Synergistic Effects of IL-17 and IL-6 on Inflammatory Chemokines: The Inflammation Amplifier

IL-6 is a classical proinflammatory cytokine first cloned in 1986 and IL-6 blockade has shown tremendous effects in the treatment of rheumatoid arthritis [1,2]. In fact, many inflammatory diseases beyond rheumatoid arthritis, including multiple sclerosis and type-1 diabetes have been genetically associated with IL-6 [3,4]. Its effects were traced to the activation of Th17 cells, a subset of activated CD4+ T cells [5,6]. We then have investigated the relationship between IL-6 and Th17 cells in autoimmune diseases using mouse models. IL-6 deficient mice are resistant to many inflammatory diseases, in particular to Th17-mediated ones such as collagen-induced arthritis and experimental autoimmune encephalomyelitis [7-10]. Additionally, in vivo experiments found that serum levels of IL-6 significantly increase after forced expression of IL-17 [9,11,12], suggesting positive feedback of IL-6 in the presence of IL-17 signaling. Further in vitro studies revealed that a combination of IL-17 and IL-6 stimulation induces much larger amounts of IL-6 and inflammatory chemokine production in type I collagen+ non-immune cells such as fibroblasts and endothelial cells, than stimulation by either cytokine alone. This combination effect is not additive but synergistic [9] (Figure 1A). We have also shown that the synergistic effect of IL-17 and IL-6 is dependent on two transcriptional factors: NF-κB and STAT3. Thus, it is possible that IL-17 from Th17 cells induces excess chemokines via an IL-6 positive-feedback loop in non-immune cells (Figure 1B). We named this phenomenon the “inflammation amplifier” and showed it to be important for the pathogenesis of the rheumatoid arthritis model, F759-arthritis, the multiple sclerosis mouse model, experimental autoimmune encephalomyelitis (EAE), and in a mouse model of chronic rejection of lung transplantation [9,13]. We further hypothesized that IL-6 acts as a fuel for the inflammation amplifier and that chemokines are functional molecules for the development of inflammation. In addition, an immunohistological study using human clinical samples revealed evidence that activation of the inflammation amplifier occurs in the affected area of bronchiolitis obliterans, a common complication via chronic inflammation after lung transplantation [14]. Additionally, genes that regulate the inflammation amplifier or are targeted by it have been functionally screened and identified. These genes were reported to have significantly high levels of genetic associations with a broad range of human diseases and disorders including autoimmune diseases, metabolic syndromes, and other inflammatory diseases [15]. Thus, the inflammation amplifier is involved not only in autoimmunity, but also other inflammatory diseases, with IL-17 being an important factor in driving the inflammation amplifier by activating NF-κB in non-immune cells [16,17].

