Process Optimization of L-Glutaminase Production; a Tumour Inhibitor from Marine Endophytic Isolate Aspergillus sp. ALAA-2000

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Abstract

L-Glutaminases have received significant attention recently owing to their potential applications. All endophytic fungi recovered from the marine soft sponge Aplysina fistularis were able to produce L-glutaminase. During screening program, Aspergillus sp. ALAA-2000 showed the highest L-glutaminase production levels. The production of L-glutaminase by Aspergillus sp. ALAA-2000 was evaluated under different fermentation modes and parameters. The L-glutaminase synthesis was increased their yield after the optimization of fermentation parameters. The hot water 40°C was the best leaching agent extracted of soy bean for L-glutaminase production (21.89 U/ml) under solid state fermentation (SSF). The highest L-glutaminase activity (91.92 U/ml) was achieved after two days incubation period under submerged fermentation (SmF), L-glutamine, dextrose, cysteine and Magnesium chloride supported the highest L-glutaminase production by Aspergillus sp. ALAA-2000 under SmF at pH 4 and 27°C. Single peak of L-glutaminase was obtained from the culture supernatant of Aspergillus sp. ALAA-2000 through ammonium sulfate precipitation and DEAE-cellulose column chromatography refer to the monomeric nature of L-glutaminase enzyme. The parameters of purified L-glutaminase were optimized as follow: pH 10, stable at 40°C to 50°C, reaction time 30 min and substrate concentration 4.38 mg/ml. Whereas the maximum activator cation is Na+ and different EDTA concentrations have no effect on L-glutaminase activity which means that L-glutaminase enzymes was represent as a non-metallic enzyme.

Keywords: L-glutaminase; Marine endophytic Aspergillus sp.; Fermentation; Optimization; Purification

Introduction

L-Glutaminase (L-glutamine amidohydrolase E.C 3.5.1.2) catalyses the hydrolysis of L-glutamine to glutamic acid and ammonia. In recent years, glutaminase has gained much attention due to their potential applications in pharmaceuticals as an anti-leukemic agent [1,2], flavor enhancing agent [3] and an efficient anti-retroviral agent [4]. Another most promising application of glutaminase is in biosensors for monitoring glutamine levels in mammalian and hybridoma cell cultures without the need of separate measurement of glutamic acid [5]. L-Glutaminase is widely distributed in animal tissues, plants and in a variety of microorganisms including bacteria, fungi and yeast [6-8] of which the most potent producers are fungi [9]. On an industrial scale, glutaminases are produced mainly by Aspergillus and Trichoderma [10-13]. The marine endophytic microflora in the coming decades will be the nature’s best source of chemicals. Natural products metabolized from endophytic microorganisms represent desirable sources for effective therapeutic enzymes [8,14,15]. Marine fungi are rich profile of biologically active metabolites, especially from genera Penicillium, Aspergillus and Fusarium have been used aiming the development of novel therapies for treating cancer. Whereas the marine fungi are least studied than terrestrial counterparts and other ecological group, they have to study due to their production of new metabolites which are not found in terrestrial fungi [16].

Different methods of fermentation technology can be applied for the production of L-glutaminase. Commercial production of L-glutaminase had been carried out using submerged fermentation (SmF) technique [17,18]. But nowadays, solid state fermentation (SSF) has been emerged as a promising technology for the development of several bioprocesses and products including the production of therapeutic enzymes on a large-scale [19]. The primary advantage of SSF is the fact that many metabolites are produced at higher concentration.

Thus the present study, focuses on the L-glutaminase production as potent anticancer agents from a potential culturable marine endophytic isolate Aspergillus sp. ALAA-2000 isolated from the internal healthy tissue of marine invertebrates as well as develop an economically viable bioprocess for production of L-glutaminase by evaluating and optimizing process parameters through manipulating the nutritional and physical parameters using low cost substrates.

