

## Stomach Content Analysis and Concentrations of Chemical Pollutants in the Clymene Dolphin (*Stenella clymene*, Gray 1846) from the Coastal Waters of Ghana

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### Abstract

Stomach content analysis constitutes an important component of fisheries management, providing insight into fish feeding patterns and quantitative assessment of food habits. Thus, a study aimed at obtaining dietary information from stomach contents analysis of the most common by-caught dolphin species, *Stenella clymene*, beached from the coastal waters of Ghana, along the Gulf of Guinea was undertaken. The stomachs of 39 by-caught clymene dolphins landed at three fisheries landing beaches along the Ghanaian coast, were analyzed. A further study correlating the chemical contaminant load in tissues of Clymene dolphins to that of their preferred prey was undertaken. The stomach contents were generally composed of digested items. Fish, cephalopods and crustaceans were identified and represented a diversity of 12 species. On taxa level, fish was the most frequent (69%) and numerically the most important prey (46.57) followed by cephalopods (3.05) with crustaceans being present in trace amounts (1.55). However, both cephalopods and fish represented a more balanced share of the diet in biomass (45% and 51%) respectively. Thus, clymene dolphins off the coastal waters of Ghana appear to rely principally on both fish and cephalopods for food. Parasites also dominated the gut contents in relative abundance (48.83%). Prey items accumulated chemical contaminants at relatively the same concentrations (50%) as the blubber, liver and muscle of the clymene dolphins, confirming that food is the main source of exposure to contaminant load for marine mammals. With regards to quantitative analysis of prey species of cetaceans, this study of diet in clymene dolphins is the first recorded in this area.

**Keywords:** Feeding habits; *S. clymene* dolphins; POPs; Parasites; Marine mammals distribution; Gulf of Guinea

### Introduction

The Clymene dolphin, *S. clymene* [1], is one of the poorest known cetaceans, as it was confirmed as a valid species only in 1981 [2]. Recent genetic analysis suggests that, it is most closely related to striped dolphin *Stenella coeruleoalba*, while it appears physically similar to both striped and spinner dolphins *Stenella longirostris* [3,4]. The species has a robust, streamlined body with a moderately short beak and a tall, falcate dorsal fin. The rounded melon is separated from the beak by a distinct crease. These dolphins are recognized by a tricolored pattern on their sides that includes a dark gray cape, moderately gray flanks, and a white or pale gray underside. They also have distinct black lips that appear similar to a "moustache" and a line that extends across the top ridge of their beak. They have 36-52 pairs of small conical teeth in each jaw that are useful for grasping prey [5]. In the western Atlantic, adult body size ranges between 1.70 m and 1.97 m, that is, 1.70 m to 1.90 m in females and 1.76 m to 1.97 m in males and they weigh from 75 kg to 90 kg [6].

Published information on the biology of the Clymene dolphin *S. clymene* were exclusively those from the Western Atlantic Ocean. The species commonly occurs in oceanic waters 250 m to 5,000 m in depth in tropical and warm temperate waters of the Atlantic Ocean [7,8].

They feed on small mesopelagic fish (e.g. myctophids) and cephalopods. Their feeding sometimes occurs at night when their prey vertically migrates towards the surface [9]. The stomach of one stranded specimen contained one pair of small beaks of an unidentified squid and over 800 very small otoliths of fishes of the families Myctophidae, Argentinidae and Bregmacerotidae [9].

The species is poorly known with regards to biology, life history, distribution and migratory habits, hence, research on all aspects of its biology is required. The paucity of information on this species has been attributed to the very low abundance, at least in coastal waters [10]. However, capture records show that *S. clymene* is the most commonly landed cetacean on Ghana's coast, which suggests a high relative abundance in that area [11-14]. In the eastern Atlantic, only a 'West African stock' is recognized, however no population studies have been undertaken [12]. By relative frequency of occurrence in catches, 34.5% in 1998-2000 and 32.1% in 2013-14 as indicator, *S. clymene* is considered the most abundant dolphin in Ghanaian coastal waters [11]. Some Ghanaian fishermen therefore refer to *S. clymene* as the common dolphin. Debrah [14] also identified Clymene dolphins to be the most dominant species caught, followed by *S. attenuata*.

Many dolphin species or populations are recognized internationally as vulnerable or threatened due to their low reproductive potential, which is, giving birth to one calf after a long gestation period of 11-12 months [15,16]. Oil spills and various fishing methods, most notably

purse-seine fishing for tuna and the use of drift and other gill nets, results in large amounts of dolphins being killed accidentally [17]. In addition, contamination of the oceans, seas, and rivers, is a critical issue of concern, especially with Persistent Organic Pollutants (POPs), heavy metals, plastics, and other industrial and agricultural pollutants. These contaminants do not disintegrate rapidly in the environment hence, dolphins gradually accumulate unusually high levels of these contaminants thus, reducing their populations [18-20]. Clymene dolphins are as well threatened as they are also incidentally captured as by-catch in fishing expeditions using drift gillnets off the coast of West Africa, where they are used as shark bait and for human consumption [21].