A Th17-dependent Model of Multiple Sclerosis

Inflammatory cytokines such as IL-1, IL-6, IL-17 and TNFα are known to play key roles in the pathogenesis of multiple sclerosis and its animal model, EAE [18]. We have reported the inflammatory amplifier is also an intrinsic part of the molecular mechanism responsible [9,19].
Using EAE as our model, we extended this work to examine how autoreactive CD4+ T cells enter the central nervous system (CNS), considering that the blood-brain barrier (BBB) tightly regulates such transport. We established a Th17-cell dependent adaptive transfer model of EAE, because we thought that more common models by using autopeptide-immunizations in the presence of complete Freund's adjuvant risk modulating a CNS basal state. Th17 cells obtained from myelin oligodendrocyte glycoprotein (MOG)-immunized mice were re-stimulated in vitro with IL-23 and infused intravenously in naive C57BL/6 recipients so that quiescence of the CNS was preserved. Mice lacking IL-6 signal transducer (gp130) or STAT3 in blood vessel endothelial cells do not show this accumulation of MOG-specific Th17 cells at the L5 cord, suggesting the inflammation amplifier in L5 dorsal vessels produces Ccl20. Intriguingly, along with the diseased state, a low-grade activation of the inflammation amplifier, namely elevated levels of Ccl20 and many other chemokines, is evident at L5 vessels even in the healthy state, regardless of pathogenic Th17 cell transfer [19]. This finding led us to hypothesize that some L5 neurons may be constantly activated. It is known that sensory neurons located in DRG at the L5 region are connected to the soleus muscles, which are the muscles primarily responsible for coping with gravity [28]. We speculated that constant stimulation of the soleus muscles by gravity might activate the inflammation amplifier via their sensory nerves. Indeed, when mice were tail suspended so that the hind legs were released from the gravity stimuli, MOG-specific Th17 cells no longer accumulated at the L5 region. Instead, these Th17 cells accumulated at cervical cords as if a new “gate” was opened by gravity stimuli to the forearm muscles. Consistently, the tail suspension suppressed Ccl20 expression in the L5 dorsal blood vessels and decreased the expression donor T cell preparation, and/or time points of sampling. The L5 entry of pathogenic Th17 cells was shown to depend on IL-17 production, as IL-17-deficient Th17 cells obtained from MOG-immunized IL-17-deficient mice did not invade the L5, while IFNγ-deficient Th17 cells did. The antigen specificity of Th17 cells is another critical determinant for CNS invasion, as CNS-irrelevant, ovalbumin-specific Th17 cells did not efficiently traverse the BBB (our unpublished data and [21]). Although how and where this antigen presentation takes place is currently unknown, we propose that cognate TCR stimulation around the L5 region promotes IL-17 secretion from the transferred pathogenic Th17 cells. The IL-17 loosens the BBB by downregulating tight junction molecules [22], allowing the Th17 invasion in the L5 cord. Even if so, this explanation does not explain why MOG-specific Th17 cells are preferentially recruited to L5.

The cervical, thoracic and lumbar cords of mouse can be divided into 7, 13 and 6 segments, respectively [23,24]. Sensory and motor neurons extend from each spinal segment to respective body regions and transmit sensations to the brain or control movements of the respective body parts. MOG-specific Th17 cells are selectively recruited to dorsal blood vessels in the L5 spinal cord at the early stages of EAE [19]. It has been reported that Ccl20 is a chemokine for Th17 cell migration and is upregulated via NF-κB activation [25-27]. Additionally, we found that NF-κB activity is increased in L5 dorsal vessels compared to vessels in the other spinal segments. Mice lacking IL-6 signal transducer (gp130) or STAT3 in blood vessel endothelial cells do not show this accumulation of MOG-specific Th17 cells at the L5 cord, suggesting the inflammation amplifier in L5 dorsal vessels produces Ccl20. Intriguingly, along with the diseased state, a low-grade activation of the inflammation amplifier, namely elevated levels of Ccl20 and many other chemokines, is evident at L5 vessels even in the healthy state, regardless of pathogenic Th17 cell transfer [19]. This finding led us to hypothesize that some L5 neurons may be constantly activated. It is known that sensory neurons located in DRG at the L5 region are connected to the soleus muscles, which are the muscles primarily responsible for coping with gravity [28]. We speculated that constant stimulation of the soleus muscles by gravity might activate the inflammation amplifier via their sensory nerves. Indeed, when mice were tail suspended so that the hind legs were released from the gravity stimuli, MOG-specific Th17 cells no longer accumulated at the L5 region. Instead, these Th17 cells accumulated at cervical cords as if a new “gate” was opened by gravity stimuli to the forearm muscles. Consistently, the tail suspension suppressed Ccl20 expression in the L5 dorsal blood vessels and decreased the expression donor T cell preparation, and/or time points of sampling. The L5 entry of pathogenic Th17 cells was shown to depend on IL-17 production, as IL-17-deficient Th17 cells obtained from MOG-immunized IL-17-deficient mice did not invade the L5, while IFNγ-deficient Th17 cells did. The antigen specificity of Th17 cells is another critical determinant for CNS invasion, as CNS-irrelevant, ovalbumin-specific Th17 cells did not efficiently traverse the BBB (our unpublished data and [21]). Although how and where this antigen presentation takes place is currently unknown, we propose that cognate TCR stimulation around the L5 region promotes IL-17 secretion from the transferred pathogenic Th17 cells. The IL-17 loosens the BBB by downregulating tight junction molecules [22], allowing the Th17 invasion in the L5 cord. Even if so, this explanation does not explain why MOG-specific Th17 cells are preferentially recruited to L5.
of the neural activation marker, c-fos, in L5 DRG. Moreover, electric stimulations to the soleus muscles of tail-suspended mice restored Ccl20 expression, Th17 accumulation and c-fos levels at the L5 cord [19] (Figure 2). These results strongly suggest that neural inputs from soleus muscles in response to gravity play a role in activating the inflammation amplifier and lead to the expression of various chemokines including Th17-attracting CCL20 in L5 dorsal blood vessels.

