

The Conjugative Transfer of the Multiresistance Gene Between Bacteria is Significantly Promoted by Nano-Alumina

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Abstract

Recently, the potential risks of nanomaterials and the spread of antibiotic resistance have become two important environmental concerns. The conjugative transfer of antibiotic resistance genes between bacteria is the most important pathway for the acquisition of antibiotic resistance by bacteria. Both environmental and genetic factors influence the conjugative transfer of antibiotic resistance genes in bacterial populations. The extent to which nanomaterials are able to bring about an increase in antibiotic resistance by regulating the conjugative transfer of antibiotic resistance genes in bacteria is still unknown. In this paper, an Orthogonal Design $L_{64}(4^{21})$ was used in duplicate to evaluate the effects of bacterial concentration, nano-alumina concentration, mating time, mating temperature and the interactions of those factors on the conjugation transfer in LB broth. The mechanisms by which nano-alumina promote the horizontal transfer of antibiotic multiresistance features were explored by morphological, biochemical, and molecular biological methods. We have shown that nano-alumina promotes the horizontal conjugative transfer multiresistance genes mediated by RP4 up to 250-fold in LB broth and 100-fold in PBS. And it would appear that the effect of promoting conjugative transfer of nano-alumina exceeds the effects of mating temperature and mating time. We also explored the mechanisms behind this phenomenon and demonstrated that nano-alumina is able to induce oxidative stress, cause the damage of bacterial cell membranes, enhance the transcriptional activity of conjugative genes and depress the global regulatory factor genes expression. The findings in this study support the notion that nano-alumina in the environment could result in ecological hazards. More important, an enhanced rate of plasmid transfer among micro-organisms may have an enormous impact on human health and environmental safety.

Introduction

Over the past few decades, microorganisms have developed a greater ability to withstand the effects of antibiotics due to several factors, which include the overuse or misuse of drugs in medicine and agriculture [1]. The development and spread of antimicrobial resistance in bacterial populations is of increasing concern, as it can lead to major difficulties in the treatment of diseases. These are exemplified by the existence of methicillin-resistant *Staphylococcus aureus* (MRSA) [2], and particularly the appearance of the new "Super Bug" in 2010 and its dissemination in all the world [3,4]. Many antibiotic resistance genes reside on plasmids. These facilitate the transfer of genetic information in a horizontal fashion between individual cells or species by plasmid exchange within their natural environment [5]. Of all the mechanisms and mobile elements that mediate horizontal gene transfer between bacteria, conjugation by self-transferable plasmids may be the most important [6]. The critical role that plasmids play in the rapid spread of antibiotic resistance genes is particularly salient [7].

Experimental data has shown that a number of environmental factors, including exposure to antimicrobial agents [8], as well as nutrient and water availability, can influence this horizontal plasmid transfer [9].

Nanomaterials, which often possess novel physical, chemical, and biological properties, are being utilised in an increasing number of fields. The use of nanoparticles (NPs) in commercial products and industrial applications has increased greatly in recent years, although understanding of the interaction mechanisms at the molecular level between NPs and biological systems is largely lacking [10]. In some of those products, such as skin creams and toothpastes, NPs can enter the water environment on a continual basis from the removal of such products, such as by hand-washing [11]. Even worse, a fatal accident during the production of engineered nanomaterials could release a

significant quantity of NPs into the water environment [12]. At the same time, scientists have also found ways of using nanomaterials in environmental remediation. Many of these are still in the testing phase [13-15]. One example is nano-alumina, which can adsorb and oxidise contaminants. Nano-alumina is already used to minimise the concentrations of contaminants in water and waste water, and to remove humic-acids from water and to improve sludge conditioning [16-19]. Their use in water treatment may result in a considerable amount of nanomaterial residue in the water [20,21]. Otherwise, nanosized materials at the same time they may, in fact, become new environmental hazards themselves.

The potential hazards associated with nanomaterials in the environment have resulted in heightened awareness and concern for their impact on health worldwide [12,22,23]. Moreover, some studies have indicated that nanomaterials can cause disruption to bacterial membranes or substantial damage to DNA, probably by the production of reactive oxygen species (ROS) [24,25] and can deliver DNA or RNA molecules into animal or plant cells [26,27]. Based on the above, we hypothesised that nanomaterials that are present in water

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may promote the horizontal transfer of multidrug-resistance genes between bacteria by acting on cell membranes and/or regulating genes involved in plasmid transfer.

In this study, we described the effect of nano-alumina on the horizontal transfer of antibiotic multiresistance genes mediated by RP4 between *Escherichia coli*. We analysed the effect of bacteria concentration, concentration of nano-alumina, mating time, and mating temperature on conjugation using orthogonal design which are fractioned factorial designs that allow testing multiple independent processes variables within a single experiment and were used by many scholars [28-31]. We also explored the mechanisms by which nano-alumina promote the horizontal transfer of antibiotic multiresistance features by morphological, biochemical, and molecular biological methods. These studies were the first to identify that the presence of nano-alumina in water resulted in a very significant increase in interspecies conjugation of the mobilisable shuttle plasmid RP4.

Experimental Section

Bacteria conjugation experiment based on orthogonal experimental design

An Orthogonal Design $L_{64}(4^2)$ was used in duplicate to evaluate the effects of bacterial concentration, nano-alumina concentration, mating time, mating temperature and the interactions of those factors on the conjugation transfer in LB broth. As our results of preliminary experiments showed that the pH value of common water environment had no significant effect on conjugal transfer, the pH value was not adopted in the orthogonal experiment. The levels of the bacteria concentration were 10^5 , 10^6 , 10^7 and 10^8 CFU/mL. The levels of nano-alumina concentration were 0, 0.5, 5 and 50 mmol/L. The levels of the mating time were 2, 4, 6 and 8 h, and the levels of the mating temperature were 4, 15, 25 and 37°C. The orthogonal experiment scheme is shown in Supplementary Table 1.

