



Brain Tumors: Epidemiology and Current Trends in Treatment

Michael J Strong^{1,2}, Juanita Garces³, Juan Carlos Vera⁴, Mansour Mathkour⁴, Noah Emerson⁵ and Marcus L Ware^{4,6*}

¹Department of Pathology, Tulane University School of Medicine, New Orleans, LA, USA

²Tulane Cancer Center, Tulane University School of Medicine, New Orleans, LA, USA

³Department of Neurological Surgery, Tulane University School of Medicine, New Orleans, LA, USA

⁴Department of Neurological Surgery, Ochsner Clinic Foundation, New Orleans, LA, USA

⁵Department of Radiology, Ochsner Clinic Foundation, New Orleans, LA, USA

⁶The University of Queensland School of Medicine, Ochsner Clinical School, New Orleans, LA, USA

*Corresponding author: Marcus L. Ware, Department of Neurological Surgery, Ochsner Clinic Foundation, 1514 Jefferson Hwy., New Orleans, LA 70121, USA, Tel: (504) 842-4033; E-mail: mware@ochsner.org

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Abstract

Background: Brain tumors represent a group of neoplasms arising from brain tissue, each with their own unique biology, prognosis, and treatment. Included in this group are neoplasms not arising from brain parenchyma, which encompass meningiomas, lymphomas, and metastatic disease from other primary sources (often referred to as secondary brain tumors). Despite the diverse group of neoplasms represented, most intracranial tumors follow similar clinical presentations and diagnostic workups.

Methods: This review focuses on primary and secondary brain tumor epidemiology, imaging, and treatment modalities. In addition, we will highlight molecular genetic advances in the field that will help shape future treatment approaches.

Results: Although tumors affecting the Central Nervous System (CNS) are relatively uncommon, they are often very difficult to treat and cause disproportionate morbidity and mortality. Many of these neoplasms are universally fatal and our ability to treat both benign and malignant tumors is still in its infancy. Our lack of effective treatment leaves many of our patients with few options.

Conclusions: The combination of poor prognosis and lack of therapeutic options make further innovation and investigation a priority to improve clinical outcomes for patients suffering from CNS malignancies.

Keywords: Brain tumors; Epidemiology; Treatment; Imaging; Neoplasm metastasis; Radiosurgery; Chemotherapy

Primary Brain Tumors

Classification

The World Health Organization (WHO) classifies brain tumors based histologic features and presumed cellular origin. In 2007, the WHO updated its Central Nervous System (CNS) classification system to reflect a grading scheme in which the histologic diagnosis directly correlates with the histologic grade of the tumor [1]. Seven major categories of tumors in the CNS have been identified by the WHO classification system (Table 1) and include tumors of neuroepithelial tissue, tumors of cranial and paraspinal nerves, tumors of the meninges, lymphomas and hematopoietic neoplasms, germ cell tumors, tumors of the sellar region, and metastatic tumors.

Epidemiology

Approximately 69,720 new cases of primary CNS tumors are expected to be diagnosed in the United States this year [1]. Of these lesions, roughly 24,620 will be malignant [1]. Although the incidence

of primary brain tumors is relatively low compared to other cancer types, primary brain tumors give rise to a disproportionate amount of morbidity and mortality, often causing debilitating impairment to patients' movement and speech [2,3]. Although primary CNS tumors comprise only 1.4% of all cancers, they are among the most aggressive tumors and result in a combined mortality rate of about 60% [2]. In fact, the five-year survival rate for primary malignant brain and central nervous system tumors is the sixth lowest among all types of cancers after pancreatic, liver & intrahepatic bile duct, lung, stomach, and esophageal [2].

The majority of primary brain tumors fall under the WHO classification scheme of tumors of neuroepithelial tissue. Malignant gliomas are the most common primary brain tumor, comprising more than 80 percent of all primary brain neoplasms [4]. Gliomas can be divided into astrocytomas, oligodendrogliomas, ependymomas, and oligo-astrocytomas (mixed gliomas). These neuroglial tumors can be further divided based on grade. Astrocytomas are subdivided into grades I-IV as follows: pilocytic, grade I; diffuse, grade II; anaplastic, grade III; and Glioblastoma Multiforme (GBM), grade IV. Ependymomas are subdivided into grades I-III. Oligodendrogliomas are typically grade II and oligoastrocytomas are usually grade III.

Tumors of Neuroepithelial Tissue	Microcystic (WHO grade I)
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Astrocytic tumors	Secretory (WHO grade I)
Pilocytic astrocytoma (WHO grade I)	Lymphoplasmacyte-rich (WHO grade I)
Diffuse astrocytoma (WHO grade II)	Metaplastic (WHO grade I)
Anaplastic astrocytoma (WHO grade III)	Chordoid (WHO grade II)
Glioblastoma (WHO grade IV)	Clear cell (WHO grade II)
Oligodendroglial tumors	Atypical (WHO grade II)
Oligodendroglioma (WHO grade II)	Papillary (WHO grade III)
Anaplastic oligodendroglioma (WHO grade III)	Rhabdoid (WHO grade III)
Ependymomas	Anaplastic meningioma (WHO grade III)
Myxopapillary (WHO grade I)	
Subependymoma (WHO grade I)	Tumors of the Sellar Region
Classic ependymoma (WHO grade II)	Pituicytoma (posterior pituitary tumor) (WHO grade I)
Cellular	Spindle cell oncocytoma of the adenohypophysis (anterior pituitary tumor) (WHO grade II)
Papillary	
Clear	Lymphomas and Haematopoietic Neoplasms
Anaplastic ependymomas (WHO grade III)	Malignant lymphomas
Mixed Gliomas	
Oligoastrocytomas (WHO grade II)	Tumors of Cranial and Paraspinal Nerves
Anaplastic oligoastrocytoma (WHO grade III)	Schwannoma
Medulloblastoma (WHO grade IV)	Neurofibroma
	Dermal (WHO grade I)
Tumors of the Meninges	Plexiform (WHO grade I)
Meningioma	
Meningothelial (WHO grade I)	Germ Cell Tumors
Fibrous (WHO grade I)	
Transitional (WHO grade I)	Metastatic Tumors
Psammomatous (WHO grade I)	
Angiomatous (WHO grade I)	

Table 1: Classification of brain tumors.

Radiation
**Ionizing
Electromagnetic radiation
Cell phones and radiofrequency radiation
Head trauma
Allergies
Diet and vitamins
N-nitroso compounds
Fat intake
Aspartame ingestion
Tobacco
Alcohol
Chemicals
Hair dyes and sprays
Traffic-related air pollution
Infection
Simian Virus 40
Human Cytomegalovirus
Polyomaviruses (e.g. JC and BK)
Toxoplasma infection
Varicella zoster – protective role
**Genetics
Neurofibromatosis type 1
Neurofibromatosis type 2
Von Hippel-Lindau syndrome
Li-Fraumeni syndrome
Turcot syndrome
Basal cell nevus syndrome
Adenomatous polyposis syndrome
Occupational Exposure
Electrical workers and electromagnetic fields
Agriculture workers exposed to pesticides, herbicides and fungicides
Other industries (vinyl chloride, petrochemical, and rubber industries)
**Only proven risk factors

Table 2: Risk factors associated with brain tumors

Of the gliomas, GBMs (WHO-grade IV) account for 60-70%, anaplastic astrocytomas (WHO-grade III) account for 10-15%, anaplastic oligodendrogliomas (WHO-grade II) and anaplastic

contrast, T2, FLAIR, Gradient Recall Echo (GRE) and Diffusion Weighted Imaging (DWI) [36]. T2 and FLAIR are fluid weighted sequences helping to identify tumoral cystic changes, necrosis, as well as cytotoxic and vasogenic edema. The DWI sequence can reveal elevated nuclear to cytoplasmic ratios. GRE capitalizes on the intrinsic artifact of magnetic substances to reveal the presence of hemorrhage or calcification even at a microscopic level. T1 pre and post contrast imaging assesses the integrity of the blood brain barrier. Cumulatively, a lesion's conventional MRI characteristics not only help in diagnosis but also in assessing tumor histological grade.

As the complexities of tumor genesis are discovered, new treatment regimes will be designed. Unfortunately, these new therapies can result in unpredictable and often confusing imaging appearances on conventional MRI sequences. Specifically with GBM therapies the phenomena coined pseudo progression and pseudo response have emerged [37]. These, as well as long known phenomena of radiation necrosis, are often indistinguishable from tumor progression on routine conventional MRI sequences. While serial follow-up imaging remains the general standard for identification of recurrent tumor and/or its progression, new and old advanced imaging techniques are being applied to help define treatment effects versus recurrent tumor. Primary amongst these are Dynamic Susceptibility Contrast (DSC) Perfusion, Permeability, and Spectroscopy.

Perfusion helps define degrees of angiogenesis by measuring the relative cerebral blood volume; Permeability can assess the leakiness of microscopic junctions; and Spectroscopy can reveal a lesion's molecular composition. Variable success has been observed with these modalities and a rising school of thought is to assess the tumor both pre and post therapy not only to have a baseline of a lesion's behavior, but also to help define a lesion's grade and therefore guide appropriate therapy decisions.

The following is an example of a GBM post resection with a large region of enhancement (Figure 1A) and vasogenic edema (Figure 1B) developing rapidly post resection. Advanced DSC perfusion identified lack of elevated blood volume compatible with pseudo progression (Figure 1C), which was confirmed post biopsy.

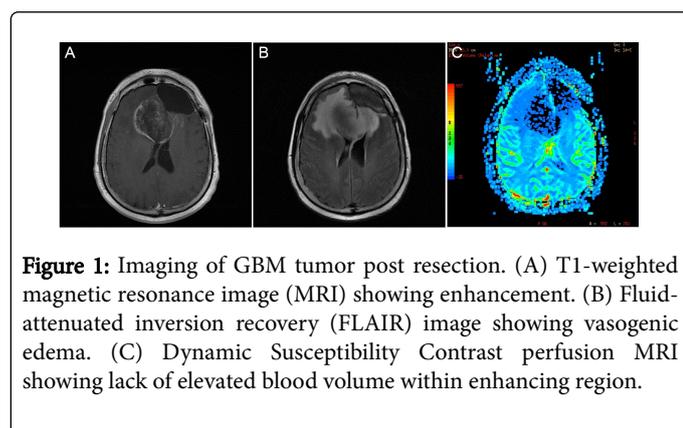


Figure 1: Imaging of GBM tumor post resection. (A) T1-weighted magnetic resonance image (MRI) showing enhancement. (B) Fluid-attenuated inversion recovery (FLAIR) image showing vasogenic edema. (C) Dynamic Susceptibility Contrast perfusion MRI showing lack of elevated blood volume within enhancing region.

Separate imaging advancements include Diffusion Tensor Imaging (DTI) with tractography and functional MRI. Although these imaging techniques are not primarily used as a diagnostic tool, they can localize a lesion precisely and define the specific affected tracks and their function [38]. This can be invaluable in guiding lesion resection and prediction of post resection functional prognosis.

Finally, the development of 3D steady state volumetric imaging has led to faster and thinner imaging than ever before. A technique which incompletely fills "K" space trades an increase in imaging speed for lesser degrees of soft tissue contrast. However, the gain in spatial resolution is profound and logical paring of this with navigational software has become routine for intra-operative stereo-tactic lesion resection.

The extent of surgical resection is an important prognostic factor in neurooncology and the goal of image-guided surgical resection is to help achieve maximum safe resection [39]. Three approaches fall within the image-guided resection modality and include intraoperative MRI (iMRI) and neuronavigation, fluorescent imaging, and intra-operative brain mapping. Intraoperative MRI allows for real-time visualization of complex three-dimensional structures of the brain to help guide the removal of brain tumors. In addition to using intra-operative structural MRI, recent efforts have been made to integration other MRI modalities, including fMRI and DTI, into neuronavigation systems [40]. This unique integration allows surgeons to account for brain shift and other anatomical changes that often affect maximal tumor debulking, particularly in eloquent areas of the brain. In addition, image quality of intra-operative ultrasound is now comparable to intra-operative MRI and is being integrated into neuronavigation systems to enable acquisition of 3D ultrasound data for direct image guidance [40,41].

Surgical approaches that can complement iMRI and help surgeons visually distinguish neoplastic tissue from healthy tissue may facilitate maximum safe resection while minimizing the amount of residual tumor cells [42].

Although several types of contrast agents are currently being developed for intraoperative surgical oncology, only a small number have been FDA approved [43]. These include several passive contrast molecules including methylene blue, Indocyanine Green (ICG), and sodium fluorescein. ICG is a water-soluble probe that binds human serum albumin and is used to visualize solid tumors [44]. ICG has proven useful in brain tumor resections as it was used intraoperatively to identify tumor boundaries in glioma surgery [45]. Fluorescein is a fluorophore that can cross capillaries and provide fluoresce in the extracellular matrix [46]. Due to this property, Fluorescein is useful in identifying infiltrative tumor margins [47].

5-Aminolevulinic acid (5-ALA) is another contrast agent that is converted to an active fluorescent state in situ. In a phase III trial, surgical resection with 5-ALA demonstrated a 1.5 month increase in progression-free survival compared to patients treated with surgical resection using white light (5.1 months vs. 3.6 months) [48].

Several studies have investigated combining structural imaging with functional imaging to study brain connectivity [49,50]. This approach is useful in understanding brain function organization. Therefore, brain mapping in eloquent regions of the brain is vital for maximum resection while at the same time preserving quality of life. The current gold standard for brain mapping is cortical and Subcortical Electrical Stimulation (CSES). However, with advances in functional imaging, the utility of fMRI and DTI-FT is being evaluated for cortical mapping. When compared to CSES, the sensitivity of fMRI in detecting sensorimotor areas ranges from 82% to 100% [51-54]. Yet, the utility of fMRI diminishes for language areas [52]. Evaluation of DTI-FT is also complicated and no clear conclusions can be drawn [52]. While we make strives in brain mapping using functional imaging, the use of CSES has allowed us to treat patients once deemed unsuitable for

surgery. CSES demonstrates high sensitivity with the ability to map the entire exposed cortical region. Also, CSES provides excellent prognosis with removal of negative cortical and subcortical areas leading to no permanent neurological impairment [55].

Neuroepithelial tumors

Although the WHO classifies neuroepithelial tumors into nine major groups, this review will only focus on the most frequent types including astrocytic, oligodendroglial, ependymal and mixed. We will also discuss the most common malignant neuroepithelial tumor in children, medulloblastomas.

Astrocytic Neoplasms

Glial-appearing cells give rise to most primary CNS neoplasms (gliomas). Of these, astrocytomas are the most common. Astrocytomas, anaplastic astrocytomas, and glioblastomas are termed diffusely infiltrating astrocytomas due to their range of diffuse infiltration. The diffuse astrocytic neoplasms are most common in the cerebrum in adults and brain stem in children [56]. They have a propensity for progression with 50%-75% of astrocytomas progressing to anaplastic astrocytomas or GBMs [57]. Therefore, all patients with astrocytomas need regular followup. GBM has the highest incidence of any primary neuroepithelial neoplasm, accounting for approximately 50% of intracranial gliomas [58].

Histology

Pilocytic astrocytoma (WHO grade I)

Macroscopically: Tumors are often cystic with discrete borders.

Microscopically: Neoplastic cells are usually bipolar with elongated hairlike processes that are arranged in a parallel fashion. Rosenthal fibers, which are tapered corkscrew shaped eosinophilic hyaline masses, are often present.

Diffuse Astrocytoma (WHO grade II)

Macroscopically: Cerebral astrocytomas diffusely expand beyond the white matter boundary oftentimes distorting the overlying gray matter. The neoplastic process is poorly demarcated.

Microscopically: Neoplastic cells show mild atypia. Fibrillary astrocytomas may appear as bare nuclei. Astrocytomas show varying degrees of astrocytic differentiation. They may exhibit prominent fibrillary strands of eosinophilic cytoplasm, or plump cell bodies in which the nucleus is displaced by homogeneous eosinophilic cytoplasm, oftentimes referred to as the gemistocytic phenotype.

Anaplastic Astrocytoma (WHO grade III)

Macroscopically: Anaplastic transformation may be associated with little macroscopic change from astrocytomas. Although on MRI, areas undergoing anaplastic progression often show contrast enhancement.