Materials and Methods

Microorganism and culture maintenance conditions

Eighteen fungal isolates used in this study were isolated from Egyptian marine sponge Aplysina fistularis, a rich source of endophytic microorganisms on the isolation media peptone yeast extract glucose agar, potato dextrose agar and malt agar [20]. The fungal cultures were purified, maintained at 4°C until use and examined for the production of L-glutaminase enzyme.

Screening of fungal isolates for L-glutaminase productivity

Screening of fungal isolates for L-glutaminase activity was performed using the modified Czapek Dox medium (glucose 2 g,..
L-glutamine 10 g, KH₂PO₄ 1.52 g, KCl 0.52 g, MgSO₄·7H₂O 0.52 g, FeSO₄·7H₂O 0.01 g, agar 20 g, and distilled water 1000 mL) for the plate assay, 3 mL of 2.5% stock solution of phenol red in ethanol (pH 6.2) was added to 1000 mL of Czapek Dox medium. After 72 h of incubation at 26 ± 1°C, the appearance of a pink zone around the fungal colony in an otherwise yellow medium indicated L-glutaminase activity. The fungus which shows highest productivity was subsequently identified and selected to study the optimal conditions for L-glutaminase production in submerged and solid state fermentation.

L-Glutaminase assay

The activity of L-glutaminase is determined by estimating the amount of NH₄⁺ liberated from glutamine. 0.5 ml of enzyme preparation was added to 0.5 ml of 0.04 M L-glutamine 0.5 ml distilled water and 0.5 ml of 0.1 M phosphate buffer (pH 8) then incubated at 37°C for 30 min. After incubation, 0.5 ml of 1.5 M trichloroacetic acid was added to stop the enzymatic reaction. Blank was run by adding the enzyme preparation after the addition of trichloroacetic acid. 0.1 ml of above mixture was taken and added to 3 ml of distilled water followed by addition of 0.2 ml Nessler's reagent. Absorbance was measured at 450 nm using a visible spectrophotometer. One international unit of L-glutaminase (U) was defined the amount of enzyme that liberates 1 μmol of ammonia from glutamate under optimum assay conditions. The enzyme yield was expressed as units/ml according to Imada et al. [17].

Protein estimation

Protein was determined by the absorbance at 280 nm using bovine serum albumin (BSA) as the standard [21]. All the sets have been performed in triplicates.

Effect of different natural wastes and leaching agents on L-glutaminase production

The production of L-glutaminase by Aspergillus sp. ALAA-2000 under solid state fermentation was estimated in 250 ml Erlenmeyer flasks containing 5 g of solid substrate; sugar cane bagasse, wheat bran, corn cobs, soy bean, kidney bean bran, wheat hay, rice bran, rice straw and corn casing separately moistened with mineral salt solution 10 ml of slates solution (glucose 0.6%, KH₂PO₄ 0.1%, MgSO₄·7H₂O 0.05 and KCl 0.05%) the flasks were autoclaved, cooled, inoculated with spore suspension and incubated under static condition for 6 days fermentation period each fermented substrate was extracted with different leaching agent [22]. The leaching out of L-glutaminase from the fermented solids was carried out with different extractants such as hot water (40°C), ethanol, acetone, Tween 80 (1%), sodium chloride (0.1%), citrate buffer pH 3, phosphate buffer pH 7, glycine buffer pH 12, and at a ratio of 1:5 (w/v).

L-Glutaminase activity under submerged fermentation

For the production of L-glutaminase by Aspergillus sp. ALAA-2000, different fermentation media as a modified Czapek Dox medium, mineral salts and starch nitrate were applied. Aspergillus sp. ALAA-2000 strain was incubated in 250 ml Erlenmeyer flasks containing 50 ml of fermentation medium supplemented with L-glutamine and incubated at 27°C and 120 rpm on a rotary shaker for 4 days. At the end of fermentation period the clear supernatant after centrifugation at 4000 rpm for 20 min was used as enzyme preparations.

