Stability of Nano Encapsulated Rice Bran Derived Bioactive Pentapeptide in Apple Juice

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Abstract

Bioactive compounds can be derived from cereal grains and their components such as rice bran. These bioactive compounds have a potency to promote health. A pentapeptide with a sequence of amino acids Glu-Gln-Arg-Pro-Arg (EQRPR) prepared from heat stabilized defatted rice bran (HDBR) has demonstrated multi-site anti-cancer properties in human cell lines. Fruit juices can be used as vehicles to incorporate the pentapeptide as a nutraceutical. To address the stability of the pentapeptide in the beverage and to prevent possible interactions with other components, nano-encapsulation with poly (lactic-co-glycolic acid) (PLGA) was used to deliver the bioactive ingredient. The nano-encapsulated pentapeptide (concentrations: 200/400/600 µg/mL) in the apple juice (model system) showed significant stability with no degradation for 2 months based on the high performance liquid chromatography analysis. The nanoparticles were uniform and stable with an effective diameter ranging between 82 and 83 nm and the results also indicated that there were no significant changes in the size over a storage period of 60 days. There was no microbial growth observed in the prepared apple juice samples. The un-encapsulated pentapeptide incorporated in the apple juice showed significant degradation after 7 days of storage. The PLGA nanoparticles showed a remarkable effect in protecting and stabilizing the bioactive compounds (pentapeptide) during the storage period at 4°C. The PLGA nanoparticles can thus be a promising carrier for the bioactive pentapeptide when incorporated into a juice medium.

Keywords: Nano-encapsulation; Polylactic-co-glycolic acid; Pentapeptide; Nutraceuticals

Introduction

Bioactive compounds, which convey nutraceutical activity, envelop broad types of functional groups. These functional groups which exist in foods can potentially affect the biological functions. These bioactive compounds consist mainly of aromatic alcohols such as polyphenols and phenolic acids, and it has been found that these compounds have an effect in alleviating the pathology of cardiovascular disease and cancer [1]. Proteins and peptides are important bioactive compounds which can impart several physiological functions in the body, such as anti-oxidant, anti-hypertensive, anti-obesity, anti-angiogenic, hypcholestremic, and immune-modulatory [1]. Rice bran is a good source of dietary protein (12-20%) and is an inexpensive co-product from rough rice-milling. A pentapeptide derived from heat stabilized defatted rice bran (HDBR) with an amino acid sequence of Glu-Gln-Arg-Pro-Arg (EQRPR) demonstrated anti-cancer activity, and has the potential of being incorporated in a food system [2].

Fruit juices can be good vehicles to incorporate nutraceuticals and bioactive compounds. Apple juice is consumed by 12.3% of Americans, second most popular juice after orange juice, and it also imparts health benefits due to the phenolic compounds [3]. In a previous study, orange juice was fortified with rice bran peptide fractions which were stable at 4°C under acidic conditions (pH 3.0-4.5) for 42 days followed by rapid degradation [4]. The results supported fruit juices as suitable vehicles to incorporate peptides and proteins; however, peptide fractions are likely to interact with other compounds in the juice matrix which affects their stability over longer storage period. The interactions among the components in fruit juices such as vitamins, minerals and phenolics can cause chemical deterioration and lead to changes in flavor and aroma of fruit juices. Nanotechnology is a fast-emerging field that provides potential benefits including enhanced stability in the controlled release and bioavailability of bioactive ingredients [5]. Nanoparticles can be defined as “a discrete entity with at least one dimension being 100 nm or less” [5]. In a study conducted using casein nanoparticles, hydrophobic nutraceuticals such as omega-3 fatty acid were delivered by preventing oxidation, odor, and off-flavor into beverages [6]. In another study caseinophosphopeptides (CPPs) were used to encapsulate micronutrients such as iron and added to fruit beverages such as grape, orange, and apricot puree [7]. This study concluded that the CPPs served as a good delivery system and contributed to a decrease in Fe deficiency by increasing bioavailability.