The study of stomach contents or feeding habits of fish constitutes an important component of fisheries and it provides the basis for understanding trophic interaction in the aquatic food web. Diets of fishes represent an integration of many important ecological components like habitat use, behavior, condition, energy intake and intra/extra specific interactions. The occurrence, distribution, stomach contents analysis and chemical pollutants have been extensively studied in northern hemisphere cetaceans [22-30]. However, there is insufficient information on population status and trends particularly, with no information available on stomach contents and contaminant loads of marine mammals found in the Gulf of Guinea including paucity of data relating to dolphins beached on the coast of Ghana. In addition, the International Union for Conservation of Nature and Natural Resources (IUCN's) Red List of Threatened Species considers *S. clymene* "Data Deficient" [31].

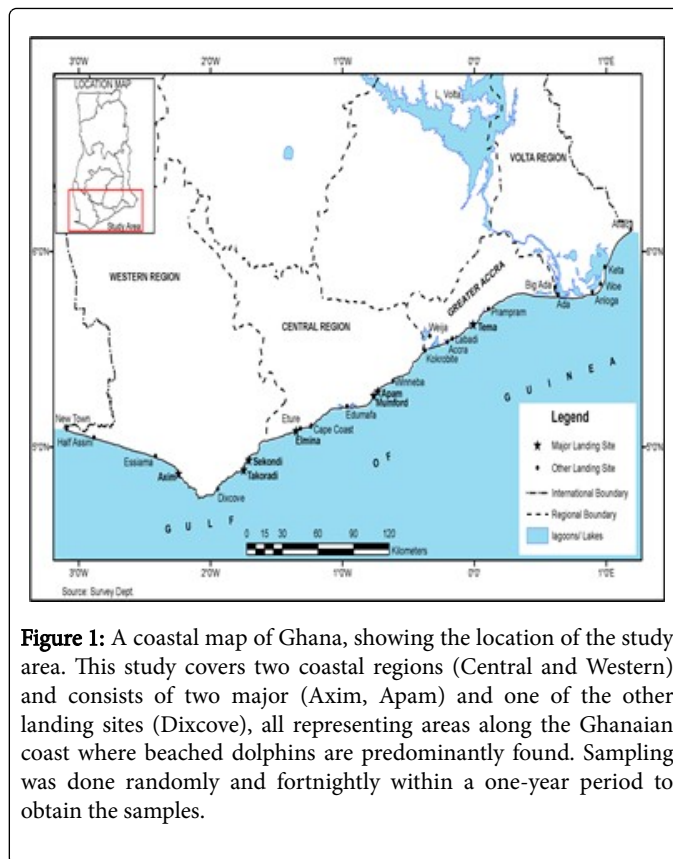
Thus, this study was undertaken to breach the gap in scientific information on this species, particularly, its feeding ecology and extent of chemical contamination.

## Materials and Methods

### Sampling

The stomachs of 39 by-caught Clymene dolphins, landed at three fish landing beaches along the Ghanaian coast (Figure 1) were analyzed. Life history data including standard body length (cm), body weight (kg), sex, tooth counts, location and date of by-catch were recorded for each specimen. Fore-, main- and pyloric stomachs were removed from freshly landed dead dolphins, fixed in 40% formalin in polythene bags to arrest post-mortem digestion and, later, examined in the laboratory for their contents, including food matter and endoparasites.

After the dissection of the whole dolphin following standard protocols [20], blubber, muscle, and liver samples were taken, stored in individual aluminium foils and then in polyethylene bags, labelled and transported on ice to the Ecological Laboratory at the University of Ghana where they were kept frozen at  $-20^{\circ}\text{C}$  until analysis was conducted.



**Figure 1:** A coastal map of Ghana, showing the location of the study area. This study covers two coastal regions (Central and Western) and consists of two major (Axim, Apam) and one of the other landing sites (Dixcove), all representing areas along the Ghanaian coast where beached dolphins are predominantly found. Sampling was done randomly and fortnightly within a one-year period to obtain the samples.

### Sample analysis

Diets were described in terms of prey occurrence, relative abundance, calculated mass and size distribution and followed standard procedures for marine top predators [32-36]. Wet weight of stomach contents was recorded in grams. Each stomach was weighed full, and then emptied into a tray. The empty stomach was weighed again, and the mass of stomach contents obtained by subtraction. The stomach content was washed through a  $0.71\ \mu\text{m}$  mesh sieve. All the diagnostic parts were recovered, and identified with the aid of microscopes and hand lenses. Fish bones and otoliths were stored dry, while's cephalopod beaks, crustacean remains as well as any remains with flesh attached were stored in 70% ethanol. Endoparasites were also stored in 70% ethanol. The items found were identified to the lowest taxonomic level possible using published guides [37,38]. Total number of food items was estimated based on paired structures (otoliths, operculum, hyomandibular, dentary and premaxillary) for fishes, upper and lower beaks for cephalopods and eyes/telsons for crustaceans. With the aid of dissecting and light microscopes and parasites identification keys [39], parasites were identified to the lowest taxonomic level possible.

The concentrations of Organochlorine pesticides (OCPs) and polychlorinated biphenyls (PCBs) in the tissues of the clymene dolphins as well as their prey items (fish, cephalopods) were analyzed following the method described by Kajiwara et al. [40]. Analysis of trace metals was also undertaken following published protocols [41].

## Description of stomach contents

The stomach contents of *S. clymene* were described based on three indices. The occurrence of a given prey taxon was the number of stomachs in which this taxon was observed [42]. Relative abundance was given by the number of individuals of the same taxon found throughout the sample set. The calculated biomass was given by the product of the average calculated body mass and the number of individuals of the same taxon in each sample, summed up throughout the sample set. These three indices were expressed by their percentage frequency respectively as, percentage frequency of occurrence (%O), percentage composition by number (%N) and percentage composition by biomass (%M).

