

Mustard Oil Cake (MOC) an Organic Fertilizer, its Toxicity and Response of *Channa punctatus*: A Review

Susanta Nath*

Department of Zoology, Government General Degree College Singur, University of Burdwan, West Bengal, India

ABSTRACT

Mustard Oil Cake (MOC) is a common bio fertilizer used widely in fish cultivation in almost all tropical countries. In sublethal concentration, the level of liver protein and lipid, hematological parameters, as well as brain Cholinesterase (ChE) activity of fish were affected due to the stress of this biofertilizer. Though *C. punctatus* has the ability to tolerate stressful situations, it is better to use MOC carefully during the preparation of the pond rather than direct application in the water for better production of the fish.

Keywords: Oil cake; Protein; Lipid; Blood; Cholinesterase

INTRODUCTION

The demand for aquatic products has increased significantly during recent decades internationally. Due to the increase in the climate crisis, biodiversity loss, and the recent pandemic, the target of making a hunger-free world is in an adverse situation. Along with the agro-food, aquatic food is also recognized as key food security and nutrition for the people globally. A total of 7% of animal protein intake daily by humans is coming from aquatic food, worldwide. The demand for aquatic products has increased significantly during the recent decades [1]. To reduce aquatic pollution, and stop aquatic biodiversity loss, attempts are made to use more organic fertilizers and organic pesticides for effective management of all fisheries and maintaining the food chain in the aquatic ecosystems. Mustard Oil Cake (MOC), an organic fertilizer, is used in pond fishery frequently. The study revealed that MOC could be utilized at 30% to 35% composition in the diet as a protein source for different herbivorous fishes including carp. Such application did not hamper the fish growth [2-4]. For the increase in production of fish, suitable and cheap locally available fish feed is required. The most Potential substitutes to fish meal in carp diets are meals containing oil seeds like MOC, linseed, and sesame oil, besides soya bean meal [5,6]. Application of the right amount of protein-rich MOC with rice bran, and vegetable wastes showed an increase in the production of fish [7,8]. Fishes are an important staple food in a large part of the world due to the presence of ample amounts of lipids and

proteins. Any effects of the xenobiotics on fish are devastating to humans [9]. Fish could respond to various biochemical and physiological strains that are secondary stress responses in comparison to the higher vertebrates [10]. Fish could recover from the stress in most cases [11]. Discreet use of biofertilizers in ponds increases productivity by producing autotrophs. Phytoplankton and algal populations increase in both rearing and stocking ponds [12-15]. However, it is essential to understand the effects on fish if they come in direct contact with MOC in the pond.

LITERATURE REVIEW

The work is conducted on the basis of previous research on the effect of MOC on fish. Different research papers and reports are considered to prepare this review.

Importance of MOC in fish farming

MOC is considered an important biofertilizer for the fish culture. 60% of the seed is converted into a byproduct as MOC during oil extraction [16]. It is used in preparing the ponds before the release of the fingerlings in the ponds. MOC contains 43% protein, 2.05% oil, 1.22% allylisothiocyanate, and 2.75% phytic acid with a considerable proportion of albumin, glutelin, and globulin [17,18]. The protein is rich in lysine and sulphurcontaining amino acids and is considered a protein supplement for human nutrition.

Correspondence to: Susanta Nath, Department of Zoology, Government General Degree College Singur, University of Burdwan, West Bengal, India, E-mail: nathsusanta2012@gmail.com

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Effects on protein and lipid content

When C. *punctatus* was exposed to water treated with sublethal doses of MOC, no mortality was observed after 35 days. Initially, the rate of increase of Total Liver Protein (TLP) was very slow, but a steady rate of increase of TLP was observed. Strength of association (ω^{3}) is estimated to measure the degree of relatedness between duration of Exposure (EP) and liver protein concentration. The computed values are 0.99 (EP=35)>0.95 (EP=07)>0.94 (EP=28)>0.82 (EP=21)>0.47 (EP=04)>0.38 (EP=14) (Table 1).

