy metals and plastic workers which will support the present investigation [44-49].

The chromate compounds are well known as human and animal carcinogens and the workers occupationally exposed to chromium compound are prone towards the increased risk of cancer. The exact mechanism for chromium mutagenicity has not been so far reported, but it has been suggested that the chromium compound give off hydroxyl, cysteinyl and thionyl radicals during cellular reductions [50-53]. These radicals can interact directly with DNA - chromatin to form DNA single strand breaks, DNA-protein cross links, chromium-DNA adducts and DNA-DNA cross links, ultimately leads to chromosomal breakage and mutations. In conclusion, the study reveals that there is a significant increase in the frequency of chromosomal aberrations in the peripheral blood lymphocytes of leather tanning industrial workers. This increase may be due to chromosomal instability and also the fact that chromium being used in tannery industry is a potent mutagen. Further this study needs an elaborate and exhaust study in order to interpret the data in a conceptual frame work. The present study facilitates to make a decision on issue of pollution control, mutagenesis and risk analysis. It is also helpful to have a solution for the damage caused by the polluting industries.

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# References

- 1. Alcedo JA, Wetterhahn KE (1990) Chromium toxicity and carcinogenesis. Int Rev Exp Pathol 31: 85-108.
- Benova D, Hadjidekova V, Hristova R, Nikolova T, Boulanova M, et al. (2002) Cytogenetic effects of hexavalent chromium in Bulgarian chromium platers. Mutat Res 514: 29-38.
- Burgaz S, Demircigil GC, Yilmazer M, Erta N, Kemaloglu Y, et al. (2002) Assessment of cytogenetic damage in lymphocytes and in exfoliated nasal cells of dental laboratory technicians exposed to chromium, cobalt, and nickel. Mutat Res 521: 47-56.
- 4. De Flora S (2000) Threshold mechanisms and site specificity in chromium(VI) carcinogenesis. Carcinogenesis 21: 533-541.
- De Flora S, Bagnasco M, Serra D, Zanacchi P (1990) Genotoxicity of chromium compounds. A review. Mutat Res 238: 99-172.
- de Jong G, van Sittert NJ, Natarajan AT (1988) Cytogenetic monitoring of industrial populations potentially exposed to genotoxic chemicals and of control populations. Mutat Res 204: 451-464.
- Dearfield KL, Auletta AE, Cimino MC (1991) Consideration in the US Environmental Protection agency's testing approach for mutagenecity. Mutat. Res 285: 259-283.
- 8. Deng CZ, Lee HH, Xian HL,Yao MC, Huang JC (1988) Chromosomal aberrations and sister chromatid exchanges of peripheral blood lymphocytes in chinese electroplating workers: effect of nickel and chromium. J Trace Elem Exp Med 1: 57-62.
- 9. Deng CZ, Ou BX, Huang JC, Zhuo ZL, Xian HL (1983) Cytogenetic effects of electroplating workers(China). Acta. Sci. Circumst 3: 267-271.
- Friberg L, Kjellstrom T, Nordberg GF (1986) Cadmium. Handbook on the toxicology of Metals, Elsevier Science, Amsterdam: 130-184.
- Gambelunghe A, Piccinini R, Ambrogi M, Villarini M, Moretti M, et al. (2003) Primary DNA damage in chrome-plating workers. Toxicology 188: 187-195.
- 12. Gennart JP, Baleux C, Verellen-Dumoulin C, Buchet JP (1993) Increased sister chromatid exchanges and tumor markers in workers exposed to

elemental chromium, Cobalt and nickel containing dusts. Mutat Res 299: 55-61.