## Statistical analysis

Concentrations of trace elements in this study were presented on a µg/g dry weight (d.w), since this represents the best bases for comparison of different tissues [43]. Descriptive statistics for mean, standard deviation and range were calculated using SPSS (version 16). Statistical analyses of chemical contaminants data were performed by Analysis of Variance (ANOVA) followed by multiple comparison tests. The significance for statistical analyses was set at  $\alpha=0.05$ . To determine the similarity of accumulation of contaminants in the different tissues, Ward's Method of tree clustering, based on analysis of variance, was employed. All analyses of stomach contents were carried out by reference analytical standard methods [44], as a measure for quality control.

## Results

### Stomach contents

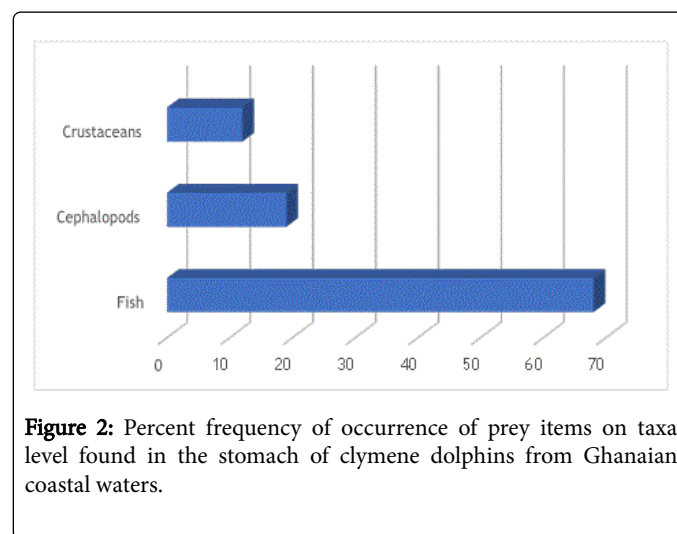
The stomach contents of 39 Clymene dolphins with an average body weight of 61 kg were examined. These comprised 20 males and 19 females. Food remains were recovered from 34 samples (87.2%), with five being empty (12.8%). The total mass of examined food material was 20323.4 g, with a mean mass of  $597.7 \text{ g} \pm 152 \text{ g}$  (range 0 g to 640 g). In general, food remains were composed of highly digested material (%O=71.8%, %N=12.6 %) and included few identifiable prey remains. Apart from the whole squids which were identified, few cephalopod beaks were found (unidentified, but possibly belonging to the squid, *Sepia* sp.). From direct count, the most important prey family in frequency and numerical terms was the cephalopod, Sepiidae (*Sepia* sp.) which accounted for 33.33% and 2.8% respectively of all prey consumed. All other items were teleosts mostly represented by their otoliths and few crustaceans. The most frequent fish prey was the cutlass fishes (Trichiuridae) which occurred in 8 (20.51%) of 34 stomachs examined as compared to the 6 stomachs in which *Decapterus* sp occurred. However, in terms of numbers the mackerel scads, *Decapterus* sp. was of high importance representing 2.6% of prey consumed as indicated in Table 1. Based on variation in morphology, shape and structure, three different otoliths were identified, suggesting the presence of three other fish species. Considering the otoliths present, a minimum of 5 fish species (within 5 families, 5 genera) and two cephalopod species were identified (Table 1). Therefore, on taxa level, fish recorded the highest frequency in terms of occurrence (Figure 2).

Fish, cephalopods, and crustaceans represented a diversity of twelve (12) different species. Average prey diversity was  $3.2 \pm 2.1$  species per sample (N=34 non-empty stomachs).

Stomach Contents	ni	N	%O	%N	%M
Otoliths	7	4	17.95	6.37	0.467
Eyes	15	34	38.46	21.04	1.71
Parasites	20	34	51.28	48.83	3.73
Trichurus lepturus	8	34	20.51	0.79	19.94
Decapterus rhoncus	6	34	15.38	2.63	3.61
Squids ( <i>Sepia</i> sp)	13	34	33.33	2.88	45.04
Shrimps	3	34	7.69	1.35	0.21
semi-digested fish (Vertebrae/skeleton)+ three unidentified fish	28	34	71.79	12.65	25.09
Fish jaws	6	34	15.38	0.46	0.23
Crustacean	3	34	7.69	0.09	0.02
Caudal fins of fish	2	34	5.13	0.06	0.0005
Amphipod	1	34	2.56	0.03	0.01

\*Where n is the number of stomachs where the prey i was found and N the total number of stomachs.

**Table 1:** General composition of items recovered from the stomachs of *S. clymene* landed from the Ghanaian coastal waters.