When C. *punctatus* was exposed to different doses of MOC, the liver showed a stress response initially. The liver showed an increase in sinusoidal space and lipidosis at its early exposure stage, followed by a recovery from the stress of MOC on the 28th day (Figure 1). After the initial setback, the growth rate i.e., weight, length, and breadth gradually increased from the 7th day, and weight gain was 9.64% on the 28th day. The muscle protein, Bone Morphogenetic Protein (BMP) of the fish also showed a similar trend. BMP was increased with the advance of days of exposure.

Effects on haematological parameters

The Total Count (TC) of RBC in non-treated and MOC-treated C. *punctatus* was $3.926 \pm 7.65 \ 10^6 \text{ mm}^3$ and $3.068 \pm 1.5 \ 106 \text{ mm}^3$ respectively after 4 days of exposure. But TC increased to $7.058 \pm 7.22 \ 10^6 \text{ mm}^3$ and $2.588 \pm 4.01 \ 10^6 \text{ mm}^3$ respectively after 28 days of exposure, which indicates a decrease in treated fish. Similarly, hemoglobin was $8.14 \pm 0.2\%$ and $7.44 \pm 0.05\%$. On 4th day in non-treated and treated fish, which increases to $12.06 \pm 0.09\%$ in non-treated and $4.94 \pm 0.09\%$ in treated fish

after 28 days. Differential Count (DC) of lymphocytes and neutrophils was found to increase after 28 days of exposure (Table 2).

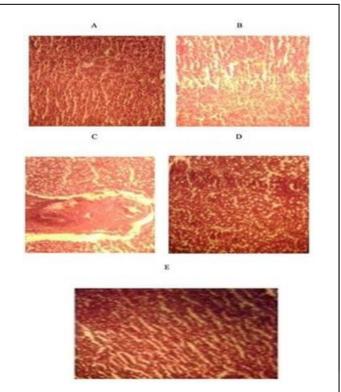


Figure 1: Liver of C. *punctatus* exposed to MOC. **Note:** (A) Control, (B) Exposed for 7 days, (C) Exposed for 14 days, (D) Exposed for 21 days, (E) Exposed for 28 days.

Statistics	Day 4	Day 7	Day 14	Day 21	Day 28	Day 35
F*	19.87	399.59	13.64	96.17	309.01	521.39
φ ²	0.47	0.95	0.38	0.82	0.94	0.99
t-test [†]	4.45	1998	3.69	9.8	17.57	72.21
Bonferroni modification	P<0.002	P<0.002	P<0.002	P<0.002	P<0.002	P<0.002

Note: *significant p<0.05; *significant p<0.001.

Table 1: Relations between control	ol and MOC treated C. punctatus [[19].
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Leucocytes	4 Days		7 Days		14 Days		21 Days		28 Days	
	С	Т	С	Т	С	Т	С	Т	С	Т
Lymphocytes	38	38	38	39	38	41	39	39	39	43
Eosinophils	38	9	10	10	12	10	12	11	13	9

Monocytes	17	17	16	15	13	13	10	10	9	14
Neutrophils	6	5	7	6	9	9	11	11	11	12
Basophils	16	16	15	15	11	11	8	9	6	9
Heterophils	9	10	9	10	11	10	12	12	13	7
Thrombocytes	4	5	5	5	6	6	8	8	9	6

Table 2: Differential count (%) showing variations in Control (C) and, MOC-Treated (T) C. punctatus.

Brain Cholinesterase (ChE) activity

ChE activity and body length have a significant correlation in C. *punctatus*, ChE activity has proportionality with the cell surface and varies with the body size of the fish. An initial increase in ChE activity in MOC-treated fish showed the tolerance of this animal against the effect of the biofertilizer used. Then ChE activity decreased, followed by an increase after 21 days and maintained up to 35 days in comparison to non-treated fishes. That was probably due to overcoming the capacity of the fish to survive in stressful situations.