Optimization of the culture condition for L-glutaminase production under submerged fermentation

Various process parameters that enhance the yield of L-glutaminase by Aspergillus sp. ALAA-2000 strain in a modified Czapek Dox broth medium under shaking were investigated. The impact of incubation time (2-14 days), initial pH (3-9, adjusted with 1 M HCl or NaOH), and incubation temperatures (20-37°C) were evaluated. Moreover, the effect of incorporation of additional various carbon sources (10 g/L) separately in modified Czapek Dox broth medium (raffinose, xylose, mannitol, mannose, sucrose, maltose, sorbose, lactose, galactose, fructose, starch, pectin, cellulose and dextrose) instead of its carbon source (glucose). Organic nitrogen sources; beef extract, yeast extract, peptone and urea as well as inorganic nitrogen sources; ammonium nitrate, ammonium sulfate, ammonium hydrogen citrate were examined for their ability separately to stimulate the enzyme production in modified Czapek Dox broth medium instead of L-glutaminase as N-base. Ten amino acids (lysine, isoleucine, glycine, thiamine, arginine, treptophan, proline, glutamic acid, cysteine and methionine) were examined as a sole nitrogen source for L-glutaminase production. Each of them was added to the medium instead of its nitrogen source in such amount that the final concentration of N-base remained unchanged. Different concentrations (0.025%, 0.05%, 0.1%, 0.15% and 0.2%) of the most suitable amino acid source were tested for their ability to enhance the L-glutaminase production for Aspergillus sp. ALAA-2000 strain. Some element supplementations (MgCl₂, LiCl, CaCl₂, K₂HPO₄, BaCl₂, and NaCl) were supplementation to the modified Czapek Dox broth medium. Different concentrations (0.025%, 0.05%, 0.1%, 0.15% and 0.2%) of the most suitable element supplementation were tested for their ability to enhance the enzyme production by Aspergillus sp. ALAA-2000 strain. All the experiments were conducted in triplicate, subsequently and the mean values are considered.

Purification and characterization of L-glutaminase

The concentrated enzyme was subjected to ammonium sulphate fractionation with concentration ranging between 20% and 100% according to the method of Gomori [23]. The precipitate of crude enzyme was dissolved in a minimum volume of 0.2 M phosphate buffer (pH 6.0) and dialyzed overnight in a dialysis bag against the same buffer at 4°C. The L-glutaminase from ammonium sulphate precipitation (80%) was loaded onto the DEAE-cellulose column chromatography which that equilibrated and eluted with 0.02 M phosphate buffer (pH 8.0). The column was washed with four to five bed volumes of 0.02 M phosphate buffer (pH 8.0). The bound protein was containing linear gradients of NaCl (0.1, 0.25, 0.5, 1.0 M). The flow rate was 0.5 ml/min. The fractions eluted at each NaCl concentration were collected, pooled and tested for protein and L-glutaminase activity [24]. The molecular weight of the purified L-glutaminase was estimated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) as described by Laemmli [25].

The optimum reaction time for the partially purified enzyme was estimated at different times 10-100 min for L-glutaminase and then assayed by the direct nesslerization method. The optimum pH was determined by measuring enzymatic activity at 37°C using different buffers (0.1 M) with various pH values as: citrate-phosphate buffer (pH 3, 4, 5 and 6), phosphate buffer (pH 6-7), Tris-HCl buffer (pH 8-9) and glycine-NaOH buffer (pH 10-12). Optimum temperature was assayed by measuring activity with varying temperatures (27, 37, 47, 57, 67, 77 and 87°C) at the optimum pH values.

Heat stability was determined by incubating the partially purified...
enzyme at various temperatures (20-90°C) for 60 min and then the residual activity was determined at optimum pH and temperature. Partially purified L-glutaminase was incubated individually with different concentrations of L-glutamine in the reaction mixture (0.29, 0.73, 1.46, 2.92, 4.38 and 5.85 mg/ml) to find out the best substrate concentration for enzymatic assay under optimized assay conditions. Different concentrations (0.05, 0.125, 0.25, 0.5, 0.75 and 1.0 ml) of the partially purified L-glutaminase were incubated individually with the optimization concentration substrate solution in the reaction mixture to determine the best enzyme concentration under the optimized assay condition.

