

## Anti-Inflammatory Activity of Methanolic Leaf Extract of *Kigelia Africana* (Lam.) Benth and Stem Bark Extract of *Acacia Hockii* De Wild in Mice

Kamau JK<sup>\*</sup>, Nthiga PM, Mwonjoria JK, Ngeranwa JJN, Ngugi MP

<sup>\*</sup>Department of Biochemistry and Biotechnology, School of Pure and Applied Sciences, Kenyatta University, P.O. Box 43844-00100, Nairobi, Kenya

<sup>\*</sup>Corresponding author: Kamau JK, Department of Biochemistry and Biotechnology, Kenyatta University, Nairobi, Kenya, Tel: +254727642699; Email: jameskimani006@gmail.com

Received date: May 24, 2016; Accepted date: June 07, 2016; Published date: June 13, 2016

Copyright: © 2016 Kamau JK. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### Abstract

Inflammation causes discomfort, suffering and lower productivity of the victims. Synthetic anti-inflammatory drugs are not readily available and have adverse side effects. Alternative herbal medicines possess bioactive compounds that are safer and efficient in the management of various diseases and disorders. The present study evaluated for the anti-inflammatory activity of methanolic extracts of *Kigelia africana* and *Acacia hockii* in mice to scientifically validate their traditional use among the Embu and Mbeere communities in Kenya. The plant samples were collected with the help of local herbalists in Embu County, Kenya and transported to Kenyatta University biochemistry and biotechnology laboratories for cleaning, air drying, milling, and extraction. Swiss albino mice of either sex were randomly divided into six groups of 5 animals each; normal control, negative control, positive control and three experimental groups. The anti-inflammatory activity was tested using carrageenan-induced hind paw edema method. The anti-inflammatory activity of the extracts was compared to reference drug diclofenac. The leaf extract of *K. africana* reduced inflamed hind paw diameter of mice by between 0.21%- 4.98% while the stem bark extract of *A. hockii* reduced inflamed hind paw diameter by between 0.6%-5.38%. The diclofenac reduced inflamed hind paw diameter by between 1.11%-4.9%. The qualitative phytochemical screening indicated the presence of saponins, flavonoid, alkaloids, terpenoids, phenolics, and cardiac glycosides. The present study demonstrated potent anti-inflammatory activities of methanolic extracts of *K. africana* and *A. hockii* in a dose-dependent manner, which supports their traditional use. The present study, therefore, recommends the ethnomedicinal use of *K. africana* and *A. hockii* in the management of inflammation.

**Keywords:** *Kigelia africana*; *Acacia hockii*; Anti-inflammatory; PGE<sub>2</sub>; Paw edema

### Introduction

Inflammation refers to body's normal protective response to tissue injury caused by physical trauma, toxic chemicals or microbiological agents [1]. The classical signs of inflammation are skin redness, swelling, pain, heat, and loss of function [2]. The process of inflammation involves changes in blood flow, destruction of tissues, increased vascular permeability and the synthesis of pro-inflammatory mediators [3]. The injured cells, lymphocytes, phagocytes, mast cells and blood proteins are the sources of inflammatory mediators. The most important inflammatory mediators include bradykinins, serotonin, histamine, tumor necrosis factor- $\alpha$ , interleukin-6, interleukin-1 $\beta$ , leukotrienes, phospholipase A<sub>2</sub>, nitric oxide (NO), lipoxygenases and cyclooxygenase 2 (COX-2) [3,4].

Inflammatory process has two phases: acute and chronic. The acute inflammation occurs a few minutes after tissue damage. It is characterized by an increase in permeability of blood vessels, extravasation of fluid and proteins and accumulation of white blood cells for a short period [5]. The primary mediators of acute inflammation include histamine, serotonin, and COX-2 [6]. The failure of the management of acute inflammation and an autoimmune response to a self-antigen lead to chronic inflammation and disease [7]. Chronic inflammation is mediated by inflammatory mediators such as PGE<sub>2</sub>, nitric oxide and lipoxygenases. Chronic inflammation

may result in ailments such as chronic peptic ulcers, rheumatoid arthritis, systemic lupus, asthma, chronic periodontitis and cancer [8].