Microscopically: Cytological and nuclear pleomorphism may be more pronounced. Nuclear to cytoplasm ratio is increased. Mitotic activity distinguishes the anaplastic astrocytoma from diffuse astrocytoma variants. However, necrosis is not present.

GBM (WHO grade IV)

Macroscopically: GBM distorts the normal anatomy of the brain. Foci of cyst formation, necrosis and hemorrhage are mixed with mucoid gray neoplastic tissue. GBMs commonly appear as spherical masses with a necrotic center, which may be seen on MRI as a ring enhancing mass. Their growth is not restricted to one hemisphere as they often track along the corpus callosum affecting the contralateral hemisphere, commonly referred to as a butterfly glioma. GBMs may also spread along CSF pathways.

Microscopically: Necrosis and a florid microvascular proliferation are the key features separating GBM from the two other diffuse astrocytic neoplasms. Thrombi are often found in these vessels and are responsible for the foci of necrosis. Cellular pleomorphism is more extreme than in anaplastic astrocytomas. Finally, palisading of cells around necrotic areas is often seen.

Molecular Genetics

Although many important genetic alterations have been known in gliomas, new technologies have shed light onto novel discoveries in recent years. These genetic alterations are currently being used as biomarkers. A biomarker is a genetic or biochemical feature that can be assessed to indicate a particular diagnosis, prognosis, or response to treatment. As technology advances along with our understanding of the complex molecular genetics underlying brain tumors, the number of biomarkers will likely increase.

O6-methylguanine methyltransferase (MGMT) promoter methylation: MGMT is a DNA repair protein that repairs the chemotherapy-induced alkylation at the O-position of guanine, the essential mediator of alkylating drug cytotoxicity, and thus counteracts the effects of alkylating chemotherapeutic drugs such as nitrosoureas or temozolomide. Hypermethylation of the MGMT gene promoter is one mechanism to silence the gene and thus reduce the protein concentration. As such, hypermethylation of MGMT is associated with 20-40% of patients with GBM [59]. Several clinical trials and cohort studies have shown that the MGMT promoter methylation phenotype is associated with prolonged progression-free and overall survival in patients with GBM who are being treated with alkylating class of chemotherapy drugs [60-62]. In a randomized clinical trial assessing radiotherapy alone with radiotherapy combined with concomitant and adjuvant treatment with temozolomide in newly diagnosed patients with glioblastoma, the benefit from chemotherapy was almost exclusively attributable to patients with tumors with a methylated MGMT gene promoter [62]. In the same study, patients with a methylated MGMT promoter showed better overall survival than patient with an unmethylated MGMT promoter [62]. These results suggest that treatment strategies should be individualized dependent on MGMT status and that MGMT status has prognostic value.

Loss of chromosomes 1p and 19q: The combined loss of chromosomal arms 1p and 19q is a cytogenetic aberration resulting from an unbalanced t(1;19)(q10;p10) translocation occurring in 50%-90% of oligodendrogliomas and 30%-50% of oligoastrocytomas [63]. Tumors with the 1p/19q deletion respond better to chemotherapy and radiotherapy resulting in prolonged progression free survival and overall survival in patients, especially with anaplastic oligodendrogliomas [64,65]. Outside of oligodendrogliomas, 1p/19q codeletions are rare and additional studies are necessary to determine whether these tumors have a less aggressive natural course [66].

Isocitrate dehydrogenase (IDH) mutation: Single point mutations in the metabolic genes IDH 1 and 2 were recently discovered in gliomas [67,68]. Approximately 80% of grade 2 and grade 3 gliomas as well as secondary GBMs harbor a single amino acid missense mutation in IDH1 at arginine 132 [69]. The IDH2 mutation at arginine 172 is less common and is mutually exclusive with mutations in IDH1 [69]. IDH1 and 2 mutations promote a neomorphic reaction in which the normal product α -ketoglutarate is converted to 2-hydroxyglutarate (2-HG) (a candidate oncometabolite) in a reaction that consumes, rather than produces, NADPH [70]. The accumulation of high concentrations of 2-HG has been shown to contribute to the formation and malignant progression of gliomas [70]. In addition, the IDH1 mutation has been associated with the CpG island methylator phenotype (G-CIMP) in gliomas, which is associated with younger patients, improved survival, and is more common in low and intermediate grade gliomas [71,72].

Epidermal Growth Factor Receptor (EGFR) variant III: EGFR is a cell surface receptor involved in the control of cell proliferation. A common alteration of the EGFR locus observed in gliomas is a genomic rearrangement with amplification of EGFR resulting in an in-frame deletion of exons 2-7 from the extracellular domain, causing a truncated mutant receptor known as EGFR variant III (EGFRvIII) [73]. This truncated mutant receptor is therefore ligand-independent and constitutively active which confers enhanced tumorigenicity on glioma cells by increasing proliferation and reducing apoptosis [74]. Overexpression of EGFR is observed in 50%-60% of GBMs with the most common EGFR mutation (EGFRvIII) expressed in 24%-67% of cases [75-79]. Since the EGFRvIII mutation creates a new surface epitope, vaccination strategies based on this unique peptide sequence have been developed. Subsequent phase 2 clinical trials have demonstrated promising results, thus warranting phase 3 trials [80,81]. The prognostic relevance of EGFR overexpression and EGFRvIII is unclear, but long-term survival might be worse in patients whose tumors carry the EGFRvIII mutation than in those who do not [75,82].

BRAF fusion or point mutation: The BRAF gene encodes a protein called B-Raf that is involved in cell signaling that promotes cell growth. Tandem duplications of BRAF and KIAA15A9 results in a gene fusion product called KIAA15A9-BRAF, which has constitutive B-Raf kinase activity. This fusion event is frequently detected in pilocytic astrocytomas, pleomorphic xanthoastrocytomas, and malignant astrocytomas [14,40]. The detection of the KIAA15A9-BRAF fusion is often used to help distinguish pilocytic astrocytomas from higher-grade astrocytic tumors. Other BRAF gene alterations including activating point mutations in BRAF (e.g., BRAF^{V600E}) have also been identified in low-grade as well as higher grade (III/IV) gliomas [83,84]. Small-molecule BRAF kinase inhibitors, such as CCT239065 and RG7204, have been evaluated in melanomas and may provide a new therapeutic approach to treat brain tumors harboring BRAF mutations [85].

Recent advances in sequencing along with the vast data from The Cancer Genome Atlas have re-classified GBMs into four distinct genetic subtypes including classical, mesenchymal, proneural, and neural [86].

Classical: The classical subtype features many of the common gene alterations observed in GBMs including chromosome 7 amplifications and chromosome 10 deletions. In addition, EGFR amplification was observed more frequently in classical GBMs than the other subtypes [86]. Other common abnormalities in genes, TP53, NF1, PDGFRA, or IDH1 are not found in this group [86]. In response to aggressive

treatment, patients in the classical group lived the longest compared to those in the other groups [86].

Mesenchymal: The mesenchymal subtype features mutations in the NF1 gene including focal hemizygous deletions of the 17q11.2 region, which contains the NF1 gene [86]. Mutations in the PTEN and TP53 tumor suppressor genes are also frequently observed in this subtype [86]. Interestingly, tumors of the mesenchymal subtype express Schwann cell markers such as the family S100A as well as microglial markers [86]. Genes associated with inflammation are enriched in this subgroup, which is evident in the observation that there is a higher overall fraction of necrosis in these tumors [86]. Patients in this subtype typically survive longer after aggressive treatment than those in the proneural and neural groups [86].

Proneural: The proneural subtype features alterations of the PDGFRA gene and point mutations in the IDH1 gene [86]. Although focal amplifications of the PDGFRA locus are seen in all GBM subtypes, proneural tumors have the highest rate. TP53 mutations and loss of heterozygosity are frequently observed in this subtype, while chromosome 7 amplification and chromosome 10 deletion are less prevalent [86]. This subtype shows high expression of oligodendrocytic development genes (e.g., PDGFRA, NKX2-2 and OLIG2), which may help explain its atypical GBM subtype status [87]. In addition, the proneural subtype contains several proneural development genes such as SOX, DCX, DLL2, ASCL1, and TCF4 [88]. This subgroup consistently represents younger patients than the other subgroups and is often associated with secondary GBMs [86]. Interestingly, patients in the proneural group who received aggressive treatment did not have a significant survival advantage compared with patients in the proneural group who did not receive aggressive treatment [86].

Neural: The neural subtype features expression of neuron markers such as NEFL, GABRA1, STY1, and SLC12A5 [86]. Tumors of this subtype are associated with neural, astrocytic and oligodendrocytic gene signatures [86]. Patients in the neural group had some improvement in survival but not as significant as those patients in the classical and mesenchymal groups [86].

Treatment and Prognosis

Although pilocytic astrocytomas commonly arise in the first two decades of life, late presentation in adults is reported and typically has a less favorable outcome [89]. If there is a high level of suspicion at the time of presentation for pilocytic astrocytoma, decision-making should consider the following: obtaining tissue diagnosis, restoring cerebrospinal fluid flow and decompressing adjacent neural structures, and non-surgical alternatives when an invasive procedure is precluded or incomplete.

Gross total resection of pilocytic astrocytomas provides the greatest clinical outcomes [90]. When risk of surgery is too high or if gross total resection cannot be performed, consideration for radio- and chemotherapy can be given. The long-term risks of radiotherapy in children suggest it be employed only in cases of recurrence or pilocytic astrocytomas with aggressive nature. While various case series have found prolonged survival after radiation therapy in children [91-93], the rate of recurrence after 5 years remains high. In a 10-year prospective trial where practitioners felt irradiation posed a high-risk for neurocognitive injury, a carboplatin and vincristine (CV) regimen was compared to a thioguanine, procarbazine, lomustine, and vincristine (TPCV) regimen. Although five-year event-free survival appeared more favorable using TPCV, toxicity observed with both

regimens presented a major limitation for long-term effectiveness in either group [94].

A multimodality approach consisting of surgical resection, radiation therapy, and chemotherapy is used in the treatment of malignant gliomas. Surgery plays a key role in the treatment of malignant gliomas as it allows for both cytoreduction and confirmation of diagnosis. Furthermore, there is growing evidence in the literature that achieving gross total resection is important in prolonging survival [95,96]. As discussed in the imaging section, advances in neuroimaging and brain mapping has allowed for increased gross total resection.

Radiation therapy is currently recommended for all patients with grade III and IV gliomas (anaplastic astrocytomas and GBM, respectively). Only infants, young moribund children, and patients declining treatment in favor of supportive care would not be recommended to receive some form of radiation therapy shortly after diagnosis. Radiation therapy alone has been shown to improve median survival from 3-4 months to 9-12 months [97,98]. Although radiation therapy has shown a clear improvement in survival for patients with primary GBMs, radiotherapy for recurrent GBMs is controversial mainly due to the serious risks associated with reirradiation including necrosis of healthy brain tissue [99]. Despite these serious risks, studies suggest there may be a benefit to a more focused radiation therapy through the use of stereotactic radiosurgery or fractionated stereotactic reirradiation [100,101].

Alkylating agents are the most frequently used chemotherapy drugs in treating anaplastic astrocytomas and GBMs. The cytotoxic effects are mediated primarily through DNA cross-linking, leading to cell death by apoptosis. Carmustine (BCNU) has been shown to prolong survival of GBM patients in two clinical trials [102,103], however, due to the post-operative complications; the addition of BCNU to a patient's treatment protocol is unclear and ultimately determined by the treating physician. In 2005, temozolomide was shown to improve median survival and increase the likelihood of long-term survival with newly diagnosed GBM when given concurrently with RT and then following RT [98]. Since this publication, the Strupp protocol, involving surgery followed by radiotherapy plus concomitant and adjuvant temozolomide for the treatment of malignant gliomas is the current gold standard [98].

Additional chemotherapeutic agents targeting specific molecules currently used in treating malignant gliomas include inhibitors of Epidermal Growth Factor Receptor (EGFR), Platelet-Derived Growth Factor Receptor (PDGFR), Vascular Endothelial Growth Factor (VEGFR), mammalian Target of Rapamycin (mTOR), Protein Kinase C (PKC), RAF-MEK-ERK pathway, and integrins [104]. With new chemotherapeutic agents being developed the gold standard for evaluating these agents remain randomized clinical trials. Recently, an anti-VEGF monoclonal antibody, bevacizumab, was evaluated in two randomized phase 3 clinical trials in which it was concluded that although there was improved progression-free survival, the overall survival in patients was not improved [105,106]. Although overall survival was not improved in these studies, progression-free survival increased 3-4 months and bevacizumab may still hold promise as an option for treating GBMs. With the development of novel chemotherapeutic agents, a combined multimodality approach including both chemotherapy and radiotherapy is needed to overcome tumor resistance through the use of multi-targeted strategies.

Since survival for patients with GBMs is relatively short, identifying prognostic indicators in order to stratify patients into risk groups may

provide valuable in determining the best therapeutic approach including enrollment into active clinical trials. Lamborn and colleagues surveyed 832 GBM patients enrolled into prospective clinical trials in order to identify potential risk groups [107]. From their analysis, they identified four risk groups in which the two lower risk groups included patients under the age of 40 with the lowest overall risk group being young patients with frontal lobe tumors only [107]. The intermediate risk group included patients aged 40-65 with KPS >70 who underwent subtotal or total resection of the tumor. Finally, the highest risk group included patients over the age of 65 or patients aged 40-65 with either KPS <80 or who only underwent biopsy [107].

In addition to the age of the patient and the KPS score, other clinical parameters serve as prognostic indicators of long-term survival. Tumor size and location are also important indicators as extent of resection is dictated not only by tumor size but also by location of the tumor. Finally, grade of tumor is an important indicator of long term survival as the higher the grade the more malignant the tumor is, which directly results in a poorer prognosis.

Oligodendroglial tumors

Historically, oligodendrogliomas (ODG) were thought to account for only about 4% of primary brain tumors [108], however, with better understanding of the tumor biology, it is thought that ODG may comprise as much as 25-33% of gliomas [109,110]. Up to 57-87% of patients with ODGs present with seizures [108,111] with 22% of patients presenting with headaches [108].

Histology

Calcifications are typically seen histologically with 73% of tumors having microscopic calcifications [112]. Histopathologic analysis of these tumors demonstrates a lucent perinuclear halo typically referred to as a "fried egg" appearance along with a "chicken-wire" vascular pattern [110]. Although these features are common to ODG, they are not pathognomonic. Since most ODGs contain microtubules and not glial filaments, they typically do not stain for Glial Fibrillary Acidic Protein (GFAP) [113]. Attempts for a grading system has been met with controversy, mainly surrounding the lack of prognostic significance [114]. Therefore, for prognostic reasons, a two classification system has been proposed that includes low grade; oligodendroglioma (WHO grade II) and high grade; anaplastic oligodendroglioma (WHO grade III) [114,115].

Treatment and Prognosis

Surgery is mainly reserved for low grade ODGs, while surgical data for high grade ODGs is less convincing. Radiation therapy for ODGs is also unclear [116] with one retrospective study showing a 10-year survival rate of 56% in those patients receiving postoperative radiation >45 Gy [117], while in another study, no difference in 5-year survival was observed in patients with or without postoperative radiation [118]. Chemotherapy is therefore the primary modality for treating ODGs as most respond to chemotherapy [119]. The 10-year survival rate for ODGs is 10-30% with pure ODGs having a higher survival rate than mixed ODGs [108,112]. The presence of calcifications on imaging as a prognostic indicator was evaluated in one series, however, additional studies are needed before conclusions can be drawn [108]. Finally, chromosomal 1p and 19q loss is associated with longer survival rates [120].

Ependymomas

Ependymomas are a rare type of glial tumor that is believed to arise from ependymal cells lining the cerebral ventricles and along the central canal of the spinal cord. A large portion of intracranial ependymomas (36-60%) occur in children, making ependymomas the second most common malignant brain tumor in this population [121]. Spinal ependymomas are more common in adults, occurring in 96% of cases [122].