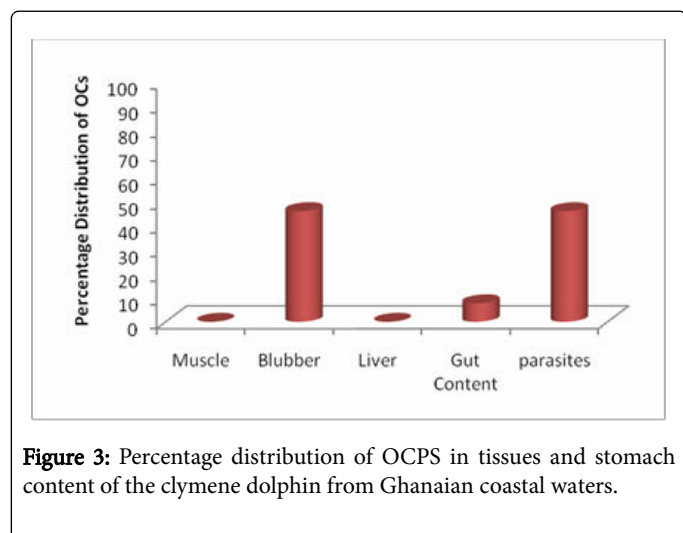
**Figure 2:** Percent frequency of occurrence of prey items on taxa level found in the stomach of clymene dolphins from Ghanaian coastal waters.

### Identification of parasites

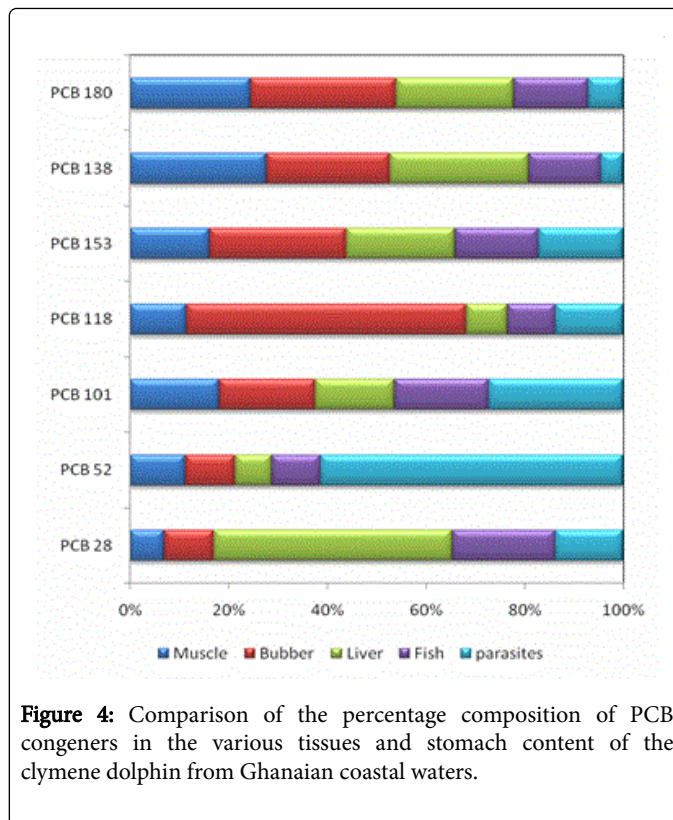
Two main types of parasites were identified in the gut of the Clymene dolphins. These were nematodes and acanthocephalans. Nematodes were accurately identified by their smooth, cylindrical, and relatively long body shape whiles the acanthocephalans were differentiated by their characteristic "thorny head", that is, their head region which is armed and encircled with numerous rows of hooks (proboscis). Parasites were identified to family level as Pomphorhynchidae. In terms of abundance, parasites (nematodes), represented equally well and were much comparable to that of fish except that due to their very small body size, it recorded just about 3.73% for its percentage composition by mass (Table 1).

### Analysis of chemical contaminants

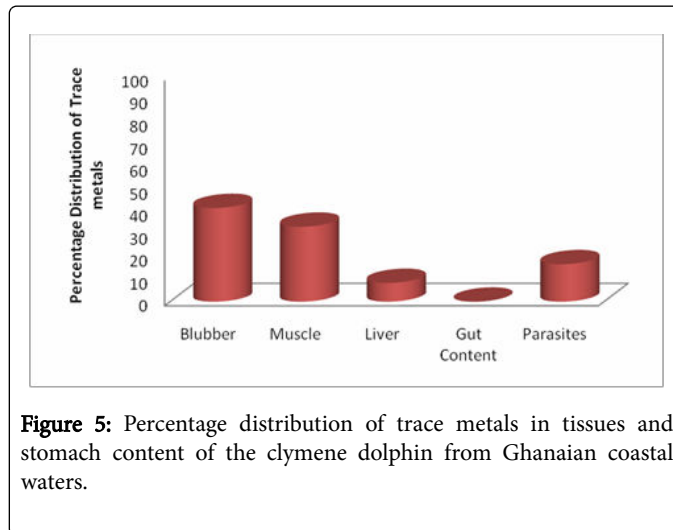
Analysis of chemical contaminants in tissues indicated a high percentage accumulation of OCPs (gamma-HCH, delta-HCH, Heptachlor, Aldrin Gamma-Chlord, A-endosulfan, p,p'-DDE, Deldrin, Endrin, p,p'-DDT) in the blubber of the dolphin and endoparasitic organisms in the gut (50% for both). Muscle and liver recorded the minimum concentrations of 0.2% respectively (Figure 3). Percentage distribution of 7 PCB congeners analyzed were highest in the blubber (42.86%) followed by both parasites and the liver with a common value of 28.57% (Figure 4). Accumulation of trace metals (As, Cu, Se, Zn, Cd, Hg, Mn, Mg, Cr, Ni, Fe) on the other hand was highest in the blubber (45%), followed by muscle (32%), parasites (20%) and then liver (10%) as shown in Figure 5. Cluster analysis using Ward's method was undertaken to provide an overview of similarities in accumulation patterns in the various tissues of dolphins, as well as the fish and parasites from the stomach content (Figure 6). Based on this analysis, accumulation pattern of OCPs, PCBs and trace metals in the liver and blubber were very similar, likewise that in the stomach contents (fish and parasites).



