

Exosomes: Extracellular Vesicles Transporting Macromolecules

A Ivana Scovassi

Istituto di Genetica Molecolare CNR, Via Abbiategrosso 207, 27100 Pavia, Italy

Extracellular vesicles (EVs), firstly identified in 1983 in the conditioned culture medium collected during the maturation of reticulocytes into erythrocytes [1,2], are released by the cells into the extracellular milieu. EVs are present in all human body fluids; their number and content are often modulated in pathological samples, thus making interesting their evaluation as possible disease biomarkers [3]. On the basis of their size (50-5000 nm) and origin (multivesicular bodies or plasma membrane), three main classes have been described within the family of EVs (Table 1), i.e., exosomes, microvesicles and apoptotic bodies [4]. EVs can be isolated from few milliliters of body fluids (mainly blood) through sequential centrifugations, filtration and sucrose gradients. Notably, they host macromolecules such as proteins, lipids and nucleic acids, and can release these molecules into recipient cells, thus becoming a vehicle for cellular communications [5,6].

The best-characterized EVs are the cup-shaped exosomes, so called in 1987 to define small vesicles surrounded by a double-layer membrane and containing organelle-free cytosol [7]. Microscopic analysis of exosomes by immunocytochemistry procedures provides the first useful information about their content; more sophisticated and miniaturized “omics” protocols can be applied to detect specific protein or RNA (especially miRNA) signatures in EVs.

Exosomes derived from cancer cells are able to transfer some modulators of tumor formation, progression and spread, such as oncogenes/oncoproteins, pro-angiogenic and anti-apoptotic factors, immunomodulators and macromolecules promoting epithelial-to-mesenchymal transition (EMT) (Figure 1). In this respect, the characterization of cancer-derived circulating exosomes (and, in general, EVs) could be useful for early cancer diagnosis and for monitoring prognosis and follow up, avoiding invasive procedures in the favor of the so-called “liquid biopsy” [8-10].

As an example of protein biomarkers, proteomic/peptidomic approaches applied to serum protein profiling of patients with different solid tumors allowed the identification of a correlation between high levels of the protein SPINK1 (serine peptidase inhibitors Kazal type) and poor prognosis [11]. Also the investigation of the exosomal protein HMGB1 (high mobility group box 1) revealed that, when expressed at

increased levels in serum from cancer patients, it is a general marker of bad outcome [12]. An active search for miRNAs packaged within EVs is done, exploiting modern technologies such as microarray profiling, PCR arrays and next generation sequencing; in fact, specific miRNA “signatures” have been found in serum from cancer patients [13,14]. However, to validate circulating EVs and their cargo as disease markers, multicenter surveys are required and standard procedures of isolation/characterization have to be developed.

EVs/exosomes field is very “hot”, not only because these extracellular structures mediate cell-to-cell communication but also because they can be considered as circulating biomarkers, in particular of cancer, where they promote proliferation, migration, invasion and metastasis [15,16]. Given the growing interest towards cell free circulating EVs, the *Journal of Extracellular Vesicles* (Co-Action Publishing) becomes the reference Journal for the scientific community; updated libraries of macromolecules included in exosomes can be found at <http://microvesicles.org> and <http://evpedia.info>. As a cautionary note, it has to be taken into account that validation of circulating biomarkers, as well as setting of reproducible sensitive/specific protocols focusing on the pre-analytical, analytical and post-analytical quality requirements for identifying and validate reliable cancer markers, are required to make “liquid biopsy” a fundamental clinical tool [17].

References

- Harding C, Heuser J, Stahl P (1983) Receptor-mediated endocytosis of transferrin and recycling of the transferrin receptor in rat reticulocytes. *J Cell Biol* 97: 329-339.
- Pan BT, Johnstone RM (1983) Fate of the transferrin receptor during maturation of sheep reticulocytes in vitro: selective externalization of the receptor. *Cell* 33: 967-978.
- van der Pol E, Böing AN, Harrison P, Sturk A, Nieuwland R (2012) Classification, functions and clinical relevance of extracellular vesicles. *Pharmacol Rev* 64: 676-705.
- Raposo G, Stoorvogel W (2013) Extracellular vesicles: Exosomes, microvesicles, and friends. *J Cell Biol* 200: 373-383.
- Shifrin DA Jr, Demory Beckler M, Coffey RJ, Tyska MJ (2013) Extracellular vesicles: Communication, coercion, and conditioning. *Mol Biol Cell* 24: 1253-1259.
- Lo Cicero A, Stahl PD, Raposo G (2015) Extracellular vesicles shuffling intercellular messages: For good or for bad. *Curr Opin Cell Biol* 35: 69-77.
- Johnstone RM, Adam M, Hammond JR, Orr L, Turbide C (1987) Vesicle formation during reticulocyte maturation. Association of plasma membrane activities with released vesicles (exosomes). *J Biol Chem* 262: 9412-9420.