Classification

Ependymomas are usually well circumscribed and benign but they have been known to be invasive. Ependymomas are divided into four major subgroups including myxopapillary (WHO grade I), subependymoma (WHO grade I), classic ependymoma (WHO grade II), and anaplastic ependymomas (WHO grade III). Within classic ependymomas there are an additional four variants including cellular, papillary, clear cell, and tanyctic. Lastly, it is worth noting that once considered a variant, ependymoblastoma, is now being regarded as a rare childhood primitive neuroectodermal tumor with abundant mitotic figures and true rosettes [123].

Histology

Similar to all brain tumors, the diagnosis of ependymoma requires histological confirmation. Classical ependymomas are characterized by dark small nuclei. They also show two cytoplasmic patterns: perivascular pseudorosettes and true rosettes. Perivascular pseudorosettes are areas of radiating neoplastic cells that lack nuclei and surround blood vessels. True rosettes are areas of ependymal tubules around a central blood vessel.

Treatment and Prognosis

Since ependymomas are highly radiosensitive, the best approach for treatment of ependymomas is gross total resection followed by radiation therapy. The role of chemotherapy for the treatment of ependymomas is currently unclear [124].

Mixed gliomas

Mixed gliomas are tumors that contain both oligodendroglioma and astrocytoma cells. These tumors can be classically divided into oligoastrocytoma (WHO grade II) and anaplastic oligoastrocytoma (WHO grade III). Oligoastrocytomas comprise 10-19% of low-grade gliomas and usually develops in middle-aged adults [125]. Treatment is usually surgical resection. If however, these tumors recur, the treatment approach is surgery followed by radiotherapy and chemotherapy.

Medulloblastoma

Medulloblastomas are the second most frequent childhood brain tumor after Pilocytic astrocytomas, and the most common malignant brain tumor in children comprising roughly 25% of intracranial tumors [126]. They occur exclusively in the posterior fossa with a peak incidence between 4 and 7 years [127]. Patients with medulloblastomas often present with symptoms of increased intracranial pressure (hydrocephalus) [128]. Since these tumors grow in the posterior fossa, gait ataxia, truncal instability, vomiting, dizziness and vision problems are also common symptoms caused by involvement of the cerebellum,

brainstem, or cranial nerves. Since metastases along the cranio-spinal axis are present in roughly 33% of patients [126], evaluation for metastases is recommended.

Classification and Histology

Although all medulloblastomas are classified as WHO grade IV, within this classification scheme, there are currently five variants including classic, desmoplastic/nodular, Medulloblastoma with Extensive Nodularity (MBEN), Large Cell (LC), and Anaplastic Medulloblastoma [129].

Molecular pathways: While the etiology is unknown, roughly 2-5% of medulloblastomas are associated with nevoid basal cell carcinoma syndrome (Gorlin syndrome), which is caused by mutations in the patched-1 gene (PTCH-1), and familial adenomatous polyposis, which is caused by inactivating mutations in the adenomatous polyposis coli gene [130,131]. In addition, great strides have been made in our understanding of the oncogenesis of medulloblastomas. Based on gene expression profiles using tissue microarrays and substantiated using whole genome and whole exome sequencing, medulloblastomas have been separated into four distinct subgroups based on their unique molecular profiles [132-135]. These groups include sonic hedgehog (SHH), wingless (WNT), group 3, and group 4.

SHH medulloblastomas: This subgroup comprises roughly 25-30% of medulloblastomas and is characterized by high desmoplasia [132,134-137]. The overexpression of SHH pathway leads to the binding and inactivation of PTCH-1, which normally blocks activation of a number of transcription factors through the inhibition of smoothened (SMO). SHH also upregulates MYCN, which is involved in the cell cycle. In addition to mutations in PTCH-1, mutations in SMO and Suppressor of Fused Homolog (SUFU) have also been observed in SHH medulloblastomas [138,139]. Chromosomal aberrations have also been associated with this subgroup including loss of 9q (accounting for 21-47%), 10q, 20p, 21p and gain of chromosome 3q and 9p [132,136,140]. The majority of SHH medulloblastomas occur in infants under the age of 3 and again in adults above age 16 [136,141]. In fact, nearly half of all adult medulloblastomas are of the SHH variant [142]. The overall survival is good in infants and intermediate in adults [143].

WNT medulloblastomas: The WNT medulloblastoma subgroup harbors mutations in essential genes of the WNT pathway including APC, β -catenin, and axin 1 [144-146]. Roughly 10-15% of medulloblastomas fall within this subgroup [137]. They are characterized by classic histology, more than 90% of WNT tumors, affecting patients above 3 years old, good prognosis, and infrequent metastasis [137]. Unlike SHH tumors, WNT tumors rarely affect infants and the overall survival is generally good [141,143].

Group 3 and 4 medulloblastomas: Both of these subgroups present with common clinical features and share similar molecular profiles. The age of onset for both groups vary with Group 3 peaking in childhood (3-10 years), while Group 4 has a more distributed age of onset from infancy to adulthood [136,141]. The majority of tumors in both groups display classical histology. Chromosomal aberrations are common to both groups with isochromosome 17q representing the most frequent structural alteration [135]. Gain of 7 and 18q along with loss of 8 and 11p are also common abnormalities [136,137,147]. MYC amplification in group 3 is the main difference between these groups since MYC amplification is rarely observed in group 4 [147,148]. Conversely, enrichment of chromosome X loss is more common in

group 4 as observed in 80% of females in this group [147]. Lastly, both groups have similar metastatic rates but group 3 has a poorer prognosis than group 4, which shows intermediate prognosis [148].

Treatment

Medulloblastomas are among the most radiosensitive tumor of the central nervous system and are moderately chemosensitive. Therefore, the recommended therapeutic approach is surgical debulking followed by radiation therapy. 36 Gy to the entire craniospinal axis with a boost to the tumor bed for a total of 55 Gy is the recommended radiation therapy dosage [149]. Medulloblastomas are moderately chemosensitive, and as such, chemotherapy is now an integral part of the treatment of these tumors, including recurrent disease [150]. Some chemotherapy agents used include lomustin, cisplatin and vincristine. Finally, placement of permanent ventriculoperitoneal shunts is required in 30-40% of patients after tumor resection of the posterior fossa [151,152].

Prognosis

Patients with medulloblastomas are classified into three risk groups that help facilitate treatment and provide predictions on prognosis. These groups include standard risk, intermediate risk, and poor risk. Patients with no residual tumor demonstrated on post-operative MRI along with negative CSF results are classified as standard risk and carry a 5-year survival rate of 73% [153]. Residual tumor measuring greater than 1.5 cm² on post-operative MRI and presence of tumor cells in the brain, spine, or CSF are characteristic of patients in the poor risk group. These patients have a poor prognosis with a 5-year disease free survival of 36-52% [154]. The intermediate risk group is poorly characterized leaving the other two groups as primary predictors. Interestingly, females have a better prognosis than males [155].

Meningioma

Meningiomas arise from the layer of tissue covering the brain and spinal cord. Meningiomas are the most common benign intracranial tumor accounting for about 13-26% of all primary brain tumors [156]. Most meningiomas are intracranial, however, spinal meningiomas may occur accounting for an estimated 7.5 to 12.7% of all meningiomas [157]. The vast majority of meningiomas rarely metastasize with a rate of less than 1 in every 1,000 meningiomas [158]. The most common sites of metastasis include lung and pleura, liver, lymph nodes, and bone [159]. Due to the slow growing nature of meningiomas, most remain asymptomatic. Meningiomas rarely affect children with an incidence rate of roughly 2.2% [160] and of these, prevalence of neurofibromatosis was 14.5% [161]. There have been reports documenting prevalence of meningiomas in roughly 50% of NF2 patients [162] with one study reporting prevalence as high as 58% [163]. If multiple meningiomas are observed, suspicion of neurofibromatosis type 2 is high. Both neurofibromatosis types 1 and 2 will be discussed in the vestibular schwannoma section.

Classification and Histology

Meningiomas have a complex and broad range of histological patterns. The WHO currently recognizes 16 different variants grouped into three grade designations (Table 1) [129]. Meningiomas falling in the WHO grade I classification include: meningothelial, fibrous, transitional, psammomatous, angiomatous, microcystic, secretory, lymphoplasmacyte-rich, and metaplastic. Although each subtype has a

distinct histological pattern, all WHO grade I meningiomas have a low recurrence rate of 9% with no evidence of brain invasion [164]. Three subtypes of meningiomas fall within WHO grade II and these include: chordoid, clear cell, and atypical. One major difference between the 2000 WHO classification scheme and the recent 2007 WHO classification scheme is the recognition that meningiomas with evidence of brain invasion should be classified as WHO grade II regardless of a benign histological appearance [165,166]. WHO grade II meningiomas have a recurrence rate of 29% [164]. Lastly, meningiomas in the WHO grade III group include papillary, rhabdoid, and anaplastic with the anaplastic variant representing the majority of WHO grade III cases [129]. All WHO grade III meningiomas have increased mitotic activity with a recurrence rate of 50% [164].

Treatment

If the tumor is not causing symptoms, tumor growth may be watched using serial MRIs. Otherwise, surgery is the standard of care for treating meningiomas. Similar to other tumor types, extent of tumor resection is beneficial for minimizing the risk of tumor recurrence. As a result, in 1957, Simpson established a classification system consisting of five subdivisions to assess extent of resection of meningiomas and to correlate postoperative recurrence rates with extent of resection [167]. In his grading system, Simpson grade I is defined as complete tumor resection with excision of the dural attachment and any abnormal bone. If the venous sinus is involved, complete resection of the sinus is also performed. Simpson grade II is defined as complete tumor resection with coagulation of the dural attachment. Simpson grade III is defined as complete tumor resection without resection or coagulation of the dural attachment. Simpson grade IV is defined as a subtotal resection. Finally Simpson grade V is defined as a simple decompression, with or without biopsy. The risk of tumor recurrence (minimum of 6 months of follow-up) for Simpson grades I, II, III, and IV were 9%, 16%, 29%, and 39%, respectively [167].

With advancements in surgical techniques and treatment options, such as radiation therapy, relying solely on the Simpson grading system of meningiomas in the modern era to predict recurrence is inconclusive. Therefore, cell proliferation markers, such as Ki-67 (MIB-1 – monoclonal antibody) are being evaluated to complement the Simpson grading system in predicting tumor recurrence [168]. In a study conducted by Oya and colleagues, they determined that the MIB-1 index could differentiate meningiomas with a high risk of recurrence [168]. Further, the authors conclude that using the MIB-1 index could be beneficial in planning optimal follow-up strategies with a shift from attempting aggressive resection to valuing the quality of the patient's life [168].

Several retrospective studies have demonstrated that radiation therapy (e.g., external beam radiation therapy and stereotactic radiosurgery) can provide improved and durable local control in selected patients with meningioma [169]. For WHO Grade I or presumed Grade I meningiomas, radiation therapy achieved long-term local control in 68% to 100% of cases at 5 to 10 years, including patients treated postoperatively, primarily, or following tumor recurrence. The use of stereotactic radiosurgery is considered most effective for patients with small meningiomas (usually less than 3 cm in diameter or 10 cm³ in volume), those with distinct margins, and those at sufficient distance from functionally important brain, nerves, and other critical structures to permit safe delivery of an adequate target dose [169].

For high-grade meningiomas, a multimodal approach using radiation therapy and/or chemotherapy is usually given. Achieving maximum resection and adjuvant radiotherapy have been shown to be independent predictors of patient survival and disease-free survival in the treatment of malignant meningioma [170]. Evaluating stereotactic radiosurgery in the setting of subtotal resection or recurrence, reported local control rates (>2 years) range from 0% to 90%, with the majority falling within 50% to 80% for WHO grade II meningiomas [169]. While some studies have suggested stereotactic radiosurgery is not indicated for malignant meningiomas [171], others have shown improved local control rates of 17% at 15 months [172]. Finally, for recurrent atypical or anaplastic meningiomas not suitable for radiosurgery, resection followed by permanent brachytherapy is a potential salvage therapy that has shown promise in the clinical setting [173,174]. In the largest series (n=21) to date examining brachytherapy for therapy for the recurrence of aggressive atypical and anaplastic meningiomas, Ware et al. reported a median survival of 1.6 years after [125] I implantation for atypical meningiomas and 2.4 years for anaplastic meningiomas [173]. Due to the high complication rates observed including radiation necrosis occurring in 27% in one study, meticulous surgical technique and medical therapies to assist with wound healing after surgery is required [173].

For meningiomas that are inoperable and/or radiation-refractory, chemotherapy is often used with little to no effect. As a result of failed chemotherapeutic approaches, several studies have investigated various chemotherapies in which all have been disappointing [175,176]. Although there is limited data, hydroxyurea, somatostatin analogues and interferon- α have all been modestly successful in patients with recurrent meningiomas [175]. Further, emerging targeted therapies including sunitinib, may prove useful in refractory meningiomas [175,176].

Prognosis

Prognosis for patients with benign meningiomas is generally good with a 5-year survival rate of 91.3% [177]. Recurrence depends on extent of surgical resection with a recurrence rate of 8% in cases with a gross total resection, a 29% in cases with a subtotal resection [178]. Atypical meningiomas have been reported to have a higher rate of local recurrence and are associated with lower survival rates compared to benign meningiomas [179]. Similar to benign meningiomas, achieving a gross total resection of atypical meningiomas was associated with a lower recurrence rate (11%) compared to achieving a subtotal resection (100%) [179]. A similar trend of increased survival is also associated with (grade I) total resection of malignant meningiomas [180]. Lastly, Al-Mefty and colleagues investigating the malignant progression in meningioma from a benign to a higher histological grade, and concluded that the presence of complex genetic alterations (e.g., increased MIB-1 staining and chromosomal aberrations such as alterations in chromosome 22 and deletion of chromosomes (1p, 14q, and 10q), even with a benign histological grade, may potentially have an aggressive phenotype and require closer follow up [181].

Pituitary Tumors

The majority of pituitary tumors are adenomas arising from the anterior pituitary gland (adenohypophysis). In rare cases, pituitary carcinomas have been described [182]. Pituitary tumors arising from the posterior pituitary gland (neurohypophyseal) are also rare [183]. Pituitary adenomas are the fourth most common intracranial tumor after gliomas, meningiomas and schwannomas [184]. Pituitary

adenomas are typically benign with even malignant pituitary tumors rarely metastasizing [184]. These tumors may secrete abnormally high amounts of hormones that may lead to physiological dysfunction resulting in patient morbidity. In addition to endocrinologic disturbances, mass effect leading to bitemporal hemianopsia is often observed in patients with pituitary neoplasms. Several risk factors have been identified including Multiple endocrine neoplasia type 1 (MEN1), Carney's complex, and Familial isolated pituitary adenoma [185].

Classification and Histology

A functional classification scheme has been developed based upon the secreted hormones and include lactotrophic adenomas (prolactinomas) which secrete prolactin, are the most common, and causes amenorrhea-galactorrhea syndrome in women and impotence in men, somatotrophic adenomas which secrete growth hormone, corticotrophic adenomas which secrete adrenocorticotrophic hormone, gonadotrophic adenomas which secrete luteinizing hormone and follicle-stimulating hormone, thyrotropin-secreting adenomas which secrete thyroid-stimulating hormone, cause thyrotoxicosis, and are rare, and null cell adenomas which do not secrete hormones. Excessive amounts of adrenocorticotrophic hormone from pituitary corticotrophic adenomas can lead to Cushing's syndrome. As a consequence of undergoing adrenalectomy for treatment of Cushing's syndrome, 8-43% of patients will develop hyperpigmentation referred to as Nelson's syndrome [186,187]. An increased concentration of growth hormone from somatotrophic adenomas can lead to acromegaly. Interestingly, more than 95% of cases of acromegaly are due to a pituitary somatotroph adenoma [188].

Treatment

MR imaging with contrast on a pituitary protocol is the gold standard for evaluating pituitary tumors. Since the normal pituitary gland also enhances, the timing of the contrast is important in achieving a high-spatial-resolution image that is able to discern normal pituitary tissue from a macroadenoma [189]. In patients with non-secreting tumors and without neurologically deficits, it is reasonable to follow these patients with serial MRIs and visual field examinations.

