

Biology of Vitamin D

Sunil Wimalawansa*

Department of Medicine, Cardiometabolic and Endocrine Institute, North Brunswick, NJ, USA

ABSTRACT

Vitamin D is both a vitamin and a hormone. It has pleotropic actions that extend beyond calcium and phosphate homeostasis, regulation of parathyroid hormone, and the prevention of osteomalacia, rickets, falls, and fractures. Over 80% of the vitamin D requirement is expected to be generated in the skin following exposure to ultraviolet B in sunlight; globally most people are under-exposed to sunlight and ingest too little vitamin D. Although overexposure to sunlight does not cause hypervitaminosis D, it can cause damage to skin cells. However, both extremes of vitamin D concentrations can be harmful. Vitamin D is essential for life, including for reproduction, fetal growth, immunity, and proper functioning of body systems. Evidence supports wider beneficial effects of vitamin D but achieving such benefits requires maintaining serum 25 dihydroxyvitamin D [25(OH)D] concentrations greater than 30 ng/mL. This article reviews biological pathways that are critical for the generation of 25(OH)D in the liver and 1,25-dihydroxyvitamin D in the kidney, and key abnormalities of vitamin D metabolism that increase the risk of common diseases and disorders, including obesity, insulin resistance, type 2 diabetes, pregnancy complications, autoimmune disorders, certain cancers, impairment of DNA repair, systemic inflammation, and oxidative stress that potentiates metabolic illnesses, such as cardiovascular disorders. Treatment of vitamin D deficiency on average costs less than 0.1% (varies between 0.2% and 0.06%) of the cost of investigations and treatment of worsening comorbidities and complications associated with hypovitaminosis D. For example, the cost of vitamin D supplementation to maintain serum 25(OH)D is \$12/year versus an average of \$6,000 to 18,000/year per affected person to manage complications. Despite the high benefits relative to cost, millions of people continue to have vitamin D deficiency. The individual and the population health can be markedly improved by maintaining serum 25(OH)D concentrations of greater than 30 ng/mL (75 nmol/L), which would improve the quality of life and reduce all-cause mortality. However, for prevention of certain diseases, such as autoimmune disorders and cancer, and to reduce all-cause mortality, serum 25(OH)D concentrations should be maintained between 40 and 60 ng/mL.

Keywords: 25(OH)D; 1,25(OH)₂D; Aging; Human diseases; Epidemiology; Morbidity and mortality; Prevention; Parathyroid hormone; Osteoporosis; Ultraviolet

INTRODUCTION

Most of the vitamin D requirement in humans can be and is supposed to be generated following exposure to sunlight. The prevalence of vitamin D deficiency increases in countries farthest from the equator. However, despite the presence of abundant sunlight, the incidence of vitamin D deficiency is high even among those who live within 1,000 km of the equator, such as the populations of India, Sri Lanka, and Far Eastern, Middle Eastern, and Persian Gulf countries [1-4]. This is attributable to the combination of individuals having darker skin color, climatic conditions, cultural habits, and sun-avoidance behaviors [5,6]. Hypovitaminosis D is a disease that can be cost-effectively prevented and treated through

the combination of adherence to specific public health guidelines and vitamin D supplementation regimens.

Vitamin D is a micronutrient that is metabolized into a multifunctional secosteroid hormone that is essential for human health, reproduction, and sustenance of life. Because dietary intake of vitamin D is low, humans are dependent on vitamin D synthesized in the skin. However, the rate of synthesis in the skin is affected by a variety of factors, including the density of melanin pigment; the use of sunscreen and ultraviolet (UV)-blocking creams and ointments, and clothing; and time of the day, month of the year, and duration of sun exposure [7-11]. Moreover, the dermal synthesis of vitamin D decreases in aging populations because of

Correspondence to: Sunil Wimalawansa, Department of Medicine, Cardiometabolic and Endocrine Institute, North Brunswick, NJ 08873, USA, Tel: 732-940-0811; E-mail: suniljw@hotmail.com

Received: December 14, 2018; **Accepted:** February 05, 2019; **Published:** February 15, 2019

Citation: Wimalawansa S (2019) Biology of Vitamin D. J steroids Horm Sci 10:198. doi: 10.24105/2157-7536.10.198

Copyright: © 2019 Wimalawansa S. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

aging or scarred skin. Meanwhile, sunlight provides vitamin D that is essential for humans and has additional multi-system benefits, many of which are less understood [12].

Most cells have vitamin D receptors and many target tissue cells have the CYP enzyme that is capable of intracellular generation of 1,25-dihydroxyvitamin D [$1,25(\text{OH})_2\text{D}$]. Moreover, several key genes that encode proteins and peptides are modulated in part by vitamin D and thus, affect musculoskeletal functions, mitochondrial respiration, cell growth, proliferation, differentiation, and apoptosis [13]. Vitamin D has many other functions in humans, including modulation of the neuromuscular actions, cell growth, inflammation, and immune functions [14,15].

LITERATURE REVIEW

Generation of vitamin D

Solar ultraviolet B (UVB) photons (approximately 290 to 310 nm) [12] are absorbed by 7-dehydrocholesterol in the epidermis of the skin and isomerized into previtamin D_3 [16], which is further isomerized to form vitamin D_3 [17]. Vitamin D binding protein (VDBP) has high affinity to previtamin D_3 and for dietary sources of vitamin D_2 and D_3 that are absorbed from the intestine; VDBP-bound vitamin D is transported *via* the circulation to the liver, where converted to $25(\text{OH})\text{D}$. In the liver, vitamin D is hydroxylated by the 25-hydroxylase (Cytochrome P450 enzymes; CYP2R1 and CYP27A1) enzymes to $25(\text{OH})\text{D}$ (caldiol), the major circulating and storage form of vitamin D [18-20].

Vitamin D is fat-soluble and naturally present in small quantities in few foods and is available as a dietary supplement. In evolutionary terms, it is supposed to be produced endogenously in the skin following exposure to ultraviolet rays of sunlight. Vitamin D obtained from sun exposure, food, and supplements is biologically inert and thus must undergo two steps of activation-hydroxylation in the body. As described, the first hydroxylation occurs in the liver, converting vitamin D to 25-hydroxyvitamin D [$25(\text{OH})\text{D}$], also known as caldiol. The second hydroxylation occurs in the kidney (and also in target tissues) *via* CYP27B1 and forms the physiologically active $1,25(\text{OH})_2\text{D}$, also known as calcitriol [21].

The physiologic way of producing vitamin D_3 is synthesis of previtamin D from 7-dehydrocholesterol in response to UVB in the skin [22]. However, any excess precursors produced in the skin are destroyed by UVB rays, preventing an accumulation of excess vitamin D in skin cells. Skin also contains the catabolic enzyme 24-hydroxylase, which catabolizes excess previtamin D to its 24-hydroxylated form, which is an inactive metabolite [23]. This feed-back process is regulated by several factors, including dose (exposure) of UVB, serum parathyroid hormone (PTH), and ionized calcium concentrations [24,25]. These inherent protective mechanisms prevent excessive retention of vitamin D in the skin [26]. Thus, sun exposure does not cause hypervitaminosis D or hypercalcemia. Figure 1 illustrates the basic activation steps of vitamin D to $25(\text{OH})\text{D}$ and further hydroxylation to $1,25(\text{OH})_2\text{D}$.