The effect of different metal ions including Na⁺, Ca²⁺, Mn²⁺, Mg²⁺, Co³⁺, Ba²⁺ and Ni²⁺ at a final concentration of 1.0 mM as well as EDTA separately purified L-glutaminase were studied. Residual activities in the presence of each chemical was assayed and compared with the control (without additions), which considered as 100% activity.

**Results and Discussion**

**Screening and production of L-glutaminase**

This study proved that Egyptian marine sponge *Aplysina fistularis* is a rich source of endophytic microorganisms. Whereas 18 fungal isolates were obtained on the isolation media peptone yeast extract glucose agar, potato dextrose agar and malt agar. All these marine endophytic isolates were evaluated as fruitful source of the therapeutic antitumor enzymes L-glutaminase. The activity of L-glutaminase enzyme was detected in all fungal strains. The fungal strain under the isolation number, ALAA-2000 was the most potent active producer. Therefore, this strain was identified and selected for further study. According to the analysis of ITS region sequence, together with its phenotypic and biochemical characteristics, the producing strain ALAA-2000 was identified as *Aspergillus* sp. and designated as *Aspergillus* sp. ALAA-2000 [20]. The screening and identification of filamentous fungi capable of secreting extracellular enzymes with biotechnological potential are activities of great importance [26]. Microbiology of sponges and corals can lead to the discovery of new enzymes. A. wentii is a species that is abundant in the marine environment. This species was selected for further study because it produced the highest L-glutaminase activity.

**Production of L-glutaminase; antitumor enzyme under different fermentation processes**

When we used different natural sources as solid substrate fermentation such as sugar cane (bagasse), wheat bran, corn cobs, soy bean, kidney bean bran, wheat hay, rice bran, rice straw and corn casing, as are available and cheap energy sources of L-glutaminase productivity, with using different leaching agents such as hot water 40°C, ethanol, acetone, sodium chloride (0.1%), phosphate buffer (pH 7), glycine buffer (pH 12), Tween 80 (1%) and citrate buffer (pH 3) shown in Table 1. The hot water 40°C was the best leaching agent extracted of soy bean for L-glutaminase production [39]. L-glutaminase production with phosphate buffer (20.14 U/ml), soy bean extracted with acetone (19.69 U/ml), wheat bran leached by NaCl (17.79 U/ml), soy bean leached with NaCl (17.65 U/ml), soy bean leached with citrate buffer (17.00 U/ml), soy been extracted with Tween 80 (16.92 U/ml), wheat bran extracted with citrate buffer (16.00 U/ml), and sugar can bagasse extracted with NaCl (15.47 U/ml), respectively. Consequently agro industrial residues proved to be promising sources for the industrial production of this therapeutic enzyme, especially soybean, wheat bran and sugar can bagasse. Fermentation technology has been widely used for the production of a wide variety of substances of industrial, medical and agriculture. Fermentation technique must be economic and environmentally friendly. Kiruthika and Saraswathy [38] supported soy bean as solid substrate in SSF for L-glutaminase production by *Vibrio aureusus* JK-79 whereas Nathiya et al. [35] reported the maximum L-glutaminase form *A. fumigatus* was produced with paddy straw; El-Sayed [10] by using different agro-industrial byproducts as solid substrates for induction of L-glutaminase by *T. koningii* found that wheat bran was the best substrate (12.1 U/mg protein). Furthermore, out of different agro industrial residues, *A. flavus* (FGNAS-7) produced the highest amount of L-glutaminase with ragi straw [39]. Moreover, Negi and Banerjee [40] tested various parameters such as leaching agent selection, amount of leaching agent, soaking time, and temperature in order to determine optimum extraction conditions of enzymes produced by *A. awamari* nakazawa MTCC 6652 and optimum conditions were achieved in a 10% glycerol. On the other hand, the effect of different three broth culture media named modified Czapek-Dox, mineral salts, and starch nitrate provided with 1% L-glutamine on L-glutaminase production by *Aspergillus* sp. ALAA-2000 were applied. Modified Czapek-Dox medium showed the highest L-glutaminase activity (91.92 U/ml), while starch nitrate medium showed the lowest L-glutaminase activity (73.24 U/ml). The highest level of L-glutaminase activity observed after two days incubation period.

