

Food Additives - Risk Factors for Renal Failure

Sivakumar J T Gowder*

College of Applied Medical Sciences, Qassim University, Buraydah, Kingdom of Saudi Arabia

The kidney is an organ of excretion, transport and metabolism. It is a complicated organ, comprising several different cell types and having a sophisticated three dimensional organization [1]. Due to structural complexity, the intact kidney is difficult to employ for adequate study of many biochemical, pharmacological and physiological processes. Cell cultures, either primary cells or established cell lines [2], have provided useful model systems for the study of renal cell functions. Primary cultures of proximal tubule cells have been considered as an appropriate model for the study of proximal tubule cell function or renal intact function [3]. Kidneys are target organs for toxicity and also many diseases and hence initiate systemic pathophysiological processes through their complex functions. Thus, kidney damage and function has been considered as a major public health hazard [4]. In developed countries, lifestyle diseases like diabetes and hypertension are the important causes of renal failure. In developing countries, kidney diseases result from even infectious conditions like cholera, malaria, schistosomiasis and hepatitis B [5,6]. Drugs and toxins are injurious to kidney and increase the risk of progression of renal failure [7]. Smoking, obesity, red meats, sodium, and sugar-sweetened beverages and food additives also cause renal failure [8].

Egyptian records of 5000 years ago describe the brewing of beer from barley. This must be the first recorded use of an additive, yeast, which was soon to be followed by other additives for it was an early practice to put in flavor principles, as hops are put in today [9]. The statutory definition of the term 'food additive' in the Federal Food, Drug and Cosmetic Act exempts any substance "generally recognized, among experts qualified by scientific training and experience to evaluate its safety, as having been adequately shown through scientific procedures to be safe under the conditions of its intended use" [10]. The Codex Alimentarius Commission, a commission which was established to develop a set of food standards to govern international trade, says: 'food additive' means any substance not normally consumed as food by itself and not normally used as a typical ingredient of the food, whether or not it has nutritive value, the addition of which to food for a technological (including organoleptic) purpose in the manufacture, processing, preparation, treatment, packing, packaging, expected to result (directly or indirectly) in it or its by-products becoming a component of or otherwise affecting the characteristics of such foods. The term does not include 'contaminants' or substances added to food for maintaining or improving nutritional qualities [11]. In 1938, Congress passed the Food, Drug, and Cosmetic Act, giving the Food and Drug Administration (FDA), power to remove from foods any chemicals found to be unsafe for human consumption. The Food Additives Amendment of 1958 further broadened that control by requiring the food industry to demonstrate the safety of any new food additives. These ended on a GRAS (Generally Recognized as Safe) list. The FDA is in the process of reevaluating all the additives on this list, to determine whether they are indeed safe [12].

Food additives have been categorized into preservatives, antioxidants, colorants, antioxidants, emulsifiers, flavors, filters and colorants [13]. Toxicological studies on food additives should be made according to necessity and adequacy. If substances belong to a well studied and unexceptionally safe group, carrying a strong initial

presumption of safety, toxicological data on each specific substance are generally not needed. But if a substance is consumed at high levels and there is no previous experience with its use or it does not occur naturally in food, more direct toxicological data are essential [10]. The interpretation of toxicological tests requires the determination of a dosage level at which no adverse effects are observed. The limitations surrounding precise estimation of the no-adverse-effect level have been discussed in an earlier report [14].

Though there are many food additives that exhibit toxicity, this article has been focused on certain widely used additives that have nephrotoxic potential. Monosodium glutamate (MSG) is quite often used in Chinese food items. Recent studies reveal involvement of MSG in distortion of renal cytoarchitecture – that is, cellular proliferation of mesangial cells and infiltration of inflammatory cells [15]. Use of melamine (an additive in milk), results in acute renal failure in thousands of Chinese infants [16]. Potassium bromate ($KBrO_3$), a food additive and also a water disinfection by-product results in kidney damage accompanied by changes in many brush border enzymes [17]. Several studies conclude nephrotoxic potential of common food additives such as cyodax, aluminium chloride, beta-myrcene, tartrazine, borax, Aloe, Sodium nitrite, Aspartame, Diazoaminobenzene, sodium metabisulphite, Methyleugenol, N-nitrosodimethylamine, sodium-o-phenylphenate, d-limonene, thiabendazole, and butylatedhydroxytoluene [18-33].