Current treatment options for symptomatic pituitary adenomas include surgical resection, radiation therapy and medication therapy (first line for treating prolactinomas). Three dopamine agonists are routinely given to treat prolactinomas and include bromocriptine, cabergoline, and pergolide. Surgery using a transsphenoidal approach is typically the first line treatment for the other subtypes of pituitary adenomas [190,191]. Medical therapy is also used for patients with somatotrophic adenomas and includes dopamine agonists (e.g., bromocriptine), somatostatin analogues (e.g., octreotide), and growth hormone antagonists (e.g., pegvisomant). Medical treatment for thyrotropin-secreting tumors typically involves somatostatin analogues (e.g., octreotide).

For the treatment of pituitary adenomas, conventional radiation therapy typically consists of 40-50 Gy administered in 20-25 fractions over 4-6 weeks [192]. Although effective, complications associated with radiation therapy are high. One of the major post-radiation complications is hypopituitarism, which is both dose- and time-dependent. In addition, injury to the optic nerves and chiasm, lethargy, memory disturbances, cranial nerve palsies, and tumor necrosis with hemorrhage and apoplexy may also occur. The effects of radiation therapy on somatotrophic adenomas is cumulative with time and may

take 10 years to reduce the growth hormone levels into a “curative” range [193,194]. In contrast, for Cushing’s disease, radiation therapy restores ACTH levels to normal range between 18 and 42 months [195].

The use of radiosurgery for the treatment of pituitary tumors is still limited to tertiary centers and protocols are not standardized. Yang et al. conducted a large aggregated

analysis of stereotactic radiosurgery treatment in patients with acromegaly and determined that the overall disease control rate was approximately 48%–53% for patients no longer taking suppressive medications after radiosurgery for acromegaly [196]. The post-radiosurgery remission rates for Cushing’s disease reported in the literature vary considerably from 0% to 100%, with most series documenting an approximately 50%–60% remission rate [197]. Long-term complications are thought to be similar to conventional radiation therapy (except for optic symptoms).

Prognosis

In cases where the tumor is compressing the optic apparatus, removal of the tumor improves vision in 90% of patients [198]. Furthermore, in one case series, only 27% of patients with prolactin-secreting tumors and 20% of patients with growth hormone-secreting macroadenomas returned to baseline hormone levels after surgical resection [198]. In the same series, the recurrent rate was roughly 13%. However, the inclusion of post-operative radiation therapy as well as degree of surgical resection of the tumor influenced the rate of recurrence. For example, for patients with partial surgical resection who did not receive post-operative radiation therapy, their recurrence rate was 50% [198]. On the other hand, there were no recurrences observed in patients with a gross total tumor removal who received postoperative radiation therapy [198].

The posterior pituitary can sometimes be damaged during surgery leading to a condition called central diabetes insipidus, which is characterized by excessive thirst and dilute urine.

Primary Central Nervous System Lymphoma

Primary Central Nervous System Lymphomas (CNS lymphomas) is a rare aggressive form of extranodal high-grade non-Hodgkin lymphoma that represents roughly 4% of intracranial neoplasms [199]. Several risk factors have been identified for primary CNS lymphomas including collagen vascular diseases (e.g., systemic lupus erythematosus and Sjogren’s syndrome), immunosuppression, including AIDS, and Epstein-Barr virus, which is associated with many lymphoproliferative disorders with nearly 100% association with primary CNS lymphomas [200], especially AIDS related [201]. On CT imaging, non-AIDS CNS lymphomas typically have a homogeneous enhancement pattern, whereas AIDS CNS lymphomas tend to have a necrotic center with the appearance of multifocal ring-enhancing lesions [202]. Surgery has a limited role in treatment and is used mainly for tumor biopsy. Radiation therapy and chemotherapy are primarily used to treat CNS lymphomas. Recent advances in treatment options have resulted in the use of high-dose chemotherapy in combination with autologous stem cell transplantation as an alternative treatment approach [199]. Whole-brain radiation therapy is often used with 40-50Gy, especially when chemotherapy is contraindicated [203].

For patients with non-AIDS CNS lymphomas the combination of radiation therapy and chemotherapy has a better overall survival than radiation therapy alone [204]. The median survival for patients not receiving treatment is 1.8-3.3 months, with radiation therapy alone median survival increases to 10 months, and with intraventricular methotrexate, the median time to recurrence increases to 41 months [205]. On the other hand, patients with AIDS-related CNS lymphomas have a much worse prognosis with a median survival of 2-5 months even after treatment [206,207].

Vestibular Schwannoma

Vestibular schwannomas, also known as acoustic neuromas, arise from cells that produce the myelin sheath covering the vestibulocochlear nerve (CN VIII). They usually originate in the superior vestibular division of CNVIII. These tumors are generally benign with an incidence rate of 1.67 per 100,000 person-years [1]. Vestibular schwannomas have been linked to loss of NF2 (tumor suppressor) on chromosome 22 [208,209]. Roughly 95% of vestibular schwannomas are unilateral, however, bilateral vestibular schwannomas is pathognomonic of neurofibromatosis type 2 (NF2) [210,211]. Most patients present with a classic clinical triad of hearing loss, tinnitus (high pitched), and disequilibrium [212]. Histologically these tumors contain Antoni A and B fibers [213]. Antoni A fibers are narrow elongated bipolar cells that are tightly packed, while Antoni B fibers are loosely packed cells with reticular fibers [213]. Verocay bodies (cellular areas surrounded by parallel arrangement of spindle shaped Schwann cells) are also seen histologically [214]. Since these tumors involve CN VIII, audiometric evaluation is part of the initial workup of this tumor and is used to help guide management. There are three approaches to treating vestibular schwannomas. These include complete surgical resection, radiation therapy, or monitoring using consecutive MRIs. With complete surgical removal, the incidence of recurrence is minimal from 0%-3% [215]. There are currently three surgical approaches used including retrosigmoid, which may preserve hearing, translabyrinthine, which sacrifices hearing but increases chances of preserving VII, and middle fossa approach, which is usually reserved for small lateral vestibular schwannomas [216]. The tumor progression rate following subtotal resection is roughly 20% [215].

Neurofibromatosis

Although there are six subtypes of neurofibromatosis, the most common are type 1 (NF1) and type 2 (NF2) and are discussed below.

Neurofibromatosis type 1

NF1 is an autosomal dominant disease that is linked with mutations in the NF1 gene on chromosome 17 which codes for neurofibromin [217]. Loss of neurofibromin, which is a negative regulator of the Ras oncogene, leads to increased growth stimulating signaling. Neurofibromatosis 1 represents more than 90% of neurofibromatosis [218]. The diagnosis of NF1 is made by two or more of the following [219]:

- 6 café au lait spots
- 2 neurofibromas of any type or one plexiform neurofibroma
- hyperpigmentation in the axillary or inguinal areas
- optic glioma
- >2 Lisch nodules (pigmented iris hamartomas)

- distinctive osseous abnormality (e.g., sphenoid dysplasia)
- a first degree relative with NF1

Neurofibromatosis type 2

NF2 is an autosomal dominant disease linked with mutations in the NF2 gene on chromosome 22, which codes for merlin (schwannomin) [211]. This tumor suppressor is typically produced in the central nervous system particularly in Schwann cells. The diagnosis of NF2 is made with either [220,221]:

- Bilateral vestibular schwannomas on imaging (MRI or CT)
- A first degree relative with NF2 and either:
- Unilateral vestibular schwannoma at age <30 years or
- Any two of the following: meningioma, glioma, posterior subcapsular lens opacity, neurofibroma

Additional criteria that carry less weight include:

- Unilateral vestibular schwannoma at age <30 and any of the following: meningioma, schwannoma, glioma, posterior subcapsular lens opacity or
- Multiple meningiomas and either of the following: unilateral vestibular schwannoma or any two of the following: glioma, neurofibroma, schwannoma, or cataract

Neurofibroma

Neurofibromas are benign nerve sheath neoplasms arising in the peripheral nervous system. They are typically found in patients with neurofibromatosis. These tumors are divided into dermal (WHO grade I) and plexiform (WHO grade I). Dermal neurofibromas are usually associated with a single peripheral nerve and do not acquire malignant features. However, plexiform neurofibromas are associated with multiple nerve bundles and although low, have the ability to transform into malignant tumors, making these tumors more difficult to treat. The clinical course usually dictates the treatment approach for neurofibromas. Since these tumors are intimately intertwined with functional nerve, surgery is only performed if there are symptoms from the neurofibroma leading to progressive morbidity [222]. Other treatment options may include antihistamines, maturation agents (e.g., retinoic acid), and antiangiogenic drugs [222]. Targeted therapeutic approaches for specific molecular pathways vital to the tumor are promising, but need to be evaluated in clinical trials [222].

Schwannoma vs. neurofibroma

Both of these tumors are very similar and hard to distinguish without histological analysis. Schwannomas are typically well circumscribed and consist of Antoni A and B fibers. On the other hand, neurofibromas are typically less cellular, not as well circumscribed, and consist of wavy collagen fibers with occasional neuritis [223]. S-100 staining is oftentimes used to help distinguish these tumors since schwannomas typically display a greater percentage of positive cells and the intensity of staining is higher. However, with both tumors expressing some degree of S-100, this stain alone is not sufficient differentiating these tumors. Other stains that have been studied with varying sensitivities and specificities to help decipher these two tumors include calretinin, CD34, CD56, glial fibrillary acidic protein (GFAP), epithelial membrane antigen (EMA), factor XIIIa, Leu-7, myelin basic protein and Glut-1 [223,224]. A combination of

immunohistochemical stains provides the greatest support in determining schwannoma versus neurofibroma.

Secondary Brain Tumors

Epidemiology

Secondary, or metastatic brain tumors (MBTs), are the most common malignancies of the central nervous system (CNS). Typically, MBTs arise from primary tumor cells that migrate hematogenously or via direct invasion of adjacent tissue. According to population studies, the estimated prevalence of MBTs in the United States is 7-14 cases per 100,000 people [225,226]. Given the considerable advancement of diagnostic imaging, preventive screening, and increasing life spans in developed countries, these national statistics likely underestimate the actual incidence [227].

In patients with previously diagnosed cancer, 10-30% will also develop a brain metastasis [228]. This is partly due to the inherent capacity of malignant tumor cells to invade and cross basement membranes and migrate to healthy tissue. While patients typically present with non-specific symptoms, the most frequently observed findings include weakness, impaired balance, headaches, and seizures. Therefore, any patient with a history of a primary malignancy, who presents with neurological symptoms, should be thoroughly evaluated for a CNS metastasis.

Data collected by CBTRUS have determined that MBTs most often originate from malignancies of the lung (39% to 56%), breast (13% to 30%), skin, colorectal, kidney (2% to 6%), or unknown primaries (2% to 14%) [229,230]. Melanoma and primary colon cancers contribute approximately 6%-11% and 3%-8%, respectively (Table 3). Of note, the malignancy potential among primary etiologies varies with respect to the propensity to metastasize to the CNS. For instance malignant melanoma, which represents only 6% of all cancers [229], has the highest propensity of all systemic malignant tumors to metastasize to the brain [231]. This is supported by incidence rates of brain metastases secondary to malignant melanoma, which vary widely from 6% to 43% in clinical series [232-234] to 12% to 90% in autopsy series [231]. Lung cancer ranks second in overall metastatic lesions with roughly 10% of lung cancer patients presenting with CNS metastases. The incidence of MBTs rises to 20% during treatment and finally, MBTs are observed in an estimated 50% of patients at autopsy [235-237].

Primary Tumor Source	Frequency
Lung	39%-56%
Breast	13%-30%
Kidney	2%-6%
Melanoma	6%-11%
Colorectal	3%-8%
Ovarian	1.20%
Unknown	2%-14%

Table 3: Primary source of brain metastasis

It is important to note that the spectrum of metastasizing primary cancers and the risk of CNS involvement varies by patient age

[238-240]. For instance, CNS metastases occur more frequently in adults, with the highest incidence seen in the fifth to seventh decades of life [238,239]. As mentioned previously, the most common primary sources of brain metastases in adults are cancers arising from the lung, breast, kidney, gastrointestinal tract, or skin, but may originate from any part of the body [229,230]. In children, the most common source of a brain metastasis is leukemia, followed by lymphomas and bone/soft tissue malignancies, including osteogenic sarcoma and rhabdomyosarcoma especially among children younger than 15 years [238]. Finally, germ cell tumors are the most frequent source of brain metastases in patients 15 to 21 years old [238].

The incidence of the primary cancers between males and females leads to differences in the sources of CNS metastasis. For example, breast cancer is the most common source of CNS metastases in women, whereas lung cancer is the most common source in men [239]. Comparing males and females diagnosed with melanoma, males are more likely to experience tumor spread to the CNS, as their primary tumors develop in locations that make it easier to spread to the brain, such as the head, neck, or trunk [231,241].

Imaging

MRI has high sensitivity to detect secondary CNS tumors. Namely, T1- and T2- weighted imaging modalities with and without contrast are used as the gold standard for initial evaluation. Generally, MBTs are detectable by gadolinium contrast enhancement and appear as round, well-circumscribed lesions surrounded by disproportionate areas of vasogenic edema. However, MBTs may have minimal or absent edema [242]. Accordingly, MBT should be included in the differential diagnosis for ring-enhancing lesions, as 30% of ring-enhancing lesions are diagnosed as secondary tumors [240].

Computed Tomography (CT) is a vital tool for initial work-up and perioperative management. On non-contrast enhanced CT (NECT), confined lesions may appear hypo-, iso-, or hyperdense relative to surrounding brain parenchyma [242]. Hyperdensity without contrast, especially at the gray-white junction and watershed zones of major vascular arteries, may suggest acute hemorrhagic lesions or CNS melanoma [243,244]. As with MRI, though, contrast enhanced CT is preferred to NECT for diagnostic purposes and is a viable alternative in cases where MRI is contraindicated.

Advanced MRI techniques such as Magnetic Resonance Spectroscopy (MRS), Magnetic Resonance Perfusion (MRP), Diffusion Weighted Imaging (DWI) and Diffusion Tensor Imaging (DTI) are employed for their specificity and to monitor treatment response. DTI, which hinges upon diffusion of water molecules along axons, is helpful to evaluate the integrity of White Matter (WM), as well as the location and orientation of WM tracts [245]. As an adjunct to other imaging, this technique has supplemented pre-surgical planning and characterization of intra-axial lesions to discern metastases from GBM [246]. MRP and DWI have utility in differentiating metastatic tumors from cerebral abscesses. In MRP studies, relative Cerebral Blood Volume (rCBV) is decreased for cerebral abscess and vice versa for metastatic lesions [247]. On DWI, restricted diffusion will be evident in the central non-enhancing portions of a cerebral abscess [248].

Treatment

Current approaches to the management of brain metastases are driven by prognostic factors, including the Karnofsky Performance Status (KPS), tumor histology, number of metastases, patient age, and

status of systemic disease [249]. These approaches include surgery, Whole Brain Radiotherapy (WBRT), Stereotactic Radiosurgery (SRS), and chemotherapy. The standard of care for treating these lesions is usually surgery and/or radiosurgery. In cases of multiple lesions, WBRT may be a viable option. Unfortunately for the vast majority of metastatic brain tumors, there are no good chemotherapeutic options.

Surgical Resection: Informed clinical judgment is imperative when considering surgical resection of MBTs. Appropriate surgical candidates should be free of systemic cancer progression, have controlled primary disease, and independent function as evident by a KPS >70. Moreover, patients with solitary brain lesions generally benefit more from surgical intervention than those with multiple brain lesions [250]. Negative predictors include extensive extracranial disease and low karnofsky performance scores (KPS<70).

As our evidence and knowledge of metastatic disease expands, surgery is now being considered as one of the primary treatments for brain metastases. As such, best practices for surgical resection are being evaluated. In two retrospective comparative studies from MD Anderson, en bloc resection was compared to piecemeal resection in patients with supratentorial and infratentorial single metastases. These studies demonstrated that there was a significant increase in the risk of leptomeningeal dissemination in patients who underwent piecemeal tumor resection compared with en bloc resection [251] and a higher risk of local tumor recurrence with piecemeal resection than with en bloc resection [252].

When combined with post-operative WBRT, surgery has been shown to dramatically improve survival, local control, and duration of independent function in selected patients with solitary tumors [253,254]. Comparing treatment strategies in patients with 1-3 MBTs, combined surgery + WBRT (2 year recurrence rate 23%-42%) has outperformed WBRT alone (recurrence rate at <1yr, 43%) [255,256].