Illustrates the activation of vitamin D to $25(\text{OH})\text{D}$ in the liver and then to $1,25(\text{OH})_2\text{D}$ in the kidneys and target tissues.

$25(\text{OH})\text{D}$, a secosteroid generated from its precursors; ergocalciferol (vitamin D_2) and cholecalciferol (vitamin D_3). The half-life of $25(\text{OH})\text{D}_2$ is 10 to 12 days and that of $25(\text{OH})\text{D}_3$ is 20 to 24 days [27]; this is a clinically important biological

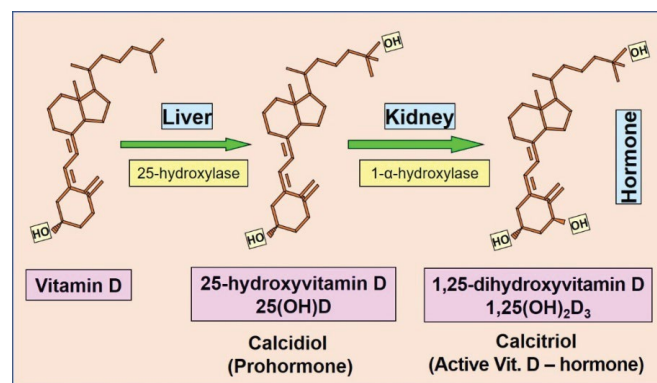


Figure 1: Essential activation steps of vitamin D to $25(\text{OH})\text{D}$ and $1,25(\text{OH})_2\text{D}$.

difference. A meta-analysis of randomized controlled clinical trials (RCTs) reported that vitamin D_3 supplements, when administered intermittently, increase and maintain serum $25(\text{OH})\text{D}$ concentrations for a significantly longer period ($p=0.001$) than do vitamin D_2 supplements [28]. Nevertheless, the 1,25-dihydroxy metabolites of these two vitamin D forms compete for the vitamin D receptor (VDR) in an equipotent manner [28,29]. Because D_3 has a longer half-life in circulation, it is considered the preferred form for supplementation [30,31]. However, D_2 which is plant or yeast-based is a good option for strict vegetarians (vegans).

Peak bone mass generally occurs during adolescence, but bone mass accrual slows toward the end of the third decade and then plateaus. Although the potential peak bone mass and aspects of skeletal development are in part determined by genetics, vitamin D, calcium, physical activity (mechanical stresses), and hormonal status influence the peak bone mass achieved and the rate of accrual of skeletal mineral content [32,33]. For those who do not live near the equator and are not regularly exposed to UVB rays, the cutaneous production of vitamin D or intake from vitamin D-rich or enriched foods occurs intermittently, especially during winter months. Thus, supplemental doses of vitamin D and sensible sun exposure are needed to prevent deficiency in many populations.

Transportation of vitamin D

Vitamin D generated in the skin binds to vitamin D binding protein (VDBP) and is transported to the liver, where it is hydroxylated to generate $25(\text{OH})\text{D}$. $25(\text{OH})\text{D}$ binds to VDBP and is transported throughout the body *via* the circulation. Upon reaching the proximal tubules of kidney and in extra-renal target tissues (see section 3.4), $25(\text{OH})\text{D}_2$ and $25(\text{OH})\text{D}_3$ are further hydroxylated to generate $1,25(\text{OH})_2\text{D}$ by the 1α -hydroxylase (CYP27B1) enzyme, the most biologically active and the hormonal form of vitamin D [18-20]. The VDBP-bound hormone calcitriol is then delivered throughout the body for its endocrine function, including to bone, intestine, and kidney, where it contributes to key physiological actions [34,35].

Following high-affinity binding of free calcitriol to the vitamin D receptors (VDRs), a ubiquitously expressed nuclear receptor in human cells, leads to physiologic actions. VDR acts as a ligand-modulated transcription factor, which belongs to a family of receptors that include steroid, thyroid, and retinoic acid receptors [36]. In addition to renal tubular cells, many extra-renal target tissues cells also convert $25(\text{OH})\text{D}$ to $1,25(\text{OH})_2\text{D}$ [37]. However, as described later, the controls for this conversion are different and the amounts generated intracellularly are difficult to quantitate.

Modulation of genes through the “VDR calcitriol complex”

The active hormone calcitriol reaches target cells through the circulation or is synthesized intra-cellularly in target tissue cells. The hormonal receptor interactions activate homodimerizations to form VDR:VDR or heterodimerization with the retinoic X receptors to form VDR:RXR complexes that are capable of binding to nuclear DNA [38]. Interactions of calcitriol with intracellular VDR leads to modulation of genes (activation or suppression), and activation of second messenger systems. At this stage, several other proteins interact with this complex as corepressors or coactivators, increasing or decreasing chromatin condensation. As a result, these complexes either enhance or suppress target gene transcription [38,39].

Mechanistically, excess vitamin A can change the ratio of RXR to retinoic acid receptor (RAR) (RXR/RAR) heterodimers. This may reduce the availability of RXR for heterodimerization with VDR and thus, dampen vitamin D signalling. This phenomenon could lead to the development of rickets even when vitamin D is replete. In part, this may be responsible for the higher fracture risks observed in older women with high serum vitamin A concentrations. Because of the differences in microcellular environments between different cells and epigenetic DNA modifications, the same activator can cause tissue-specific, different responses through the nuclear receptors. This process activates numerous genes, such as osteocalcin, bone sialoprotein, osteopontin, CYP24A1 and CYP27B1, TRPV6, PTH, PTH-related peptide, and the calcium-binding protein calbindin [40,41]. The rapid nongenomic effects of vitamin D do not depend on VDR-mediated activation [42-44] but respond to swift changes in intracellular calcium [45].

In addition to its well-known musculoskeletal benefits, 25(OH)D adequacy decreases risks and the severity of many extra-skeletal diseases and disorders, including insulin resistance, severity of type 2 diabetes mellitus (T2D), prediabetes, metabolic syndrome, inflammation, and autoimmunity. In addition to its endocrine effects, calcitriol also exerts autocrine and paracrine effects at target tissues and may modulate effects through epigenetic processes [46].