The whole fermentation broth and mycelium of *Aspergillus* sp. ALAA-2000 strain were subjected to different release and chemical treatments, and the results are shown in Table 1.
Table 2: Effect of different organic solvents on L-glutaminase yield obtained from whole broth and mycelium of Aspergillus sp. ALAA-2000 after submerged fermentation for different incubation period.

<table>
<thead>
<tr>
<th>pH value</th>
<th>20°C</th>
<th>27°C</th>
<th>37°C</th>
</tr>
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<tbody>
<tr>
<td>3</td>
<td>131.02</td>
<td>139.19</td>
<td>126.06</td>
</tr>
<tr>
<td>4</td>
<td>115.26</td>
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</tr>
<tr>
<td>9</td>
<td>86.23</td>
<td>80.25</td>
<td>81.56</td>
</tr>
</tbody>
</table>

Table 2: Effect of different organic solvents on L-glutaminase yield obtained from whole broth and mycelium of Aspergillus sp. ALAA-2000 after submerged fermentation for different incubation period.

<table>
<thead>
<tr>
<th>Incubation period (day)</th>
<th>Treatment</th>
<th>L-Glutaminase (U/ml)</th>
<th>Ethyl acetate</th>
<th>Butanol</th>
</tr>
</thead>
<tbody>
<tr>
<td>2</td>
<td>91.92</td>
<td>1.02</td>
<td>2.48</td>
<td>1.17</td>
</tr>
<tr>
<td>4</td>
<td>74.99</td>
<td>2.33</td>
<td>0.15</td>
<td>13.28</td>
</tr>
<tr>
<td>6</td>
<td>81.70</td>
<td>3.06</td>
<td>0.44</td>
<td>11.67</td>
</tr>
<tr>
<td>8</td>
<td>83.31</td>
<td>1.46</td>
<td>9.19</td>
<td>11.67</td>
</tr>
<tr>
<td>10</td>
<td>84.62</td>
<td>1.31</td>
<td>1.02</td>
<td>13.13</td>
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<tr>
<td>12</td>
<td>75.87</td>
<td>0.88</td>
<td>7.29</td>
<td>0.73</td>
</tr>
<tr>
<td>14</td>
<td>71.49</td>
<td>0.73</td>
<td>11.67</td>
<td>0.44</td>
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</table>

Table 3: Effect of different concentrations of substrate, nitrogen and carbon sources, supplementation elements as well as amino acids on L-glutaminase production.