- Gibb HJ, Lees PS, Pinsky PF, Rooney BC (2000) Lung cancer among workers in chromium chemical production. Am J Ind Med 38: 115-126.
- 14. Hodges NJ, Adams B, Lee AJ, Cross HJ, Chipman JK (2001) Induction of DNA-strand breaks in human peripheral blood lymphocytes and A549 lung cells by sodium dichromate: association with 8-oxo-2deoxyguanosine formation and interindividual variability. Mutagenesis 16: 467-474.
- 15. Huvinen M, Makitie A, Jarventaus H, Wolff H, Stjernvall T (2002) Nasal cells micronuclei, cytology and clinical symptoms in stainless steel production workers exposed to chromium. Mutagenesis 17: 425-429.
- 16. [No authors listed] (1993) Beryllium, cadmium, mercury, and exposures in the glass manufacturing industry. Working Group views and expert opinions, Lyon, 9-16 February 1993. IARC Monogr Eval Carcinog Risks Hum 58: 1-415.
- 17. IARC monograph (1987) Monographs on the evaluation of carcinogenic risks to Human.Supplement 7 IARC, Lyon.
- 18. IARC (1990) Chromium, Nickel and welding. IARC monographs on elevation of the carcinogenic risk to humans.
- 19. Izzotti A, Bagnasco M, Camoirano A, Orlando M, De Flora S (1998) DNA fragmentation, DNA-protein crosslinks, postlabeled nucleotidic modifications, and 8-hydroxy-2'-deoxyguanosine in the lung but not in the liver of rats receiving intratracheal instillations of chromium(VI). Chemoprevention by oral N-acetylcysteine. Mutat Res 400: 233-244.
- Jelmert O, Hansteen IL, Langård S (1994) Chromosome damage in lymphocytes of stainless steel welders related to past and current exposure to manual metal arc welding fumes. Mutat Res 320: 223-233.
- Koshi K, Yagami T, Nakanishi Y (1984) Cytogenetic analysis of peripheral blood lymphocytes from stainless steel welders. Ind Health 22: 305-318.
- Langård S1 (1990) One hundred years of chromium and cancer: a review of epidemiological evidence and selected case reports. Am J Ind Med 17: 189-215.
- Linnainmaa K (1983) Sister chromatid exchanges among workers occupationally exposed to phenoxy acid herbicides 2,4-D and MCPA. Teratog Carcinog Mutagen 3: 269-279.
- 24. Liu S, Dixon K (1996) Induction of mutagenic DNA damage by chromium (VI) and glutathione. Environ Mol Mutagen 28: 71-79.
- 25. Major J, Jakab MG, Tompa A (1998) Genotoxicological monitoring of 175 subjects living in the green belts, inner town or near chemical industrial estates in Greater Budapest agglomeration, Hungary. Mutat Res 412: 9-16.
- Manning FC, Xu J, Patierno SR (1992) Transcriptional inhibition by carcinogenic chromate: relationship to DNA damage. Mol Carcinog 6: 270-279.
- 27. Montaldi A, Zentilin L, Paglialunga S, Levis AG (1987) Solubilization by nitrilotriacetic acid (NTA) of genetically active Cr(VI) and Pb(II) from insoluble metal compounds. J Toxicol Environ Health 21: 387-394.
- Moorhead PS, Nowell PC, Mellman WJ, Battips DM, Hungerford DA (1960) Chromosome preparations of leukocytes cultured from human peripheral blood. Exp Cell Res 20: 613-616.
- 29. Nanda NV, Baekavathy DM (1986) Adverse health effects of tannery and chromate industrial workers exposed to occupational environment: An environmental epidemiological study. Env Rep 12: 51-60.
- Newton MF, Lilly LJ (1986) Tissue-specific clastogenic effects of chromium and selenium salts in vivo. Mutat Res 169: 61-69.
- O'Brien TJ, Ceryak S, Patierno SR (2003) Complexities of chromium carcinogenesis: role of cellular response, repair and recovery mechanisms. Mutat Res 533: 3-36.
- 32. Petering HG, Yeager DW, Witherup SO (1973) Trace metal content of hair. II. Cadmium and lead of human hair in relation to age and sex. Arch Environ Health 27: 327-330.