The objectives of this study were to prepare and encapsulate pentapeptide and investigate the shelf life stability of nano-encapsulated pentapeptide in apple juice at 4°C, and to evaluate the physical attributes for 60 days.

Materials and Methods

Materials

A pentapeptide with a sequence of amino acids Glu-Gln-Arg-Pro-Arg (EQRPR) was purchased from Biomatik USA, LLC (Wilmington, DE). Apple juice concentrate was purchased from a local grocery store. Poly (lactic-co-glycolic acid) (PLGA) was purchased from Boehringer Ingelheim Chemicals, Inc. (Ingelheim, Germany). Dimethylsulfoxide (DMSO) was purchased from Fisher Scientific (Fair Lawn, NJ) and

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polystyrene and polystyrene microbeads (120,000xg) for 30 min and 4°C to separate the nanoparticles from the control of nanoparticles in water were ultra-centrifuged using an ultracentrifuge Model J2-21, Beckman Inc. (Brea, CA) at 60,000 rpm for 30 min at 4°C. To investigate the stability of nanoparticles [10-12]. The sample of nano-encapsulated pentapeptide incorporated into apple juice was dried by placing a drop of the sample on a gold coated double-sided tape attached on a metallic sample stand for 48 hours at room temperature before the analysis. The images of the nanoparticles in SEM were obtained using the microscope column and the sample was placed in a specimen camber under a low pressure (vacuum) conditions. Specimen position controllers were used to expose an area of beam/specimen interaction, which produced the image of nanoparticles by generating several detectable signals.

**Testing physical properties of pasteurized apple juice with nanoparticles contained pentapeptide**

The samples of pentapeptide in water, pentapeptide in apple juice, apple juice alone, nano-encapsulated pentapeptide in DI water (pH 3.7), and in apple juice (at 200/400/600 µg/mL) were tested for color and microbial attributes. All the measurements were conducted in triplicate using the following procedures [13].

**Color:** A Chroma-meter from Minolta Inc. (Osaka, Japan) was used to measure the color using a white tile to calibrate the instrument. The apple juice samples were measured in triplicate and the values for ‘L’, ‘a’, and ‘b’ were recorded. The ‘L’ value is a measure of brightness or whiteness, which ranges from 0 to 100 (if L=100, the sample is white, and if L=0, the sample is black). The parameter ‘a’ is a measure of redness, which ranges from -a to +a (green=-a and red=+a) and ‘b’ is a measure of yellowness, which ranges from -b to +b (blue=-b and yellow=+b). The differences of color were calculated using the following equations:

\[
\Delta E_{ab} = \sqrt{\Delta L^2 + \Delta a^2 + \Delta b^2}
\]

Where \(\Delta E_{ab}\) is color different between nano-encapsulated and un-encapsulated pentapeptides.

**Total soluble solids (TSS) and acidity (pH):** TSS is an important parameter that defines the consistency and solubility of apple juice. A refractometer from Atago Inc. (Osaka, Japan) was used to measure total soluble solids of apple juice at ambient temperature. The recommended TSS for apple juice is 11.00 ºBrix. The pH of the solutions was determined using a pH-meter with epoxy pH electrode (VWR Symphony model

Enumeration of microbial survivors: Microbiology tests were conducted to determine the effect of pasteurization. Microbial inactivation and growth during the storage period were determined using TPC and yeast and mold colony counts. Sterile peptone water (0.1% w/v) was used to make serial dilutions. TSA pour plates were prepared to determine the TPC and observed for colony counts after 48 hours of incubation at 30˚C after plating. PDA pour plates were used to determine yeast and mold growth. Observations were made for microbial growth after incubating the plates at 22˚C for 48 hours. Both the TPC and yeast and mold growth were continually observed until 60 days.

Statistical analysis

The statistical analysis was conducted using the GLM procedure of SAS 9.3 (SAS Institute, Cary, NC) and the differences among treatments for the stability, physical properties, and enumeration of the microbial survivors of the nano-encapsulated and un-encapsulated pentapeptides were determined statistically at a 5% significance level. All values are reported as means of triplicate ± SD.