Due to inadequate data on the nephrotoxicity of the food additive (flavor) cinnamaldehyde, we have designed our study to evaluate the effect of cinnamaldehyde on kidney. The occurrence of cinnamaldehyde is widely noticed in most of the food, medicinal and cosmetic products. Thus, cinnamaldehyde has a high potential for human consumption in the world. Cinnamaldehyde treated rats equivalent to the WHO suggested ADI level (73.5 mg/kg body weight/day) showed many histopathological changes of kidney accompanied by an increased activity of marker enzymes and an imbalance in the antioxidant status [34,35]. We have concluded that cinnamaldehyde induced renal damage, is due to the reactive oxygen species that formed while in the free radical scavenging reactions [36]. Thus, the toxic food additives may induce oxidative stress and thereby result in renal damage or failure. Antioxidants (vitamins) play a significant role to ameliorate toxicity. Thus, fruits and vegetables in routine diet might protect human health from toxic hazards at certain extent [37].

*Corresponding author: Sivakumar J T Gowder, College of Applied Medical Sciences, Qassim University, Buraydah 51452, Kingdom of Saudi Arabia, Tel: +966566873969; Fax: +96663802268; E-mail: sivakumargowder@yahoo.com

Received December 18, 2013; Accepted December 19, 2013; Published December 23, 2013

Citation: Gowder SJT (2013) Food Additives - Risk Factors for Renal Failure. J Socialomics 3: e122. doi:10.4172/2167-0358.1000e122

Copyright: © 2013 Gowder SJT. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

