

Effects of Ethanol Extract and its Different Fractions of *Phrynium imbricatum* (Roxb.) Leaves on *In Vitro* Anthelmintic and their Condensed Tannin Content

Mohammad Shah Hafez Kabir¹, Abu Monsur Dinar Md², Mohammed Munawar Hossain¹, Muhammad Abdulla Al Noman³, Fahima Zaheed³, Rabiul Hossain Md³, Abul Hasanat¹, Raju Dash⁴, Mominur Rahman Md¹, and Zahid Hosen SM⁵*

¹Department of Pharmacy, International Islamic University Chittagong, Chittagong, Bangladesh

²Department of Pharmacy, East West University, Dhaka, Bangladesh

³Department of Pharmacy, University of Science and Technology Chittagong, Chittagong, Bangladesh

⁴Department of Pharmacy, BGC Trust University Bangladesh, Chittagong, Bangladesh

⁵Pharmacology Research Division, BCSIR, Chittagong, Bangladesh

Abstract

Background: To identify the therapeutic effects of ethanol extract and its different fractions of *Phrynium imbricatum* leaves in anthelmintic (in vitro) and to determine their total condensed tannin content.

Methods: Leaves of *Phrynium imbricatum* was extracted with pure ethanol and fractioned with chloroform (CHFPI), petroleum ether (PEFPI), n-hexane (NHFPPI) and ethyl acetate (EAFPI), which are tested for anthelmintic activity on aquarium worm *Tubifex tubifex* by using three concentrations viz., 5, 10 and 20 mg/ml of each. Total condensed tannin content determined based on the procedure of Oyedemi et al.

Results: Among the various crude extract, ethanol extract *Phrynium imbricatum* (EEPI) exhibited strong anthelmintic activity in vitro. Where it paralyzed (3.69 ± 0.25 min; P<0.001) and produced death (14.28 ± 0.44 min; P<0.001) of the *Tubifex tubifex* at 20 mg/ml dose near the value of the standard, Levamisole (3.3 ± 0.38 min and 6.5 ± 0.76 min) at 1 mg/ml. The content of condensed tannin good at EEPI and its all fractions, but EEPI (168.44 ± 0.87 mg catechin/g) contained highest among them. For both of experiment, activity found as follows, EEPI>CHFPI>EAFPI>NHFPPI>PEFPI.

Conclusion: These findings suggest that the plant may be a potential source for the development of new anthelmintic and condensed tannin may one of such phytochemical, which exhibit anthelmintic activity.

Keywords: *Phrynium imbricatum*; *In vitro*; Anthelmintic; *Tubifex tubifex*; Condensed tannin

Introduction

Plant materials have been utilized for the treatment of serious diseases all through the world before the approach of advanced clinical medications. The utilization of therapeutic plants still assumes an essential part to cover the fundamental wellbeing needs in the developed countries [1]. Most of the Phytochemical, secondary metabolites of plants, are physiologically active [2] and this metabolites are known as to give an achieve wellspring of natural, anthelmintic, antibacterial and insecticides [3]. Helminth infestations are now being recognized as a cause of chronic ill health and sluggishness amongst the children. World Health Organization estimated 2 billion people infected with helminthes and it was also estimated that 100% of all age group of school children are at risk of morbidity [4]. The major phyla of helminthes are nematodes (round worms) which are soil transmitted helminthes that mostly cause the intestinal infection, filarial worms cause the onchocerciasis and lymphatic filariasis, while plathelminthes (flatworms) also known as trematodes like schistosomes and cestodes causes cyticerosis [5,6]. Current estimates suggest that over half of the world population is infected with intestinal helminthes, such as *Ascaris*, hookworms, *Trichuris*, *Enterobius*, *Strongyloides*, and tapeworms, and that most of these infected people live in remote rural areas in the developing countries [6,7]. In case of other animals also gastrointestinal parasites causes infections that diminish the animal survival, growth rates and reproductive performance [8]. Morbidity from nematodes is common with diabetes and lung cancer. The helminthes parasites mainly subsist in human body in intestinal tract, but they are also found in tissue, as their larvae migrate towards them [9]. Chemical control of helminthes coupled with improved management has been

the important worm control strategy throughout the world. Side effects of anthelmintic commonly include intestinal gastro-intestinal disturbances nausea and giddiness, while various studies and reviews have showed the resistance to anthelmintic is increasing day to day [10]. Henceforth it is important to look for alternative strategies against gastrointestinal nematodes, which have led to the proposal of screening medicinal plants for their anthelmintic activity.