The *E. coli* strain HB101 [32], which contains the RP4 plasmid carrying kanamycin resistance, served as donor bacteria for conjugation. *E. coli* K12 (ATCC 47076) acquired a nalidixic acid-resistant mutant, K12Nal [33,34], which was used as the recipient strain. Strains were grown at 37°C in LB broth with appropriate antibiotics, as follows: kanamycin (Km, product number K1377-25G [Sigma-Aldrich, 3050 Spruce St. St. Louis, MO 63103, USA]), 60 mg/L for donor *E. coli* HB101 bearing RP4 plasmid, and nalidixic acid (Nal, product number N8878-25G, Sigma-Aldrich) 100 g/L was used for the *E. coli* K12.

Sixty-four sterile glass tubes (Diameter: 18 mm, and Length: 180 mm) were marked with a number from 1 to 64, respectively. Each number corresponded to an experimental condition for one group in the orthogonal experiment. For each experiment, donor and recipient strains were grown separately over night at 37°C in LB with the appropriate antibiotics. Equal amounts of donor and recipient cells were then mixed. To prepare this initial 50:50 cell mixture, the optical densities (OD600) of the overnight cultures were measured and appropriate culture volumes were taken to ensure the cells concentration fit the experimental conditions. The bacteria cells were centrifuged for 1 min at 10,000 g at room temperature, and the resulting pellets were re-suspended in 1 ml of saline solution (NaCl, 8.5 g/L) and re-centrifuged. The new pellets were re-suspended in LB in a volume determined by the average of the initial culture volumes and then mixed together. Then, 9.5 ml of the suspension was aliquoted into sterile glass tubes marked with the number that corresponded to the experimental condition. Then, 0.5 mL dispersions of nano-alumina

(product number 642991, 100 mL, Sigma-Aldrich, particle size is <50 nm (TEM), diluted with sterile water to a concentration corresponded to the experimental condition) or sterile water was aliquoted into the glass tubes. The culture was vigorously mixed by vortexing.

After all the 64 group experiments were prepared, the cultures were incubated in different temperature incubators and different times, according to the experimental design. The specified cultures were vigorously mixed and appropriate dilutions were spread on LB selection plates (containing 60 mg/L Km and 100 mg/L Nal). After overnight incubation of the plates at 37°C, the colonies were counted and the results were presented as colony forming units per milliliter culture (CFU/mL).

In parallel, the HB101 and K12 bacteria were also plated onto LB selection plates (containing 60 mg Km and 100 mg Nal) to ensure that the bacteria growing on the double-antibiotic agar medium were transconjugants to rule out the spontaneous mutation of HB101 and K12. A further control group was established using bulk alumina (product number H7881-10G, particle size is mean 5 μ m Sigma-Aldrich) in order to confirm the effects of the nano-structure of alumina on the conjugative transfer of RP4. Finally, HB101 bacteria bearing a non-conjugative plasmid pCB182 [35] was used as the donor bacteria to observe the transformation of free plasmids dissolved from died bacteria in the presence of nano-alumina.

A further orthogonal experimental was carried out to evaluate the effects of bacterial concentration, nano-alumina concentration, mating time, mating temperature and the interactions of those factors on the conjugation transfer in Phosphate Buffered Saline (PBS, Ingredients: NaCl 137 mmol/L, KCl 2.7mmol/L, Na_2HPO_4 10mmol/L, KH_2PO_4 2mmol/L, pH7.4). The experimental design and experimental procedures were the same as the experiment carried out in LB broth, but the media was changed to PBS. All of the above-described control groups were also carried out in parallel.

All the strains and plasmid used in this study are shown in Supplementary Table 2.

Samples preparation for TEM, anti-oxidant systems detection, transcriptional activity detection and real-time PCR

To prepare samples for further experiments, the bacteria mating experiments based on Factorial Design with two factors was performed. The protocols were as follows: all the strains were incubated and treated as described in the "Bacteria conjugation experiment based on orthogonal experimental design" section. The cell density was adjusted to an optical density at 600 nm (OD600) of 0.1, and then donor and recipients were mixed 50:50 in LB medium. In the same time, a specified concentration and volume dispersions of nano-alumina or bulk alumina were aliquoted into the LB medium, and the final concentrations of alumina were 0, 0.05, 0.5, 5 and 50 mmol/L. After incubated at 37°C for 8 h, the mating mixtures were centrifuged at 1,000 g for 20 min at 4°C. Then, the supernatant of each sample was completely removed, and the cell pellet was used for further experiments. In order to prepare samples for the detection of transcriptional activity, the donor strains used in the conjugation experiments were HB101 (RP4+PCB267.1) and HB101 (RP4+PCB267.2).