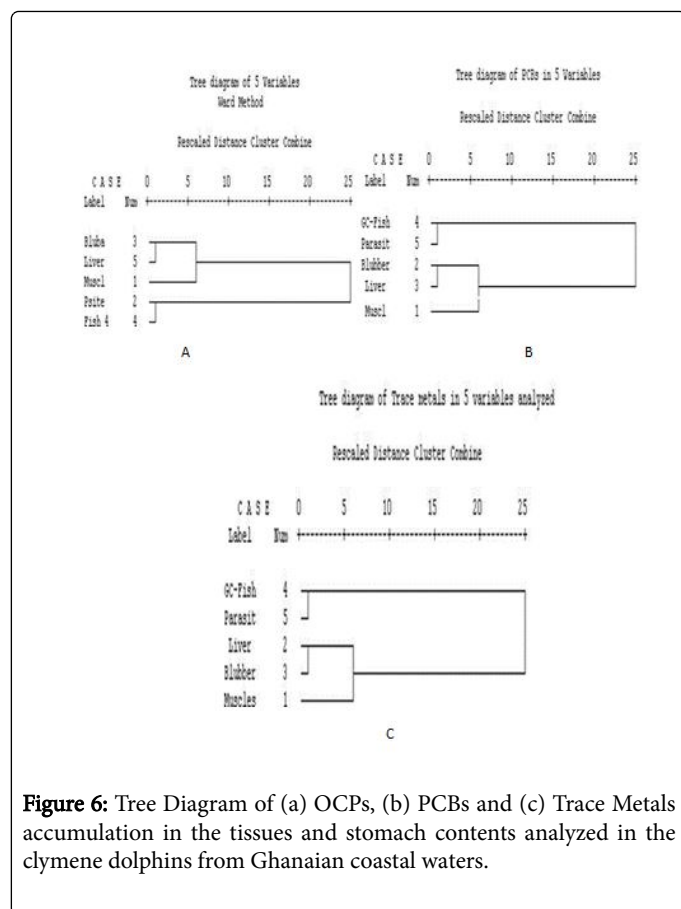
**Figure 3:** Percentage distribution of OCPs in tissues and stomach content of the clymene dolphin from Ghanaian coastal waters.



**Figure 4:** Comparison of the percentage composition of PCB congeners in the various tissues and stomach content of the clymene dolphin from Ghanaian coastal waters.



**Figure 5:** Percentage distribution of trace metals in tissues and stomach content of the clymene dolphin from Ghanaian coastal waters.



**Figure 6:** Tree Diagram of (a) OCPs, (b) PCBs and (c) Trace Metals accumulation in the tissues and stomach contents analyzed in the clymene dolphins from Ghanaian coastal waters.

## Discussion

This study again confirms *S. clymene* as the most commonly beached dolphin from the coastal waters of Ghana, as all the samples encountered at the landing sites during the one-year sampling period were predominantly of this species. This confirms the preference of this species of dolphins to temperate and tropic waters. From the stomach contents analysis of the Clymene dolphins, fish constituted a major portion of their food, making it the most preferred food item whereas cephalopods constituted an important but minor portion of their diet. It can however be said that both fish and cephalopods represented a more balanced share of the diet in biomass (45.04% and 50.98%) respectively by comparing the number of items that make up the total mass of fish species to the single cephalopod item recorded from each stomach. This is as a result of the generally greater individual body mass of cephalopods relative to fish. Crustaceans accounted for a negligible portion of the diet, by number and by mass. It was observed that the content also included prey of prey, certainly from the fact that many fish species predate on other species. These however constituted the bulk of the digested food. Clymene dolphins off the coastal waters of Ghana thus, appear to rely predominantly on both fish and cephalopods for food. Important prey included typical pelagic groups like the cutlass fishes as well as the bottom-dwelling species, observed from the positioning of the mouth of the unidentified fishes. Studies conducted by Barros et al., [45], in Hong Kong waters indicated that the cutlass fish (*Trichiurus sp*) is consumed by humpback dolphins, finless porpoises and bottlenose dolphins [46], implying they may constitute an important resource for resident and transient cetaceans.

This seems to have been confirmed in this study with reference to the frequency at which these species occurred in the diet of Clymene dolphins in Ghanaian coastal waters.

## Parasitic infestation of clymene dolphins

In Ghana, Debrah [14] identified some helminth parasites in the fore and main stomachs of dolphins landed on Ghanaian coast, although the numbers recorded were too small for any statistical analysis. Contrary to that research, this study identified, two types of parasites; acanthocephalans, and nematodes, found mainly in fish, and thus may have occurred secondary from ingested prey. This constituted about 48.83 % of the total number of items found in the gut of 20 Clymene dolphins, thus, dominating the gut in relative abundance. Adult parasites especially round worms, live in the stomachs of marine mammals. Large fish tend to be more heavily infested by parasites than small fish of the same species [47]. This is because large fish eat more, and therefore ingest greater numbers of parasites, and also because the larval worms, although inactive, can survive for a long time in fish, and consequently their numbers accumulate as the fish grows older. A similar situation might have occurred in the clymene dolphins and could account for the high percentage of parasites in the guts. Although parasites are a natural occurrence, with small numbers often well tolerated and may not necessarily be contamination, their presence particularly in high amounts as recorded in this study, which can result in excessive interactions, irritations, ulceration or perforations of the stomach hence giving opportunity for secondary infection by bacteria and fungi, can cause significant pathology in the dolphins, likewise in humans through consumption. As there have been cases of human illness [47], caused by the ingestion of some live genus of *nematodes*, *Phocanema* or *Anisakis* larvae, in countries where raw or lightly cured fish is commonly eaten.