Results and Discussion

Stability of pentapeptide incorporated nanoparticles in pasteurized apple juice by HPLC

The pH of DI water was adjusted to 3.7 to match that of apple juice for comparison of the stability of nano-encapsulated pentapeptide over a storage period. The percentage degradation of pentapeptide at three different concentrations (in water and apple juice) over the storage period was calculated using the equation (R²=0.99) derived from the standard curve plot. Pure apple juice (reconstituted from concentrate) was spiked into the HPLC column to show the absence of any peptides that have the same retention time as the pentapeptide. The apple juice containing known concentrations of pentapeptide (200/400/600 µg/mL) was spiked to confirm the retention time of the pentapeptide which was between 11 and 13 minutes from the start of the run. Figure 1 is the HPLC profiles of the un-encapsulated pentapeptide in water (pH 3.7) showing degradation over time. Hence, the HPLC profiles for only 400 µg/mL are shown. The HPLC analysis showed significant degradation (P value <0.001) of un-encapsulated pentapeptide (at three different concentrations) in water from the 0 day: (200 µg/mL-87.8%, 400 µg/mL-97.4%, 600 µg/mL-91.9%) to the 60th day: (200 µg/mL-41.8%, 400 µg/mL-60.8%, 600 µg/mL-55.8%) of storage. This could be due to the low pH (3.7) of the solution and possible interactions between the pentapeptide and water molecules which have been reported in previous studies [14].

Figure 1 also shows the HPLC profile of apple juice alone, which is composed of organic acids including quinic acid, citric acid, galacturonic acid, and malic acid, and amino acids including aspartic, asparagine and glutamic acids [15]. The retention times of the HPLC analysis of apple juice is representative of these components. The HPLC profiles in Figure 3 shows the un-encapsulated pentapeptide at three different concentrations in apple juice with significant degradation over storage period 0 day: (200 µg/mL-96.6%, 400 µg/mL-98.1%, 600 µg/mL-94.5%) to the 60th day: (200 µg/mL-60.6%, 400 µg/mL-67.5%, 600 µg/mL-59.1%). The effect of pH and the possible interactions with the components of apple juice (organic and amino acids) are the possible reasons for the degradation of the pentapeptide which is shown in Figure 2. The degradation rate of pentapeptide over 60 day study was similar in both water and apple juice and hence can be attributed to the low pH.

Figure 3 show the HPLC profiles of nano-encapsulated pentapeptide incorporated water at pH of 3.7 and apple juice are stable through the duration of the experiment. In this case, Figure 4 shows an illustration of nanoparticles and pentapeptide attached through electrostatic interaction. The electrostatic interactions occur when the nanoparticles that have the negatively/positively charged surface attach to the peptide which has the opposite charge. In a study conducted by Khairallah, peptide fractions derived from rice bran were incorporated in orange juice to test the storage stability [4]. The peptide fractions were found to degrade significantly during the 0 to 42 day storage period. The researcher indicated that the degradation might have occurred due to interactions between peptide fractions and the orange juice components. Therefore, encapsulating the pentapeptide can considerably reduce the interactions and potentially lead to prolonged stability.

Particle size of nano-encapsulated pentapeptide in pasteurized apple juice

Particle size is the most important characteristic that should be monitored for successful encapsulation, loading, and release of pentapeptide. This can also affect nanoparticle stability and the delivery of bioactives. Particle size of pasteurized nano-encapsulated pentapeptide in water (pH of 3.7) at varying concentrations of pentapeptide (200/400/600 µg/mL) was measured from day 0 to day 60 using particle size analyzer. Figure 5 shows the particle size of the nanoparticles which ranged from 82 to 83 nm and remained stable during the storage period (P value <0.05). The uniformity of the particle diameter indicated that the prepared nanoparticles were of even size and the pasteurization had no deleterious effect. Particle size for nano-encapsulated pentapeptide in apple juice with varying concentrations of pentapeptide (200/400/600 µg/mL) was measured from 0 to 60 days, and the diameter of particle size ranged from 82 to 83 nm. The results from the particle size analysis showed successful stability of pentapeptide since they had large surface area to the volume ratio.