DISCUSSION

MOC is rich in protein. The liver increases the synthesis of protein during prolonged exposure after overcoming the initial low rate of increase. The quantity of protein in the body depends upon the synthesis or degradation of proteins. Improper incorporation of the amino acids in the polypeptides also affects the quantity of proteins [20]. The protein level in the body also decreases due to the inhibitory activity of alkaline phosphatase [21]. The increase in the protein level in the body muscle indicates the fish's ability to compete with the initial opposing stress situation. This is due to more protein being used from the MOC of the medium to meet the increase in energy demand, and protein synthesis increases [22].

Survival mainly depends on the protein synthetic potential of animals [23]. An initial decrease in muscle and liver protein indicates an increase in proteolytic activity that is probably due to an increase in catabolic activity and a decrease in the anabolism of protein [24,25]. Such a response of fish is known as a secondary stress response in contrast to higher vertebrates [10]. Hepatic lipid level was found to decrease after 21 days of exposure in C. punctatus. Lipids provide energy in the metabolic process and energy reservoir of the body. Decreased liver lipid indicates the utilization of stored lipids to mitigate the demand during stressful situations [26]. However, liver lipid levels after 35 days indicate the recovery from the MOC effect [27]. Any variation in lipid profile will cause structural change in the cell membrane [28]. Any alteration of the chemical composition of the natural environment affects the physiological system of the aquatic inhabitants including fish [29,30].

Reduced TC of RBC was probably due to degeneration of erythropoietic tissue in the body or may be due to haemodilution [31,32]. Such reduction of RBC also reduces the Hb percentage in the blood. Solomon, et al. [33], also observed noticeable changes in haematological parameters in C. striata indicating slow recovery from the disadvantageous condition for fish. A gradual decrease in erythrocyte count and Hb indicates anaemia because of the breakdown of RBC due to the influx of MOC into erythrocytes as was observed in urea-treated H. fossilis [11]. MOC contains Allyl Isothiocyanate (AITA) and phytic acid. AITA helps the plant to defend itself from herbivores and phytic acid chelates multivalent metal ions like iron. Salt is produced which does not absorb properly from the intestine and the body suffers from the availability of minerals [34]. The decreased level of haemoglobin in MOC-treated fish is also probably due to less availability of iron. Variation of MCH in MOC-treated fish is probably due to swelling of erythrocytes or release of young RBC containing less haemoglobin in the blood circulation [35-37].

A decrease in the ChE activity in fish exposed to xenobiotics is not suitable for survival. 70 to 80% inhibition of ChE is considered to be critical for fishes [38,39]. C. *punctatus* has the ability to tolerate MOC as was observed in organophosphate-treated fish and such fast recovery may be due to the continuous synthesis of essential enzymes [40-43].

CONCLUSION

The study revealed that the application of Mustard Oil Cake (MOC) has a significant stress-inducing impact on *C. punctatus*. So, direct application in the pond water is not advisable, because of its impact on the different physiological parameters on fish bodies. Though *C. punctatus* has the ability to recover from stressful situations, but scrupulous use of MOC during the preparation of ponds for fish culture may increase the production, provided that its potential adverse effects on the fish's well-being are appropriately managed and monitored.

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CONFLICT OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