## Chemical contaminants in clymene dolphins

Chemical contaminants (OCPs, PCBs, Trace metals) were found in all tissues of the Clymene dolphin analyzed, as well as in fish, cephalopods and parasites recovered from the stomach. Chemicals enter the marine environment in many ways, typically through fertilizer run offs and use of pesticides in agriculture and home gardening; industrial waste, dumping and airborne emissions, as well as flame retardants used in everyday products. These chemicals are carried around the globe by wind and ocean currents. Regardless the usefulness of these xenobiotics, their increased released into the environment in recent years are causing serious pollution problems.

Marine mammals have an increased likelihood of accumulating these toxins, basically because of high rate of biomagnification due to their elevated trophic level in the marine food chain, their high metabolic rate and long life-span and thus they may serve as a potential indicator of contaminants [18,20,48]. Accumulation of all contaminants analyzed in this study was predominant in the blubber of the dolphin. This is consistent with records of chemical accumulation in marine mammal species [49-51], and justifies the fact that these xenobiotics are lipophilic in nature, thus get deposited in the thick fatty layer of their skin (blubber).

Uptake of most of these chemical pollutants has been reported to be through the lung, skin, diet, placental transfer and milk [52]. However, many studies suggest that diet is the major entry route of elements in marine mammals [18,55]. This is consistent with results from this study, as concentrations of chemical contaminants in the body tissues

of the Clymene dolphin were comparable to that recorded from the stomach content. This is particularly so because, dolphins usually consume a wide range of prey species, such as sciaenids available within their geographic ranges [53]. Aside other biological and ecological factors, including geographic area, age, gender, tissue type, metabolic rate and temporal distribution which may influence potential accumulation, feeding strategies also greatly influence the concentrations of chemical contaminants found in dolphins. For instance, particularly high concentrations of trace elements are often encountered in those feeding mainly on cephalopods, which are known to accumulate Cadmium (Cd) in higher levels than fish. Cephalopods are thus, considered to be an important vector of this element to top marine predators [41].

Also, of particular interest in this study, was the high concentration of the organochlorine pesticides in the parasites, relative to the other tissues especially the blubber ( $P < 0.05$ ). High concentrations of copper and selenium were also found in the parasite. This supports recent studies which have demonstrated that particular fish parasites can accumulate toxic contaminants (especially metals) from aquatic environment to concentrations that surpass those in the best-established accumulation indicators like the sessile mussel [56-58]. This in a way provides evidence that parasites could be important indicators of contaminant load in marine mammals and might be valuable as environmental indicators, principally for assessing water quality.

## Conclusion

The results presented in this study provide evidence of the presence and abundance of *S. clymene* in Ghanaian coastal waters. Fish and cephalopods constitute the most preferred prey items for the Clymene dolphins with high endoparasitic infestation. Analysis confirmed the presence of chemical contaminants in the tissues of the Clymene dolphins as well as in their prey items, at comparable concentrations, suggesting diet as possible route of entry. Based on this study, further investigations on the parasitology of the Clymene dolphins are recommended.

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## Competing Interests

The authors have declared that no competing interests exist.