Scanning electron microscopy (SEM) of nano-encapsulated pentapeptide in pasteurized apple juice

Scanning electron microscopy was used to determine the form and stability of nanoparticles during the storage period. Images of the nano-encapsulated pentapeptide (400 µg/mL) in apple juice were taken at 0, 2, 7, 14, 30, and 60 days of storage period using the SEM. Figure 6 shows the SEM images of the nano-encapsulated pentapeptide in apple juice (400 µg/mL) from 0 day and 60 day storage. The nanoparticles showed spheroidal shape, which was stable over the storage period. Thus, it can be concluded that the nano-encapsulation provided protection from degradation to the pentapeptide in the apple juice environment.

Physical properties of nano-encapsulated pentapeptide in pasteurized apple juice

Color: Color is an important parameter which can provide an image for the consumer about the quality and acceptability of a food product. The analysis of the apple juice color was based on the lightness (L), redness (a), and yellowness (b). Chroma is a significant parameter of color which indicates the intensity and relates to consumer appeal. Hue of a food product indicates the actual color of the material and contributes to the overall color expectation from the product. Total color difference, ΔE, is the magnitude of overall color difference
compared between the apple juice alone (control) and the nano-encapsulated pentapeptide in apple juice [14]. Figure 7 show Chroma, Hue, and ΔE for the two beverage samples over the storage period of 0 to 60 days.

Chroma of apple juice alone did not show any significant changes during the storage period. For the apple juice incorporated with varying concentrations of pentapeptide (200/400/600 µg/mL) Chroma measurements over the storage period (0-60 days) ranged from 0.5 to 0.3. The results indicated that there was no significant change in the Chroma values among the three concentrations of pentapeptide in apple juice (controls). The Chroma of nano-encapsulated pentapeptide in apple juice during the storage period ranged from 3.2 to 2.8.

The Hue angle of the apple juice alone was stable over the storage period of 60 days, and it was intensely colored (yellow). The Hue angle of apple juice containing varying concentrations of pentapeptide (200/400/600 µg/mL) ranged from -0.1 to 0.1. There is no difference in the Hue among the three concentrations of pentapeptide incorporated apple juice. In a previous study investigating the stability of peptide fractions in orange juice, the changes in the color of orange juice control samples and the orange juice with rice bran peptide fractions was similar over the storage time [4]. These results are consistent with our findings, however, the addition of nanoparticles to the apple juice changed its color to light yellow and the Hue measurements were significantly different over the storage period. The results indicated that the Hue of the nano-encapsulated pentapeptide in apple juice was stable over the storage period which showed Light yellow color. The negative values for Hue angles might be due to the negative values of “a” since the Hue angle is a function of “a” and “b” values.

Total color difference (ΔE) between the control and the sample throughout the storage period between the apple juice alone and the nano-encapsulated pentapeptide incorporated apple juice ranged from 15.2 to 14.2. The results from the color studies indicated significant differences between the apple juice alone (control) and apple juice with the nano-encapsulated pentapeptide. The difference in color remained consistent during the storage period, of 0 to 60 days which is attributed to the addition of nanoparticles which were white in color. These changes in color are significant between the treatments (apple juice and nano-encapsulated pentapeptide); while there was no significant changes over the storage time. The pasteurization temperature of
71°C for 30 seconds did not affect the color during the storage period. Pasteurization and low storage temperature of 4°C were effective in preventing significant changes or deterioration of color during storage which is consistent with a published research [16].