Radiation: Radiation therapy is commonly used to treat cerebral metastases. The addition of WBRT increases patient survival to 4 to 7 months. Some studies have shown that focal radiation treatment in addition to WBRT can increase survival for up to 12 months [250]. A Radiation Therapy Oncology Group trial (RTOG 9508) concluded several important aspects of radiation therapy in the treatment of cerebral metastases [250,255]. For instance, in the case of a single brain metastasis, SRS in addition to WBRT was shown to increase survival [250]. In the case of multiple brain metastases (limited in this case to 1-3), The RTOG 95-08 trial concluded that SRS in addition to WBRT provides palliative benefits and should be used for patients who have a KPS >70 [250,255]. For patients with more than 3 brain metastases, SRS has not been shown effective [250].

Chemotherapy: Several chemotherapeutic agents have been studied in combination with WBRT for patients with brain metastases. Long-term benefits from treating MBTs using chemotherapy are yet to be seen [249], though, as positive clinical responses from anti-cancer agents are quickly undermined by drug toxicity. This negatively impacts quality of life and results in minimal benefit to overall survival [249]. Incidentally, no chemotherapeutic agents have been approved for treatment of MBTs.

A major challenge to chemotherapeutic success is drug-delivery across the blood-brain barrier (BBB). Novel chemotherapeutic agents capable of crossing the BBB may promote survival and ultimately improve the bleak outlook posed by MBTs. Preliminary data from an inhibitor of topoisomerase I, topotecan, which crosses the BBB, has shown to be effective in treating brain metastases from small cell lung

and breast cancer [257]. An alkylating agent approved for treatment of malignant gliomas, temozolomide (TMZ), also has the capability to cross the BBB and may be effective against brain metastases secondary to melanoma [258]. Additionally, TMZ may be combined with cisplatin to treat metastases from non-small cell lung cancer, breast cancer, and melanomas [259]. Finally, a novel lipid-conjugated compound, DM-CHOC-PEN, has shown promising results in Phase I clinical trials [260-262]. After crossing the BBB, DM-CHOC-PEN is preferentially taken up by tumor cells, resulting in cytotoxicity via pseudo-alkylation of N7-guanine, while sparing normal brain parenchyma.

Prognosis

Until recently, the mainstay of MBT treatment had been whole-brain radiation therapy (WBRT) for symptom relief. When considering all patients with CNS metastases, WBRT alone improved median survival after diagnosis from 1 month to 3-6 months [228]. Given the poor prognosis and the peak incidence in elderly populations, most plans center on palliative options. To address this, the RTOG, a collaborative group funded by the National Cancer Institute established to improve clinical outcomes and quality of life in cancer patients, established the Recursive Partitioning Analysis (RPA) classification to guide selection of patients who would benefit from intervention. The reliable predictors to determine short-term vs long-term survival include extracranial disease activity, number of CNS foci (e.g., solitary vs. multiple), and performance status (i.e., KPS).

Without treatment, metastatic brain disease confers median survival of about one month. Whole-Brain Radiation Therapy (WBRT), however, only improves survival to 3-6 months, with most individuals succumbing to systemic illness rather than to CNS metastasis.

The RPA classification has been validated in patients with breast, lung (NSCLC and SCLC), malignant melanoma, and unknown primaries. Additionally, it has been applied to cases where surgical resection and SRS have been main local treatment modalities. Table 4 represents the 3 classes defined by RPA.

RPA Class	Description
I	Controlled primary tumor Age < 65 years; Karnofsky Performance Status (KPS) > 70 Absence of non-CNS metastases
II	Patients that do not fit classes I or II
III	KPS < 70

Table 4: Recursive partitioning analysis classification scheme.

Patients who received WBRT after surgery, which in general had favorable prognosis (RPA Class I) prior to any treatment, represent the most homogeneous cohorts studied. Differences in survival outcomes among clinical trials, however, have been large. Overall median survival in RPA Class I ranged from 15–29 months; Class II was 5.5–11 months; Class III was 1.4–9 months [263-267].

Stereotactic Radiosurgery (SRS) may be another option for treatment of solitary or multiple tumors less than 3 cm or as an adjunct to other therapies, respectively. Retrospective reports evaluating the benefits of SRS in treating solitary metastasis have demonstrated a high

rate of local control at one year (80%-95%) and a reduced risk for developing late side effects and severe neurologic complications (<10%) when administered without WBRT [268,269]. In a multi-center, retrospective analysis of radiosurgical outcomes from patients treated with WBRT followed by adjunct radiosurgery, Auchter et al. observed a 86% local control rate, with an actuarial median survival of 56 weeks and duration of functional independence (i.e., KPS >70) of 44 weeks [270].

Though strides have been made to improve survival and prolong independent function in patients with CNS metastases, the best treatment modality or regimen for a large, heterogeneous group of MBT patients is unknown [271]. Moreover, few options are available to patients with significantly impaired KPS. Indeed some retrospective studies have shown prolonged survival in patients with KPS <70 using SRS as an adjunct to WBRT (median survival 8.7 mo) compared to WBRT-alone [272], but no prospective trials involving this patient population have been performed. In most cases, the conservative approach is taken, which involves prescription of corticosteroids, namely dexamethasone, to mitigate the neurological symptoms secondary to intracranial mass effect. In patients treated with corticosteroids-alone, survival rates may be extended up to two to three months [273,274].

Conclusions

Brain metastases are a devastating complication of advanced systemic cancer and remain the most common intracranial tumor type. Patients develop metastatic disease in the setting of advanced pathology and thus palliative care is often the primary therapy offered. However, there has been a change in recent years in diagnostics and treatment strategies for systemic cancer with associated CNS involvement, in which, more effective treatments are allowing patients to experience limited or stable systemic disease. Currently, surgery and SRS offer the best outcomes, but their combined impact on survival leaves much to be desired. WBRT is still an important treatment modality for patients with multiple brain metastases as well as patients who cannot tolerate surgery and SRS largely due to the palliative effectiveness in improving quality of life of patients with brain metastasis. Continued research and development of novel chemotherapeutic agents may offer therapeutic advantages in the future, but to date none have led to significant gains for improving clinical outcomes.

References

1. <http://www.cbtrus.org/factsheet/factsheet.html>
2. <http://www.cancer.org/acs/groups/content/@editorial/documents/document/acspc-044552.pdf>
3. Macdonald D, Kiebert G, Prados M, Yung A, Olson J (2005) Benefit of temozolomide compared to procarbazine in treatment of glioblastoma multiforme at first relapse: effect on neurological functioning, performance status, and health related quality of life. *Cancer Invest* 23:138-144.
4. Chandana SR, Movva S, Arora M, Singh T (2008) Primary brain tumors in adults. *Am Fam Physician* 77: 1423-1430.
5. Wen PY, Kesari S (2008) Malignant gliomas in adults. *N Engl J Med* 359: 492-507.
6. Fisher JL, Schwartzbaum JA, Wrensch M, Berger MS (2006) Evaluation of epidemiologic evidence for primary adult brain tumor risk factors using evidence-based medicine. *Prog Neurol Surg* 19: 54-79.

7. Wrensch M, Minn Y, Chew T, Bondy M, Berger MS (2002) Epidemiology of primary brain tumors: current concepts and review of the literature. *Neuro Oncol* 4: 278-299.
8. Bohnen NI, Kurland LT (1995) Brain tumor and exposure to pesticides in humans: a review of the epidemiologic data. *J Neurol Sci* 132: 110-121.
9. Fisher SG, Weber L, Carbone M (1999) Cancer risk associated with simian virus 40 contaminated polio vaccine. *Anticancer Res* 19: 2173-2180.
10. Huang H, Reis R, Yonekawa Y, Lopes JM, Kleihues P, et al. (1999) Identification in human brain tumors of DNA sequences specific for SV40 large T antigen. *Brain Pathol* 9: 33-42.
11. Strickler HD, Rosenberg PS, Devesa SS, Hertel J, Fraumeni JF Jr, et al. (1998) Contamination of poliovirus vaccines with simian virus 40 (1955-1963) and subsequent cancer rates. *JAMA* 279: 292-295.
12. Cobbs CS, Harkins L, Samanta M, Gillespie GY, Bharara S, et al. (2002) Human cytomegalovirus infection and expression in human malignant glioma. *Cancer Res* 62: 3347-3350.
13. Lau SK, Chen YY, Chen WG, Diamond DJ, Mamelak AN, et al. (2005) Lack of association of cytomegalovirus with human brain tumors. *Mod Pathol* 18: 838-843.
14. Mitchell DA, Xie W, Schmittling R, Learn C, Friedman A, et al (2008) Sensitive detection of human cytomegalovirus in tumors and peripheral blood of patients diagnosed with glioblastoma. *Neuro-Oncology* 10: 10-18.
15. Poltermann S, Schlehofer B, Steindorf K, Schnitzler P, Geletneký K, et al. (2006) Lack of association of herpesviruses with brain tumors. *J Neurovirol* 12: 90-99.
16. Saddawi-Konefka R, Crawford J (2010) Chronic Viral Infection and Primary Central Nervous System Malignancy. *J Neuroimmune Pharmacol* 5: 387-403.
17. Scheurer ME, Bondy ML, Aldape KD, Albrecht T, El-Zein R (2008) Detection of human cytomegalovirus in different histological types of gliomas. *Acta Neuropathol* 116: 79-86.
18. Bondy M, Wiencke J, Wrensch M, Kyritsis AP (1994) Genetics of primary brain tumors: a review. *J Neurooncol* 18: 69-81.
19. Crino PB, Nathanson KL, Henske EP (2006) The tuberous sclerosis complex. *N Engl J Med* 355: 1345-1356.
20. Kijima C, Miyashita T, Suzuki M, Oka H, Fujii K (2012) Two cases of nevoid basal cell carcinoma syndrome associated with meningioma caused by a PTCH1 or SUFU germline mutation. *Fam Cancer* 11: 565-570.
21. Hamilton SR, Liu B, Parsons RE, Papadopoulos N, Jen J, et al. (1995) The molecular basis of Turcot's syndrome. *N Engl J Med* 332: 839-847.
22. Kamihara J, Rana HQ, Garber JE (2014) Germline TP53 mutations and the changing landscape of Li-Fraumeni syndrome. *Hum Mutat* 35: 654-662.
23. Kyritsis AP, Bondy ML, Xiao M, Berman EL, Cunningham JE, et al. (1994) Germline p53 gene mutations in subsets of glioma patients. *J Natl Cancer Inst* 86: 344-349.
24. Maher ER, Neumann HP, Richard S (2011) von Hippel-Lindau disease: a clinical and scientific review. *Eur J Hum Genet* 19: 617-623.
25. Wrensch M, Lee M, Miike R, Newman B, Bargar G, et al (1997) Familial and Personal Medical History of Cancer and Nervous System Conditions among Adults with Glioma and Controls. *American Journal of Epidemiology* 145: 581-593.
26. Malmer B, Grönberg H, Bergenheim AT, Lenner P, Henriksson R (1999) Familial aggregation of astrocytoma in northern Sweden: an epidemiological cohort study. *Int J Cancer* 81: 366-370.
27. Hemminki K, Li X, Vaittinen P, Dong C (2000) Cancers in the first-degree relatives of children with brain tumours. *Br J Cancer* 83: 407-411.
28. Malmer B, Iselius L, Holmberg E, Collins A, Henriksson R, et al. (2001) Genetic epidemiology of glioma. *Br J Cancer* 84: 429-434.
29. Grossman SA, Osman M, Hruban R, Piantadosi S (1999) Central nervous system cancers in first-degree relatives and spouses. *Cancer Invest* 17: 299-308.
30. Chang SM, Parney IF, Huang W, Anderson FA Jr, Asher AL, et al. (2005) Patterns of care for adults with newly diagnosed malignant glioma. *JAMA* 293: 557-564.
31. Forsyth PA, Posner JB (1993) Headaches in patients with brain tumors: a study of 111 patients. *Neurology* 43: 1678-1683.
32. FRANKEL SA, GERMAN WJ (1958) Glioblastoma multiforme; review of 219 cases with regard to natural history, pathology, diagnostic methods, and treatment. *J Neurosurg* 15: 489-503.
33. ROTH JG, ELVIDGE AR (1960) Glioblastoma multiforme: a clinical survey. *J Neurosurg* 17: 736-750.
34. DeAngelis LM1 (2001) Brain tumors. *N Engl J Med* 344: 114-123.
35. Liigant A, Haldre S, Oun A, Linnamägi U, Saar A, et al. (2001) Seizure disorders in patients with brain tumors. *Eur Neurol* 45: 46-51.
36. Cha SI (2009) Neuroimaging in neuro-oncology. *Neurotherapeutics* 6: 465-477.
37. Brandsma D, Stalpers L, Taal W, Sminia P, van den Bent MJ (2008) Clinical features, mechanisms, and management of pseudoprogression in malignant gliomas. *Lancet Oncol* 9: 453-461.
38. Ito R, Mori S, Melhem ER (2002) Diffusion tensor brain imaging and tractography. *Neuroimaging Clin N Am* 12: 1-19.
39. Barone DG, Lawrie TA, Hart MG (2014) Image guided surgery for the resection of brain tumours. *Cochrane Database Syst Rev* 1: CD009685.
40. Rasmussen IA Jr, Lindseth F, Rygh OM, Berntsen EM, Selbekk T, et al. (2007) Functional neuronavigation combined with intra-operative 3D ultrasound: Initial experiences during surgical resections close to eloquent brain areas and future directions in automatic brain shift compensation of preoperative data. *Acta Neurochirurgica* 149: 365-378.
41. Unsgaard G, Rygh OM, Selbekk T, Müller TB, Kolstad F, et al. (2006) Intra-operative 3D ultrasound in neurosurgery. *Acta Neurochir (Wien)* 148: 235-253.
42. Stummer W, Novotny A, Stepp H, Goetz C, Bise K, et al. (2000) Fluorescence-guided resection of glioblastoma multiforme by using 5-aminolevulinic acid-induced porphyrins: a prospective study in 52 consecutive patients. *J Neurosurg* 93: 1003-1013.
43. Liu JT, Meza D, Sanai N (2014) Trends in fluorescence image-guided surgery for gliomas. *Neurosurgery* 75: 61-71.
44. Petrovsky A, Schellenberger E, Josephson L, Weissleder R, Bogdanov A Jr (2003) Near-infrared fluorescent imaging of tumor apoptosis. *Cancer Res* 63: 1936-1942.
45. Behbahani M, Martirosyan NL, Georges J, Udovich JA, Kalani MY, et al. (2013) Intraoperative fluorescent imaging of intracranial tumors: a review. *Clin Neurol Neurosurg* 115: 517-528.
46. Neumann H, Kiesslich R, Wallace MB, Neurath MF (2010) Confocal laser endomicroscopy: technical advances and clinical applications. *Gastroenterology* 139: 388-392, 392.
47. Sankar T, Delaney PM, Ryan RW, Eschbacher J, Abdelwahab M, et al. (2010) Miniaturized Handheld Confocal Microscopy for Neurosurgery: Results in an Experimental Glioblastoma Model. *Neurosurgery* 66: 410-418.
48. Stummer W, Pichlmeier U, Meinert T, Wiestler OD, Zanella F, et al. (2006) Fluorescence-guided surgery with 5-aminolevulinic acid for resection of malignant glioma: a randomised controlled multicentre phase III trial. *Lancet Oncol* 7: 392-401.
49. Ramnani N, Behrens TE, Penny W, Matthews PM (2004) New approaches for exploring anatomical and functional connectivity in the human brain. *Biol Psychiatry* 56: 613-619.
50. Rykhlevskaia E, Gratton G, Fabiani M (2008) Combining structural and functional neuroimaging data for studying brain connectivity: a review. *Psychophysiology* 45: 173-187.
51. Fandino J, Kollias SS, Wieser HG, Valavanis A, Yonekawa Y (1999) Intraoperative validation of functional magnetic resonance imaging and cortical reorganization patterns in patients with brain tumors involving the primary motor cortex. *Journal of Neurosurgery* 91: 238-250.
52. Spena G, Nava A, Cassini F, Pepoli A, Bruno M, D'Agata F, et al: Preoperative and intraoperative brain mapping for the resection of