Circulatory 25(OH)D and cellular internalization

Serum 25(OH)D is the most sensitive biomarker for establishing vitamin D status [47]; serum concentrations of more than 30 ng/mL (75 nmol/L) are defined as vitamin D adequacy. However, some consider adequate serum concentration as above 20 ng/mL. The latter is adequate for skeletal physiology but not for other tissues. VDBP containing 25(OH)D is internalized by renal tubular and muscle cells through a megalin/cubilin-dependent plasma membrane transport mechanism [30,40,48]. In muscle cells, the internalized VDBP binds to actin, which contains high-affinity binding sites for 25(OH)D. When the VDBP undergoes proteolytic degradation in target tissue cells, 25(OH)D is released intracellularly [30], where it can be activated to calcitriol by intracellular CYP27B1 [40,49,50]. The feedback actions of calcitriol modify VDBP-dependent internalization and intracellular release of 25(OH)D, another mechanism for maximizing the utilization of vitamin D in skeletal muscle cells [51,52]. The vitamin D-DBP as well as vitamin D-albumin complexes are filtered through the glomeruli and re-uptake by megalin in the proximal tubule. Consequently, in kidney diseases that are characterized by tubular damage, vitamin D-DBP complexes are lost *via* urine. Thus, measurement of these can be used as a non-specific marker for renal tubular diseases. The

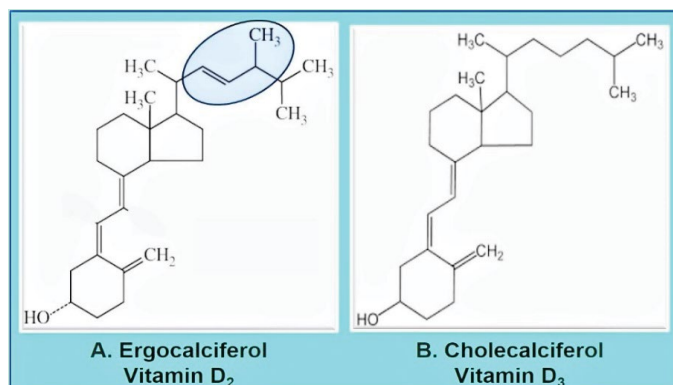


Figure 2: Molecular structures of vitamin D₂ and D₃.

structure of D₂ differs from D₃ only by having a c22-23 double bond and a methyl group at c24 (Figure 2), but the half-life of 25(OH)D₃ is almost double that of the D₂ metabolite. It is likely that a combination of mechanisms contributes to the longer half-life of 25(OH)D₃ than of 25(OH)D₂ [48].

Illustrates that the structures of D₂ differs from D₃ by having a double bond between c22-23 and a methyl group in c24 of the basic vitamin D molecule.

Skin is the organ generating vitamin D₃

In humans, skin thickness and the color have changed gradually, over thousands of years to maximize survival and protect humans from diseases [53]. When people started to migrate northward, from Africa, insufficient sunlight (thus, less UVB exposure) and less generation of vitamin D in the skin, negatively affected fertility and reproduction and the survival. During the process of migration northward, those who developed lighter skin color through mutagenesis of melanin-generating genes, had an overwhelming survival advantage in the less sunny climates (the beginning of the Caucasians). In evolutionary terms, in northern latitudes, having a lighter skin pigmentation created a favorable condition for generating optimal quantities of vitamin D, facilitating a balance between protecting dermal cells from UV damage, maximizing vitamin D production, and avoiding vitamin D toxicity [54-56].

In our equatorial ancestors, dark skin evolved to protect against sunburn and skin cancer. But, as humans moved away from the equator, paler skin facilitated increased synthesis of vitamin D in the skin, but it increased the risk of sunburn and skin cancer. Especially in those with skin-freckles, high UV exposure enhances the cell division and DNA damage, and leads to increased need for DNA repair. Folate is an essential component for DNA repair, so those with paler skin have an increased need for folate in those with darker skin. The latter group has enhanced melanogenesis but fewer dermal cell divisions, thus, a reduced the need for folate [54,55].

Biological activities of vitamin D

Key functions of vitamin D include the facilitation of calcium and phosphate absorption *via* the intestine and the regulation of bone metabolism. Together with PTH, vitamin D plays a key role in tightly maintaining serum ionized calcium concentrations [56]; this is exemplified by the negative correlation of serum 25(OH)D with PTH concentrations [25,57].

An understanding of the biochemistry and biology of this key steroid hormone and its generation and physiological actions, will help clinicians determine the best way to guide patients to obtain

improved clinical outcomes. In target tissues, $1,25(\text{OH})_2\text{D}$ is mostly generated intracellularly and acts as an autocrine signal: a cell produces and secretes a hormone or messenger that has its effects within that cell. In paracrine signaling, hormones and chemical messengers are secreted by a cell or a group of cells leading to local effects around the secretory cells. Figure 3 illustrates the cycle of generation of vitamin D in the skin, together with need-based activation of $25(\text{OH})\text{D}$ and calcitriol in target tissues and the control of serum ionized calcium concentrations through intestinal calcium absorption and increased bone turnover.

Vitamin D deficiency leads to increased secretion of PTH (i.e., secondary hyperparathyroidism), higher bone turnover, and the consequent loss of calcium and bone mineral content [58,59]. Moreover, suboptimal $1,25(\text{OH})_2\text{D}$ concentrations decrease intestinal calcium absorption and increase urinary calcium loss [60]; causing an overall negative calcium balance.

Skin- and oral/dietary-derived vitamin D is activated to its hormonal form, $1,25(\text{OH})_2\text{D}$ through a common path. Although 25-hydroxylase (CYP27B1) activity is exclusive to the liver, conversion of $25(\text{OH})\text{D}$ to $1,25(\text{OH})_2\text{D}$ via the 1α -hydroxylase enzyme occurs predominantly in renal tubules but also in target tissue cells. Also illustrated is the control of serum ionized calcium levels through intestinal absorption and bone turnover, together with the PTH-mediated increased renal tubular absorption of calcium.

⊕ Activated or upregulated; ⊖ Suppressed or downregulated.

Better vitamin D repletion is associated with reductions in the incidence and severity of several non-musculoskeletal disorders [61], including diabetes mellitus (T1D and T2D), insulin resistance, and metabolic syndrome [62-65], depression, infectious diseases, autoimmune diseases, cardiovascular diseases (CVDs), neurocognitive dysfunction, and specific cancers [66-82]. Prospective epidemiological studies conducted with stable, long-term $25(\text{OH})\text{D}$ concentrations demonstrated reduced risks of these conditions with higher baseline vitamin D status. However, many such findings have not been substantiated through RCTs [15]. This is in part due to design failures of RCTs, inherent bias, and the lack of properly designed RCTs with vitamin D-related primary outcomes.

Catabolism of vitamin D

Although extrarenal target tissue cells generate $1,25(\text{OH})_2\text{D}$, the concentrations achieved are unclear because it remains within the target tissue cells. Nevertheless, such extra-renal production of hormone provides biologically and physiologically important autocrine and paracrine functions. The amounts of $1,25(\text{OH})_2\text{D}$ generated in renal tubules and target cells can vary from person to person and day to day. In addition, the catabolic activity of 24-hydroxylase in target tissues plays a part in regulating intracellular concentrations of vitamin D metabolites, and thus, the availability of active, free hormone (Figure 4).