<table>
<thead>
<tr>
<th>L-Glutamine (%)</th>
<th>L-Glutaminase (U/ml)</th>
<th>Nitrogen sources</th>
<th>L-Glutaminase (U/ml)</th>
<th>Carbon source</th>
<th>L-Glutaminase (U/ml)</th>
<th>Supplementation elements</th>
<th>L-Glutaminase (U/ml)</th>
<th>Amino acids</th>
<th>L-Glutaminase (U/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.5</td>
<td>32.09</td>
<td>Without nitrogen</td>
<td>4.52</td>
<td>Without carbon</td>
<td>119.64</td>
<td>Control</td>
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<td>Control</td>
<td>86.08</td>
</tr>
<tr>
<td>1.0</td>
<td>52.52</td>
<td>Peptone</td>
<td>69.30</td>
<td>Xylose</td>
<td>100.67</td>
<td>Lysine</td>
<td>134.67</td>
<td>Lysine</td>
<td>134.67</td>
</tr>
<tr>
<td>1.6</td>
<td>73.63</td>
<td>Beef extract</td>
<td>31.66</td>
<td>Mannitol</td>
<td>104.90</td>
<td>Isoleucine</td>
<td>124.16</td>
<td>Isoleucine</td>
<td>124.16</td>
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<td>2.0</td>
<td>75.87</td>
<td>Yeast extract</td>
<td>59.09</td>
<td>Mannose</td>
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<td>Glycine</td>
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<td>126.93</td>
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<tr>
<td>2.5</td>
<td>78.79</td>
<td>Ammonium sulphate</td>
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<td>Sucrose</td>
<td>119.20</td>
<td>Thiamine</td>
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<td></td>
<td></td>
<td>Diammonium citrate</td>
<td>77.32</td>
<td>Maltose</td>
<td>99.07</td>
<td>Arginine</td>
<td>127.37</td>
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<td></td>
<td></td>
<td>Sodium nitrate</td>
<td>93.67</td>
<td>Sorbose</td>
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<td>Tryptophosphate</td>
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<td></td>
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<td>Ammonium nitrate</td>
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<td>Lactose</td>
<td>103.44</td>
<td>Proline</td>
<td>132.12</td>
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<td>132.12</td>
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<tr>
<td></td>
<td></td>
<td>L-Glutamine</td>
<td>91.92</td>
<td>Galactose</td>
<td>109.72</td>
<td>Glutamic acid</td>
<td>117.01</td>
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<td></td>
<td>109.43</td>
<td>Fructose</td>
<td>100.67</td>
<td>Cystine</td>
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<td></td>
<td></td>
<td>Starch</td>
<td>107.97</td>
<td>Methionine</td>
<td>125.91</td>
<td>Methionine</td>
<td>125.91</td>
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</table>

Table 4: Effect of different concentrations of substrate, nitrogen and carbon sources, supplementation elements as well as amino acids on L-glutaminase production.
Aspergillus sp. ALAA-2000 strain was required the inducer L-glutamine for the best biosyntheses of L-glutaminase (109.43 U/ml). However beef extract and sodium nitrate showed lowest production of L-glutaminase (31.66 and 7.59 U/ml, respectively) by Aspergillus sp. ALAA-2000. Katikala et al. [48] reported that marine bacterial isolate LG24 gave the highest yield of extracellular L-glutaminase 22.68 U/ml in 120 h when L-glutamine supplemented as sole carbon and nitrogen source in the media.

On the other hand, to determine the best carbon sources for therapeutic enzyme production L-glutaminase produced by Aspergillus sp. ALAA-2000 grown in modified Dox medium supplemented by different carbon sources are represented in Table 4. It was obvious that dextrose increased the L-glutaminase production by Aspergillus sp. ALAA-2000 to 124.02 U/ml followed by mannose, sucrose, sorbose, pectin, galactose, cellulose, starch, mannotol, lactose, xylose, maltose and raffinose (121.09, 119.20, 115.26, 113.80, 109.72, 107.97, 107.97, 104.90, 103.44, 100.67, 99.07 and 84.62 U/ml), respectively. Overall, all carbon sources did not exhibited much significant increase in L-glutaminase production due to Dox modify medium free carbon relatively yielded a high amount of L-glutaminase (119.64 U/ml). On the contrary, the yield of L-glutaminase from S. griseus was increased to 26.3 U/ml by utilized the galactose as the carbon source [47].

**Effect of different amino acids on L-glutaminase production:** Data illustrated in Table 4 showed effect lysine, isoleucine, glycine, thiamine, arginine, treptophan, proline, glutamic acid, cysteine, and methionine on L-glutaminase production. All amino acids had stimulation effect on L-glutaminase production with different ratio ranged between 57.8% to 124.02 U/ml enhanced L-glutaminase production by the marine endophytic strain Aspergillus sp. ALAA-2000 was studied. Out of this concentration cysteine at a concentration of 0.1% supported the highest L-glutaminase production 135.98 U/ml. Amino acids were served as source of energy and carbon in addition to nitrogen as previously reported for L-glutaminase production by Fusarium sp. [46].