In this regard, *Phrynium imbricatum* (Family: Marantaceae) is a rigid herb, which is commonly known as Pitulpata (Bangladesh) [11-13]. Leaves large, oblong, Spikes oblong, bracts oblong with obtuse, minutely toothed tips. Fruits usually 3-seeded. A paste prepared from leaves of *Phrynium imbricatum*, *Blumea clarkei* and an unidentified species (locally called Khedom gas) is applied to affected areas and bandaged for the treatment of fractures (Chakma). Leaves of *P. imbricatum* has activities like antiarthritic and membrane stabilizing [14]. Occurs in the forests of Chittagong, Chittagong Hill Tracts, Cox's Bazar and Sylhet.

*Corresponding authors: Zahid Hosen SM, Pharmacology Research Division, Bangladesh Council of Scientific and Industrial Research (BCSIR), Chittagong-4220, Bangladesh, Tel: +8801777447192; E-mail: smzahidhosen@bcsir.gov.bd

Received November 28, 2015; Accepted December 17, 2015; Published December 19, 2015

Citation: Kabir MSH, Md AMD, Hossain MM, Noman MAA, Zaheed F, et al. (2015) Effects of Ethanol Extract and its Different Fractions of *Phrynium imbricatum* (Roxb.) Leaves on *In Vitro* Anthelmintic and their Condensed Tannin Content. Pharm Anal Chem Open Access 1: 109. doi:10.4172/2471-2698.1000109

Copyright: © 2015 Kabir MSH, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

The aim of the present study was to identify the anthelmintic activity and total tannin content of ethanol extract and its fractions of leaves of *Phrynium imbricatum*.

Methods and Materials

Plant collection and identification

Leaves of *Phrynium imbricatum* (Accession No. 1315 CTGUH) were collected from Alu tila, khagrachari, Chittagong, Bangladesh in the month of September 2014 at the last time of its flowering. It is authenticated by Dr. Shaikh BU, Associate Professor, Department of Botany, University of Chittagong, Chittagong, Bangladesh.

Extraction and fractionation

Leaves were cleaned with fresh water and dried for a period of 10 days under shade and then powdered with a mechanical grinder, passing through sieve #40, and stored in a tight container. The powdered of whole plant (850 g) of *Phrynium imbricatum* was soaked in 1.5 L ethanol for 7 days with occasional shaking and stirring and filtered through a cotton plug followed by Whatman filter paper number-1. The extract was then concentrated by using a rotary evaporator at reduced temperature and pressure. A portion (55 g) of the concentrated ethanol extract (EEPI) was fractionated by the modified Kupchan partitioning method [15,16] into chloroform, CHFPI (8 g), *n*-hexane, NHFPI (6 g), ethyl acetate, EAFPI (9 g) and pet ether, PEFPI (14 g).

Chemicals

All chemicals used were of analytical reagent grade. Ethanol, methanol, chloroform, pet ether, ethyl acetate, *n*-hexane and hydrochloric acid were purchased from Merck, Germany. Levamisole was purchased from ACI Limited, Bangladesh. Vanillin was purchased from Sigma Chemicals Co (PO Box 14508, St. Louis, MO 63178 USA). Catechin was purchased from BDH Chemicals (BDH Chemicals Ltd. Poole, England).

In-vitro anthelmintic assay

The anthelmintic activity of ethanol extract and its fractions of leaves of *Phrynium imbricatum* were carried out as per the procedure of Ajaiyeoba *et al.* [17] with some minor modifications. The aquarium worm *Tubifex tubifex* were used in the present study because it has anatomical similarity and belongs to the same group of intestinal worm i.e., annelida [18-20]. The worms were collected from the local market of Chittagong, average size of worms 2-2.5 cm. in length were used for the study. The standard drug Levamisole and three different concentrations of EEPI and its different fractions (5, 10 and 20 mg/ml) in double distilled water [21,22] were prepared freshly and used for the study of anthelmintic activity. One group was composed of water and it was considered as controlled group. The anthelmintic activity was determine at two different stage 'time of paralysis' and 'time of death' of the worms. Time for paralysis was noted when no movement of any sort could be observed except when the worms were shaken vigorously. Death was concluded when the worms lost their motility followed with fading away of their body colors [23]. Death was also confirmed by dipping the worms in slightly warm water. The mortality of parasite was assumed to have occurred when all signs of movement had ceased [24].