Although calcitriol in the circulation is modulated by PTH and the serum ionized calcium concentration [25], the intracellular content is regulated largely through serum $25(\text{OH})\text{D}$ (substrate) availability and, calcidiol and calcitriol catabolism through hydroxylation at C-24 and C-23 by a specific 24-hydroxylase (CYP24A1). Unlike in renal tubular cells, extrarenal tissues' (target tissue cells and macrophage) conversion of $25(\text{OH})\text{D}$ to $1,25(\text{OH})_2\text{D}$ is sensitive to

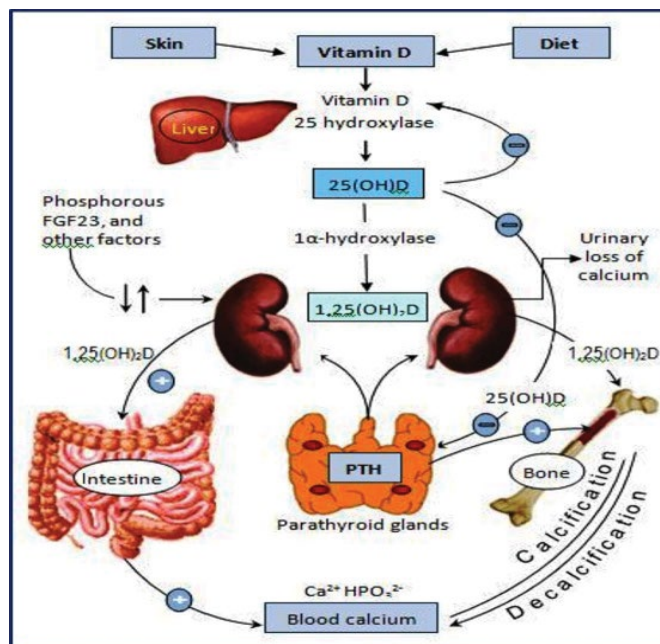


Figure 3: The interactions and pathways for generating vitamin D, $25(\text{OH})\text{D}$, and $1,25(\text{OH})_2\text{D}$.

medications, such as glucocorticoids. The renal $25(\text{OH})\text{D}$ -1 alpha-hydroxylase (CYP27B1) is not sensitive to hydroxychloroquine, ketoconazole or glucocorticoids [83]. Thus, when hypercalcemia is caused by extra-renal over production of calcitriol, as occurs with granulomatous diseases such as sarcoidosis, it can be controlled at least temporarily with these medications.

Catabolic pathways of vitamin D: The 24-hydroxylase (CYP24A1) enzyme generates biologically inactive, 24-hydroxylated products that are excreted as calcitroic acids through the biliary tract; whereas, the 23-hydroxylase pathway products are excreted as 1,25-26,23 lactone. The importance of biologically active calcitriol concentrations has been demonstrated in a study with CYP24A1 knockout mice, in which the animals developed impaired bone mineralization and hypercalcemia and had perinatal death rates of approximately 50% [84,85]. However, these characteristics do not exist in CYP24A1/VDR double knockout mice, suggesting that increased calcitriol levels, but not the absence of 24- or 23-hydroxylated vitamin D metabolites, are responsible for this abnormal phenotype.

A protective biofeedback mechanism is also present in the liver; when excess $25(\text{OH})\text{D}$ is formed, it is catabolized to $24(\text{OH})\text{D}$ or $24,25(\text{OH})_2\text{D}$, both of which are biologically inert [86]. Similarly, in the renal tubular cells and target cells, if and when excess $1,25(\text{OH})_2\text{D}$ is generated, it is catabolized into $1,24,25(\text{OH})_3\text{D}$, another biologically inactive vitamin D metabolite. Depending on the target tissue, calcitriol and its products can be metabolized into additional inactive metabolites, but this happens in smaller quantities and current methodologies preclude quantitating them (Figure 4).

The paths of catabolism/inactivation of active vitamin D metabolites. Excess $25(\text{OH})\text{D}$ and $1,25(\text{OH})_2\text{D}$ are catabolized to inactive forms $24(\text{OH})\text{D}$, $24,25(\text{OH})_2\text{D}$ (mainly in liver and in target tissues), and $1,24,25(\text{OH})_3\text{D}$ (renal cells and target tissues) [86]. The threshold and modes of inactivation vary among tissues.

Broader effects of vitamin D: Hypovitaminosis D can have significant musculoskeletal consequences, such as rickets in

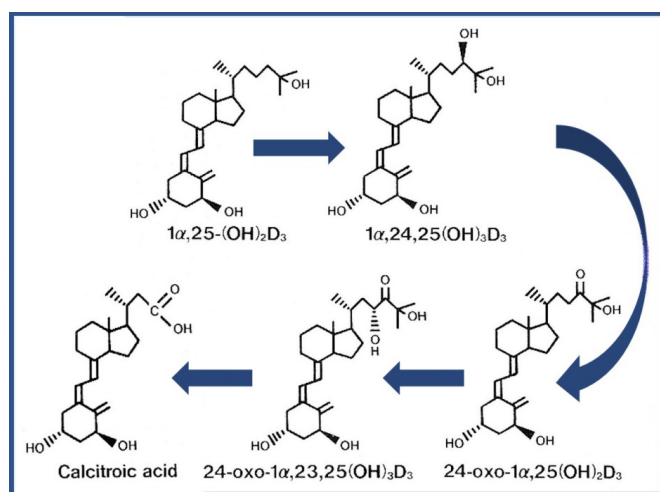


Figure 4: Catabolism of vitamin D.

children and osteomalacia in adults; many such patients present with the inability to raise arms above the head and difficulty in getting up from a chair (i.e., signs of proximal myopathy) [87]. Vitamin D influences many physiologic activities, including maintaining calcium and phosphorus homeostasis, subduing autoimmunity and infections, controlling cell growth, innate and adaptive immunity, mitochondrial respiration, and metabolic activities [88-91]. Although, diabetes can lead to renal failure and the resultant lowering of the generation of 1,25(OH)₂D, hypovitaminosis D does not cause renal impairment. Renal failure causes deficiency of 1,25(OH)₂D and an array of other disorders. In addition, hypovitaminosis is known to worsen several metabolic disorders, including metabolic syndrome, insulin resistance, and diabetes.

Common causes of vitamin D deficiency: Those with dark skin, those who have less exposure to sunlight, and older persons, all have less capacity to generate vitamin D compared with a healthy adult. In addition, stored vitamin D quantity decreases with advancing age, especially during winter months, irrespective of living location. Having gastrointestinal diseases that cause fat malabsorption, such as celiac and Crohn's disease, short bowel syndrome including gastric bypass, and cystic fibrosis also reduces the capacity to absorb vitamin D.

To be biologically effective, vitamin D molecules requires activation. Because activation (*via* hydroxylation) occurs in the liver and kidney, failure of either of these organs (because of reduced levels of activating enzymes) leads to lesser amounts of vitamin D. Thus, people with chronic liver or kidney diseases are at a high risk of having low concentrations of 25(OH)D and/or 1,25(OH)₂D.

Genetic causes of low vitamin D serum concentrations include lack or impairment of generation of active vitamin D (or formation of inappropriate amounts of biologically inactive forms or enhanced catabolism), abnormalities of vitamin D-binding protein and/or VDR abnormalities. Any of these abnormalities or syndromes can present with low vitamin D activity, but these disorders are rare; however, they need to be considered when assessing the vitamin D deficiency of an individual person.