- eloquent-area tumors. A prospective analysis of methodology, correlation, and usefulness based on clinical outcomes. *Acta Neurochirurgica* 152:1835-1846, 2010
53. Lehericy S, Duffau H, Cornu P, Capelle L, Pidoux B, Carpentier A, et al: Correspondence between functional magnetic resonance imaging somatotopy and individual brain anatomy of the central region: comparison with intraoperative stimulation in patients with brain tumors. *Journal of Neurosurgery* 92:589-598, 2000
54. Roux FE, Ibarrola D, Tremoulet M, Lazorthes Y, Henry P, et al. (2001) Methodological and technical issues for integrating functional magnetic resonance imaging data in a neuronavigational system. *Neurosurgery* 49: 1145-1156.
55. Kim SS, McCutcheon IE, Suki D, Weinberg JS, Sawaya R, et al. (2009) Awake Craniotomy for Brain Tumors Near Eloquent Cortex: Correlation of Intraoperative Cortical Mapping With Neurological Outcomes in 309 Consecutive Patients. *Neurosurgery* 64: 836-846.
56. Ohgaki H, Kleihues P (2005) Population-Based Studies on Incidence, Survival Rates, and Genetic Alterations in Astrocytic and Oligodendroglial Gliomas. *J Neuropathol Exp Neurol* 64: 479-489.
57. Chaichana KL, McGirt MJ, Latta J, Olivi A, Quiñones-Hinojosa A (2010) Recurrence and malignant degeneration after resection of adult hemispheric low-grade gliomas. *J Neurosurg* 112: 10-17.
58. Adamson C, Kanu OO, Mehta AI, Di C, Lin N, et al. (2009) Glioblastoma multiforme: a review of where we have been and where we are going. *Expert Opin Investig Drugs* 18: 1061-1083.
59. Ostrom Q, Cohen ML, Ondracek A, Sloan A, Barnholtz-Sloan J (2013) Gene markers in brain tumors: what the epileptologist should know. *Epilepsia* 54 Suppl 9: 25-29.
60. Esteller M, Garcia-Foncillas J, Andion E, Goodman SN, Hidalgo OF, et al. (2000) Inactivation of the DNA-Repair Gene MGMT and the Clinical Response of Gliomas to Alkylating Agents. *N Engl J Med* 343: 1350-1354.
61. Hegi ME, Diserens A-C, Godard S, Dietrich P-Y, Regli L et al. (2004) Clinical Trial Substantiates the Predictive Value of O-6-Methylguanine-DNA Methyltransferase Promoter Methylation in Glioblastoma Patients Treated with Temozolomide. *Clin Cancer Res* 10: 1871-1874.
62. Hegi ME, Diserens AC, Gorlia T, Hamou MF, de Tribolet N, et al. (2005) MGMT gene silencing and benefit from temozolomide in glioblastoma. *N Engl J Med* 352: 997-1003.
63. Mur P, Mollejo M, Ruano Y, de Lope AR, Fiaño C, et al. (2013) Codeletion of 1p and 19q determines distinct gene methylation and expression profiles in IDH-mutated oligodendroglial tumors. *Acta Neuropathol* 126: 277-289.
64. Cairncross G, Berkey B, Shaw E, Jenkins R, Scheithauer B, et al. (2006) Phase III Trial of Chemotherapy Plus Radiotherapy Compared With Radiotherapy Alone for Pure and Mixed Anaplastic Oligodendroglioma: Intergroup Radiation Therapy Oncology Group Trial 9402. *J Clin Oncol* 24: 2707-2714.
65. van den Bent M, Carpentier A, Brandes A, Sanson M, Taphoorn M, et al. (2006) Adjuvant Procarbazine, Lomustine, and Vincristine Improves Progression-Free Survival but Not Overall Survival in Newly Diagnosed Anaplastic Oligodendrogliomas and Oligoastrocytomas: A Randomized European Organisation for Research and Treatment of Cancer Phase III Trial. *J Clin Oncol* 24: 2715-2722.
66. Weller M, Stupp R, Hegi ME, van den Bent M, Tonn JC, et al. (2012) Personalized care in neuro-oncology coming of age: why we need MGMT and 1p/19q testing for malignant glioma patients in clinical practice. *Neuro Oncol* 14 Suppl 4: iv100-108.
67. Yan H, Parsons DW, Jin G, McLendon R, Rasheed BA, et al. (2009) IDH1 and IDH2 mutations in gliomas. *N Engl J Med* 360: 765-773.
68. Parsons DW, Jones S, Zhang X, Lin JC, Leary RJ, et al. (2008) An integrated genomic analysis of human glioblastoma multiforme. *Science* 321: 1807-1812.
69. Cohen AL, Holmen SL, Colman H (2013) IDH1 and IDH2 mutations in gliomas. *Curr Neurol Neurosci Rep* 13: 345.
70. Dang L, White DW, Gross S, Bennett BD, Bittinger MA, et al. (2009) Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. *Nature* 462: 739-744.
71. Noushmehr H, Weisenberger DJ, Diefes K, Phillips HS, Pujara K, et al. (2010) Identification of a CpG island methylator phenotype that defines a distinct subgroup of glioma. *Cancer Cell* 17: 510-522.
72. Turcan S, Rohle D, Goenka A, Walsh LA, Fang F, et al. (2012) IDH1 mutation is sufficient to establish the glioma hypermethylator phenotype. *Nature* 483: 479-483.
73. Del Vecchio CA, Giacomini CP, Vogel H, Jensen KC, Florio T, et al. (2013) EGFRvIII gene rearrangement is an early event in glioblastoma tumorigenesis and expression defines a hierarchy modulated by epigenetic mechanisms. *Oncogene* 32: 2670-2681.
74. Batra S, Castelino-Prabhu S, Wikstrand C, Zhu X, Humphrey P, et al. (1995) Epidermal growth factor ligand-independent, unregulated, cell-transforming potential of a naturally occurring human mutant EGFRvIII gene. *Cell Growth Differ* 6: 1251-1259.
75. Heimberger AB, Hlatky R, Suki D, Yang D, Weinberg J, et al. (2005) Prognostic Effect of Epidermal Growth Factor Receptor and EGFRvIII in Glioblastoma Multiforme Patients. *Clin Cancer Res* 11: 1462-1466.
76. Ekstrand AJ, James CD, Cavenee WK, Seliger B, Pettersson RF, et al. (1991) Genes for Epidermal Growth Factor Receptor, Transforming Growth Factor α , and Epidermal Growth Factor and Their Expression in Human Gliomas in Vivo. *Cancer Research* 51: 2164-2172.
77. Wikstrand CJ, McLendon RE, Friedman AH, Bigner DD (1997) Cell surface localization and density of the tumor-associated variant of the epidermal growth factor receptor, EGFRvIII. *Cancer Res* 57: 4130-4140.
78. Hegi ME, Rajakannu P, Weller M (2012) Epidermal growth factor receptor: a re-emerging target in glioblastoma. *Curr Opin Neurol* 25: 774-779.
79. Gan HK, Kaye AH, Luwor RB (2009) The EGFRvIII variant in glioblastoma multiforme. *J Clin Neurosci* 16: 748-754.
80. Sampson JH, Heimberger AB, Archer GE, Aldape KD, Friedman AH, et al. (2010) Immunologic Escape After Prolonged Progression-Free Survival With Epidermal Growth Factor Receptor Variant III Peptide Vaccination in Patients With Newly Diagnosed Glioblastoma. *J Clin Oncol* 28: 4722-4729.
81. Sampson JH, Aldape KD, Archer GE, Coan A, Desjardins A, et al. (2011) Greater chemotherapy-induced lymphopenia enhances tumor-specific immune responses that eliminate EGFRvIII-expressing tumor cells in patients with glioblastoma. *Neuro-Oncol* 13: 324-333.
82. Weller M, Kaulich K, Hentschel B, Felsberg J, Gramatzki D, et al. (2014) Assessment and prognostic significance of the epidermal growth factor receptor vIII mutation in glioblastoma patients treated with concurrent and adjuvant temozolomide radiochemotherapy. *Int J Cancer* 134: 2437-2447.
83. Schindler G, Capper D, Meyer J, Janzarik W, Omran H, et al. (2011) Analysis of BRAF V600E mutation in 1,320 nervous system tumors reveals high mutation frequencies in pleomorphic xanthoastrocytoma, ganglioglioma and extra-cerebellar pilocytic astrocytoma. *Acta Neuropathol* 121: 397-405.
84. Nicolaidis TP, Li H, Solomon DA, Hariono S, Hashizume R, et al. (2011) Targeted Therapy for BRAFV600E Malignant Astrocytoma. *Clin Cancer Res* 17: 7595-7604.
85. Whittaker S, Ménard D, Kirk R, Ogilvie L, Hedley D, et al. (2010) A novel, selective, and efficacious nanomolar pyridopyrazinone inhibitor of V600EBRAF. *Cancer Res* 70: 8036-8044.
86. Verhaak RG, Hoadley KA, Purdom E, Wang V, Qi Y, et al. (2010) Integrated genomic analysis identifies clinically relevant subtypes of glioblastoma characterized by abnormalities in PDGFRA, IDH, EGFR, and NF1. *Cancer Cell* 17: 98-110.
87. Noble M, Pröschel C, Mayer-Pröschel M (2004) Getting a GR(i)P on oligodendrocyte development. *Dev Biol* 265: 33-52.
88. Phillips HS, Kharbanda S, Chen R, Forrest WF, Soriano RH, Wu TD, et al: Molecular subclasses of high-grade glioma predict prognosis, delineate a

- pattern of disease progression, and resemble stages in neurogenesis. *Cancer Cell* 9:157-173, 2006
89. Ye JM, Ye MJ2, Kranz S3, Lo P4 (2014) A 10 year retrospective study of surgical outcomes of adult intracranial pilocytic astrocytoma. *J Clin Neurosci* 21: 2160-2164.
90. Due-Tønnessen BJ, Helseth E, Scheie D, Skullerud K, Aamodt G, et al. (2002) Long-Term Outcome after Resection of Benign Cerebellar Astrocytomas in Children and Young Adults (0–19 Years): Report of 110 Consecutive Cases. *Pediatr Neurosurg* 37: 71-80.
91. Viano JC, Herrera EJ, Suárez JC (2001) Cerebellar astrocytomas: a 24-year experience. *Childs Nerv Syst* 17: 607-610.
92. Dirven CMF, Mooij JJA, Molenaar WM (1997) Cerebellar pilocytic astrocytoma: a treatment protocol based upon analysis of 73 cases and a review of the literature. *Childs Nerv Syst* 13: 17-23.
93. Hadjipanayis CG, Kondziolka D, Gardner P, Niranjana A, Dagam S, et al. (2002) Stereotactic radiosurgery for pilocytic astrocytomas when multimodal therapy is necessary. *J Neurosurg* 97: 56-64.
94. Ater JL, Zhou T, Holmes E, Mazewski CM, Booth TN, et al. (2012) Randomized Study of Two Chemotherapy Regimens for Treatment of Low-Grade Glioma in Young Children: A Report From the Children's Oncology Group. *J Clin Oncol* 30: 2641-2647.
95. Kramm CM, Wagner S, Van Gool S, Schmid H, Strater R, et al. (2006) Improved Survival after Gross Total Resection of Malignant Gliomas in Pediatric Patients from the HIT-GBM Studies. *Anticancer Research* 26: 3773-3779.
96. Hentschel SJ, Sawaya R (2003) Optimizing outcomes with maximal surgical resection of malignant gliomas. *Cancer Control* 10: 109-114.
97. Walker MD, Alexander E Jr, Hunt WE, MacCarty CS, Mahaley MS Jr, et al. (1978) Evaluation of BCNU and/or radiotherapy in the treatment of anaplastic gliomas. A cooperative clinical trial. *J Neurosurg* 49: 333-343.
98. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, et al. (2005) Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* 352: 987-996.
99. Butowski NA, Sneed PK, Chang SM (2006) Diagnosis and treatment of recurrent high-grade astrocytoma. *J Clin Oncol* 24: 1273-1280.
100. Tsao M, Mehta M, Whelan T, Morris D, Hayman J, et al. (2005) The American Society for Therapeutic Radiology and Oncology (ASTRO) evidence-based review of the role of radiosurgery for malignant glioma. *International journal of radiation oncology, biology, physics* 63: 47-55.
101. Combs SE, Thilmann C, Edler L, Debus J, Schulz-Ertner D (2005) Efficacy of Fractionated Stereotactic Irradiation in Recurrent Gliomas: Long-Term Results in 172 Patients Treated in a Single Institution. *J Clin Oncol* 23: 8863-8869.
102. Brem H, Piantadosi S, Burger PC, Walker M, Selker R, et al. (1995) Placebo-controlled trial of safety and efficacy of intraoperative controlled delivery by biodegradable polymers of chemotherapy for recurrent gliomas. *The Lancet* 345: 1008-1012.
103. Westphal M, Hilt DC, Bortey E, Delavault P, Olivares R, et al. (2003) A phase 3 trial of local chemotherapy with biodegradable carmustine (BCNU) wafers (Gliadel wafers) in patients with primary malignant glioma. *Neuro Oncol* 5: 79-88.
104. Minniti G, Muni R, Lanzetta G, Marchetti P, Enrici RM (2009) Chemotherapy for glioblastoma: current treatment and future perspectives for cytotoxic and targeted agents. *Anticancer Res* 29: 5171-5184.
105. Gilbert MR, Dignam JJ, Armstrong TS, Wefel JS, Blumenthal DT, et al. (2014) A randomized trial of bevacizumab for newly diagnosed glioblastoma. *N Engl J Med* 370: 699-708.
106. Chinot OL, Wick W, Mason W, Henriksson R, Saran F, et al. (2014) Bevacizumab plus radiotherapy-temozolomide for newly diagnosed glioblastoma. *N Engl J Med* 370: 709-722.
107. Lamborn KR, Chang SM, Prados MD (2004) Prognostic factors for survival of patients with glioblastoma: recursive partitioning analysis. *Neuro Oncol* 6: 227-235.
108. Mørk SJ, Lindegaard KF, Halvorsen TB, Lehmann EH, Solgaard T, et al. (1985) Oligodendroglioma: incidence and biological behavior in a defined population. *J Neurosurg* 63: 881-889.
109. Coons SW, Johnson PC, Scheithauer BW, Yates AJ, Pearl DK (1997) Improving diagnostic accuracy and interobserver concordance in the classification and grading of primary gliomas. *Cancer* 79: 1381-1393.
110. Daumas-Duport C, Varlet P, Tucker ML, Beuvon F, Cervera P, et al. (1997) Oligodendrogliomas. Part I: Patterns of growth, histological diagnosis, clinical and imaging correlations: a study of 153 cases. *J Neurooncol* 34: 37-59.
111. Chin HW, Hazel JJ, Kim TH, Webster JH (1980) Oligodendrogliomas. I. A clinical study of cerebral oligodendrogliomas. *Cancer* 45: 1458-1466.
112. Roberts M, German WJ (1966) A long term study of patients with oligodendrogliomas. Follow-up of 50 cases, including Dr. Harvey Cushing's series. *J Neurosurg* 24: 697-700.
113. Rutka JT, Murakami M, Dirks PB, Hubbard SL, Becker LE, et al. (1997) Role of glial filaments in cells and tumors of glial origin: a review. *J Neurosurg* 87: 420-430.
114. Fortin D, Cairncross GJ, Hammond RR (1999) Oligodendroglioma: an appraisal of recent data pertaining to diagnosis and treatment. *Neurosurgery* 45: 1279-1291.
115. Daumas-Duport C, Tucker ML, Kolles H, Cervera P, Beuvon F, et al. (1997) Oligodendrogliomas. Part II: A new grading system based on morphological and imaging criteria. *J Neurooncol* 34: 61-78.
116. Paleologos NA, Cairncross JG (1999) Treatment of oligodendroglioma: an update. *Neuro Oncol* 1: 61-68.
117. Wallner KE, Gonzales M, Sheline GE (1988) Treatment of oligodendrogliomas with or without postoperative irradiation. *J Neurosurg* 68: 684-688.
118. Reedy DP, Bay JW, Hahn JF (1983) Role of radiation therapy in the treatment of cerebral oligodendroglioma: an analysis of 57 cases and a literature review. *Neurosurgery* 13: 499-503.
119. Cairncross G, Macdonald D, Ludwin S, Lee D, Cascino T, et al. (1994) Chemotherapy for anaplastic oligodendroglioma. National Cancer Institute of Canada Clinical Trials Group. *J Clin Oncol* 12: 2013-2021.
120. Smith J, Perry A, Borell T, Lee H, O'Fallon J, et al. (2000) Alterations of Chromosome Arms 1p and 19q as Predictors of Survival in Oligodendrogliomas, Astrocytomas, and Mixed Oligoastrocytomas. *J Clin Oncol* 18: 636.
121. Dohrmann GJ, Farwell JR, Flannery JT (1976) Ependymomas and ependymoblastomas in children. *J Neurosurg* 45: 273-283.
122. Mørk SJ, Loken AC (1977) Ependymoma: a follow-up study of 101 cases. *Cancer* 40: 907-915.
123. Mørk SJ, Rubinstein LJ (1985) Ependymoblastoma. A reappraisal of a rare embryonal tumor. *Cancer* 55: 1536-1542.
124. Souweidane MM, Bouffet E, Finlay J (1998) The role of chemotherapy in newly diagnosed ependymoma of childhood. *Pediatr Neurosurg* 28: 273-278.
125. Shaw EG, Scheithauer BW, O'Fallon JR, Davis DH (1994) Mixed oligoastrocytomas: a survival and prognostic factor analysis. *Neurosurgery* 34: 577-582.
126. Meyers SP, Wildenhain SL, Chang JK, Bourekas EC, Beattie PF, et al. (2000) Postoperative Evaluation for Disseminated Medulloblastoma Involving the Spine: Contrast-enhanced MR Findings, CSF Cytologic Analysis, Timing of Disease Occurrence, and Patient Outcomes. *AJNR Am J Neuroradiol* 21: 1757-1765.
127. Gerber NU, Mynarek M2, von Hoff K3, Friedrich C4, Resch A5, et al. (2014) Recent developments and current concepts in medulloblastoma. *Cancer Treat Rev* 40: 356-365.
128. Park TS, Hoffman HJ, Hendrick EB, Humphreys RP, Becker LE (1983) Medulloblastoma: clinical presentation and management. Experience at the hospital for sick children, Toronto, 1950-1980. *J Neurosurg* 58: 543-552.