- 1. FAO. The state of world fisheries and aquaculture. Towards Blue Transformation. 2022.
- Paul BN, Sarkar S, Mukhopadhyay PK. Comparative effect of varied dietary protein sources on growth performance of rohu (*Labeo rohita* Hamilton). J Aquac. 1999;7:11-15.
- 3. Gangadhara B, Nandeesha MC, Keshavanath P, Singh KP, Manissery JK. Evaluation of rapeseed meal as a feed ingredient in catla (*Catla catla*) diets. J Aquac Trop. 2002;17:261-72.
- Mohanta KN, Mohanty SN, Jena J, Sahu NP. Effect of three different oil cake-based diets on pond production performance of silver barb, *Puntius gonionotus* (Bleeker). Aquac Res. 2008;39(11): 1131-1140.
- 5. El-Sayed AF. Alternative dietary protein sources for farmed tilapia, Oreochromis spp. Aquac. 1999;179(1-4):149-168.
- Jahan DA, Hussain L, Islam MA, Khan M. Comparative study of mustard oil cake and soybean meal based artificial diet for Rohu, *Labeo rohita* (Ham.) fingerlings. Agric. 2013;11(1):61-66.
- Ghosh K, Mandal S. Nutritional evaluation of groundnut oil cake in formulated diets for *rohu*, *Labeo* rohita (Hamilton) fingerlings after solid state fermentation with a tannase producing yeast, *Pichia kudriavzevii* (GU939629) isolated from fish gut. Aquac Rep. 2015;2:82-90.
- Bhandari S, Kaphle K, Lamsal RK. Local feeds in aquaculture and their feeding efficiency: Review from Nepal. Int J Vet Sci Anim Husb. 2019;4:6-9.
- Srivastava P, Singh A, Pandey AK. Pesticides toxicity in fishes: Biochemical, physiological and genotoxic aspects. Biochem Cell Arch. 2016;16(2):199-218.
- Mazeaud M, Mazeaud F. Adrenergic responses to stress in fish. In: Pickring AD (Eds.) stress and fish. Academic Press, New York and London.1981.
- 11. Maitra S, Nath S. Toxic impacts of urea on the hematological parameters of air breathing fish *Heteropneustes fossilis* (Bloch). Am Eur J Agricul Environ Sci. 2014;14(4):336-342.
- Hepher B, Pruginin Y. Commercial fish farming with special reference to fish culture in Israel. NewYork: JohnWiley and Sons. 1981.
- New M, Fedoruk K. Reports on the regional workshop on aquaculture planning in Asia, Bangkok, Thialand. ACDP/REP/ 76/2; 2003.
- Bhakta JN, Bandyopadhyay PK, Jana BB. Effect of different doses of mixed fertilizer on some biogeochemical cycling bacterial population in carp culture pond. Turk J Fish Aquat Sci. 2006;6(2).
- 15. Palafox JTP, Figueroa JLA, Vargasmachuca SGC, Chavez GER, Valle AUB, De Dios MAR, et al. The effect of chemical and organic fertilization on Phytoplankton and fish production in carp (*Cyprinidae*) polyculture system. CONACYT. 2010.
- Sehwag S, Das M. A brief overview: Present status on utilization of mustard oil and cake. NIScPR. 2015.
- 17. Niazi AH. Improvement in the nutritive value of mustard seed cake. Doctoral dissertation, Pakistan. 1986.