- Prasad MH, Reddy PP (1993) Rubber industry toxicity of work environment. In: Hand book of hazardous material. Academic press. Inc, USA: 639-648.
- Quievryn G, Peterson E, Messer J, Zhitkovich A (2003) Genotoxicity and mutagenicity of chromium(VI)/ascorbate-generated DNA adducts in human and bacterial cells. Biochemistry 42: 1062-1070.
- Rita P, Reddy PP, Reddy SV (1987) Monitoring of workers occupationally exposed to pesticides in grape gardens of Andhra Pradesh. Environ Res 44: 1-5.
- 36. Rowbotham AL, Levy LS, Shuker LK (2000) Chromium in the environment: an evaluation of exposure of the UK general population and possible adverse health effects. J Toxicol Environ Health B Crit Rev 3: 145-178.
- Sarto F, Cominato I, Bianchi V, Levis AG (1982) Increased incidence of chromosomal aberrations and sister chromatid exchanges in workers exposed to chromic acid in electroplating factories. Carcinogenesis 9: 1011-1016.
- Sathwara NG, Patel KG, Vyas JB, Patel S, Trivedi MR, et al. (2007) Chromium exposure study in chemical based industry. J Environ Biol 28: 405-408.
- 39. Snow ET (1992) Metal carcinogenesis: mechanistic implications. Pharmacol Ther 53: 31-65.
- 40. Sorsa M, Falck K, Mäki-Paakkanen J, Vainio H (1983) Genotoxic hazards in the rubber industry. Scand J Work Environ Health 9: 103-107.
- Stearns DM, Silveira SM, Wolf KK, Luke AM (2002) Chromium(III) tris(picolinate) is mutagenic at the hypoxanthine (guanine) phosphoribosyltransferase locus in Chinese hamster ovary cells. Mutat Res 513: 135-142.
- Stella M, Montaldi A, Rossi R, Rossi G, Levis AG (1982) Clastogenic effect of chromium on human lymphocytes invitro and invivo. Mutat. Res 101: 151-164.
- Stearn DM, Wetterhahn KE (1994) Reaction of chromium (VI) with ascorbate produces chromium (V), chromium (IV) and carbon-based radicals. Chem Res Toxicol 7, 219-230.
- 44. Sugiyama M, Tsuzuki K, Ogura R (1991) Effect of ascorbic acid on DNA damage, Cytotoxicity, glutathione reductase and formation of paramagnetic chromium in Chinese hamster V-79 cells treated with Sodium chromate(VI). J.Biol Chem 26: 3383-3386.
- 45. Sunderman FW Jr. (1984) Recent advances in metal carcinogenesis. Ann Clin Lab Sci 14: 93-122.
- 46. Vaglenov A, Nosko M (1999) Genotoxicity and radio resistance in electroplating workers exposed to chromium. Mutat Res 446: 23-34.
- Van-Hummelen P, Severi M, Pauwels W, Roosels D, Veuleman H (1994) Cytogentic analysis of lymphocytes from fiber glass reinforced plastic workers occupationally exposed to styrene. Mutat Res 310: 157-165.
- 48. Wei Q, Gu J (1996) Benzo (a) pyrene diol epoxide induced chromosomal aberration and risk of lung cancer. Cancer Res 56: 3975-3979.
- 49. Werfel U, Langen V, Eickhoff I, Schoonbrood J, Vahrenholz C (1998) Elevated DNA singlestrand breakage frequencies in lymphocytes of welders exposed to chromium and nickel. Carcinogenesis 19: 413-418.
- 50. Wise JP, Loenard JC, Patierno SR (1992) Clastogenecity of lead chromate particles in hamster and human cells. Mutat Res 278: 69-79.
- 51. Wu FY, Tsai FJ, Kuo HW, Tsai CH, Wu WY (2000) Cytogenetic study of workers exposed to chromium compounds. Mutat Res 464: 289-296.
- 52. Xu J, Wise JP, Patierno SR (1992) DNA damage induced by carcinogenic lead chromate particles in cultured mammalian cells. Mutat Res 280: 129-136.
- Zhitkovich A, Lukanova A, Popov T, Taioli E, Cohen H (1996) DNAprotein crosslinks in peripheral lymphocytes of individuals exposed to hexavalent chromium compounds. Biomarkers 1: 86-93.