The cell pellet was re-suspended gently with the pre-cooled 2.5% glutaraldehyde fixative. Then the samples were fixed at 4°C for 2 hours. Next, the liquid was centrifuged at 1,000 g for 20 min, after which the clarified supernatant was removed. The cell pellet was washed twice with

0.2 mol/L citric acid solution and then centrifuged again. The samples were then fixed with 2% OsO₄ in anhydrous acetone that contained 8% dimethoxypropane for two days at 4°C, followed by dehydration using gradient ethanol method. Afterwards, the precipitate was treated with propylene oxide, and then embedded in Epon812 resin. The samples were sectioned into slices about 50 nm with ultra microtome (LEICA ULTRACUT-R). Following post-staining with 2% aqueous uranyl acetate for 15 min and Reynold's lead citrate for 5 min, the samples were examined and photographed with a HITACHI-7500 transmission electron microscope and Megaview digitalisation electron microscope photograph system. The elements in the bacterial slice were analysed by high-resolution TEM (Philips TECNAI G2F20) with energy dispersive analysis system of X-ray (GENESIS).

Detection of bacterial anti-oxidant systems

The cell precipitation which contained both donor and recipient was washed with PBS twice. Then cell precipitation was re-suspended with 1 mL PBS. The cell suspension was broken into pieces with ultrasonic cracker (JY92-IIN). The suspension was centrifuged at 10,000 g for 10 min at 4°C. Then, the protein fraction of the supernatant was quantified with a protein quantification kit (SK3051, Shanghai Sangon Biological Engineering Technology & Services Co., Ltd.), and the levels of OH•, T-AOC, SOD, CAT and GR were analysed. The activity levels of these enzymes were tested with appropriate kits from the Nanjing Jiancheng Bioengineering Institute (Nanjing, China).

Construction of transcriptional fusions and detection of transcriptional activity

The *trbBp* and *trfAp* regions of RP4 plasmid were amplified by PCR using primers that contained *Bgl*III restriction sites (Supplementary Table 3), upon which *Bgl*III (Takara, Kyoto, Japan) was added. The primers and reaction conditions for the amplification of *trbBp* and *trfAp* were according to a previous study [36].

These two PCR products were cloned into the pTZ57R/T vector (Fermentas, USA). After verifying the correct promoter regions through PCR amplification and sequence analysis, these plasmids were called pTZ57R.1 (containing *trbBp*) and pTZ57R.2 (containing *trfAp*), respectively.

The method of cloning *trbBp* and *trfAp* into pCB267 [37] vector was as follows. The pTZ57R.1 and pTZ57R.2 were cultured overnight in LB culture medium that contained 50 mg/L ampicillin (Amp). The plasmids were then extracted, purified, and restricted at 37°C for 1 hour with *Bgl*III. Subsequently, 5 µL of the target gene sequence and plasmid vector that were acquired after restriction and purification were ligated overnight. As a consequence, we were able to obtain pCB267.1 (containing *trbBp*) and pCB267.2 (containing *trfAp*). Samples that contained 5 µL aliquots of purified pCB267.1 or pCB267.2 were transformed into competent *E. coli* HB101 (RP4), shaken for 1 hour at 37°C, transferred onto LB plates with 60 mg/L Km and 50 mg/L Amp, and then cultured overnight at 37°C. Red colonies which pCB267.1 or pCB267.2 were transformed successfully were selected and cultured in LB medium that contained 60 mg/L Km and 50 mg/L Amp. Positive clones were identified by amplification through PCR of the *trbBp* and *trfAp* genes from the plasmids. The two strains containing two plasmid called HB101 (RP4+PCB267.1) and HB101 (RP4+PCB267.2).

The cell pellets were used to detect the transcriptional activities of *trbBp* and *trfAp* with a kit (GMS60003.3 V.A GENMED SCIENTIFICS INC. U.S.A).

Detection of mRNA expression of global regulatory genes

The total bacterial RNA was extracted from the cell pellet with a RNA extraction kit (Invitrogen, USA). Subsequently, the RNA was reverse-transcribed into DNA by RT-PCR with a reverse transcription kit (Invitrogen, USA). Real-time PCR was carried out using SYBR Green I (TaKaRa, Japan), and the absolute quantification method was used to quantify the global regulatory genes: *KorA*, *KorB* and *trbA* of RP4. The 16s rRNA was used as the internal control. The primers were designed specifically for this study using Oligo 6.0, and primer sequences are listed in Supplementary Table 4. Regulatory gene primers and 16s rRNA primers were synthesised by Invitrogen. The reaction conditions used for the two-step PCR procedure are as follows: an initial denaturation at 95°C for 30 sec, a PCR step of 40 cycles of 95°C for 5 s and 60°C for 31 s, and a dissociation stage of 95°C for 15 s, 60°C for 1 min, and 95°C for 15 s. The real-time PCR equipment was Applied Biosystems 7300 Real-Time PCR System, and the software was SDS v1.3.2.

Statistical analyses

All data were analysed using SAS 9.2 software version for windows (SAS (r) Proprietary Software 9.2 (TS2M2), Licensed to Academy of Military Medical Sciences, Site 11201240). The bacterial conjugation experimental results were analysed by the "analysis of variance of univariate quantitative data of the orthogonal design" method. All of the mechanistic experiments were conducted using Factorial Design with two factors for at least three separate experiments in duplicate, and the results were analysed by the "analysis of variance of univariate quantitative data of the two-factor factorial design" method. P-values less than 0.05 were considered statistically significant.