## References

1. Gray Jr LE, J Ostby, C Wolf, DB Miller, WR Kelce, et al. (1995) Functional Developmental Toxicity of Low Doses of 2,3,7,8- Tetrachlorodibenzo-p-Dioxin and a Dioxin-Like PCB (169) in Long Evans Rats and Syrian Hamsters: Reproductive, Behavioral and Thermoregulatory Alterations. *Organohalogen Compounds* 25: 33.
2. Perrin WF, Mitchell ED, Mead JG, Caldwell DK, Van Bree PJH (1981) *Stenella clymene*, a rediscovered tropical dolphin of the Atlantic. *J Mammal* 62: 583-598.
3. Amaral AR, Jackson JA, Möller LM, Beheregaray LB, Manuela Coelho M (2012) Species tree of a recent radiation: the subfamily Delphininae (Cetacea, Mammalia). *Mol Phylogenet Evol* 64: 243-253.
4. Perrin WF, Rosel PE, Cipriano F (2013) How to contend with paraphyly in the taxonomy of the delphinine cetaceans?. *Mar Mam Sci* 29.4: 567-588.
5. Jefferson TA (1996) Estimates of abundance of cetaceans in offshore waters of the northwestern Gulf of Mexico 1992-1993. *Southwestern Naturalist* 41: 279-287.
6. Jefferson TA (2002) Clymene dolphin-*Stenella clymene*. In: *Encyclopedia of marine mammals* (Perrin WF, Würsig B, Thewissen JGM, eds.) Academic Press. San Diego 235-236.
7. Weir CR, Coles P, Ferguson A, May D, Baines M, et al. (2014) Clymene dolphins (*Stenella clymene*) in the eastern tropical Atlantic: distribution, group size, and pigmentation pattern. *J Mammal* 95: 1289-1298.
8. Fertl D, Jefferson TA, Moreno IB, Zerbini AN, Mullin KD (2003) Distribution of the Clymene dolphin *Stenella clymene*. *Mammal Rev* 33: 253-271.
9. Shirihai H, Jarrett B (2006) Whales, Dolphins and Other Marine Mammals of the World. Princeton pp: 184-186.
10. Perrin WF, Mead JG (1994) Clymene dolphin - *Stenella clymene*. In: *Handbook of Marine Mammals* (Ridgway SH, Harrison SR editors.) The first book of dolphins. Academic Press, London. 5: 161-172.
11. Van Waerebeek K, Ofori-Danson PK, Debrah J (2009) The Cetaceans of Ghana, a Validated Faunal Checklist. *West African Journal of Applied Ecology* 15: 61-90.
12. Van Waerebeek K, Ndiaye E, Djiba A, Diallo M, Murphy P, et al. (2000) A survey of the conservation status of cetaceans in Senegal, the Gambia and Guinea-Bissau. UNEP/CMS Secretariat, Bonn, Germany pp: 80.
13. Ofori-Danson PK, Van Waerebeek K, Debrah S (2003) A survey for the conservation of dolphins in Ghanaian coastal waters. *J Ghana Sci Assoc* 5: 45-54.
14. Debrah JS (2000) Taxonomy, exploitation and conservation of dolphins in the marine waters of Ghana MPhil Thesis. Dept of Oceanography and Fisheries, University of Ghana, Legon pp: 86.
15. Robinson KP, Sim TMC, Culloch RM, Bean TS, Aguilar IC, et al. (2017) Female reproductive success and calf survival in a North Sea coastal bottlenose dolphin (*Tursiops truncatus*) population. *PLoS ONE* 12: e0185000.
16. Clutton-Brock TH (1988) Reproductive success: studies of individual variation in contrasting breeding systems. University of Chicago Press: Chicago.
17. Clover C (2004) *The End of the Line: How overfishing is changing the world and what we eat*. Ebury Press, London. ISBN 0-09-189780-7.
18. Das K, Siebert U, Fontaine M, Jauniaux T, Holsbeek L, et al. (2003) Ecological and pathological factors related to trace metal concentrations in harbour porpoises *Phocoena phocoena* from the North Sea and adjacent areas. *Marine Ecology Progress Series* 281: 283-295.
19. Wells RS, Rhinehart HL, Hansen LJ, Sweeny JC, Townsend FI, et al. (2004) Bottlenose dolphins as marine ecosystem sentinels: developing a health monitoring system. *Eco-Health* 1: 246-254.
20. Bossart GD (2006) Marine mammals as sentinel species for oceans and human health. *Oceanography* 19: 134-137.
21. Jefferson TA, Webber MA, Pitman RL (2008) *Marine Mammals of the World, A Comprehensive Guide to their Identification*. Amsterdam, Elsevier pp: 238-240.
22. Jarman WM, Norstorm RJ, Muir DCG, Rosenberg B, Simon M, et al. (1996) Levels of organochlorine compounds, including PCDDs and PCDFs, in the blubber of cetaceans from the west coast of North America. *Mar Pollut Bull* 32: 426-436.
23. Law RJ (1996) Metals in marine mammals. In: Beyer WN, Heinz GH, Redmon-Norwood AW (editors). *Environmental contaminants in*