What mechanisms do afferent sensory neurons from the soleus muscle use to regulate the status of blood vessels at L5? Although a precise neural network remains elusive, we have shown sympathetic nerves to be involved. Blood flow speeds at L5 dorsal vessels become slower when mice are tail suspended, while electronic stimulation of the soleus muscles increases the flow speed suggesting that autonomic nerves including sympathetic ones are involved in the response. On the other hand, blood flow speeds in blood vessels other than the L5 region, such as femoral vessels, brain surface vessels and the portal vein, are not affected by tail suspension. Furthermore, treatment with atenolol, an α1 adrenergic receptor antagonist, or prazosin, a β1 adrenergic receptor antagonist, significantly inhibits Ccl20 mRNA expression, NF-κB activation and MOG-reactive Th17 accumulation at L5 vessels and also suppresses clinical signs of EAE [19]. It is also known that the main adrenergic receptors associated with blood vessels are α1 and β2 ones. We hypothesize that the β2 receptor might play a role just like the β1 receptor, because both β1 and β2 receptors activate NFκB pathway [29-31], which is the main pathway of the inflammation amplifier activation. Therefore, it is important to check whether the β2 receptor might play a role by using the inhibitors in our system.

Consistent with these in vivo results, the addition of norepinephrine, which is a neurotransmitter from sympathetic neurons, to a culture of endothelial cell lines, enhances the inflammation amplifier based on IL-6 and Ccl20 expressions. Thus, anti-gravity responses of the soleus muscles lead to sympathetic nerve stimulation, creating a gateway for immune cells to pass through the CNS via L5 dorsal vessels [19]. However, we do not comprehensively describe the link between the sensory activation and the L5 vascular changes, which makes it unclear if the locally released neurotransmitters directly affect the BBB integrity or the Th17 activity. Based on these findings, we proposed that MOG-reactive, disease-causing Th17 cells make use of the L5 gateway to infiltrate the CNS and induce local inflammation by producing cytokines like IL-17, which further induces chemokines via the inflammation amplifier in parenchymal non-immune cells and results in chronic inflammation in the CNS (Figure 3).

**The Gate Theory**

Thigh muscles including the quadriceps are known to be regulated by L3 DRG neurons. Interestingly, electronic stimulation of these muscles led to an increased expression of Ccl20 in L3 cord vessels in mice. In a similar fashion, chemokine levels in the fifth cervical to fifth thoracic (C5-T5), third lumbar (L3), and fifth lumbar (L5) cords, respectively. This effect offers possible targets for the manipulation of immune cell migration in vivo.

**Figure 4: The gate theory**

Electric pulses to the upper arm, thigh, and soleus muscles induce activation of the inflammation amplifier: chemokine upregulation, opening a gate in the dorsal vessels of the fifth cervical to fifth thoracic (C5-T5), third lumbar (L3), and fifth lumbar (L5) cords, respectively. This effect offers possible targets for the manipulation of immune cell migration in vivo.

**References**
Complementary DNA for a novel human interleukin (BSF-2) that induces B lymphocytes to produce immunoglobulin. Nature 324: 73-76.