DISCUSSION

Although adequate vitamin D is important for proper muscle functioning and skeletal development and maintenance, evidence suggests that vitamin D facilitates the prevention of several

diseases, including diabetes mellitus, hypertension, autoimmune diseases, and certain common cancers. Consequently, in the presence of insufficient vitamin D, the body's systems are unlikely to work optimally. Epidemiological data reported a high prevalence of vitamin D inadequacy among children, the elderly, and those with osteoporosis. Low sunlight exposure, age-related decreases in cutaneous synthesis, and diets low in vitamin D, contribute to the high prevalence of vitamin D inadequacy worldwide [12].

The proper functioning of the biology of vitamin D-endocrine, paracrine, and autocrine systems is essential for most physiological activities. Normal serum concentrations of 25(OH)D and the intracellular concentration of 1,25(OH)₂D are essential for optimal musculoskeletal and soft tissue health. Vitamin D deficiency, as determined by serum 25(OH)D concentrations of less than 30 ng/mL, is associated with increased risks of illnesses and disorders and increased all-cause mortality even among apparently healthy individuals [92,93]. In addition, having lower serum 25(OH)D concentrations can cause dysfunctions in many systems, despite the presence of physiologic concentrations of calcitriol.

The minimum recommended steady-state, serum 25(OH)D concentration is 30 ng/mL (75 nmol/L). In general, the range between 30 and 60 ng/mL is considered as healthy (physiological); no known adverse effects related to vitamin D occur till the serum 25(OH)D concentration exceeds 125 ng/mL (375 nmol/L). However, persons with certain disorders, such as obesity, metabolic disorders, autoimmunity and cancer, may require higher levels (concentrations between 40 and 60 ng/mL). To achieve such concentrations, adequate exposure to UVB rays and/or vitamin D₃ supplements of between 2,000 and 6,000 IU per day are required. It has been demonstrated that daily intake of 10,000 IU is safe [94]. The elderly, the obese, those who are taking medications that activate hepatic cytochrome P450 enzymes that enhance catabolism of vitamin, and pregnant women and those who are lactating require higher intakes (i.e., vitamin D₃, 6,000 IU/day) of vitamin D.

Vitamin D adequacy can be assessed only through the measurement of serum 25(OH)D. Recent data from epidemiological, cross-sectional, and longitudinal studies support that having physiological serum concentrations of 25(OH)D (i.e., more than 30 ng/mL) leads to a reduced incidence of many extra-musculoskeletal disorders, including diabetes [95-97], osteoporosis [98,99], multiple sclerosis [100], rheumatoid arthritis [101], and certain types of cancer [102-104].

Having adequate serum 25(OH)D concentrations allows vitamin D to generate its active hormone, 1,25(OH)₂D (calcitriol) in renal tubules and in target tissues with some feedback controls and facilitates its intended positive or negative modulatory effects. These include enzymatic reactions; synthesis and secretion of hormones, such as insulin and PTH; and modulating the renin-angiotensin-aldosterone and FGF23-Klotho systems [105]. Meanwhile, data from metabolomics and transcriptomics promise the generation of improved longer-term extra-skeletal clinically meaningful outcomes.

CONCLUSION

Vitamin D metabolism and actions are influenced by many medications, environmental pollutants, and physical activities and lifestyles; some of these also modulate the balance between energy intake and expenditure. Cumulative evidence supports biological associations of vitamin D adequacy with disease risk

reduction and improved physical and mental well-being. In this regard, few investigations were done on CYP27B1-mediated “target tissue” production of 1,25(OH)₂D until recently; thus, expansion of research into this area is important. In addition to the endocrine role of vitamin D, the paracrine and autocrine functions of calcitriol are essential for full biological activity of vitamin D but not yet fully understood.

Although many diseases and disorders are related to vitamin D deficiency, the costs of investigating and managing the complications associated with hypovitaminosis D-induced metabolic disorders are extremely high. Maintaining serum 25(OH)D concentrations between 30 and 60 ng/mL would significantly reduce the severity of and complications associated with several common diseases. Because of the favorable cost-benefit ratio, this approach is highly beneficial. The positive impact on benefits in humans and on the economy by following public health approaches would exceed the benefits derived from the combined targeting of infectious and parasitic diseases globally.