129. Louis DN, Ohgaki H, Wiestler OD, Cavenee WK, Burger PC, et al. (2007) The 2007 WHO classification of tumours of the central nervous system. *Acta Neuropathol* 114: 97-109.
130. Farnndon PA, Del Mastro RG, Evans DG, Kilpatrick MW (1992) Location of gene for Gorlin syndrome. *Lancet* 339: 581-582.
131. Wicking C, Shanley S, Smyth I, Gillies S, Negus K, et al. (1997) Most germ-line mutations in the nevoid basal cell carcinoma syndrome lead to a premature termination of the PATCHED protein, and no genotype-phenotype correlations are evident. *Am J Hum Genet* 60: 21-26.
132. Pugh TJ, Weeraratne SD, Archer TC, Pomeranz Krummel DA, Auclair D, et al. (2012) Medulloblastoma exome sequencing uncovers subtype-specific somatic mutations. *Nature* 488: 106-110.
133. Northcott PA, Jones DT, Kool M, Robinson GW, Gilbertson RJ, et al. (2012) Medulloblastomas: the end of the beginning. *Nat Rev Cancer* 12: 818-834.
134. Robinson G, Parker M, Kranenburg TA, Lu C, Chen X, et al. (2012) Novel mutations target distinct subgroups of medulloblastoma. *Nature* 488: 43-48.
135. Jones DT, Jäger N, Kool M, Zichner T, Hutter B, et al. (2012) Dissecting the genomic complexity underlying medulloblastoma. *Nature* 488: 100-105.
136. Kool M, Korshunov A, Remke M, Jones DT, Schlanstein M, et al. (2012) Molecular subgroups of medulloblastoma: an international meta-analysis of transcriptome, genetic aberrations, and clinical data of WNT, SHH, Group 3, and Group 4 medulloblastomas. *Acta Neuropathol* 123: 473-484.
137. Li KK, Lau KM, Ng HK (2013) Signaling pathway and molecular subgroups of medulloblastoma. *Int J Clin Exp Pathol* 6: 1211-1222.
138. Taylor MD, Liu L, Raffel C, Hui CC, Mainprize TG, et al. (2002) Mutations in SUFU predispose to medulloblastoma. *Nat Genet* 31: 306-310.
139. Reifenberger J, Wolter M, Weber RG, Megahed M, Ruzicka T, et al. (1998) Missense Mutations in SMOH in Sporadic Basal Cell Carcinomas of the Skin and Primitive Neuroectodermal Tumors of the Central Nervous System. *Cancer Research* 58: 1798-1803.
140. Kool M, Koster J, Bunt J, Hasselt NE, Lakeman A, et al. (2008) Integrated genomics identifies five medulloblastoma subtypes with distinct genetic profiles, pathway signatures and clinicopathological features. *PLoS One* 3: e3088.
141. Northcott PA, Korshunov A, Witt H, Hielscher T, Eberhart CG, et al. (2011) Medulloblastoma comprises four distinct molecular variants. *J Clin Oncol* 29: 1408-1414.
142. Al-Halabi H, Nantel A, Klekner A, Guiot MC, Albrecht S, et al. (2011) Preponderance of sonic hedgehog pathway activation characterizes adult medulloblastoma. *Acta Neuropathol* 121: 229-239.
143. Remke M, Hielscher T, Northcott PA, Witt H, Ryzhova M, et al. (2011) Adult medulloblastoma comprises three major molecular variants. *J Clin Oncol* 29: 2717-2723.
144. Baeza N, Masuoka J, Kleihues P, Ohgaki H (2003) AXIN1 mutations but not deletions in cerebellar medulloblastomas. *Oncogene* 22: 632-636.
145. Eberhart CG, Tihan T, Burger PC (2000) Nuclear localization and mutation of beta-catenin in medulloblastomas. *J Neuropathol Exp Neurol* 59: 333-337.
146. Huang H, Mahler-Araujo BM, Sankila A, Chimelli L, Yonekawa Y, et al. (2000) APC mutations in sporadic medulloblastomas. *Am J Pathol* 156: 433-437.
147. Taylor M, Northcott P, Korshunov A, Remke M, Cho YJ, et al. (2012) Molecular subgroups of medulloblastoma: the current consensus. *Acta Neuropathol* 123: 465-472.
148. Northcott PA, Korshunov A, Pfister SM, Taylor MD (2012) The clinical implications of medulloblastoma subgroups. *Nat Rev Neurol* 8: 340-351.
149. Packer RJ, Goldwein J, Nicholson HS, Vezina LG, Allen JC, et al. (1999) Treatment of children with medulloblastomas with reduced-dose craniospinal radiation therapy and adjuvant chemotherapy: A Children's Cancer Group Study. *J Clin Oncol* 17: 2127-2136.
150. Packer RJ, Finlay JL (1996) Chemotherapy for Childhood Medulloblastoma and Primitive Neuroectodermal Tumors. *Oncologist* 1: 381-393.
151. Culley DJ, Berger MS, Shaw D, Geyer R (1994) An Analysis of Factors Determining the Need for Ventriculoperitoneal Shunts after Posterior Fossa Tumor Surgery in Children. *Neurosurgery* 34: 402-408.
152. Gopalakrishnan CV, Dhakoji A, Menon G, Nair S (2012) Factors predicting the need for cerebrospinal fluid diversion following posterior fossa tumor surgery in children. *Pediatr Neurosurg* 48: 93-101.
153. David KM, Casey AT, Hayward RD, Harkness WF, Phipps K, et al. (1997) Medulloblastoma: is the 5-year survival rate improving? A review of 80 cases from a single institution. *J Neurosurg* 86: 13-21.
154. Evans AE, Jenkin RD, Sposto R, Ortega JA, Wilson CB, et al. (1990) The treatment of medulloblastoma. Results of a prospective randomized trial of radiation therapy with and without CCNU, vincristine, and prednisone. *J Neurosurg* 72: 572-582.
155. Weil MD, Lamborn K, Edwards MS, Wara WM (1998) Influence of a child's sex on medulloblastoma outcome. *JAMA* 279: 1474-1476.
156. Whittle IR, Smith C, Navoo P, Collie D (2004) Meningiomas. *Lancet* 363: 1535-1543.
157. Solero CL, Fornari M, Giombini S, Lasio G, Oliveri G, et al. (1989) Spinal meningiomas: review of 174 operated cases. *Neurosurgery* 25: 153-160.
158. Som PM, Sacher M, Strenger SW, Biller HF, Malis LI (1987) "Benign" metastasizing meningiomas. *AJNR Am J Neuroradiol* 8: 127-130.
159. Karasick JL, Mullan SF (1974) A survey of metastatic meningiomas. *J Neurosurg* 40: 206-212.
160. Sheikh BY, Siqueira E, Dayel F (1996) Meningioma in children: a report of nine cases and a review of the literature. *Surg Neurol* 45: 328-335.
161. Kotecha RS, Pascoe EM, Rushing EJ, Rorke-Adams LB, Zwerdling T, et al. (2011) Meningiomas in children and adolescents: a meta-analysis of individual patient data. *Lancet Oncol* 12: 1229-1239.
162. Goutagny S, Kalamarides M (2010) Meningiomas and neurofibromatosis. *J Neurooncol* 99: 341-347.
163. Mautner VF, Lindenau M, Baser ME, Hazim W, Tatagiba M, et al. (1996) The neuroimaging and clinical spectrum of neurofibromatosis 2. *Neurosurgery* 38: 880-885.
164. Kolles H, Niedermayer I, Schmitt C, Henn W, Feld R, et al. (1995) Triple approach for diagnosis and grading of meningiomas: histology, morphometry of Ki-67/Feulgen stainings, and cytogenetics. *Acta Neurochir (Wien)* 137: 174-181.
165. Perry A, Scheithauer BW, Stafford SL, Lohse CM, Wollan PC (1999) "Malignancy" in meningiomas: a clinicopathologic study of 116 patients, with grading implications. *Cancer* 85: 2046-2056.
166. Backer-Grøndahl T, Moen BH, Torp SH (2012) The histopathological spectrum of human meningiomas. *Int J Clin Exp Pathol* 5: 231-242.
167. SIMPSON D (1957) The recurrence of intracranial meningiomas after surgical treatment. *J Neurol Neurosurg Psychiatry* 20: 22-39.
168. Oya S, Kawai K, Nakatomi H, Saito N (2012) Significance of Simpson grading system in modern meningioma surgery: integration of the grade with MIB-1 labeling index as a key to predict the recurrence of WHO Grade I meningiomas. *J Neurosurg* 117: 121-128.
169. Rogers L, Barani I, Chamberlain M, Kaley TJ, McDermott M, et al. (2015) Meningiomas: knowledge base, treatment outcomes, and uncertainties. A RANO review. *J Neurosurg* 122: 4-23.
170. Dziuk TW, Woo S, Butler EB, Thornby J, Grossman R, et al. (1998) Malignant meningioma: an indication for initial aggressive surgery and adjuvant radiotherapy. *J Neurooncol* 37: 177-188.
171. Modha A, Gutin PH (2005) Diagnosis and treatment of atypical and anaplastic meningiomas: a review. *Neurosurgery* 57: 538-550.
172. Kondziolka D, Mathieu D, Lunsford LD, Martin JJ, Madhok R, et al. (2008) Radiosurgery as definitive management of intracranial meningiomas. *Neurosurgery* 62: 53-58.
173. Ware ML, Larson DA, Sneed PK, Wara WW, McDermott MW (2004) Surgical resection and permanent brachytherapy for recurrent atypical and malignant meningioma. *Neurosurgery* 54: 55-63.