Acidity (pH): The pH of apple juice is an important factor that can determine the stability of pentapeptide. The pH of the nano-encapsulated pentapeptide in apple juice samples and the controls from zero to 30 days did not show any significant change. The results indicated that the pH remained stable for the samples at 3.7, which is within the range of apple juice pH (3.00 to 4.00) that prevented any degradation of the nutritional components. After 60 days the pH of the (controls) pentapeptide in apple juice with varying concentrations (200/400/600 µg/mL) dropped to 3.3, which was not significant. This change in pH might be due to the glutamic acid in the pentapeptide, which is negatively charged that maintained the pH in the acidic range [4]. In a previous study peptide fractions showed rapid degradation when incorporated into orange juice at pH 3.5 in comparison to orange juice stored at pH 7.2. This was purported because of the presence of negatively charged glutamic and aspartic acid in the peptide fractions.
[4]. The addition of pentapeptide did not affect the pH of apple juice since there was no significant difference between apple juice and pentapeptide incorporated apple juice. Similar observations were recorded by previous researchers on the stability of proteins and protein hydrolysates in acidic beverages [4,17].

**Total soluble solid** Total soluble solid (TSS) is a significant factor to determine the consistency of color, clarity and solubility of apple juice. The TSS of apple juice was adjusted to 11 °Brix which is consistent with that of commercially available apple juice. The total soluble solids of the control samples, apple juice alone and pentapeptide incorporated apple juice, at 0, 2, 7, 14, 30, and 60 days were 11 and 11.3 °Brix respectively. The incorporation of pentapeptide in the apple juice did not significantly affect the TSS. The TSS of nano-encapsulated pentapeptide in apple juice at 0, 2, 7, 14, 30 and 60 days was 13 °Brix. Hence, there was a significant difference in TSS between the control samples and the nano-encapsulated pentapeptide in apple juice which was due to the addition of hydrophobic PLGA nanoparticles.

**Enumeration of microbial survivors**

The microbial growth was determined during the storage period using TSA plates for the TPC. The PDA was used to count the yeast and mold colonies. The TSA and PDA plates were prepared to enumerate the microbial colonies during the entire shelf life of the control samples, pentapeptide incorporated in water and apple juice, and nano-encapsulated pentapeptide in apple juice. No microbial growth was observed on both TSA and PDA plates during the storage period of 60 days, which indicated that the pasteurization of apple juice at 71°C was effective in inhibiting microbial growth. Furthermore, apple juice is rich in phytochemicals, which can be defined as non-nutrient plant components including polyphenols, flavonoids, hydroxycinnamic acids, dihydrochalcones, flavonols (quercetin glycosides), catechins and oligomeric procyanidins which are known for antioxidant activities [18]. These phytochemicals have also shown significant antimicrobial activity against human microorganisms [19].

**Conclusion**

PLGA nanoparticles are used to encapsulate the pentapeptide due to its biodegradability and compatibility. PLGA has been extensively used to deliver proteins and peptide and it is approved by FDA (20). PLGA can be degraded in vivo enzymatically or non-enzymatically producing toxicologically safe monomers [20]. Nano precipitation method was used to prepare nano-encapsulated pentapeptide since it is a simple method of preparation and fabrication of polymeric nanoparticles such as PLGA [21]. The formation of nanoparticles by nano-precipitation does not require any external input such as high shearing, sonication, or homogenization [21]. It is a procedure with low energy cost and modest equipment, and a variety of solvents can be used such as DMSO or acetone. Furthermore, this method aids in preparing nano-dispersion in one step, decreasing energy input, and increasing encapsulation yield. Peptides provide several unique features in comparison to proteins and lipids; thus, they are more efficient for therapeutic delivery applications. Peptides can be easily sequenced and synthesized in the laboratory; therefore, they are economical. Peptides can be very specific and bind with high affinity to their cognate receptors, since they are naturally occurring from protein precursors [22]. Furthermore, multifunctional groups of peptides can be incorporated into the nanoparticles to produce a substance containing individual nanoparticle with multiple purposes [22]. Peptides are very attractive and useful molecules for the evolution of nanomaterials; however, maintaining biactivity and stability of peptides are two main challenges facing the food industry. Nano-encapsulation...
of these bioactives can potentially maintain the stability and prevent degradation of bioactive peptides during storage period [22].

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References