**Effect of addition and elimination of different supplementation elements on L-glutaminase production:** Magnesium chloride enhanced L-glutaminase production by Aspergillus sp. ALAA-2000 to 97.75% but LiCl, CaCl, K2HPO, BaCl, and NaCl decrease it to 74.41, 71.49, 57.92, 77.62 and 78.79 U/ml, respectively (Table 4). Therefore, the effect different concentrations of MgCl2 on L-glutaminase production were studied. L-glutaminase activity increased with the increasing of MgCl2 concentration till 0.1% after that decreased. The main components of the modified Dox medium are KCl, MgSO4·7H2O, and KH2PO4. Elimination of these components from culture medium did not strongly inhibited L-glutaminase production. Inhibition ratio reached up to 6%, 3% and 22% with the elimination of KCl, MgSO4·7H2O, and KH2PO4, respectively. Na+ enhanced of L-glutaminase production by A. fumigatus and A. oryzae NCIM 1212 [35,44].

**Partially purification of L-glutaminase with DEAE-cellulose**

The purification steps from the crude extract of Aspergillus sp. ALAA-2000 are summarized in Table 5 and Figure 1. L-Glutaminase was purified from the culture filtrate by 80% saturation of ammonium sulfate (the fraction showed the highest glutaminase activity), resulted in specific activity of 9.52 U/mg protein, 8.89 purification folds followed by DEAE-cellulose column chromatography. The purification of the glutaminase was increased 36.72 fold with overall yields of 37.42 %. The partially purified of L-glutaminase produced by P. brevicompactum NRC 829 has total activity 321.6 U, total protein 0.37 mg, Specific activity 869.08 (U/mg), purification 162.75-fold and yield 48.21% after Sephadex G-200 [42]. While Ali and Mohamed [49] reported that the total activity 44 U, total protein 0.33 mg, specific activity 133 U/mg, purification 230-fold and yield 25% of L-glutaminase produced by P. politans NRC510.

<table>
<thead>
<tr>
<th>Purification Step</th>
<th>Total activity (U)</th>
<th>Total protein (mg)</th>
<th>Specific activity (U/mg)</th>
<th>Yield (%)</th>
<th>Fold</th>
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</thead>
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<td>Filtrate</td>
<td>210</td>
<td>195</td>
<td>1.07</td>
<td>100</td>
<td>1</td>
</tr>
<tr>
<td>(NH4)2SO4 precipitation</td>
<td>120</td>
<td>12.6</td>
<td>9.52</td>
<td>57.14</td>
<td>8.89</td>
</tr>
<tr>
<td>DEAE-cellulose</td>
<td>78.6</td>
<td>2.0</td>
<td>39.3</td>
<td>37.42</td>
<td>36.72</td>
</tr>
</tbody>
</table>

**Table 5:** Summary of the purification steps of L-glutaminase.

![Figure 1: Elution diagram of L-glutaminase using DEAE-cellulose column chromatography.](image)
Characterization of partially purified L-glutaminase activity

Due to the purification of L-glutaminase showed the maximum single peak of L-glutaminase relative activity (107.2%) produced by Aspergillus sp. ALAA-2000 at 47°C refer to the mono meric nature of L-glutaminase enzymes. The thermo stability of L-glutaminase activities showed a wide range of L-glutaminase thermo stability from (30°C to 60°C) and higher than this temperature stability gradually decreased. Maximum thermo stability of L-glutaminase activity at 40°C and 50°C (100 and 100%), respectively and sharply decreases the activity (41%) at 90°C (Table 6). The maximum activity of purified L-glutaminase from P. brevicompactum NRC 829 and S. variabilis were insured at incubation temperatures 30°C and 50°C, respectively [42,46]. Furthermore, Elshafei et al. [42] reported that L-glutaminase was stable at 50°C to 60°C for 60 min and it retained about 92% and 66% of its initial activity after incubation at 70°C for 30 min and 80°C for 5 min, respectively without the substrate.