Results

Effects of nano-alumina on the conjugative transfer of the RP4 plasmid

In order to ensure the accuracy of the experiment, we carried out two batches orthogonal test in Luria-Bertani (LB) broth to determine the effects of the bacterial concentration, the nano-alumina concentrations, mating time and mating temperature on the formation of transconjugations. The results are shown in Figure 1. The statistically analysed results are shown in the Supplementary SAS Output. The statistical results shown that, every one of these four factors has a significant effect on the formation of transconjugations, while there

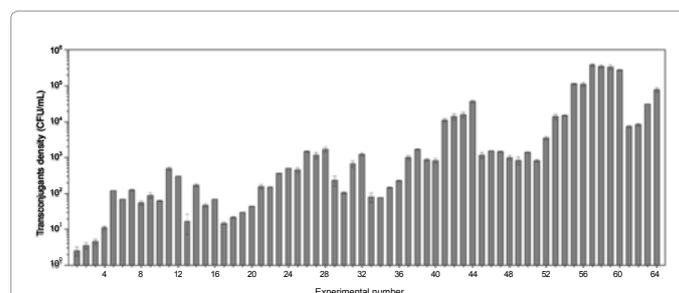


Figure 1: The results of the Orthogonal Design Experiments L₆₄(4²¹) in LB broth. The results of conjugation obtained with the non-conjugative plasmid pCB182 were zero. This revealed that the uptake of naked DNA does not occur by transformation. Furthermore, the results of conjugations induced by bulk alumina were almost the same as the results when the nano-alumina concentrations were 0 mmol/L.

was an interaction effect between the bacteria concentration and nano-alumina concentration.

The ranking of these factors in the order of importance of affecting the formation of transconjugations was as follows: bacteria concentration > nano-alumina concentration > mating time > mating temperature > the interaction between bacterial concentration and the nano-alumina concentration.

After further analyses of the effects of different levels of each factor on the formation of transconjugations, we found that the number of transconjugants increased with increasing bacterial concentration, mating time and mating temperature. The mating time could be divided into two sub-groups: 2-4 h, and 6-8 h. There was no significant difference between the two time periods in all sub-groups, but there was a significant difference between the two groups. The levels of mating temperatures could be divided into three sub-groups. There was no significant difference in terms of the influence of temperature on the formation of transconjugations between 15°C and 25°C. However, it was very significantly higher ($P < 0.05$) than that at 4°C and significantly lower than that at 37°C.

We focused more on the effect of nano-alumina on the formation of transconjugations. Our results showed that the effects of different levels of nano-alumina on the formation of transconjugations were not identical with the other three factors. With increasing concentrations of nano-alumina, the number of transconjugants was significantly elevated; at 5 mmol/L, the number of transconjugants peaked and was 2.4 orders of magnitude (250-fold) higher than no nano-alumina or bulk alumina treatment. When the nano-alumina concentration was 50 mmol/L, the number of transconjugants was lower than that at 5 mmol/L.

We also investigated the effect of nano-alumina on the formation of transconjugants in PBS instead of LB broth in order to simulate the RP4 transfer process in the natural water environments without nutrition. The results were consistent with those observed in LB broth. When the nano-alumina concentration was 5 mmol/L, the highest number of transconjugants were noted, which were increased by more 100 times than those in the control groups (Supplementary Figure 1).

The above results indicate that nano-alumina can significantly promote the formation of transconjugants of RP4, while bulk alumina had no effect on the formation of transconjugants of RP4. The ability of nano-alumina to promote conjugative transfer far exceeded the effects of mating temperature and mating time.

The effect of nano-alumina on cell membranes and its distribution in cells

We observed the morphology of the parent bacteria using transmission electron microscopy (TEM), and the results indicate that the cell membranes in the control group (0 mmol/L) and the groups with different concentrations of bulk alumina were intact with cell borders that were clearly visible. The cytoplasm was compact and distributed evenly (Figures 2a, b and c). When observing the bacteria in the 5 mmol/L nano-alumina treatment group, we found that the cytoplasm of the bacteria was agglomerated and that parts of the cell membranes were undefined (Figure 2d). When the concentration of nano-alumina was increased to 50 mmol/L, portions of the cell membranes of the parent bacteria were severely damaged, and the borders of the cell membranes were also unclear (Figure 2e). Furthermore, a number of spherical, highly dense particles were observed in these bacterial cells (Figures 2d). Subsequently, we used energy dispersive analysis (EDS) technology to analyse the elements that were present in the

regions around the highly dense particles in the ultra-thin bacterial slices (Figure 2f). The EDS results showed that there were high levels of aluminium in this region. It indicates that the nano-alumina not only damaged the structure of the cell membrane of bacteria, but also entered into bacterial cells.

The effects of nano-alumina on bacteria conjugation

Conjugation resulted in the formation of a compact electron-dense area between the outer cell membranes of the donor and recipient, as visualised by TEM and the outer cell membranes of the two cells appeared to be fused [32]. We selected 6 to 10 visual fields at random in a single sample; fields were magnified by 20,000 times to observe the conjugants. We found that there were few junctions observed in the control group (0 mmol/L) and bulk alumina groups (Figure 3a, b, c, d), but conjugants occurred between numerous bacteria in the nano-

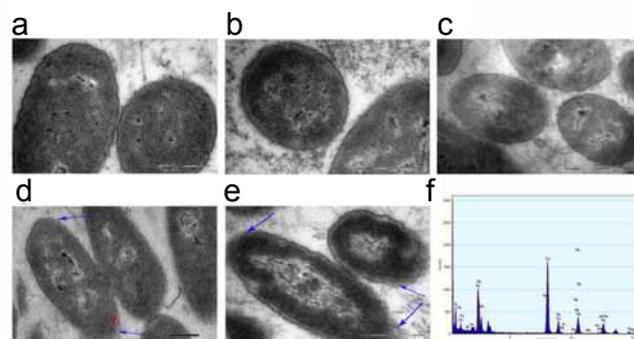


Figure 2: Detection of *E. coli* in ultrafine slices by TEM and elements analysis. a-c) In the control group (a, 0 mmol/L) and bulk alumina groups (b, 5 mmol/L; c, 50 mmol/L), the cell membranes are distinct, and the cytoplasm is compact; d and e) the cell membranes of bacteria, which were treated with different concentrations of nano-alumina (d, 5 mmol/L; e, 50 mmol/L) were damaged (indicated with blue arrows), and the extent of damage increased with increasing concentrations of nano-alumina. There were also many highly dense particles in cells (red astragal zone); f) the composition of chemical elements in the bacterial red astragal zone. Scale bars: 500 nm.