- Klockeman DM, Toledo R, Sims KA. Isolation and characterization of defatted canola meal protein. J Agricul Food Chem. 1997;45(10): 3867-3870.
- 19. Nath S, Rakshit P, Matozzo V. Effects of mustard oil cake on liver proteins of (Bloch). Interdiscip Toxicol. 2018;11(3):200-203.
- Singh A, Singh DK, Misra TN, Agarwal RA. Molluscicides of plant origin. Biol Agric Hortic. 1996;13(3):205-252.
- Ibrahim AM, Higazi MG, Demian ES. Histochemical localization of alkaline phosphatase activity in the alimentary tract of the snail *Marisa comuarietis* (L.). Bull Zool Soc Egypt. 1974;26:94-105.
- Mondal S, Das R, Das AC. A comparative study on the decomposition of edible and non-edible oil cakes in the gangetic alluvial soil of West Bengal. Environ Monit Assess. 2014;186:5199-5207.
- Rohankar P, Zade V, Dabhadkar D, Labhsetwar N. Evaluation of impact of phosphamidon on protein status of freshwater fish *Channa punctatus*. Indian J Sci Res. 2012;3(1):123-126.
- Mastan SA, Rammayya PJ. Biochemical profile of Channa gachua (Ham) exposed to sublethal doses of Dichlorovas (DDVP). Int J Toxicol. 2010;8(1):27-32.
- 25. Ghosh D, Bhattacharya S, Mazumder S. Perturbations in the catfish immune responses by arsenic: Organ and cell specific effects. Comp Biochem Physiol C Toxicol Pharmacol. 2006;143(4):455-463.
- Shruti SG, Tantarpale VT. Effects of cypermethrin on lipid and cholesterol contents of freshwater fish *Channa orientalis* (Bloch). Ind J Res. 2014a.;3(8):200-201.
- Nath S, Saha C, Bhowmick HS, Matozzo V. Effects of organic fertilizer on hepatic lipid levels and cholinesterase activity in *Channa punctatus* (Bloch). Philippine J Sci. 2016;145(4):413-418.
- Javed M, Usmani N. Stress response of biomolecules (carbohydrate, protein and lipid profiles) in fish *Channa punctatus* inhabiting river polluted by thermal power plant effluent. Saudi J Biol Sci. 2015;22(2):237-242.
- Radhaiah V, Girija M, Rao KJ. Changes in selected biochemical parameters in the kidney and blood of the fish, *Tilapia mossambica* (Peters), exposed to heptachlor. Bull Environ Contam Toxicol. 1987;39(6):1006-1011.
- Nath S, Prasad S, Matozzo V. Effects of mustard oil cake on haematological parameters of the freshwater fish Channa punctatus. Intl J Appl Environ Sci. 2017;12(5):839-848.
- 31. Hota S. Toxic effect of arsenic on haemato biochemical abnormalities in *Channa punctatus* (Bloch). J Ecotoxicol Environ Monit. 1995;5(4):249-255.
- 32. Sampath K, Velamman S, Kennedy IJ, James R. Haematological changes and their recovery in *Oreochromis mossambicus* as a function of exposure period and sublethal levels of Ekalux. Acta Hydrobiologica. 1993;1(35):73-83.
- Solomon SG, Okomoda VT. Effect of photoperiod on some biological parameters of *Clarias gariepinus* juvenile. J Stress Physiol Biochem. 2012;8(4):47-54.
- 34. Zhou JR, Erdman Jr JW. Phytic acid in health and disease. Crit Rev Food Sci Nutr. 1995;35(6):495-508.
- 35. Sobecka E. Changes in the iron level in the organs and tissues of wels catfish, *Silurus glanis* L. caused by nickel. Acta ichthyologica et piscatoria. 2001;31(2):127-143.
- Pathiratne AL, Chandrasekera LW, De Seram PK. Effects of biological and technical factors on brain and muscle cholinesterases in Nile tilapia, *Oreochromis niloticus*: Implications for biomonitoring neurotoxic contaminations. Arch Environ Contam Toxicol. 2008;54:309-317.
- 37. Koenig S, Solé M. Muscular cholinesterase and lactate dehydrogenase activities in deep-sea fish from the NW Mediterranean. Mar Environ Res. 2014;94:16-23.

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- Coppage DL, Matthews E, Cook GH, Knight J. Brain acetylcholinesterase inhibition in fish as a diagnosis of environmental poisoning by malathion, O, O-dimethyl S-(1,2-dicarbethoxyethyl) phosphorodithioate. Pestic Biochem Physiol. 1975;5(6):536-542.
- 39. Das BK, Mukherjee SC. Toxicity of cypermethrin in *Labeo rohita* fingerlings: Biochemical, enzymatic and haematological consequences. Comp Biochem Physiol C Toxicol Pharmacol. 2003;134(1): 109-121.
- 40. Fulton MH, Key PB. Acetylcholinesterase inhibition in estuarine fish and invertebrates as an indicator of organophosphorus insecticide exposure and effects. Environ Toxicol Chem. 2001;20(1):37:45.
- 41. Ferrari A, Venturino A, de D'Angelo AM. Time course of brain cholinesterase inhibition and recovery following acute and subacute azinphosmethyl, parathion and carbaryl exposure in the goldfish (*Carassius auratus*). Ecotoxicol Environ Saf. 2004;57(3):420-425.
- Murphy LA, Lewbart GA, Meerdink GL, Bargren GL. Wholeblood and plasma cholinesterase levels in normal koi (*Cyprinus carpio*). J Vet Diagn Invest. 2005;17(1):74-75.
- Nath S, Matozzo V, Bhandari D, Faggio C. Growth and liver histology of *Channa punctatus* exposed to a common biofertilizer. Nat Prod Res. 2019;33(11):1591-1598.