The L-glutaminase produced by marine endophytic Aspergillus sp. ALAA-2000 strain exhibited classical pattern of pH activity relationship with pH optimum at pH10 (Table 6). Elshafei et al. [42] and Dura et al. [24] reported that the pH 7 and 8.5 were increased L-glutaminase activity purified from P. brevicompactum NRC 829 and Debaryomyces sp. CECT 11815, respectively. Figure 2 showed that, the activity of partially purified L-glutaminase was tested at 10, 20, 30, 40, 50, 60, 70, 80, 90 and 100 min incubation times. The results showed on peak of activity after 30 min of incubation and it was 100%. The best incubation time for L-glutaminase production by P. brevicompactum NRC 829 and S. variabilis was 60 min [42,46].

The effect of different concentrations of L-glutamine 0.29, 0.73, 1.46, 2.92, 4.38 and 5.85 mg/ml on the activity of L-glutaminase were detected. The activity of L-glutaminase was increased with increasing substrate concentration ratio to 4.38 mg/ml of L-glutamine (114.1 U/ml). Increasing substrate concentration resulted decreased in the activity of L-glutaminase. Different partially purified enzymes concentrations, 0.05, 0.125, 0.25, 0.5, 0.75 and 1 ml of L-glutaminase were attempted in the enzymatic reaction with glutamine at concentration of 0.75 mg/ml. The suitable enzyme concentration in the reaction mixture that supported the highest activity was 113.4% for L-glutaminase produced by 1ml. Elshafei et al. [42] and Ali and Mohamed [49] reported the high affinity of L-glutaminase activity produced by P. brevicompactum NRC 829 and P. politans NRC510 with 1.66 mM and 10 µmol concentrations of L-glutamine, respectively.

The effect of Ni²⁺, Ba²⁺, Ca²⁺, Na⁺, Co²⁺, Mn²⁺ and Mg²⁺ with final concentration of 0.5 mM of each cation on partially purified L-glutaminase activities was studied. Compared to control Ba²⁺, Ca²⁺, Na⁺, Co²⁺, Mn²⁺ and Mg²⁺ increase the L-glutaminase activity by 15.7, 6.8, 41.7, 15.4, 9.4 and 15.8, respectively but Ni²⁺ achieved decrease enzyme activity. The maximum activator cation is Na⁺ by 41.7% compared with control. On the other hand, the effect of different EDTA concentrations (0.0001, 0.001, 0.005, 0.01, 0.05 and 0.1 M) on the activity of partially purified L-glutaminase was studied. Results indicated that different EDTA concentrations have no effect on L-glutaminase activity which means that L-glutaminase enzymes was represent as a non metalloenzyme. Ali and Mohamed [49] and Elshafei et al. [42] reported that sodium chloride was the best activator for L-glutaminase activity from P. politans NRC 510 and P. brevicompactum NRC 829. Also, Elshafei et al. [42] reported L-glutaminase produced by P. brevicompactum NRC 829 as non metalloenzyme but Abd-Alla et al. [46] reported L-glutaminase produced by S. variabilis as metallo enzyme.

Conclusion

The potential of isolated marine endophytic strain Aspergillus sp. ALAA-2000 for L-glutaminase production was analyzed under different fermentation modes with different process parameters. Maximum production was supported with L-glutamine, dextrose,

<table>
<thead>
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<th>Thermo stability</th>
<th>L-Glutaminase relative activity (%)</th>
<th>pH</th>
<th>L-Glutaminase relative activity (%)</th>
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Table 6: Effect of thermo stability and different pH values on L-gluatminase activity.

![Figure 2: Effect of reaction time on L-glutaminase activity.](image-url)
cysteine, and Magnesium chloride under SmF at pH 4 and 27°C. The parameters of purified L-glutaminase were optimized as follow: pH 10, stable at 40°C to 50°C, reaction time 30 min, and L-glutamine 4.38 mg/ ml whereas the maximum activator cation is Na+ and different EDTA concentrations have no effect on L-glutaminase activity.

References