174. Walcott BP, Nahed BV, Brastianos PK, Loeffler JS (2013) Radiation Treatment for WHO Grade II and III Meningiomas. *Front Oncol* 3: 227.
175. Chamberlain MC (2012) The role of chemotherapy and targeted therapy in the treatment of intracranial meningioma. *Curr Opin Oncol* 24: 666-671.
176. Kaley TJ, Wen P, Schiff D, Ligon K, Haidar S, et al. (2015) Phase II trial of sunitinib for recurrent and progressive atypical and anaplastic meningioma. *Neuro Oncol* 17: 116-121.
177. Mahaley MS Jr, Mettlin C, Natarajan N, Laws ER Jr, Peace BB (1989) National survey of patterns of care for brain-tumor patients. *J Neurosurg* 71: 826-836.
178. Yamashita J, Handa H, Iwaki K, Abe M (1980) Recurrence of intracranial meningiomas, with special reference to radiotherapy. *Surg Neurol* 14: 33-40.
179. Ko KW, Nam DH, Kong DS, Lee JI, Park K, et al. (2007) Relationship between malignant subtypes of meningioma and clinical outcome. *J Clin Neurosci* 14: 747-753.
180. Palma L, Celli P, Franco C, Cervoni L, Cantore G (1997) Long-term prognosis for atypical and malignant meningiomas: a study of 71 surgical cases. *J Neurosurg* 86: 793-800.
181. Al-Mefty O, Kadri PA, Pravdenkova S, Sawyer JR, Stangeby C, et al. (2004) Malignant progression in meningioma: documentation of a series and analysis of cytogenetic findings. *J Neurosurg* 101: 210-218.
182. Ragel BT, Couldwell WT (2004) Pituitary carcinoma: a review of the literature. *Neurosurg Focus* 16: E7.
183. Harris RD, Sung JH, Seljeskog EL (1979) Transnasal excision of a neurohypophyseal tumor. *Surg Neurol* 11: 53-56.
184. <http://www.aans.org>
185. Beckers A, Daly AF (2007) The clinical, pathological, and genetic features of familial isolated pituitary adenomas. *Eur J Endocrinol* 157: 371-382.
186. Nagesser SK, van Seters AP, Kievit J, Hermans J, Krans HM, et al. (2000) Long-term results of total adrenalectomy for Cushing's disease. *World J Surg* 24: 108-113.
187. Barber TM, Adams E, Ansong O, Byrne JV, Karavitaki N, et al. (2010) Nelson's syndrome. *Eur J Endocrinol* 163: 495-507.
188. Chanson P, Salenave S (2008) Acromegaly. *Orphanet J Rare Dis* 3: 17.
189. Chaudhary V, Bano S (2011) Imaging of the pituitary: Recent advances. *Indian J Endocrinol Metab* 15 Suppl 3: S216-223.
190. Sanno N, Teramoto A, Osamura RY (2000) Long-term surgical outcome in 16 patients with thyrotropin pituitary adenoma. *J Neurosurg* 93: 194-200.
191. Melmed S (2006) Medical progress: Acromegaly. *N Engl J Med* 355: 2558-2573.
192. O'Halloran DJ, Shalet SM (1996) Radiotherapy for pituitary adenomas: an endocrinologist's perspective. *Clin Oncol (R Coll Radiol)* 8: 79-84.
193. Eastman RC, Gorden P, Roth J (1979) Conventional supervoltage irradiation is an effective treatment for acromegaly. *J Clin Endocrinol Metab* 48: 931-940.
194. Roth J, Gorden P, Brace K (1970) Efficacy of conventional pituitary irradiation in acromegaly. *N Engl J Med* 282: 1385-1391.
195. Loeffler JS, Shih HA (2011) Radiation therapy in the management of pituitary adenomas. *J Clin Endocrinol Metab* 96: 1992-2003.
196. Yang I, Kim W, De Salles A, Bergsneider M (2010) A systematic analysis of disease control in acromegaly treated with radiosurgery. *Neurosurg Focus* 29: E13.
197. Sheehan JP, Xu Z, Salvetti DJ, Schmitt PJ, Vance ML (2013) Results of gamma knife surgery for Cushing's disease. *J Neurosurg* 119: 1486-1492.
198. Ciric I, Mikhael M, Stafford T, Lawson L, Garces R (1983) Transphenoidal microsurgery of pituitary macroadenomas with long-term follow-up results. *J Neurosurg* 59: 395-401.
199. Ferreri AJ, Marturano E (2012) Primary CNS lymphoma. *Best Pract Res Clin Haematol* 25: 119-130.
200. Hochberg FH, Miller G, Schooley RT, Hirsch MS, Feorino P, et al. (1983) Central-nervous-system lymphoma related to Epstein-Barr virus. *N Engl J Med* 309: 745-748.
201. Guterman K, Hair L, Morgello S (1996) Epstein-Barr virus and AIDS-related primary central nervous system lymphoma. Viral detection by immunohistochemistry, RNA in situ hybridization, and polymerase chain reaction. *Clin Neuropathol* 15: 79-86.
202. Poon T, Matoso I, Tchertkoff V, Weitzner I, Gade M (1989) CT features of primary cerebral lymphoma in AIDS and non-AIDS patients. *J Comput Assist Tomogr* 13: 6-9.
203. Ferreri AJ (2011) How I treat primary CNS lymphoma. *Blood* 118: 510-522.
204. DeAngelis LM, Yahalom J, Heinemann MH, Cirincione C, Thaler HT, et al. (1990) Primary CNS lymphoma: combined treatment with chemotherapy and radiotherapy. *Neurology* 40: 80-86.
205. DeAngelis LM, Yahalom J, Thaler HT, Kher U (1992) Combined modality therapy for primary CNS lymphoma. *J Clin Oncol* 10: 635-643.
206. Goldstein JD, Dickson DW, Moser FG, Hirschfeld AD, Freeman K, et al. (1991) Primary central nervous system lymphoma in acquired immune deficiency syndrome. A clinical and pathologic study with results of treatment with radiation. *Cancer* 67: 2756-2765.
207. Formenti SC, Gill PS, Lean E, Rarick M, Meyer PR, et al. (1989) Primary central nervous system lymphoma in AIDS. Results of radiation therapy. *Cancer* 63: 1101-1107.
208. Fong B, Barkhoudarian G, Pezeshkian P, Parsa AT, Gopen Q, et al. (2011) The molecular biology and novel treatments of vestibular schwannomas. *J Neurosurg* 115: 906-914.
209. Wolff RK, Frazer KA, Jackler RK, Lanser MJ, Pitts LH, et al. (1992) Analysis of chromosome 22 deletions in neurofibromatosis type 2-related tumors. *Am J Hum Genet* 51: 478-485.
210. Rouleau GA, Wertelecki W, Haines JL, Hobbs WJ, Trofatter JA, et al. (1987) Genetic linkage of bilateral acoustic neurofibromatosis to a DNA marker on chromosome 22. *Nature* 329: 246-248.
211. Asthagiri AR, Parry DM, Butman JA, Kim HJ, Tsilou ET, et al. (2009) Neurofibromatosis type 2. *Lancet* 373: 1974-1986.
212. Harner SG, Laws ER Jr (1983) Clinical findings in patients with acoustic neurinoma. *Mayo Clin Proc* 58: 721-728.
213. Wippold FJ, Lubner M, Perrin RJ, Lammle M, Perry A (2007) *Neuropathology for the Neuroradiologist: Antoni A and Antoni B Tissue Patterns*. *AJNR* 28: 1633-1638.
214. Ramzy I (1977) Benign schwannoma: demonstration of Verocay bodies using fine needle aspiration. *Acta Cytol* 21: 316-319.
215. Pollock BE, Lunsford LD, Flickinger JC, Clyde BL, Kondziolka D (1998) Vestibular schwannoma management. Part I. Failed microsurgery and the role of delayed stereotactic radiosurgery. *J Neurosurg* 89: 944-948.
216. Silk PS, Lane JI, Driscoll CL (2009) Surgical approaches to vestibular schwannomas: what the radiologist needs to know. *Radiographics* 29: 1955-1970.
217. Karnes PS (1998) Neurofibromatosis: a common neurocutaneous disorder. *Mayo Clin Proc* 73: 1071-1076.
218. Singhal S, Birch JM, Kerr B, Lashford L, Evans DG (2002) Neurofibromatosis type 1 and sporadic optic gliomas. *Arch Dis Child* 87: 65-70.
219. [No authors listed] (1988) Neurofibromatosis. Conference statement. National Institutes of Health Consensus Development Conference. *Arch Neurol* 45: 575-578.
220. Evans DG (2009) Neurofibromatosis type 2 (NF2): a clinical and molecular review. *Orphanet J Rare Dis* 4: 16.
221. Parry DM, Eldridge R, Kaiser-Kupfer MI, Bouzas EA, Pikus A, et al. (1994) Neurofibromatosis 2 (NF2): clinical characteristics of 63 affected individuals and clinical evidence for heterogeneity. *Am J Med Genet* 52: 450-461.
222. Packer RJ, Gutmann DH, Rubenstein A, Viskochil D, Zimmerman RA, et al. (2002) Plexiform neurofibromas in NF1: toward biologic-based therapy. *Neurology* 58: 1461-1470.

223. Fine SW, McClain SA, Li M (2004) Immunohistochemical staining for calretinin is useful for differentiating schwannomas from neurofibromas. *Am J Clin Pathol* 122: 552-559.
224. Park JY, Park H, Park NJ, Park JS, Sung HJ, et al. (2011) Use of Calretinin, CD56, and CD34 for Differential Diagnosis of Schwannoma and Neurofibroma. *Korean J Pathol* 45: 30-35.
225. Owonikoko TK, Arbiser J2, Zelnak A, Shu HK3, Shim H3, et al. (2014) Current approaches to the treatment of metastatic brain tumours. *Nat Rev Clin Oncol* 11: 203-222.
226. Fox BD, Cheung VJ, Patel AJ, Suki D, Rao G (2011) Epidemiology of metastatic brain tumors. *Neurosurg Clin N Am* 22: 1-6, v.
227. Gavrilovic IT, Posner JB (2005) Brain metastases: epidemiology and pathophysiology. *J Neurooncol* 75: 5-14.
228. Norden AD, Wen PY, Kesari S (2005) Brain metastases. *Curr Opin Neurol* 18: 654-661.
229. Nayak L, Lee EQ, Wen PY (2012) Epidemiology of brain metastases. *Curr Oncol Rep* 14: 48-54.
230. Pectasides D, Aravantinos G, Fountzilias G, Kalofonos C, Efstathiou E, et al. (2005) Brain Metastases from Epithelial Ovarian Cancer. The Hellenic Cooperative Oncology Group (HeCOG) Experience and Review of the Literature. *Anticancer Res* 25: 3553-3558.
231. Amer MH, Al-Sarraf M, Baker LH, Vaitkevicius VK (1978) Malignant melanoma and central nervous system metastases. Incidence, diagnosis, treatment and survival. *Cancer* 42: 660-668.
232. Amer MH, Al-Sarraf M, Vaitkevicius VK (1979) Clinical presentation, natural history and prognostic factors in advanced malignant melanoma. *Surg Gynecol Obstet* 149: 687-692.
233. Atkinson L (1978) Melanoma of the central nervous system. *Aust N Z J Surg* 48: 14-16.
234. McNeer G, Das Gupta T (1965) Problem of recurrence in the management of melanoma. *CA Cancer J Clin* 15: 270-274.
235. Postmus PE, Smit EF (1999) Chemotherapy for brain metastases of lung cancer: a review. *Ann Oncol* 10: 753-759.
236. Burt M, Wronski M, Arbit E, Galicich JH (1992) Resection of brain metastases from non-small-cell lung carcinoma. Results of therapy. Memorial Sloan-Kettering Cancer Center Thoracic Surgical Staff. *J Thorac Cardiovasc Surg* 103: 399-410.
237. Nugent JL, Bunn PA Jr, Matthews MJ, Ihde DC, Cohen MH, et al. (1979) CNS metastases in small cell bronchogenic carcinoma: increasing frequency and changing pattern with lengthening survival. *Cancer* 44: 1885-1893.
238. Graus F, Walker RW, Allen JC (1983) Brain metastases in children. *J Pediatr* 103: 558-561.
239. Walker AE, Robins M, Weinfeld FD (1985) Epidemiology of brain tumors: the national survey of intracranial neoplasms. *Neurology* 35: 219-226.
240. Garg RK, Sinha MK (2010) Multiple ring-enhancing lesions of the brain. *J Postgrad Med* 56: 307-316.
241. Joosse A, de Vries E, Eckel R, Nijsten T, Eggermont AM, et al. (2011) Gender differences in melanoma survival: female patients have a decreased risk of metastasis. *J Invest Dermatol* 131: 719-726.
242. Potts DG, Abbott GF, von Sneidern JV (1980) National Cancer Institute study: evaluation of computed tomography in the diagnosis of intracranial neoplasms. III. Metastatic tumors. *Radiology* 136: 657-664.
243. Delattre JY, Krol G, Thaler HT, Posner JB (1988) Distribution of brain metastases. *Arch Neurol* 45: 741-744.
244. Hwang TL, Close TP, Grego JM, Brannon WL, Gonzales F (1996) Predilection of brain metastasis in gray and white matter junction and vascular border zones. *Cancer* 77: 1551-1555.
245. Douek P, Turner R, Pekar J, Patronas N, Le Bihan D (1991) MR color mapping of myelin fiber orientation. *J Comput Assist Tomogr* 15: 923-929.
246. Lu S, Ahn D, Johnson G, Cha S (2003) Peritumoral diffusion tensor imaging of high-grade gliomas and metastatic brain tumors. *AJNR Am J Neuroradiol* 24: 937-941.
247. Hakyemez B, Erdogan C, Bolca N, Yildirim N, Gokalp G, et al. (2006) Evaluation of different cerebral mass lesions by perfusion-weighted MR imaging. *J Magn Reson Imaging* 24: 817-824.
248. Nadal Desbarats L, Herlidou S, de Marco G, Gondry-Jouet C, Le Gars D, et al. (2003) Differential MRI diagnosis between brain abscesses and necrotic or cystic brain tumors using the apparent diffusion coefficient and normalized diffusion-weighted images. *Magn Reson Imaging* 21: 645-650.
249. Eichler AF, Loeffler JS (2007) Multidisciplinary management of brain metastases. *Oncologist* 12: 884-898.
250. Hazard LJ, Jensen RL, Shrieve DC (2005) Role of stereotactic radiosurgery in the treatment of brain metastases. *Am J Clin Oncol* 28: 403-410.
251. Suki D, Hatiboglu MA, Patel AJ, Weinberg JS, Groves MD, et al. (2009) Comparative risk of leptomeningeal dissemination of cancer after surgery or stereotactic radiosurgery for a single supratentorial solid tumor metastasis. *Neurosurgery* 64: 664-676.
252. Patel AJ, Suki D, Hatiboglu MA, Abouassi H, Shi W, et al. (2010) Factors influencing the risk of local recurrence after resection of a single brain metastasis. *J Neurosurg* 113: 181-189.
253. Vecht CJ, Haaxma-Reiche H, Noordijk EM, Padberg GW, Voormolen JH, et al. (1993) Treatment of single brain metastasis: radiotherapy alone or combined with neurosurgery? *Ann Neurol* 33: 583-590.
254. Patchell RA, Tibbs PA, Walsh JW, Dempsey RJ, Maruyama Y, et al. (1990) A randomized trial of surgery in the treatment of single metastases to the brain. *N Engl J Med* 322: 494-500.
255. Andrews DW, Scott CB, Sperduto PW, Flanders AE, Gaspar LE, et al. (2004) Whole brain radiation therapy with or without stereotactic radiosurgery boost for patients with one to three brain metastases: phase III results of the RTOG 9508 randomised trial. *Lancet* 363: 1665-1672.
256. Kocher M, Soffiotti R, Abacioglu U, Villà S, Fauchon F, et al. (2011) Adjuvant whole-brain radiotherapy versus observation after radiosurgery or surgical resection of one to three cerebral metastases: results of the EORTC 22952-26001 study. *J Clin Oncol* 29: 134-141.
257. Wong ET, Berkenblit A (2004) The role of topotecan in the treatment of brain metastases. *Oncologist* 9: 68-79.
258. Agarwala SS, Kirkwood JM, Gore M, Dreno B, Thatcher N, et al. (2004) Temozolomide for the treatment of brain metastases associated with metastatic melanoma: a phase II study. *J Clin Oncol* 22: 2101-2107.
259. Christodoulou C, Bafaloukos D, Linardou H, Aravantinos G, Bamias A, et al. (2005) Temozolomide (TMZ) combined with cisplatin (CDDP) in patients with brain metastases from solid tumors: a Hellenic Cooperative Oncology Group (HeCOG) Phase II study. *J Neurooncol* 71: 61-65.
260. Morgan L, Struck R, Waud W, LeBlanc B, Rodgers A, et al. (2009) Carbonate and carbamate derivatives of 4-demethylpenclomedine as novel anticancer agents. *Cancer Chemother Pharmacol* 64: 829-835.
261. Weiner R, Ware M, Friedlander P, Gordon C, Saenger Y, et al. (2013) A first-in-humans Phase I cancer clinical trial for 4-demethyl-4-cholesteryloxy-carbonylpenclomedine (DM-CHOC-PEN) in humans. *Cancer Res* 73: 73.
262. Weiner RS, Ware ML, Bastian G, Urien S, Rodgers AH, et al. (2012) Comparative pharmacokinetics for 4-demethyl-4-cholesteryloxy-carbonylpenclomedine (DM-CHOC-PEN) in humans. Proceedings of the 103rd Annual Meeting of the American Association for Cancer Research, Chicago, IL, USA.
263. Agboola O, Benoit B, Cross P, Da Silva V, Esche B, et al. (1998) Prognostic factors derived from recursive partition analysis (RPA) of radiation therapy oncology group (RTOG) brain metastases trials applied to surgically resected and irradiated brain metastatic cases. *International Journal of Radiation Oncology • Biology • Physics* 42: 155-159.
264. Nieder C, Geinitz H, Molls M (2008) Validation of the graded prognostic assessment index for surgically treated patients with brain metastases. *Anticancer Res* 28: 3015-3017.
265. Paek SH, Audu PB, Sperling MR, Cho J, Andrews DW (2005) Reevaluation of surgery for the treatment of brain metastases: review of 208 patients with single or multiple brain metastases treated at one

- institution with modern neurosurgical techniques. *Neurosurgery* 56: 1021-1034.
266. Rades D, Pluemer A, Veninga T, Dunst J, Schild SE (2007) A boost in addition to whole-brain radiotherapy improves patient outcome after resection of 1 or 2 brain metastases in recursive partitioning analysis class 1 and 2 patients. *Cancer* 110: 1551-1559.
267. Tendulkar RD, Liu SW, Barnett GH, Vogelbaum MA, Toms SA, et al. (2006) RPA classification has prognostic significance for surgically resected single brain metastasis. *Int J Radiat Oncol Biol Phys* 66: 810-817.
268. Serizawa T1 (2009) Radiosurgery for metastatic brain tumors. *Int J Clin Oncol* 14: 289-298.
269. Tsao M, Xu W, Sahgal A (2012) A meta-analysis evaluating stereotactic radiosurgery, whole-brain radiotherapy, or both for patients presenting with a limited number of brain metastases. *Cancer* 118: 2486-2493.
270. Auchter RM, Lamond JP, Alexander Iii E, Buatti JM, Chappell R, et al. (1996) A multiinstitutional outcome and prognostic factor analysis of radiosurgery for resectable single brain metastasis. *Int J Radiat Oncol Biol Phys* 35: 27-35.
271. Tsao MN, Sultanem K, Chiu D, Copps F, Dixon P, et al. (2003) Supportive care management of brain metastases: what is known and what we need to know. Conference proceedings of the National Cancer Institute of Canada (NCIC) Workshop on Symptom Control in Radiation Oncology. *Clin Oncol (R Coll Radiol)* 15: 429-434.
272. Linskey M, Andrews D, Asher A, Burri S, Kondziolka D, et al. (2010) The role of stereotactic radiosurgery in the management of patients with newly diagnosed brain metastases: a systematic review and evidence-based clinical practice guideline. *J Neurooncol* 96: 45-68.
273. kofman S, Garvin JS, Nagamani D, Taylor SG 3rd (1957) Treatment of cerebral metastases from breast carcinoma with prednisolone. *J Am Med Assoc* 163: 1473-1476.
274. Ruderman NB, Hall TC (1965) use of glucocorticoids in the palliative treatment of metastatic brain tumors. *Cancer* 18: 298-306.