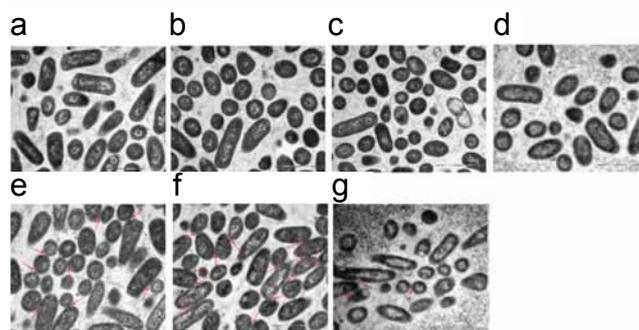


Figure 3: TEM micrographs of the conjugative junctions of mating cells. a-d) The conjugative junctions of mating cells in the control (a, 0 mmol/L) and bulk alumina groups (b: 0.5 mmol/L, c: 5 mmol/L, d: 50 mmol/L). No conjugation was found; e-g) the conjugative junctions of mating cells in the nano-alumina groups (e, 0.5 mmol/L; f, 5.0 mmol/L; g, 50.0 mmol/L). The red arrows indicate that conjugations occurred between two bacteria or multiple bacteria. The numbers of conjugations of nano-alumina groups were much greater in these groups than in the controls and the bulk alumina groups, and the 5 mmol/L nano-alumina group had the greatest number of conjugations. Scale bars: 2 μ m.

alumina groups (Figure 3e, f, g), and a single bacterium might even be conjugated with multiple other bacteria in a single view. The number of the junctions in the 5 mmol/L nano-alumina groups was the highest in all of the nano-alumina groups. Moreover, the aggregation of nano-particles was observed to some extent in the 50 mmol/L nano-alumina groups.

We used the fused promoter to detect the transcriptional activity of the RP4 conjugation genes, *trbBp* and *trfAp*, which regulated the formation of conjugation bridges and regulated RP4 processing into a single chain, transferring from the donor to recipients and replicating from single chain to double chains in recipient, respectively. Results indicated that β -glycosidase activities from these two reporters were relatively low in the control group (0 mmol/L) and the bulk alumina groups (Figure 4), and there were no significant differences between those groups. In contrast, the activities were significantly higher in all the nano-alumina groups compared to the control group and the bulk alumina groups. With increasing the concentration of nano-alumina, the transcriptional activities of both *trbBp* and *trfAp* increased significantly. Our results indicate that high concentrations of nano-alumina can enhance the transcriptional activities of the *trbBp* and *trfAp* of RP4 conjugation genes to make more conjugants forming.

The effect of nano-alumina on bacterial oxidative stress-response systems

We assessed whether nano-alumina affects the bacterial oxidative stress-response systems under these conditions. We found that the levels of hydroxyl free radicals ($\text{OH}\cdot$) were significantly higher at all of the nano-alumina concentrations than those in the control group and bulk alumina groups. The $\text{OH}\cdot$ yield of the bacteria treated with nano-alumina increased in a manner that was positively correlated with increasing nano-alumina concentrations (Figure 5a). Simultaneously, there was a significant increase in the total anti-oxidative capacity (T-AOC), as well as the catalase (CAT) superoxide dismutase (SOD) and glutathione reductase (GR) activities of the bacteria at all of the

nano-alumina concentrations compared to the control and bulk alumina groups (Figure 5b, c, d, e). Furthermore, with increasing concentrations of nano-alumina, almost all of the levels of oxidative stress were significantly increased. Comparatively, all of the levels of oxidative stress indicators did not differ significantly between the bulk alumina groups and the control group. The results indicate that nano-alumina can affect the oxidative stress-response systems of bacteria in contrast to bulk alumina.

Effects of nano-alumina on the global regulation of RP4 gene expression

We conducted real-time PCR to estimate the mRNA expression of the major global regulatory genes *korA*, *korB*, and *trbA* of the RP4 plasmid under the different concentrations of nano-alumina and bulk alumina. The results did not show any significant difference between the mRNA expression of the three genes in the control group and the bulk alumina groups (Figure 6). However, nano-alumina induced a statistically significant repression in mRNA expression at all nano-alumina concentrations compared to the results of the control and bulk alumina groups. With increasing concentrations of nano-alumina, the mRNA expression of *korA*, *korB* and *trbA* decreased significantly. These results show that nano-alumina can significantly repress the expression of the global regulatory genes of the RP4 plasmid, *korA*, *korB* and *trbA*.