Total condensed tannins

Condensed tannins (proanthocyanidin) was determined based on the procedure of Oyedemi *et al.* [25]. To 0.5 ml of 1 mg/ml of the extract

solution was added 3 ml of vanillin-methanol (4% v/v) and 1.5 ml of hydrochloric acid was added and vortexed. The mixture was allowed to stand for 15 min at room temperature and the absorbance was measured at 500 nm. Total proanthocyanidin content was evaluated at a concentration of 0.1 mg/ml and expressed as catechin equivalent (mg/g) using the calibration curve equation: $Y=0.5825x$, $R^2=0.9277$, where x is the absorbance and Y is the catechin equivalent.

Statistical Analysis

The data on *in vitro* studies were reported as mean \pm S.E.M (n=3). Data were analyzed using one way factorial ANOVA tests using SPSS followed by Dennett's tests on each group except control for anthelmintic. Regression analysis was performed to calculate total tannin content. $P<0.05$ and $P<0.001$ were considered as statistically significant. Statistical program used was Graphpad Prism[®] (version 6.00; GraphPad Software Inc., San Diego, CA, USA) and Microsoft Excel, 2007, used for graphical presentation.

Results

In vitro anthelmintic activity

Results of study were recorded as shown in Table 1 and Figure 1 as in the form of time required getting consecutive attacks of paralysis and at the end time required for complete death of parasite. From the observations made, higher concentration of extract and fractions produced paralytic effect much earlier and the time to death was shorter for all worms. From the above study it was seen that the ethanol extract showed dose dependent anthelmintic activity as compared to a standard drug Levamisole. Different treatment showed different anthelmintic activity. But ethanol extract of *P. imbricatum* showed highest anthelmintic activity. Where it paralyzed (3.69 ± 0.25 min; $P<0.001$) and produced death (14.28 ± 0.44 min; $P<0.001$) of the *Tubifex tubifex* at 20 mg/ml dose near the value of the standard Levamisole (3.3 ± 0.38 min and 6.5 ± 0.76 min) at 1 mg/ml. EAFPI showed the lowest anthelmintic activity. It's paralyzing and death time of *Tubifex tubifex* is 28.32 ± 0.63 min and 61.45 ± 1.14 min at dose 5 mg/ml. So the anthelmintic activities of ethanol and its fractions of *P. imbricatum* leaves are as follows, EEPI>CHFPI>EAFPI>NHFPI>PEFPI

Ethanol extract of *Phrynium imbricatum* (EEPI) leaves showed highest anthelmintic activity, which indicated by arrow mark in this graph.

Total condensed tannin content

The total phenol contents of the extracts are shown in Table 2. The total condensed tannin content of *Phrynium imbricatum* leaves was higher in plants at ethanol extract, which was 168.44 ± 0.87 mg catechin/g. Fractions of ethanol extract of *Phrynium imbricatum* contain good amounts of condensed tannin, ranging from 152.42 to 114.85 mg catechin/g. So condensed tannin content of ethanol and its fractions of *P. imbricatum* leaves are as follows, EEPI>CHFPI>EAFPI>NHFPI>PEFPI

Discussion

Consumption of high concentrations of condensed tannins (>7% of DM) had a number of detrimental effects on ruminants, such as reduction in food intake, growth inhibition and interference with the morphology and the proteolytic activity of microbes in the rumen, low or moderate concentrations of condensed tannins (<6% of DM) have resulted in the positive effects on. Levamisole works as a nicotinic acetylcholine receptor agonist that causes continued stimulation of the