During the inflammatory response, the PGE<sub>2</sub> are at low levels in tissues with no inflammation and increase immediately in acute inflammation. As immune cells infiltrate the tissues, further increases in PGE<sub>2</sub> levels is observed [9]. The non-steroidal anti-inflammatory drugs (NSAIDs) such as naproxen, indomethacin, ibuprofen, diclofenac, and ketoprofen are the most commonly used conventional medicinal products in the treatment of inflammation [10]. The NSAIDs inhibit the expression of cyclooxygenase 2 (COX-2) enzyme responsible for the production of PGE<sub>2</sub> which induces pyrexia [11]. However, the prolonged use of NSAIDs is linked with severe effects on the gastrointestinal tract, kidney, and cardiovascular system [12].

The demand for herbal medicine is increasing due to the growing recognition of natural products having fewer side effects, easily available, better cultural acceptability and being comparatively affordable [13]. *Kigelia africana* (Lam) Benth and *Acacia hockii* De Wild are used traditionally to manage inflammation among Embu and Mbeere communities in Embu County Kenya but lack validated scientific data to confirm their use [14]. The present study was, therefore, designed to evaluate for the anti-inflammatory potential of the two extracts to act as a preliminary step towards the development of more efficient plant-derived anti-inflammatory agents.

## Materials and Methods

### Collection and preparation of plant materials

The fresh leaves of *K. africana* and stem bark of *A. hockii* were collected in Mbeere North sub-county, Embu County, Kenya with the help of local herbalists in August, 2015. The plant samples were availed to an acknowledged taxonomist for botanical authentication and a sample voucher deposited at Kenyatta University herbarium. The plant samples were transported in polyethene bags to biochemistry and biotechnology laboratories at Kenyatta University, where they were sorted out, cleaned with tap water and rinsed with distilled water. The plant materials were separately chopped into small pieces, and air dried at room temperature until dry. The dried sample materials were ground into fine homogenous powder using an electric mill.

### Extraction

For each sample, 400 grams of powder was soaked in 2 litres of methanol, stirred for four hours and left for 48 hours. The extracts were then filtered using Whatman's No.1 filter paper. The filtrate was concentrated to dryness under reduced pressure using a rotary evaporator at a maximum temperature of 64°C. The concentrate was put in an airtight container and stored at 40°C until use in the bioassay.

### Experimental design

**Laboratory animals:** Swiss albino aged between 2-3 months and weighing between 19-25 grams were used in this study. The animals breeding colonies were acquired and bred in the animal breeding and experimentation facility in the department of biochemistry and biotechnology, Kenyatta University. The animals were allowed to acclimatize for two days prior to experimentation. The experimental animals were kept in the standard cages in the animal house maintained under standard laboratory condition of an ambient temperature of 25°C with 12 hours daylight and 12 hours darkness cycles. The experimental animals were fed on standard rodent pellets and provided with water ad libitum [15].

**Evaluation of anti-inflammatory activity:** Thirty Swiss albino mice of either sex were divided randomly into six groups of five mice each and treated as follows; Group I (normal control) was not induced with inflammation but received 4% DMSO. Group II (negative control) was induced with inflammation and received 4% DMSO. Group III (positive control) was induced with inflammation and received diclofenac (reference drug) at a dose of 15 mg/kg body weight. Groups IV, V and VI (experimental groups) were induced with inflammation and received the extracts at the dose levels of 50 mg/kg, 100 mg/kg and 150 mg/kg body weight. This design is summarized in Table 1.

The anti-inflammatory activity of the extracts was assessed using carrageenan-induced right paw edema in mice as described by Winter. Acute inflammation was induced by sub-plantar injection of 0.05 ml 1% carrageenan (sigma-type I) in normal saline 30 minutes after treatment. The change in paw diameter was measured using a digital vernier caliper 30 minutes before injection of carrageenan and at 1, 2, 3 and 4 hours after induction of inflammation [17]. The percentages inhibition in inflammation was calculated using the formula described by Uma mageswari A [18], as follows;

$$\text{Inhibition (\%)} = \frac{C_t - T_t}{C_t} \times 100 \text{ Where,}$$

Ct=Paw diameter at 1 hour after carrageenan administration

Tt=Paw diameter after Treatment

Group	Status	Treatment
I	Control	DMSO
II	Negative control	Carrageenan+DMSO
III	Positive control	Carrageenan+15 mg/kg/bw diclofenac
IV	Experimental group A	Carrageenan+DMSO+50 mg/kg bw extract
V	Experimental group B	Carrageenan+DMSO+100 mg/kg bw extract
VI	Experimental group C	Carrageenan+DMSO+150 mg/kg bw extract

**Table 1:** Treatment protocol for evaluation of anti-inflammatory activities of methanolic extracts of *Kigelia africana* (Lam) Benth and *Acacia hockii* De Wild in Swiss albino mice; Carrageenan=1%, DMSO=4%.