Discussion

Conjugation is one of several mechanisms of horizontal gene transfer by plasmids, which code for traits such as antimicrobial resistance and can be transferred between bacteria. RP4 is a typical IncP α -type plasmid, with multidrug-resistant properties that can be transferred by conjugation to other bacteria in either the solid or liquid phase [38, 39]. The conjugal transfer of RP4 is influenced by many factors, such as cell number density, mating time, and mating temperature, amongst others. Our studies show that a new type environmental pollutant, nano-alumina, participates in the conjugation

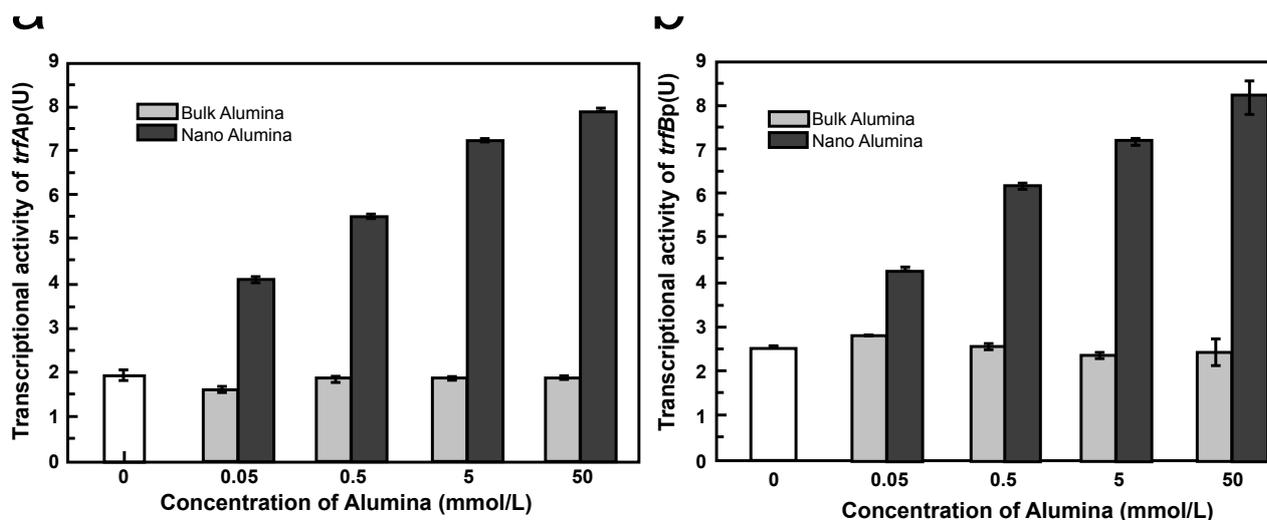
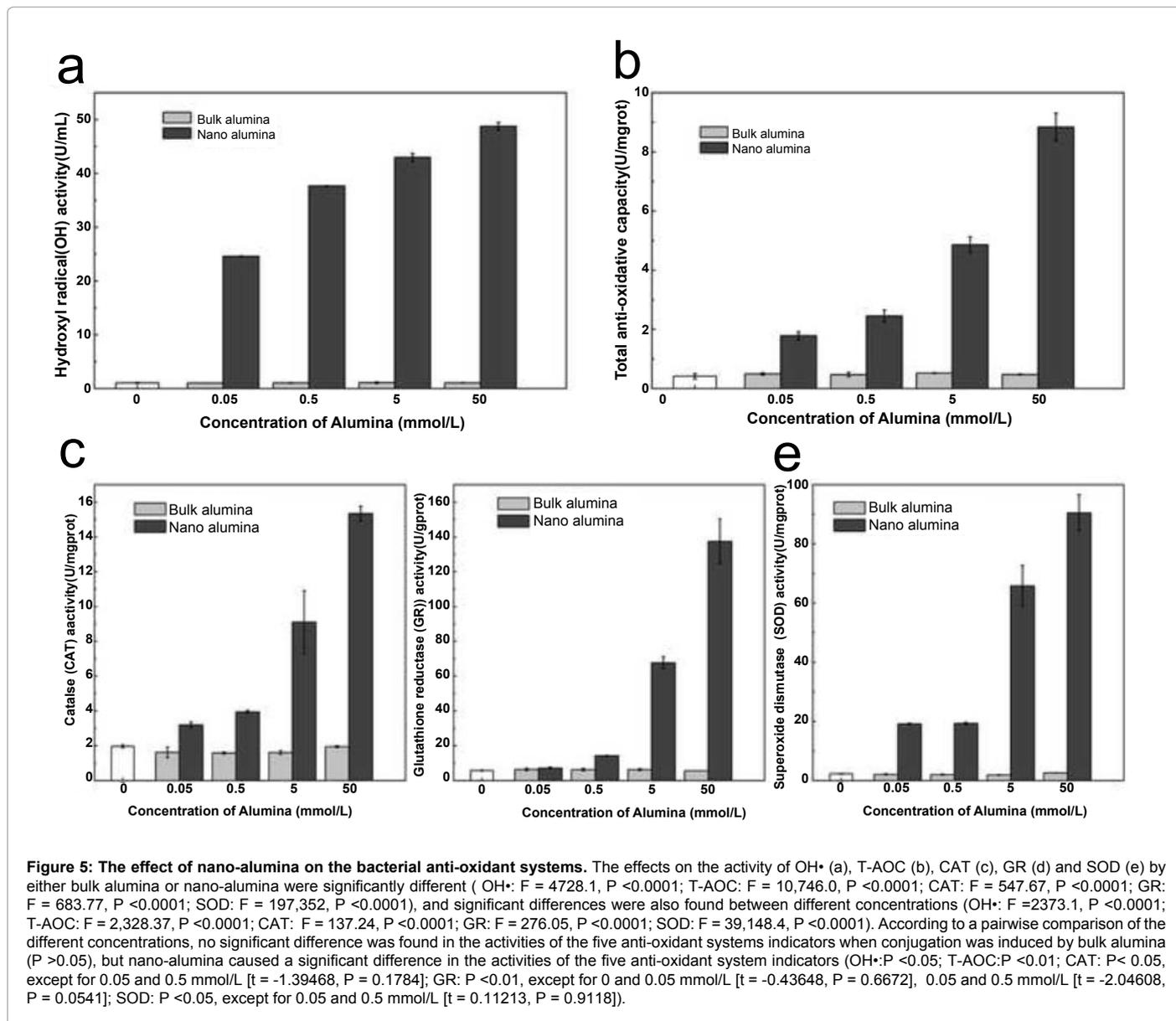