REFERENCES

1. Van Schoor NM, Lips P. Worldwide vitamin D status. *Best Pract Res Clin Endocrinol Metab.* 2011;25:671-680.
2. Eggemoen AR, Knutsen KV, Dalen I, Jenum AK. Vitamin D status in recently arrived immigrants from Africa and Asia: a cross-sectional study from Norway of children, adolescents and adults. *BMJ Open.* 2013;3:e003293.
3. Garland CF, Gorham ED, Mohr SB, Garland FC. Vitamin D for cancer prevention: global perspective. *Ann Epidemiol.* 2009;19:468-83.
4. Hilger J, Friedel A, Herr R, Rausch T, Roos F, Wahl DA, et al. A systematic review of vitamin D status in populations worldwide. *Br J Nutr.* 2014;111:23-45.
5. Haq A, Wimalawansa SJ, Carlberg C. Highlights from the 5th International Conference on Vitamin D Deficiency, Nutrition and Human Health, Abu Dhabi, United Arab Emirates, March 24-25, 2016. *J Steroid Biochem Mol Biol.* 2018;175:1-3.
6. Pludowski P, Holick MF, Grant WB, Konstantynowicz J, Mascarenhas MR, Haq A, et al. Vitamin D supplementation guidelines. *J Steroid Biochem Mol Biol.* 2018;175:125-135.
7. Mithal A, Wahl DA, Bonjour JP, Burckhardt P, Dawson-Hughes B, Eisman JA, et al. Global vitamin D status and determinants of hypovitaminosis D. *Osteoporos Int.* 2009;20:1807-1820.
8. Wimalawansa SJ. Vitamin D in the new millennium. *Curr Osteoporos Rep.* 2012;10:4-15.
9. Grant WB, Wimalawansa SJ, Holick MF. Vitamin D supplements and reasonable solar UVB should be recommended to prevent escalating incidence of chronic diseases. *British Medical Journal.* 2015;350:h321.
10. Holick MF. High prevalence of vitamin D inadequacy and implications for health. *Mayo Clin Proc.* 2006;81:353-373.
11. Mark KA, Dumas KJ, Bhaumik D, Schilling B, Davis S, Oron TR, et al. Vitamin D promotes protein homeostasis and longevity via the stress response pathway genes *skn-1*, *ire-1*, and *xbp-1*. *Cell Rep.* 2016;17:1227-1237.
12. Wacker M, Holick MF. Sunlight and vitamin D: A global perspective for health. *Dermato-endocrinology.* 2013;5:51-108.
13. Molina P, Carrero JJ, Bover J, Chauveau P, Mazzaferro S, Torres PU, et al. Vitamin D, a modulator of musculoskeletal health in chronic kidney disease. *J Cachexia Sarcopenia Muscle.* 2017;8:686-701.
14. Sintov AC, Yarmolinsky L, Dahan A, Ben-Shabat S. Pharmacological effects of vitamin D and its analogs: recent developments. *Drug Discov Today.* 2014;19:1769-1774.
15. Wimalawansa SJ. Non-musculoskeletal benefits of vitamin D. *J Steroid Biochem Mol Biol.* 2018;175:60-81.
16. Jeon SM, Shin EA. Exploring vitamin D metabolism and function in cancer. *Exp Mol Med.* 2018;50:20.
17. Holick MF. Sunlight and vitamin D for bone health and prevention of autoimmune diseases, cancers, and cardiovascular disease. *Am J Clin Nutr.* 2004;80:1678S-1688S.
18. Jones G, Prosser DE, Kaufmann M. Cytochrome P450-mediated metabolism of vitamin D. *J Lipid Res.* 2014;55:13-31.
19. Masuda S, Gao M, Zhang A, Kaufmann M, Jones G. Importance of cytochrome P450-mediated metabolism in the mechanism of action of vitamin D analogs. *Recent Results Cancer Res.* 2003;164:189-202.
20. Schuster I. Cytochromes P450 are essential players in the vitamin D signaling system. *Biochim Biophys Acta.* 2011;1814:186-199.
21. Chow EC, Quach HP, Vieth R, Pang KS. Temporal changes in tissue 1 α ,25-dihydroxyvitamin D₃, vitamin D receptor target genes, and calcium and PTH levels after 1,25(OH)₂D₃ treatment in mice. *Am J Physiol Endocrinol Metab.* 2013;304:E977-989.
22. Holick MF, Frommer JE, McNeill SC, Richtand NM, Henley JW, Potts JT Jr. Photometabolism of 7-dehydrocholesterol to previtamin D₃ in skin. *Biochem Biophys Res Commun.* 1977;76:107-114.
23. Webb AR, DeCosta BR, Holick MF. Sunlight regulates the cutaneous production of vitamin D₃ by causing its photodegradation. *J Clin Endocrinol Metab.* 1989;68:882-887.
24. Armbrecht HJ, Hodam TL, Boltz MA, Partridge NC, Brown AJ, Kumar VB. Induction of the vitamin D 24-hydroxylase (CYP24) by 1,25-dihydroxyvitamin D₃ is regulated by parathyroid hormone in UMR106 osteoblastic cells. *Endocrinology.* 1998;139:3375-3381.
25. Kroll MH, Bi C, Garber CC, Kaufman HW, Liu D, Caston-Balderrama A, et al. Temporal relationship between vitamin D status and parathyroid hormone in the United States. *PLoS One.* 2015;10:e0118108.
26. Miller WL. Genetic disorders of Vitamin D biosynthesis and degradation. *J Steroid Biochem Mol Biol.* 2017;165:101-108.
27. Chow EC, Durk MR, Maeng HJ, Pang KS. Comparative effects of 1 α -hydroxyvitamin D₃ and 1,25-dihydroxyvitamin D₃ on transporters and enzymes in *fxr* (+/+) and *fxr* (-/-) mice. *Biopharm Drug Dispos.* 2013;34:402-416.
28. Tripkovic L, Lambert H, Hart K, Smith CP, Bucca G, Penson S, et al. Comparison of vitamin D(2) and vitamin D(3) supplementation in raising serum 25-hydroxyvitamin D status: a systematic review and meta-analysis. *The American J Clin Nutri.* 2012;95:1357-1364.
29. Chun RF, Hernandez I, Pereira R, Swinkles L, Huijs T, Zhou R, et al. Differential responses to vitamin d2 and vitamin d3 are associated with variations in free 25-hydroxyvitamin d. *Endocrinology.* 2016;157:3420-3430.
30. Abboud M, Rybchyn MS, Ning YJ, Brennan-Speranza TC, Girgis CM, Gunton JE, et al. 1,25-Dihydroxycholecalciferol (calcitriol) modifies uptake and release of 25-hydroxycholecalciferol in skeletal muscle cells in culture. *J Steroid Biochem Mol Biol.* 2018;177:109-115.
31. Mocanu V, Vieth R. Three-year follow-up of serum 25-hydroxyvitamin D, parathyroid hormone, and bone mineral density in nursing home residents who had received 12 months of daily bread fortification with 125 mug of vitamin D(3). *Nutr J.* 2013;0.595138889.
32. Slemenda CW, Peacock M, Hui S, Zhou L, Johnston CC. Reduced rates of skeletal remodeling are associated with increased bone mineral density during the development of peak skeletal mass. *J*