### Qualitative phytochemical screening

The plant extracts were subjected to qualitative phytochemical screening to identify the absence or the presence of various phytochemicals using methods of analysis described by Ref. [19,20]. Phytochemicals tested include alkaloids, terpenoids, saponins, flavonoids, phenolics, cardiac glycosides and steroids. These phytochemicals are reported to possess anti-inflammatory activity [21].

### Data management and statistical analysis

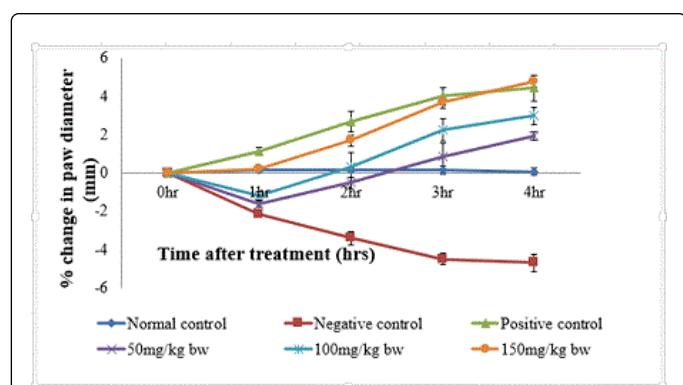
Paw diameter measured was recorded and tabulated on spreadsheet. The data was exported to Minitab statistical software version 17.0 (State College, Pennsylvania) for analysis. The data was subjected to descriptive statistics and expressed as mean  $\pm$  standard error of mean (SEM). One-way analysis of variance (ANOVA) was used to determine whether there was any significant difference between the means of different groups. This was followed by Turkey's tests to separate means and obtain the specific significant differences among the various treatment groups. Unpaired student t-test was done to compare between the mean activities of the two extracts. The values of  $p \leq 0.05$  were considered significant. The data was presented in tables and graphs.

## Results

### Anti-inflammatory activity of methanolic leaf extract of *Kigelia africana* (Lam.) Benth on carrageenan-induced inflammation in Swiss albino mice

The methanolic leaf extract of *K. africana* showed significant anti-inflammatory activity on carrageenan-induced paw edema, which was demonstrated by the reduction in inflamed hind paw diameter after extract administration (Figure 1; Table 2). In the first hour after treatment, the leaf extract of *K. africana* at the dose level of 150 mg/kg and the diclofenac (reference drug) at the dosage of 15 mg/kg body weight showed anti-inflammatory effect by reducing hind paw diameter by 0.21% and 1.10% respectively (Figure 1; Table 2). However,

the extract at the dose levels of 50 mg/kg and 100 mg/kg body weight never showed anti-inflammatory activity at this hour (Figure 1; Table 2). The anti-inflammatory activity of the extract at the dose levels of 50 mg/kg, 100 mg/kg and 150 mg/kg body weight showed no significant difference and were comparable to normal control ( $p>0.05$ ; Table 2). In addition, the anti-inflammatory activity of extract at the dose level of 150 mg/kg body weight was comparable to aspirin (reference drug) ( $p>0.05$ ; Table 2).



**Figure 1:** Anti-inflammatory effects of methanolic leaf extract of *Kigelia africana* (Lam) Benth on carrageenan-induced inflammation in Swiss albino mice.

In the second hour, the leaf extract of *A. hockii* at the dosage of 100 mg/kg and 150 mg/kg body weight as well as the diclofenac (reference

drug) reduced inflamed paw diameter by 0.42%, 1.42% and 2.8% respectively (Figure 1; Table 2). However, the extract at the dosage of 50 mg/kg body weight never showed anti-inflammatory activity at this hour (Figure 1; Table 2). The anti-inflammatory activity of the extract at the dose level of 50 mg/kg and 100 mg/kg showed no significant difference ( $p>0.05$ ; Table 2). Besides, the anti-inflammatory activity of the extract at the dose level of 150 mg/kg body weight was comparable to diclofenac (reference drug) ( $p>0.05$ ; Table 2).