- wildlife. Interpreting tissue concentrations. Boca Raton, USA: CRC Press Inc pp: 357-376.
24. McKenzie C, Rogan E, Reid RJ, Wells DE (1997) Concentrations and patterns of organic contamination in Atlantic white-sided dolphins (*Lagenorhynchus acutus*) from Irish and Scottish coastal waters. *Environ Pollut* 98: 15-27.
  25. Parsons ECM, Chan HM, Kinoshita R (1999) Trace metal and organochlorine concentrations in a pygmy Bryde's whale (*Balaenoptera edeni*) from the South China Sea. *Mar Pollut Bull* 38: 51-55.
  26. Siebert U, Joiris C, Holsbeek L, Benke H, Failing K, et al. (1999) Potential relation between mercury concentrations and necropsy findings in cetaceans from German waters of the North and Baltic seas. *Mar Pollut Bull* 38: 285-295.
  27. Watanabe M, Tanabe S, Miyazaki N, Petrov EA, Jarman WM (1999) Contamination of Tris (4-Chlorophenyl) methane and Tris (Chlorophenyl) methanol in marine mammals from Russia and Japan: body distribution, bioaccumulation and contamination. *Mar Pollut Bull* 39: 393-398.
  28. Frodello JP, Romeo M, Viale D (2000) Distribution of mercury in the organs and tissues of five toothed-whale species of the Mediterranean. *Environ Pollut* 108: 447-452.
  29. Hernandez F, Serrano R, Roig-Navarro AF, Martinez-Bravo Y, Lopez FJ (2000) Persistent organochlorines and organophosphorus compounds and heavy elements in common whale, *Balaenoptera physalus*, from the Western Mediterranean Sea. *Mar Pollut Bull* 40: 426-433.
  30. Hobbs KE, Muir CG, Mitchell E (2001) Temporal and biogeographic comparisons of PCBs and persistent organochlorine pollutants in the blubber of fin whales from Eastern Canada in 1971-1991. *Environ Pollut* 114: 243-254.
  31. Berrow SD, Mchugh B, Glynn D, MCGovern E, Parsons KM, et al. (2002) Organochloride concentrations in resident bottlenose dolphins (*Tursiops truncatus*) in the Shannon Estuary, Ireland. *Mar Pollut Bull* 44: 1296-1313.
  32. Hammond PS, Bearzi G, Bjørge A, Forney KA, Karkzmarzski L, Kasuya T, et al. (2012) *Stenella clymene*. The IUCN Red List of Threatened Species 2012: e.T20730A17840531.
  33. Fair PA, Adams JD, Zolman E, McCulloch SD, Goldstein JD, et al. (2006) Protocols for conducting dolphin capture-release health assessment studies. NOAA tech memo NOS NCCOS 49.
  34. Pierce GJ, Boyle PR (1991) A review of methods for diet analysis in piscivorous marine mammals. *Oceanogr. Mar Biol Ann Rev* 29: 409-486.
  35. Croxall JP (1993) Diet. In: Laws, RM (editors.), *Antarctic Seals*. Cambridge University Press, Cambridge pp: 268-290.
  36. Ridoux V (1994) The diets and dietary segregation of seabirds at the subantarctic Crozet Islands. *Mar Ornithol.* 22: 1-192.
  37. Clarke MR (1986) *A Handbook for the Identification of Cephalopod Beaks*. Clarendon Press, Oxford.
  38. Harkonen TJ (1986) Guide to the otoliths of the bony fishes of the northeast Atlantic. *Danbiu ApS, Hellerup*.
  39. Mignucci-Giannoni AA, Hoberg EP, Siegel-Causey D, Williams Jr. EH (1998) Metazoan Parasites and Other Symbionts of Cetaceans in the Caribbean by The American Society of Parasitologists. *J Parasitol* 84: 939-946.
  40. Kajiwara N, Ueno D, Monirith I, Tanabe S, Pourkazemi M, et al. (2003) Contamination by organochlorine compounds in sturgeons from Caspian Sea during 2001 and 2002. *Marine Pollution Bulletin* 46: 741-747.
  41. Bustamante P, Caurant F, Fowler SW, Miramand P (1998) Cephalopods as a vector for the transfer of cadmium to top marine predators in the north-east Atlantic Ocean. *Science Total Environment* 220: 71-80.
  42. Hyslop EJ (1980) Stomach content analysis, a review of methods and their application. *J Fish Biol* 17: 411-430.
  43. Clark RB (2003) *Marine Pollution*. 5th edition. Oxford University UK pp: 236.
  44. Alpha (1980) Standard methods for the examination of water and waste water. *Am Public Health Assoc Washington DC* pp: 1500.
  45. Barros NB, Jefferson TA, Parsons ECM (2002) Food habits of finless porpoises (*Neophocaena phocaenoides*) in Hong Kong waters. *Raffles Bulletin of Zoology* 10: 115-123.
  46. Barros NB, Parsons ECM, Jefferson TA (2000) Prey of offshore bottlenose dolphins from the South China Sea. *Aquatic Mammals* 26: 2-6.
  47. FAO (2009) Round Worm in Fish. The Marine Laboratory of the Department of Agriculture and Fisheries for Scotland and the Torry Research Station of the Ministry of Agriculture, Fisheries and Food.
  48. Fair PA, Becker PR (2000) Review of stress in marine mammals. *J Aquat Ecosyst Stress Recovery* 7: 335.
  49. Boon JP, Lewis WE, Choy MRAT, Allchin CR, Law RJ, et al. (2002) Levels of polybrominated diphenyl ether (PBDE) flame retardants in animals representing different trophic levels of the North Sea food web. *Environ Sci Technol* 36:4025-4032.
  50. Kalantzi OI, Hall AJ, Thomas GO, Jones KC (2005) Polybrominated diphenyl ethers and selected organochlorine chemicals in grey seals (*Halichoerus grypus*) in the North Sea. *Chemosphere* 58: 345-354.
  51. Kannan K, Ramu K, Kajiwara N, Sinha RK, Tanabe S (2005) Organochlorine pesticides, polychlorinated biphenyls, and polybrominated diphenyl ethers in Irrawaddy dolphins from India. *Arch Environ Contam Toxicol* 49: 415-420.
  52. Dehn LA, Follmann EH, Thomas DL, Sheffield GG, Rosa C, et al. (2006) Trophic relationships in an Arctic food web and implications for trace metal transfer. *Sci Total Environ* 362: 103-123.
  53. Barros NB, Odell DK (1990) Food Habits of Bottlenose Dolphins in the Southeastern United States. In *The Bottlenose Dolphin*, edited by Stephen Leatherwood and Randall R. Reeves pp: 309-328.
  54. Sures B, Taraschewski H, Siddall R (1997) Heavy metal concentration in adult acanthocephalans and cestodes compared to their fish hosts and to established free-living bioindicators. *Parassitologia* 39: 213-218.
  55. Fair PA, Becker PR (2000) Review of stress in marine mammals. *J Aquat Ecosyst Stress Recovery* 7: 335.
  56. FAO-WHO (1997) Codex maximum residue limits for pesticides. FAO, Rome.
  57. Jefferson TA, Curry BE, Leatherwood S, Powell JA (1997) Dolphins and porpoises of West Africa: a review of records (Cetacea: Delphinidae, Phocoenidae). *Mammalia* 61: 87-108.
  58. Klinowska M (1991) *Dolphins, Porpoises, and Whales of the World*. The IUCN Red Data Book. Cambridge, United Kingdom: IUCN/World Conservation Union.