Treatment	Time taken for paralysis (min)	Time taken for Death (min)
Control(Water)	0	0.00
Levamisole (1 mg/ml)	3.3 ± 0.38	6.5 ± 0.76
EEPI (20 mg/ml)	3.69 ± 0.25^b	14.28 ± 0.44^b
EEPI (10 mg/ml)	10.37 ± 0.67 ^a	18.61 ± 0.8 ^a
EEPI (5 mg/ml)	12.3 ± 0.67 ^a	23.72 ± 0.92 ^a
CHFPI (20 mg/ml)	9.15 ± 0.67 ^b	15.01 ± 0.96
CHFPI (10 mg/ml)	14.92 ± 0.9 ^a	29.07 ± 1.04 ^a
CHFPI (5 mg/ml)	21.75 ± 0.87 ^a	45.9 ± 1.2 ^a
EAFPI (20 mg/ml)	7.51 ± 0.55	15.29 ± 0.87 ^a
EAFPI (10 mg/ml)	12.62 ± 0.79 ^a	23.92 ± 0.81 ^a
EAFPI (5 mg/ml)	18.35 ± 1.17	39.12 ± 1.24 ^a
PEFPI (20 mg/ml)	13.69 ± 0.82 ^b	25.12 ± 1.57 ^a
PEFPI (10 mg/ml)	18.61 ± 0.53 ^a	34.62 ± 1.11 ^a
PEFPI (5 mg/ml)	28.32 ± 0.63 ^a	61.45 ± 1.14 ^a
NHFPI (20 mg/ml)	10.61 ± 0.78	19.09 ± 0.97 ^a
NHFPI (10 mg/ml)	14.01 ± 0.90 ^a	27.07 ± 0.85 ^a
NHFPI (5 mg/ml)	22.46 ± 0.58 ^a	49.11 ± 1.28 ^a

Values are mean ± SEM (n=3); ^aP<0.05, ^bP<0.001, Dennett's test as compared to positive control (Levamisole, 1 mg/ml). Statistical representation of the effective paralysis and dead time by *Phrynium imbricatum* ethanol extract and its fractions, positive anthelmintic control (Levamisole, 1 mg/ml) processed by paired t-test analysis (Dennett's test). Bold text indicates the highest anthelmintic activity of ethanol extract of *Phrynium imbricatum* (EEPI). Data were processed by paired t-test analysis by using SPSS for windows, version 16.0.

Table 1: Anthelmintic activity of ethanol extract and its different fractions of leaves of *Phrynium imbricatum*.

Sample	Total proanthocyanidin (mg catechin/g)
EEPI	168.44 ± 0.87^b
CHFPI	152.42 ± 1.02 ^a
PEFPI	114.85 ± 0.89 ^b
NHFPI	120.5 ± 1.02 ^b
EAFPI	136.67 ± 1.47 ^a

Values are mean ± SEM (n=3). Bold text indicates the highest tannin content of ethanol extract of *Phrynium imbricatum* (EEPI). The different superscripted (a,b) values have significantly different (^aP<0.05, ^bP<0.001) from the other sample in same column.

Table 2: Contents of condensed tannin (expressed as mg catechin/g dry weight) in ethanol extract and its fractions of *Phrynium imbricatum* leaves.

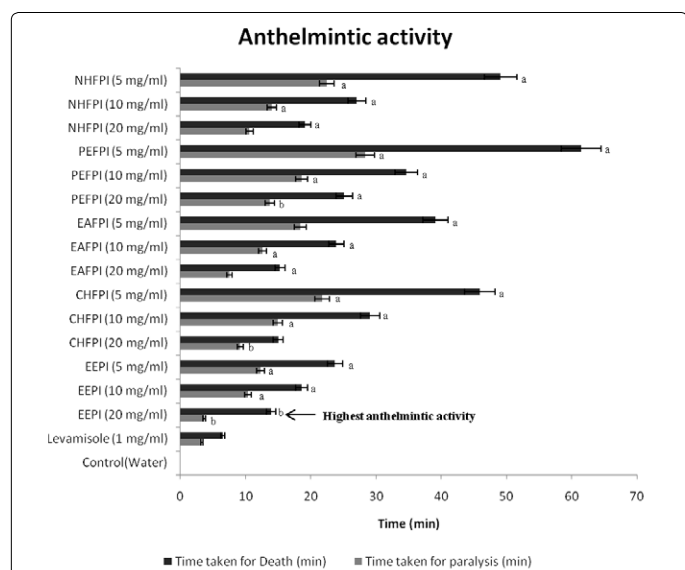


Figure 1: Anthelmintic activity of ethanol extract and its different fractions of leaves of *Phrynium imbricatum*.