In the third hour after treatment, the extract at the dose levels of 50 mg/kg, 100 mg/kg, and 150 mg/kg body weight, as well as the diclofenac (reference drug) reduced the inflamed hind paw diameter by 0.86%, 2.25%, 3.41% and 4.02% respectively (Figure 1; Table 2). The anti-inflammatory activity of the extract at the dosage of 50 mg/kg and 100 mg/kg and 150 mg/kg body weight showed no significant difference and were comparable to diclofenac (reference drug) ( $p>0.05$ ; Table 2).

In the fourth hour after treatment, the leaf extract of *K. africana* at the dose levels of 50 mg/kg, 100 mg/kg, and 150 mg/kg body weight reduced inflamed hind paw diameter by 1.95%, 2.98% and 4.98% respectively (Figure 1; Table 2). Similarly, the diclofenac (reference drug) reduced the inflamed paw diameter by 4.43% at this hour (Figure 1; Table 2). The anti-inflammatory activity of the extract at the dosages of 50 mg/kg and 100 mg/kg body weight showed no significant difference ( $p>0.05$ ; Table 2). In addition, the antipyretic activity of the extract at the dose level of 150 mg/kg body weight was comparable to diclofenac (reference drug) ( $p>0.05$ ; Table 2).

Group	Treatment	% change in paw diameter (mm) after treatment				
		0 hr	1 hr	2 hr	3 hr	4 hr
Normal Control	10% DMSO	100 ± 0.00 (0.00)	99.84 ± 0.10b (0.16)	99.84 ± 0.10b (0.16)	99.84 ± 0.10b (0.16)	99.93 ± 0.07b (0.07)
		100 ± 0.00 (0.00)	102.13 ± 0.20a(2.13)	103.61 ± 0.08a (3.61)	104.60 ± 0.30a (4.6)	104.95 ± 0.48a (4.95)
Positive Control	Carrageenan +diclofenac+DMSO	100 ± 0.00 (0.00)	98.89 ± 0.22c(1.11)	97.20 ± 0.53e (2.8)	95.98 ± 0.44bc (4.02)	95.57 ± 0.47de(4.43)
Methanolic Extracts	Carrageenan+50 mg/kg bw +DMSO	100 ± 0.00 (0.00)	101.74 ± 0.15b (1.74)	100.37 ± 0.26bc (0.37)	99.14 ± 0.18b (0.86)	98.05 ± 0.21c (1.95)
	Carrageenan+100 mg/kg bw +DMSO	100 ± 0.00 (0.00)	101.10 ± 0.25b (1.1)	99.58 ± 0.77cd (0.42)	97.75 ± 0.60b (2.25)	97.02 ± 0.45cd (2.98)
	Carrageenan+150 mg/kg bw +DMSO	100 ± 0.00 (0.00)	99.79 ± 0.14bc (0.21)	98.59 ± 0.25de (1.42)	96.59 ± 0.26b (3.41)	95.02 ± 0.20e (4.98)

**Table 2:** Effects of intraperitoneal administration of methanolic leaf extract of *Kigelia africana* (Lam) Benth on carrageenan-induced inflammation in Swiss albino mice. Values expressed as Mean ± SEM for five animals per group. Values with the same superscript letter are not significantly different by one-way ANOVA followed by Turkey's test ( $p>0.05$ ). Percentage reduction in rectal temperature is given within parenthesis. Carrageenan=1%; 15 mg/kg bw diclofenac and 4% DMSO.