- Bone Miner Res. 1997;12:676-682.
33. Baxter-Jones AD, Faulkner RA, Forwood MR, Mirwald RL, Bailey DA. Bone mineral accrual from 8 to 30 years of age: an estimation of peak bone mass. *J Bone Miner Res.* 2011;26:1729-1739.
 34. Basit S. Vitamin D in health and disease: A literature review. *Br J Biomed Sci.* 2013;70:161-172.
 35. Heaney RP. Vitamin D in health and disease. *Clin J Am Soc Nephrol.* 2008;3:1535-1541.
 36. Liberman UA. Disorders in Vitamin D Action. 2014. In: De Groot LJ, Chrousos G, Dungan K, et al. eds, South Dartmouth (MA): MDText.com, Inc.; 2000.
 37. Zehnder D, Bland R, Williams MC, McNinch RW, Howie AJ, Stewart PM, et al. Extrarenal expression of 25-hydroxyvitamin d(3)-1 alpha-hydroxylase. *J Clin Endocrinol Metab.* 2001;86:888-894.
 38. Valdivielso JM. The physiology of vitamin D receptor activation. *Contrib Nephrol.* 2009;163:206-212.
 39. Kawahara M, Iwasaki Y, Sakaguchi K, Taguchi T, Nishiyama M, Nigawara T, et al. Predominant role of 25OHD in the negative regulation of PTH expression: clinical relevance for hypovitaminosis D. *Life Sci.* 2008;82:677-683.
 40. Pike JW, Meyer MB. The vitamin D receptor: new paradigms for the regulation of gene expression by 1,25-dihydroxyvitamin D(3). *Endocrinol Metab Clin North Am.* 2010;39:255-269.
 41. Gallieni M, Cozzolino M, Fallabrino G, Pasho S, Olivi L, Brancaccio D. Vitamin D: physiology and pathophysiology. *Int J Artif Organs.* 2009;32:87-94.
 42. Blomberg Jensen M. Vitamin D and male reproduction. *Nat Rev Endocrinol.* 2014;10:175-186.
 43. Bikle DD. Vitamin D metabolism, mechanism of action, and clinical applications. *Chem Biol.* 2014;21:319-329.
 44. Wimalawansa SJ. Vitamin D: An essential component for skeletal health. *Annals of NYAS.* 2012;1240:90-98.
 45. Gibson CC, Davis CT, Zhu W, Bowman-Kirigin JA, Walker AE, Zhengfu Tai, et al. Dietary vitamin D and its metabolites non-genomically stabilize the endothelium. *PLoS One.* 2015;10:e0140370.
 46. Kato S, Nishimura KI, Mori JI. Update on recent progress in vitamin D research. Molecular basis of epigenetic regulation by vitamin D via its nuclear receptor. *Clin Calcium.* 2017;27:1543-1550.
 47. Hollis BW. Circulating 25-hydroxyvitamin D levels indicative of vitamin D sufficiency: implications for establishing a new effective dietary intake recommendation for vitamin D. *J Nutr.* 2005;135:317-22.
 48. Christensen EI, Birn H. Megalin and cubilin: Multifunctional endocytic receptors. *Nature Reviews Molecular Cell Biology.* 2002;3:258.
 49. Gekle M, Knaus P, Nielsen R, Mildenerger S, Freudinger R, Wohlfarth V, et al. Transforming growth factor-beta1 reduces megalin- and cubilin-mediated endocytosis of albumin in proximal-tubule-derived opossum kidney cells. *J Physiol.* 2003;552:471-481.
 50. Liu D, Wen Y, Tang TT, Lv LL, Tang RN, Liu H, et al. Megalin/Cubulin-Lysosome-mediated Albumin Reabsorption Is Involved in the Tubular Cell Activation of NLRP3 Inflammasome and Tubulointerstitial Inflammation. *J Biol Chem.* 2015;290:18018-18028.
 51. Pojednic RM, Ceglia L. The emerging biomolecular role of vitamin D in skeletal muscle. *Exerc Sport Sci Rev.* 2014;42:76-81.
 52. Ceglia L, Harris SS. Vitamin D and its role in skeletal muscle. *Calcif Tissue Int.* 2013;92:151-162.
 53. Jablonski NG, Chaplin G. Epidermal pigmentation in the human lineage is an adaptation to ultraviolet radiation. *J Hum Evol.* 2013;65:671-675.
 54. Chaplin G, Jablonski NG. Vitamin D and the evolution of human depigmentation. *Am J Phys Anthropol.* 2009;139:451-461.
 55. Yuen AW, Jablonski NG. Vitamin D: in the evolution of human skin colour. *Med Hypotheses.* 2010;74:39-44.
 56. Wimalawansa SJ, Razaque DMS, Al-Daghri NM. Calcium and vitamin d in human health: hype or real? *J Steroid Biochem Mol Biol.* 2018;180:4-14.
 57. Wimalawansa SJ. Extra-skeletal benefits, endocrine functions, and toxicity of vitamin D. *J Endocrinol Diab.* 2016;3:1-5.
 58. Singla R, Gurung P, Aggarwal N, Dutta U, Kochhar R. Relationship between preeclampsia and vitamin D deficiency: a case control study. *Arch Gynecol Obstet.* 2015;291:1247-1251.
 59. Lenders CM, Feldman HA, Von Scheven E, Merewood A, Sweeney C, Wilson DM, et al. Relation of body fat indexes to vitamin D status and deficiency among obese adolescents. *Am J Clin Nutr.* 2009;90:459-467.
 60. Findling JW, Adams ND, Lemann J Jr, Gray RW, Thomas CJ, Tyrrell JB. Vitamin D metabolites and parathyroid hormone in Cushing's syndrome: relationship to calcium and phosphorus homeostasis. *J Clin Endocrinol Metab.* 1982;54:1039-1044.
 61. Wimalawansa SJ. Vitamin D; What clinicians would like to know. *Sri Lanka Journal of Diabetes, Endocrinology and Metabolism.* 2012;1:73-88.
 62. Conrado T, Miranda-Filho Dde B, Bandeira F. Vitamin D deficiency in HIV-infected individuals: one more risk factor for bone loss and cardiovascular disease? *Arq Bras Endocrinol Metabol.* 2010;54:118-122.
 63. Pacifico L, Anania C, Osborn JF, Ferraro F, Bonci E, Olivero E, et al. Low 25(OH)D3 levels are associated with total adiposity, metabolic syndrome, and hypertension in Caucasian children and adolescents. *Eur J Endocrinol.* 2011;165:603-611.
 64. Tepper S, Shahar DR, Geva D, Ish-Shalom S. Differences in homeostatic model assessment (HOMA) values and insulin levels after vitamin D supplementation in healthy men: a double-blind randomized controlled trial. *Diabetes Obes Metab.* 2016;18:633-637.
 65. Cheng Q, Boucher BJ, Leung PS. Modulation of hypovitaminosis D-induced islet dysfunction and insulin resistance through direct suppression of the pancreatic islet renin-angiotensin system in mice. *Diabetologia.* 2013;56:553-562.
 66. Garland CF, Garland FC, Gorham ED, Lipkin M, Newmark H, Mohr SB, et al. The role of vitamin D in cancer prevention. *Am J Public Health* 2006;96:252-261.
 67. Giovannucci E, Liu Y, Willett WC. Cancer incidence and mortality and vitamin D in black and white male health professionals. *Cancer Epidemiol Biomarkers Prev.* 2006;15:2467-2472.
 68. Gross MD. Vitamin D and calcium in the prevention of prostate and colon cancer: new approaches for the identification of needs. *J Nutr.* 2005;135:326-331.
 69. Vieth R. Why the optimal requirement for Vitamin D3 is probably much higher than what is officially recommended for adults. *J Steroid Biochem Mol Biol.* 2004;89:575-579.
 70. George PS, Pearson ER, Witham MD. Effect of vitamin D supplementation on glycaemic control and insulin resistance: a systematic review and meta-analysis. *Diabet Med.* 2012;29:e142-150.
 71. Manousopoulou A, Al-Daghri NM, Garbis SD, Chrousos GP. Vitamin D and cardiovascular risk among adults with obesity: a systematic review and meta-analysis. *Eur J Clin Invest.* 2015;45:1113-1126.