parasitic worm muscles, leading to paralysis. The literature have been reported that the presence of flavonoids, tannins and polyphenolic compounds show anthelmintic activity, [26,27] as they can bind to free protein in the gastrointestinal tract of host animal or glycoprotein on the cuticle of the parasite and thereby causes death [28]. Some synthetic phenol anthelmintics e.g., niclosamide, oxclozanide and bithionol are shown effects to interfere with energy generation in antihelminthic parasites by uncoupling oxidative phosphorylation and phosphorylation [29]. Finally, study concludes that the plant under study has found to possess significant anthelmintic activity in dose dependent manner. The plant might have potential to be developed as useful economic and safe anthelmintic alternative, but it demands more thorough study to find out the exact chemical responsible for anthelmintic activity of plant so as to isolate and extract it separately so as to improve the potency.

Similarly, condensed tannins (CTs) have high relevance for livestock production as tannin-rich plants have a long tradition of use not only as forages but also as 'green' control of gastrointestinal nematode infections. Several excellent reviews deal with the various aspects of feeding of small ruminants with forages containing tannin-rich plants or even fodder trees [30-33]. They pointed that bioactive tanniniferous plants represent a valuable option as an alternative to commercial drugs for the control of gastro-intestinal nematodes (GINs) as consumption of these plants has been associated with antiparasitic and anthelmintic effects: reductions in nematode numbers, worm fecundity, and nematode eggs excretion. The principle risk to the utilization of exclusively substance medications is the quick advancement of imperviousness to any anthelmintic medication in worm populaces after commercialization [34] and the spread of anthelmintic resistance within worm populations [35]. Within the last decade a number of studies focused on isolation of condensed tannins and sesquiterpene lactones from various legume forages and plants with the aim to reveal their effects *in vitro* and *in vivo* on various species and developmental stages of nematodes. Differentiated action of condensed tannins on parasite stages was observed by Athanasiadou *et al.* [36,37] which were more effective against larvae than adults. This can also be explained by the distinction between the cuticular components of the pre-parasitic stages (eggs to L3) and the parasitic stages (L4 and adults), as demonstrated by the study of Stepek *et al.* [38,39].

So present studies suggested that condensed tannin really responsible for anthelmintic activity. Because highest condensed tannin containing extract gave highest anthelmintic effect and lowest one gave lowest anthelmintic activity.

Conclusion

Our aim was to determine the anthelmintic activity and condensed tannin content. However, we find out that according to condensed tannin content, extracts giving their anthelmintic activity. This suggested that specific, key processes in the parasite life cycle can be disrupted by condensed tannin. These data encourage further investigations to determine *in vivo* efficacy in animal model. In addition, further mechanistic studies, such as the relationship between the fine structure of condensed tannin molecules and anthelmintic activity, are also a high priority.

Acknowledgements

Authors are greatly indebted to the management of the Department of Pharmacy, International Islamic University Chittagong, Bangladesh, for providing all the laboratory facilities and support with a research grant to accomplish the study (Grant No. Pharm-P&D 71/09-15/36). Authors would like to thank Mr. Mostafa Kamal ATM, Assistant Professor, International Islamic University Chittagong,

Bangladesh, for his help in experiment and supervision. The authors are also thankful to all members of GUSTO (A research group), for their kind help in the experiment.

Conflict of Interest

The authors declare that they have no competing interests.