### Anti-inflammatory activity of methanolic stem bark extract of *Acacia hockii* De Wild on carrageenan-induced inflammation in Swiss albino mice

The methanolic stem bark extract of *A. hockii* demonstrated anti-inflammatory activity on carrageenan-induced paw edema in mice, which was indicated by the reduction in paw edema after extract

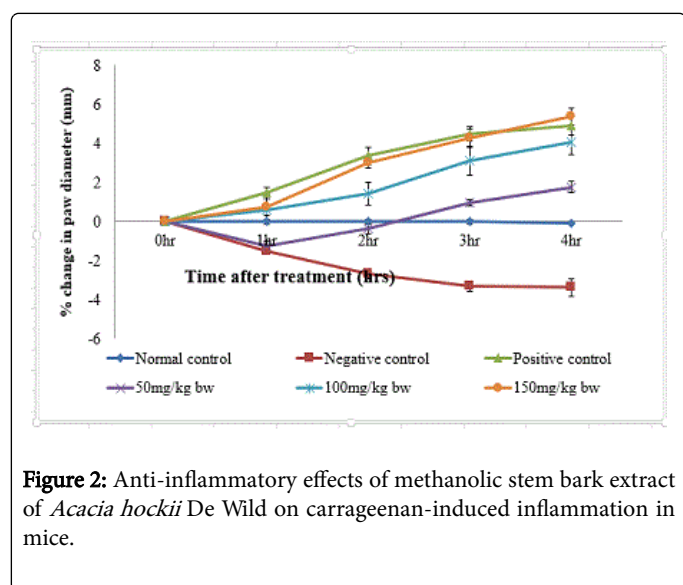
administration (Figure 2 and Table 3). In the first hour after treatment, the stem bark extract of *A. hockii* at the dose levels of 100 mg/kg and 150 mg/kg body weight as well as the diclofenac (reference drug) at the dosage of 15 mg/kg body weight reduced inflamed hind paw diameter by 0.6%, 0.77% and 1.48% respectively. However, the extract at the dose level of 50 mg/kg body weight never showed anti-inflammatory activity at this hour (Figure 2; Table 3).

Group	Treatment	% change in paw diameter (mm) after treatment				
		0 hr	1 hr	2 hr	3 hr	4 hr
Normal Control	DMSO	100 ± 0.00(0.00)	99.99 ± 0.12 <sup>bc</sup> (0.01)	99.99 ± 0.12 <sup>bc</sup> (0.01)	99.99 ± 0.12 <sup>b</sup> (0.01)	100.08 ± 0.08 <sup>b</sup> (0.08)
Negative Control	Carrageenan+DMSO	100 ± 0.00(0.00)	101.49 ± 0.17 <sup>a</sup> (1.49)	102.66 ± 0.26 <sup>a</sup> (2.66)	103.30 ± 0.26 <sup>a</sup> (3.30)	103.37 ± 0.45 <sup>a</sup> (3.37)
Positive Control	Carrageenan +Diclofenac +DMSO	100 ± 0.00(0.00)	98.52 ± 0.28 <sup>d</sup> (1.48)	96.63 ± 0.44 <sup>d</sup> (3.37)	95.50 ± 0.36 <sup>c</sup> (4.5)	95.10 ± 0.47 <sup>c</sup> (4.9)
Methanolic Extracts	Carrageenan +50 mg/kg bw+DMSO	100 ± 0.00(0.00)	101.24 ± 0.24 <sup>ab</sup> (1.24)	100.35 ± 0.25 <sup>b</sup> (0.35)	99.04 ± 0.17 <sup>b</sup> (0.96)	98.22 ± 0.29 <sup>b</sup> (1.78)
	Carrageenan +100 mg/kg bw+DMSO	100 ± 0.00(0.00)	99.40 ± 0.56 <sup>cd</sup> (0.6)	98.57 ± 0.58 <sup>c</sup> (1.43)	96.89 ± 0.74 <sup>c</sup> (3.11)	95.93 ± 0.63 <sup>c</sup> (4.07)
	Carrageenan +150 mg/kg bw+DMSO	100 ± 0.00(0.00)	99.23 ± 0.41 <sup>cd</sup> (0.77)	96.97 ± 0.28 <sup>d</sup> (3.03)	95.72 ± 0.47 <sup>c</sup> (4.28)	94.62 ± 0.41 <sup>c</sup> (5.38)

**Table 3:** Effects of intraperitoneal administration of methanolic stem bark extracts of *Acacia hockii* De Wild on carrageenan-induced inflammation in Swiss albino mice. Values expressed as Mean ± SEM for five animals per group. Values with the same superscript letter are not significantly different by one-way ANOVA followed by Turkey's test (p>0.05). Percentage reduction in rectal temperature is given within parenthesis. Carrageenan=1%; 15 mg/kg bw diclofenac and 4% DMSO.