References

1. Yokoo T, Ohashi T, Shen JS, Sakurai K, Miyazaki Y, et al. (2005) Human mesenchymal stem cells in rodent whole-embryo culture are reprogrammed to contribute to kidney tissues. *ProcNatlAcadSci USA* 102:3296-3300.
2. Rops AL, van der Vlag J, Jacobs CW, Dijkman HB, Lensen JF, et al. (2004) Isolation and characterization of conditionally immortalized mouse glomerular endothelial cell lines. *Kidney Int* 66:2193-201.
3. Gowder SJT, McMartin KE (2010) Development of a primary culture system of rat kidney proximal tubule cells for transport studies. *J Epithelial BiolPharmacol* 3:15-19.
4. Eckardt KU, Coresh J, Devuyst O, Johnson RJ, Köttgen A, et al. (2013) Evolving importance of kidney disease: from subspecialty to global health burden. *Lancet* 382:158-69.
5. Broeck DV, De Wolf MJS (2012) Brefeldin A and Exo1 Completely Release the Block of Cholera Toxin Action by a Dipeptide Metalloendoprotease Substrate. *Cholera*, SivakumarJoghiThathaGowder, Intech Publications, Croatia.
6. WHO (2002) Reducing Risks, Promoting Healthy Life. In *The World Health Report 2002*, Geneva: WHO.
7. CDC (2010) National Chronic Kidney Disease Fact Sheet: General Information and National Estimates on Chronic Kidney Disease in the United States. *Diabetes Public Health Resource*.
8. Chang A, Van Horn L, Jacobs DR Jr, Liu K, Muntner P, et al. (2013) Lifestyle-related factors, obesity, and incident microalbuminuria: the CARDIA (Coronary Artery Risk Development in Young Adults) study. *Am J Kidney Dis* 62:267-75.
9. Taylor RJ (1980) *Food additives*. John Wiley and Sons, New York, USA.
10. Oser BL, Hall RL (1977) Criteria employed by the expert panel of FEMA for the GRASEvaluation of flavouring substances. *Food CosmetToxicol* 15: 457- 466.
11. JECFA (FAO-WHO Joint Expert Committee on Food Additives) (1975) *General Principles*. Codex Alimentarius Commission Procedural Manual. Food and Agricultural Organization, Rome, Italy.
12. Lecos C (1984) Food preservatives: a fresh report. *FDA Consum* 4: 23-25.
13. Weber RW (1993) Food additives and allergy. *Ann Allergy* 70: 183-190.
14. NAS-NRC (National Academy of Sciences- National Research Council) (1975) *Principles for Evaluating Chemicals in the Environment*. Am J Public Health.
15. Dixit SG, Rani P, Anand A, Khatri K, Chauhan R, et al. (2013) To study the effect of monosodium glutamate on histomorphometry of cortex of kidney in adult albino rats. *RenFail*.
16. Yang L, Huo D, Jiang Y, Hou C, Zhang S (2013) Monitoring the adulteration of milk with melamine: a visualised sensor array approach. *Food AdditContam Part A Chem Anal Control Expo Risk Assess* 30:786-95.
17. Ahmad MK, Khan AA, Mahmood R (2013) Taurine ameliorates potassium bromate-induced kidney damage in rats. *Amino Acids* 45:1109-1121.
18. Huang C, Lei H, Zhao X, Tang H, Wang Y (2013) Metabolic influence of acute cyadox exposure on Kunming mice. *J Proteome Res* 12:537-45.
19. El-Kenawy AE, Hussein Osman HE, Daghestani MH (2012) Role of propolis (bee glue) in improving histopathological changes of the kidney of rat treated with aluminum chloride. *Environ Toxicol*.
20. Chan PC, Cesta MF, Sills RC, Bishop JB, Bristol DW, et al. (2010) NTP technical report on the toxicology and carcinogenesis studies of beta-myrcene (CAS No. 123-35-3) in F344/N rats and B6C3F1 mice (Gavage studies). *NatlToxicol Program Tech Rep Ser* 557:1-163.
21. Amin KA, Abdel Hameid H, AbdElsttar AH (2010) Effect of food azo dyes tartrazine and carmoisine on biochemical parameters related to renal, hepatic function and oxidative stress biomarkers in young male rats. *Food ChemToxicol* 48:2994-2994.
22. Pongsavee M (2009) Effect of borax on immune cell proliferation and sister chromatid exchange in human chromosomes. *J Occup Med Toxicol* 10:4:27.
23. Yokohira M, Matsuda Y, Suzuki S, Hosokawa K, Yamakawa K, et al. (2009) Equivocal colonic carcinogenicity of *Aloe arborescens* Miller var. *natalensis* Berger at high-dose level in a Wistar Hannover rat 2-y study. *J Food Sci* 74:T24-30.
24. Hassan HA, El-Agmy SM, Gaur RL, Fernando A, Raj MH, et al. (2009) In vivo evidence of hepato- and reno-protective effect of garlic oil against sodium nitrite-induced oxidative stress. *Int J BiolSci* 5:249-55.
25. Belpoggi F, Soffritti M, Padovani M, DegliEsposti D, Lauriola M, et al. (2006) Results of long-term carcinogenicity bioassay on Sprague-Dawley rats exposed to aspartame administered in feed. *Ann N Y AcadSci* 1076:559-577.
26. Ress NB (2002) NTP Technical Report on the metabolism, toxicity and predicted carcinogenicity of diazoaminobenzene (CAS No. 136-35-6). *Toxic Rep Ser*.
27. Ribera D, Jonker D, Narbonne JF, O'Brien J, Antignac E (2001) Absence of adverse effects of sodium metabisulphite in manufactured biscuits: results of subacute (28-days) and subchronic (85-days) feeding studies in rats. *Food AdditContam* 18:103-14.
28. NTP (2000) NTP Toxicology and Carcinogenesis Studies of Methyleugenol (CAS NO. 93-15-2) in F344/N Rats and B6C3F1 Mice (Gavage Studies). *NatlToxicol Program Tech Rep Ser* 491:1-412.
29. Noronha RF (1997) Decreased incidence of renal tumors in DMN treated Balb/C mice after orchietomy. *J SurgOncol* 9:463-468.
30. Hiraga K, Fujii T (1983) Induction of tumours of the urinary system in F344 rats by dietary administration of sodium o-phenylphenate. *Food CosmetToxicol* 19:303-310.
31. Webb DR, Ridder GM, Alden CL (1989) Acute and subchronic nephrotoxicity of d- limonene in Fischer 344 rats. *Food ChemToxicol* 27:639-649.
32. Mizutani T, Yoshida K, Ito K, Kawazoe S (1992) Sex difference in the nephrotoxicity of thiabendazole in mice depleted of glutathione by treatment with DL-buthioninesulphoximine. *Food ChemToxicol* 30:247-250.
33. Takahashi O (1992) Haemorrhages due to defective blood coagulation do not occur in mice and guinea-pigs fed butylatedhydroxytoluene, but nephrotoxicity is found in mice. *Food ChemToxicol* 30: 89-97.
34. Gowder S JT, Devaraj H (2008) Food flavor cinnamaldehyde - induced biochemical and histological changes in the kidney of male albino wistar rat. *Environ Toxicol Pharma* 26:68-74.
35. Gowder S JT, Devaraj H (2006) Effect of food flavor cinnamaldehyde on the antioxidant status of rat kidney. *B ClinPharmacolToxicol* 99: 379-382.
36. Gowder S, Devaraj H (2010) A review on the nephrotoxicity of food flavor Cinnamaldehyde. *Current Bioactive Compounds* 6: 106-117.
37. Gowder S JT (2013) An updated review of toxicity of bisphenol A (BPA) with special reference to the kidney. *Current Molecular Pharmacology* [accepted; volume -6].