Figure 4: The transcriptional activities of *trfAp* and *trbBp*. The transcriptional activities of *trfAp* (a) and *trbBp* (b) between the bulk alumina and nano-alumina groups were significantly different between each other (*trfAp*: $F = 51018.1$, $P < 0.0001$; *trbBp*: $F = 288422$, $P < 0.0001$), and at different levels (*trfAp*: $F = 4931.41$, $P < 0.0001$; *trbBp*: $F = 28606.1$, $P < 0.0001$). When the process of conjugation was induced by bulk alumina, the transcriptional activities of *trfAp* and *trbBp* between the different levels were not significantly different, according to pairwise comparisons ($P > 0.05$). When the conjugal process were induced by nano-alumina, the transcriptional activities of *trfAp* and *trbBp* between pairwise comparisons of the different levels were statistically significant different ($P < 0.01$).



process of multidrug-resistance plasmids. This has been shown to be the most important factors that influence RP4 conjugal transfer, with the exception of bacterial concentration.

The presence of nano-alumina could promote the transfer of RP4 significantly. The conjugations of drug-resistant plasmids increased with increasing concentrations of nano-alumina and reached a peak at 5 mmol/L nano-alumina, at which point the number of conjugations increased by an order of magnitude of almost 2.4 (more than 250 times) in LB broth and 100 times in PBS compared to the bacteria in 0 mmol/L nano-alumina. When the concentration of nano-alumina was higher than 5 mmol/L, the number of transconjugants decreased, but the number of transconjugants still far exceeded those in the control group. The reason why this promoting effect decreased may be that the particles became aggregated, as showed in the TEM photograph. Therefore, the nano-alumina particles would become relatively less bioavailable than in dispersion at lower concentrations, which is a common phenomenon in all kinds of nano-particles [40]. Alternatively,

higher concentrations of nano-alumina (e.g., 50 mmol/L) could damage the membrane severely, as was seen from the results of TEM, and inhibited the growth of bacteria (Supplementary Figure 2) in LB broth or kill the majority of bacteria in PBS (Supplementary Table 5). Comprehensive consideration of the above factors, it resulted in a reduction of the conjugative transfer rate. In contrast, no remarkable effect on the rate of RP4 conjugation transfer was observed with the same concentrations of bulk alumina. It would appear that the effect of promoting conjugative transfer of nano-alumina exceeds the effects of mating temperature and mating time, although some reports have indicated that temperature and time may affect the conjugative transfer of drug resistance traits [41].

The effect of nano-alumina on the conjugative transfer of RP4 may be explained via the following potential mechanisms:

(1) It is known that oxidative stress (the SOS response) can damage cell membranes and may promote the horizontal transfer of antibiotic

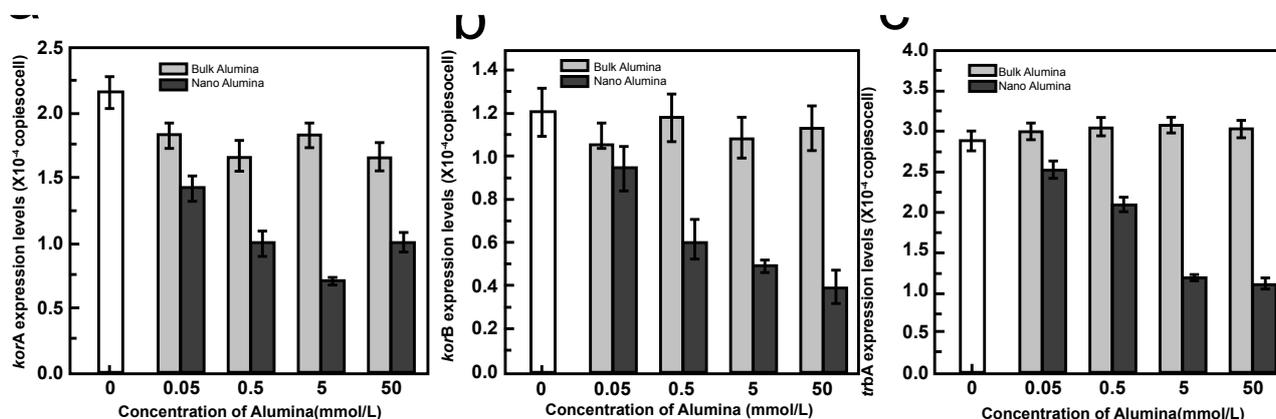


Figure 6: The mRNA expression of global regulation genes of RP4. The expression levels of *korA* (a), *korB* (b) and *trbA* (c) between bulk alumina groups and nano-alumina groups were significantly different (*korA*: $F = 1,769.68$, $P < 0.0001$; *KorB*: $F = 87$, $P < 0.05$; *trbA*: $F = 10046.8$, $P < 0.0001$); at different concentrations, the mRNA expression levels of the three genes were also significantly different (*korA*: $F = 181.23$, $P < 0.0001$; *korB*: $F = 102.4$, $P < 0.05$; *trbA*: $F = 615.81$, $P < 0.0001$). When conjugation was induced by bulk alumina, the three mRNA expression levels between pairwise comparisons of the different levels of treatment were not significant different ($P > 0.05$), but different concentrations of nano-alumina produced significant differences in the levels of three mRNA expression (*korA*: $P < 0.05$, except for 0.5 and 50 mmol/L [$t = -0.11034$, $P = 0.9132$]; *korB*: $P < 0.05$, except for 0.5 and 5 mmol/L [$t = 0.008474$, $P = 0.9933$], 0.5 and 50 mmol/L [$t = 0.1565$, $P = 0.9877$], 5 and 50 mmol/L [$t = 0.00717$, $P = 0.9943$]; *trbA*: $P < 0.05$, except for 5 and 50 mmol/L [$t = -0.7057$, $P = 0.4885$]).