References

- Hidayathulla S, Chandra K, Chandrashekar K (2011) Phytochemical evaluation and antibacterial activity of *Pterospermum diversifolium* Blume. *International Journal of Pharmacy and Pharmaceutical Sciences* 3: 165-167.
- Oliveira DM, Melo FG, Balogun SO, Flach A, de Souza ECA, et al. (2015) Antibacterial mode of action of the hydroethanolic extract of *Leonotis nepetifolia* (L.) R. Br. involves bacterial membrane perturbations. *Journal of Ethnopharmacology* 172: 356-363.
- Suman A, Kumar DG, Kumar BD, Raj CR, Matushree VB (2011) Preliminary phytochemical investigation and anthelmintic activity of *Acacia suma* (Roxb) barks. *Int Res J Phar* 2: 136-141.
- World Health Organization (2012) Eliminating soil-transmitted helminthiasis as a public health problem in children: progress report 2001-2010 and strategic plan 2011-2020. WHO Library, Geneva. p: 78.
- Liu C, Luo R, Yi H, Zhang L, Li S, et al. (2015) Soil-Transmitted Helminths in Southwestern China: A Cross-Sectional Study of Links to Cognitive Ability, Nutrition, and School Performance among Children. *PLoS Negl Trop Dis* 9: 1-16.
- Smith AF, Semeniuk C, Kutz SJ, Massolo A (2014) Dog-walking behaviours affect gastrointestinal parasitism in park-attending dogs. *Parasites and vectors* 7: 1-10.
- Reznick JS (2014) Embracing the Future as Stewards of the Past: Charting a Course Forward for Historical Medical Libraries and Archives. *RBM: A Journal of Rare Books, Manuscripts and Cultural Heritage* 15: 111-123.
- Tripathi KP (2003) Essentials of medical pharmacology. 5th edn. Jaypee Brothers Medical Publishers Pvt. Ltd., New Delhi, India. p: 759.
- Tu Z, Zhang H (2015) Controlled Veterinary Drug Delivery Systems against Parasitic Infection. *Zhongguo ji sheng chong xue yu ji sheng chong bing za zhi Chinese journal of parasitology and parasitic diseases* 33: 58-63.
- Mejia-Carmona G, Gosselink K, Pérez-Ishiwara G, Martínez-Martínez A (2015) Oxidant/antioxidant effects of chronic exposure to predator odor in prefrontal cortex, amygdala, and hypothalamus. *Molecular and cellular biochemistry* 406: 121-129.
- Haslam E (1989) Plant Polyphenols vegetable tannins revisited. Cambridge University Press, Cambridge, UK.
- Ping L, Brosse N, Chrusciel L, Navarrete P, Pizzi A (2011) Extraction of condensed tannins from grape pomace for use as wood adhesives. *Industrial Crops and Products* 33: 253-257.
- Bangladesh Ethnobotany Online Database (BEOD) (2015) *Phrynium imbricatum* Roxb.
- Hossain MM, Kabir MSH, Hasanat A, Kabir MI, Chowdhury TA, et al. (2015) Investigation of in vitro anti-arthritis and membrane stabilizing activity of ethanol extracts of three Bangladeshi plants. *The Pharma Innovation J* 4: 76-80.
- Islam MK, Eti IZ, Chowdhury JA (2010) Phytochemical and antimicrobial analysis on the extract of *Oroxylum indicum* Linn. Stem-Bark. *Iranian journal of pharmacology & therapeutics* 9: 25-28.
- Bulbul IJ, Nahar L, Haque M (2011) Antibacterial, cytotoxic and antioxidant activity of chloroform, n-hexane and ethyl acetate extract of plant *Coccinia cordifolia*. *Agriculture and Biology journal of north America* 2: 713-719.
- Ajaiyeoba E, Onocha P, Olarenwaju O (2001) *In vitro* anthelmintic properties of *Buchholzia coriacea* and *Gynandropsis gynandra* extracts. *Pharmaceutical Biology* 39: 217-220.
- Verma VK, Sarwa K, Kumar A (2013) Anthelmintic Activity of Fruit Peel and Root Extracts of *Trapa natans* L. var. *bispinosa* Roxb. *Academic Journal of Plant Sciences* 6: 73-76.
- Debebe Y, Tefera M, Mekonnen W, Abebe D, Woldekidan S, et al. (2015) Evaluation of anthelmintic potential of the Ethiopian medicinal plant *Embellia schimperii* Vatke in vivo and in vitro against some intestinal parasites. *BMC complementary and alternative medicine* 15: 1-6.
- Hossain MM, Kabir MSH, Chowdhury TA, Hasanat A, Chakrabarty N (2015) Anthelmintic Effects of Different Extracts of *Hopea odorata* Leaves on Tubifex tubifex Worm Using in vitro Method and their Condensed Tannin Content. *British Journal of Pharmaceutical Research* 8: 1-7.