72. Khan H, Kunutsor S, Franco OH, Chowdhury R. Vitamin D, type 2 diabetes and other metabolic outcomes: a systematic review and meta-analysis of prospective studies. *Proc Nutr Soc.* 2013;72:89-97.
73. Pilz S, Kienreich K, Rutters F, de Jongh R, van Ballegooijen AJ, Grübler M, et al. Role of vitamin D in the development of insulin resistance and type 2 diabetes. *Curr Diab Rep.* 2013;13:261-270.
74. Sung CC, Liao MT, Lu KC, Wu CC. Role of vitamin D in insulin resistance. *J Biomed Biotechnol.* 2012;634195.
75. Takiishi T, Gysemans C, Bouillon R, Mathieu C. Vitamin D and diabetes. *Rheum Dis Clin North Am.* 2012;38:179-206.
76. Luxwolda MF, Kuipers RS, Kema IP, van der Veer E, Dijk-Brouwer DAJ, Muskiet FA. Vitamin D status indicators in indigenous populations in East Africa. *Eur J Nutr.* 2013;52:1115-1125.
77. Al Nozha OM. Vitamin D and extra-skeletal health: Causality or consequence. *Int J Health Sci (Qassim).* 2016;10:443-452.
78. Body JJ, Bergmann P, Boonen S, Devogelaer JP, Gielen E, Goemaere S, et al. Extraskeletal benefits and risks of calcium, vitamin D and anti-osteoporosis medications. *Osteoporos Int 23 Suppl.* 2012;1:S1-23.
79. Cangoz S, Chang YY, Chempakaseril SJ, Guduru RC, Huynh LM, John JS, et al. Vitamin D and type 2 diabetes mellitus. *J Clin Pharm Ther.* 2013;38:81-84.
80. Cianferotti L, Bertoldo F, Bischoff-Ferrari HA, Bruyere O, Cooper C, Cutolo M, et al. Vitamin D supplementation in the prevention and management of major chronic diseases not related to mineral homeostasis in adults: research for evidence and a scientific statement from the European society for clinical and economic aspects of osteoporosis and osteoarthritis (ESCEO). *Endocrine.* 2017;56:245-61.
81. Pilz S, Gaksch M, Hartaigh BO, Tomaschitz A, Marz W. Vitamin D in preventive medicine. *Anticancer Res.* 2015;35:1161-1170.
82. Weinert LS, Silveiro SP. Maternal-fetal impact of vitamin D deficiency: a critical review. *Matern Child Health J.* 2015;19:94-101.
83. Adams JS, Hewison M. Extrarenal expression of the 25-hydroxyvitamin D-1-hydroxylase. *Arch Biochem Biophys.* 2012;523:95-102.
84. Masuda S, Byford V, Arabian A, Sakai Y, Demay MB, St-Arnaud R, et al. Altered pharmacokinetics of 1 α ,25-dihydroxyvitamin D₃ and 25-hydroxyvitamin D₃ in the blood and tissues of the 25-hydroxyvitamin D-24-hydroxylase (Cyp24a1) null mouse. *Endocrinology.* 2005;146:825-834.
85. St-Arnaud R, Arabian A, Travers R, Barletta F, Raval-Pandya M, Chapin K, et al. Deficient mineralization of intramembranous bone in vitamin D-24-hydroxylase-ablated mice is due to elevated 1,25-dihydroxyvitamin D and not to the absence of 24,25-dihydroxyvitamin D. *Endocrinology.* 2000;141:2658-2666.
86. Matsumoto T, Ikeda K, Yamato H, Morita K, Ezawa I, Fukushima M, et al. Effect of 24,25-dihydroxyvitamin D₃ on 1,25-dihydroxyvitamin D₃ metabolism in calcium-deficient rats. *Biochem J.* 1988;250:671-677.
87. Uday S, Hogler W. Prevention of rickets and osteomalacia in the UK: political action overdue. *Arch Dis Child.* 2018;103:901-906.
88. Verstuyf A, Carmeliet G, Bouillon R, Mathieu C. Vitamin D: A pleiotropic hormone. *Kidney Int.* 2010;78:140-145.
89. Cancela L, Nemere I, Norman AW. 1 α ,25(OH)₂ vitamin D₃: A steroid hormone capable of producing pleiotropic receptor-mediated biological responses by both genomic and nongenomic mechanisms. *J Steroid Biochem.* 1988;30:33-39.
90. Bouillon R, Carmeliet G, Verlinden L, van Etten E, Verstuyf A, Luderer HF, et al. Vitamin D and human health: lessons from vitamin D receptor null mice. *Endocr Rev.* 2008;29:726-776.
91. Feldman D, Krishnan AV, Swami S, Giovannucci E, Feldman BJ. The role of vitamin D in reducing cancer risk and progression. *Nat Rev Cancer.* 2014;14:342-357.
92. Garland CF, Kim JJ, Mohr SB, Gorham ED, Grant WB, Giovannucci EL, et al. Meta-analysis of all-cause mortality according to serum 25-hydroxyvitamin D. *Am J Public Health.* 2014;104:e43-50.
93. Sun YQ, Langhammer A, Skorpen F, Chen Y, Mai XM. Serum 25-hydroxyvitamin D level, chronic diseases and all-cause mortality in a population-based prospective cohort: the HUNT Study, Norway. *BMJ Open.* 2017;7:e017256.
94. Amir E, Simmons CE, Freedman OC, Dranitsaris G, Cole DE, Vieth R, et al. A phase 2 trial exploring the effects of high-dose (10,000 IU/day) vitamin D(3) in breast cancer patients with bone metastases. *Cancer.* 2010;116:284-291.
95. Chiu KC, Chu A, Go VL, Saad MF. Hypovitaminosis D is associated with insulin resistance and beta cell dysfunction. *Am J Clin Nutr.* 2004;79:820-825.
96. Gupta AK, Brashear MM, Johnson WD. Prediabetes and prehypertension in healthy adults are associated with low vitamin D levels. *Diabetes Care.* 2011;34:658-660.
97. Hamed EA, Abu Faddan NH, Adb Elhafeez HA, Sayed D. Parathormone - 25(OH)-vitamin D axis and bone status in children and adolescents with type 1 diabetes mellitus. *Pediatr Diabetes.* 2011;12:536-546.
98. Feskanich D, Willett WC, Colditz GA. Calcium, vitamin D, milk consumption, and hip fractures: a prospective study among postmenopausal women. *Am J Clin Nutr.* 2003;77:504-511.
99. Meier C, Woitge HW, Witte K, Lemmer B, Seibel MJ. Supplementation with oral vitamin D₃ and calcium during winter prevents seasonal bone loss: a randomized controlled open-label prospective trial. *J Bone Miner Res.* 2003;19:1221-1230.
100. Munger KL, Zhang SM, O'Reilly E, Hernan MA, Olek MJ, Willett WC, et al. Vitamin D intake and incidence of multiple sclerosis. *Neurology.* 2004;62:60-65.
101. Merlino LA, Curtis J, Mikuls TR, Cerhan JR, Criswell LA, Saag KG, et al. Vitamin D intake is inversely associated with rheumatoid arthritis: results from the Iowa Women's Health Study. *Arthritis Rheum.* 2004;50:72-77.
102. Lieberman DA, Prindiville S, Weiss DG, Willett W. Risk factors for advanced colonic neoplasia and hyperplastic polyps in asymptomatic individuals. *JAMA.* 2003;290:2959-2967.
103. McCullough ML, Robertson AS, Rodriguez C, Jacobs EJ, Chao A, Carolyn J, et al. Calcium, vitamin D, dairy products, and risk of colorectal cancer in the Cancer Prevention Study II Nutrition Cohort (United States). *Cancer Causes Control.* 2003;14:1-12.
104. Tretli S, Schwartz GG, Torjesen PA, Røsbak TE. Serum levels of 25-hydroxyvitamin D and survival in Norwegian patients with cancer of breast, colon, lung, and lymphoma: A population-based study. *Cancer Causes Control.* 2012;23:363-370.
105. Zaheer S, Taquechel K, Brown JM, Adler GK, Williams JS, Vaidya A. A randomized intervention study to evaluate the effect of calcitriol therapy on the renin-angiotensin system in diabetes. *J Renin Angiotensin Aldosterone Syst.* 2018;19:1470320317754178.