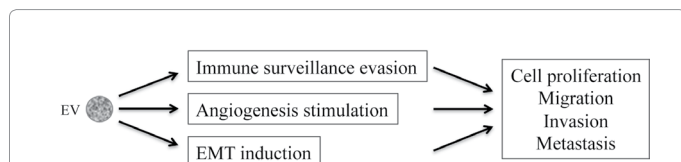


Figure 1: Effects of extracellular vesicles (EVs) secreted by cancer cells. In recipient cells, EVs stimulate angiogenesis, affect the immune system and promotes epithelial-to-mesenchymal transition (EMT). These changes, in turn, favor cancer cell proliferation, migration and invasion, leading to metastasis formation.

EV type	Origin	Shape	Size (nm)	Density (g/cm ³)
Exosomes	Multivesicular bodies	Cup-shaped	50-150	1.13-1.19
Microvesicles	Plasma membrane	Heterogeneous	50-2000	?
Apoptotic bodies	Plasma membrane	Heterogeneous	50-5000	1.16-1.28

Table 1: Distinctive features of the different EVs.

***Corresponding author:** A Ivana Scovassi, Istituto di Genetica Molecolare CNR, Via Abbiategrosso 207, 27100 Pavia, Italy, Tel +39-0382-546334; E-mail: scovassi@igm.cnr.it

Received April 06, 2016; Accepted April 08, 2016; Published April 13, 2016

Citation: Scovassi AI (2016) Exosomes: Extracellular Vesicles Transporting Macromolecules. *Biochem Pharmacol* (Los Angel) 5: e181. doi:10.4172/2167-0501.1000e181

Copyright: © 2016 Scovassi AI. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

8. EL Andaloussi S, Mäger I, Breakefield XO, Wood MJ (2013) Extracellular vesicles: biology and emerging therapeutic opportunities. *Nat Rev Drug Discov* 12: 347-357.
9. Verma M, Lam TK, Hebert E, Divi RL (2015) Extracellular vesicles: potential applications in cancer diagnosis, prognosis, and epidemiology. *BMC Clin Pathol* 15: 6.
10. Fujita Y, Yoshioka Y, Ochiya T (2016) Extracellular vesicle transfer of cancer pathogenic components. *Cancer Sci* DOI: 10.1111/cas.12896.
11. Räsänen K, Itkonen O, Koistinen H, Stenman UH (2016) Emerging roles of SPINK1 in cancer. *Clin Chem* 62: 449-457.
12. Pilzweiger C, Holdenrieder S (2015) Circulating HMGB1 and RAGE as clinical biomarkers in malignant and autoimmune diseases. *Diagnostics (Basel)* 5: 219-253.
13. He Y, Lin J, Kong D, Huang M, Xu C, et al. (2015) Current state of circulating microRNAs as cancer biomarkers. *Clin Chem* 61: 1138-1155.
14. Wang J, Chen J, Sen S (2016) MicroRNA as biomarkers and diagnostics. *J Cell Physiol* 231: 25-30.
15. Ciardiello C, Cavallini L, Spinelli C (2016) Focus on extracellular vesicles: New frontiers of cell-to-cell communication in cancer. *Int J Mol Sci* 17.
16. Kosaka N, Yoshioka Y, Fujita Y, Ochiya T (2016) Versatile roles of extracellular vesicles in cancer. *J Clin Invest* 126: 1163-1172.
17. López E, Madero L, López-Pascual J, Latterich M (2012) Clinical proteomics and OMICS clues useful in translational medicine research. *Proteome Sci* 10: 35.