- Kosalge SB, Fursule RA (2009) Investigation of in vitro anthelmintic activity of *Thespesia lampas* (Cav.). *Asian Journal of Pharmaceutical and Clinical Research* 2: 69-71.
- Kabir MSH, Ahmad S, Mahamoud MS, Al MA, Adnan M (2015) Evaluation of Total condensed tannin content and anthelmintic activities of organic extracts of four Bangladeshi plants on *Tubifex tubifex* worm using in vitro method. *International Journal of Pharmacy* 5: 903-910.
- McMullen II JG, Stock SP (2014) In vivo and In vitro Rearing of Entomopathogenic Nematodes (Steinernematidae and Heterorhabditidae). *JoVE* 91: e52096.
- Yadav AK (2005) Anticestodal efficacy of folklore medicinal plants of Naga tribes in north-east India. *African Journal of Traditional, Complementary and Alternative Medicines* 2: 190-194.
- Oyedemi S, Bradley G, Afolayan A (2010) In vitro and in vivo antioxidant activities of aqueous extract of *Strychnos henningsii* Gilg. *Afr J Pharm Pharmacol* 4: 70-78.
- Thorn GW, Adams RD, Braunwald E, Isselbacher KJM, Petersdorf RG (1977) *Harrison's Principles of Internal Medicine*. 8th edn. McGraw Hill Company, New York. pp: 501-519.
- Sreejith M, Kannappan N, Santhiagu A, Mathew AP (2013) Phytochemical, Anti-oxidant and Anthelmintic activities of various leaf extracts of *Flacourtia sepriaria* Roxb. *Asian Pacific journal of tropical biomedicine* 3: 947-953.
- Williams AR, Ropiak HM, Fryganas C, Desrués O, Mueller-Harvey I, et al. (2014) Assessment of the anthelmintic activity of medicinal plant extracts and purified condensed tannins against free-living and parasitic stages of *Oesophagostomum dentatum*. *Parasites & vectors* 7: 1-12.
- Burns AR, Luciani GM, Musso G, Bagg R, Yeo M, et al. (2015) *Caenorhabditis elegans* is a useful model for anthelmintic discovery. *Nature communications* 6: 1-11.
- Tedeschi L, Ramírez-Restrepo C, Muir J (2014) Developing a conceptual model of possible benefits of condensed tannins for ruminant production. *Animal* 8: 1095-1105.
- Hoste H, Martínez-Ortiz-De-Montellano C, Manolaraki F, Brunet S, Ojeda-Robertos N, et al. (2012) Direct and indirect effects of bioactive tannin-rich tropical and temperate legumes against nematode infections. *Veterinary parasitology* 186: 18-27.
- Hoste H, Torres-Acosta J (2011) Non chemical control of helminths in ruminants: adapting solutions for changing worms in a changing world. *Veterinary parasitology* 180: 144-154.
- Sandoval-Castro C, Torres-Acosta J, Hoste H, Salem A, Chan-Pérez J (2012) Using plant bioactive materials to control gastrointestinal tract helminths in livestock. *Animal Feed Science and Technology* 176: 192-201.
- Waller P (2006) From discovery to development: current industry perspectives for the development of novel methods of helminth control in livestock. *Veterinary parasitology* 139: 1-14.
- Zanzani SA, Gazzonis AL, Di Cerbo A, Varady M, Manfredi MT (2014) Gastrointestinal nematodes of dairy goats, anthelmintic resistance and practices of parasite control in Northern Italy. *BMC veterinary research* 10: 1-10.
- Ríos-Hoyo A, Cortés MJ, Ríos-Ontiveros H, Meaney E, Ceballos G, et al. (2014) Obesity, Metabolic Syndrome, and Dietary Therapeutic Approaches with a Special Focus on Nutraceuticals (Polyphenols): A Mini-Review. *Int J Vitam Nutr Res* 84: 113-123.
- Athanasiadou S, Kyriazakis I, Jackson F, Coop R (2001) Direct anthelmintic effects of condensed tannins towards different gastrointestinal nematodes of sheep: in vitro and in vivo studies. *Veterinary parasitology* 99: 205-219.
- Stepek G, Lowe A, Buttle D, Duce I, Behnke J (2007) Anthelmintic action of plant cysteine proteinases against the rodent stomach nematode, *Protophysa muricola*, in vitro and in vivo. *Parasitology* 134: 103-112.
- Stepek G, Lowe AE, Buttle DJ, Duce IR, Behnke JM (2007) In vitro anthelmintic effects of cysteine proteinases from plants against intestinal helminths of rodents. *Journal of helminthology* 81: 353-360.