resistance genes [42]. Nano-alumina increased the yield of OH^\bullet , which results in cell damage and attacks membranes directly [43]. Oxidative damage to membranes can arise through either damage to lipids or membrane proteins. It can cause the peroxidation of lipids both *in vitro* and *in vivo* [44]. When fatty acid chains become shortened or gain ionic charges, their ability to rotate within the membrane is altered and the membrane may become more fluid [45]. An increase in membrane fluidity results in a loss of structural integrity, which is required for the transport of most nutrients and DNA [46]. It has been found that the SOS response can promote the horizontal transfer of antibiotic resistance genes [42]. Our study showed that the conjugative transfer of RP4 increased with increasing concentrations of nano-alumina.

(2) Nano-alumina can enhance the conjugation efficiency through regulation the transcriptional activity of conjugative gene. The conjunctive transfer of RP4 can be divided into two processes. The first process is that the donor strain and recipient strain form conjugation bridges under the regulation of mating pair formation (Mpf) system gene *trbBp*. The second process is under the DNA transfer and replication (Dtr) system gene *trfAp* regulation, DNA is processed into a single chain, transferred from the donor to recipients and replicated from single chain to double chains in recipients. The conjugative transfer of IncP plasmids, such as broad-host-range plasmid RP4 and RK2, depends on the productions (protein) of the *trfAp* and *trbBp* genes, which had been certified by many researchers [36, 47-51]. The transcriptional activities of the RP4 conjugation genes imply the amount of the productions (protein) of *trfAp* and *trbBp* genes. The transcriptional activity of *trbBp* was enhanced after treatment with nano-alumina. It suggested that nano-alumina could promote expression of *trbB* protein which could interact with the first step of conjugative transfer, i.e. forming conjugation bridges. And this result was consistent with the results of TEM. In addition, nano-alumina could promote the transcriptional activity of *trfAp*. It presented that more *trfA* protein which could interact with the second step of conjugative transfer was produced under nano-alumina induced.

(3) Nano-alumina repressed the expression of global regulatory

genes which coded the global repressors of RP4. The whole nucleotide sequence of RP4 has been compiled [38] and many studies have certified that there is a very complex regulatory circuit controlled by global repressors, *KorA*, *KorB* and *TrbA*, in the regulation of the plasmid genes for replication, transfer and stable inheritance [44, 48-52]. As Kostelidou et al. and Pansegrau et al. [36,38] described that the presence of both *KorA* and *KorB* led to severely decrease the activity of promoter *trfAp*, and the presence of both *KorB* and *TrbA* led to severely decrease the activity of promoter *trbBp*. Our results showed that the mRNA expression of *korA*, *korB* and *trbA* were repressed by nano-alumina. It made the presence of *KorA*, *KorB* and *TrbA* decreased, as a result of the promoter *trbBp* and *trfAp* which were activated to promote the conjugative transfer of RP4.

Alumina plays an important role in the regulation of the composition of soil water, sediment water, and other natural water systems [57]. Nano-alumina, due to its huge surface area, has several applications as an adsorbent and a catalyst [16-19]. Nano-alumina has now become one of the two market leaders in the USA for nanosized materials [58]. The dosage of nano-alumina used in a previous research project was 34.2 g/kg suspended solid [19], which was much higher than the concentration used in our present study. Kaegia et al. [20] found that the density of nanoparticles that were less than 20 nm in diameter in drinking water after conventional water treatment was between 7 and 10×10^8 particles/mL. We calculated the density of nano-alumina particles and found that the number of nano-alumina particles in our study was approximately 2.08×10^6 particles/mL (0.5 mmol/L), which significantly promoted the transfer of the RP4 plasmid. If nanomaterials are used in water treatment in the future, the residual amount of nanoparticles in drinking water may be even greater than this.

Once the nano-alumina and the bacteria that carry the plasmid encoding resistance traits meet within a water environment contaminated by pathogenic bacteria or opportunist pathogens, the horizontal transfer of drug-resistant plasmids among bacteria can be significantly promoted by nano-alumina. This function may be

especially useful in waters with high localised bacterial concentrations, such as waste water and hospital sewage water.

Conclusions

In conclusion, we described the effect of nano-alumina on the horizontal transfer of antibiotic resistance genes by conjugation of the RP4 plasmid between bacteria. Nano-alumina was able to promote the horizontal transfer of multidrug-resistant genes by the conjugation of the RP4 plasmid. The transfer of drug-resistant genes may be explained by the actions of nano-alumina, which include the involvement of oxidative stresses, enhancing the transcription activities of conjugative genes, and repressing of the global regulatory factor genes for RP4 conjugation. The findings in this study support the notion that nano-alumina in the environment could result in ecological hazards. More important, an enhanced rate of plasmid transfer among microorganisms may have an enormous impact on human health and environmental safety.

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