The anti-inflammation activity of the extract at the dosage of 100 mg/kg and 150 mg/kg body weight showed no significant difference and was comparable to diclofenac (reference drug) (P>0.05; Table 3). Besides, the anti-inflammatory activity of the extract at the dose level of 50 mg/kg body weight was significantly different (p<0.05; Table 4) from 100 mg/kg and 150 mg/kg body weight and comparable to negative control (p<0.05; Table 3).

3.03% and 3.37% respectively (Figure 2; Table 3). However, the extract at the dose level of 50 mg/kg body weight never showed the anti-inflammatory effect at this hour (Figure 2; Table 3). The anti-inflammatory activity of the extract of at the dose levels of 50 mg/kg, 100mg/kg and 150 mg/kg body weight were significantly different (p<0.05; Table 3). However, the anti-inflammatory activity of the extract of at the dosage of 150 mg/kg body weight was comparable to the diclofenac (reference drug) (p>0.05; Table 3).



**Figure 2:** Anti-inflammatory effects of methanolic stem bark extract of *Acacia hockii* De Wild on carrageenan-induced inflammation in mice.

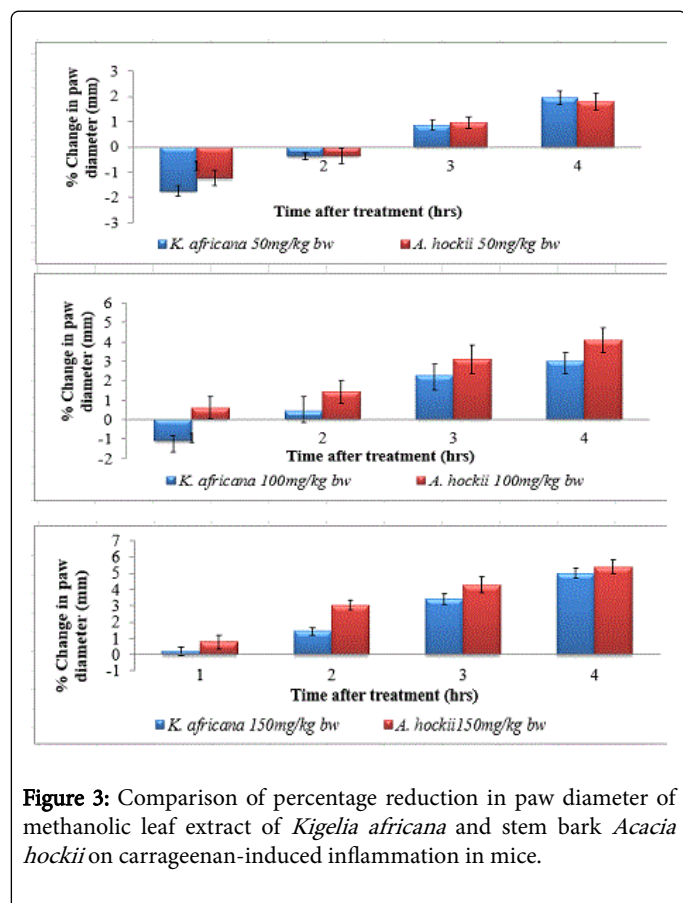
In the second hour after treatment, the extract at the dose levels of 100 mg/kg and 150 mg/kg body weight, as well as the diclofenac (reference drug) reduced inflamed paw diameter of mice by 1.43% and

In the third hour after treatment, the extract at dose levels of 50 mg/kg, 100 mg/kg and 150 mg/kg body weight, as well as the diclofenac (reference drug) reduced the inflamed hind paw diameter by 0.96%, 3.11%, 4.28% and 4.5% respectively (Figure 2; Table 3). The anti-inflammatory activity of the extract at the dosage of 100 mg/kg and 150 mg/kg body weight demonstrated no significant difference and were comparable to diclofenac (reference drug) (p>0.05; Table 3). However, the anti-inflammatory activity of the extract at the dose level of 50 mg/kg body weight was significantly different from 100 mg/kg and 150 mg/kg body weight (p<0.05; Table 3) and comparable to normal control (p>0.05; Table 3).

In the fourth hour, the extract at the dose levels of 50 mg/kg, 100 mg/kg and 150 mg/kg body weight, as well as the diclofenac reduced inflamed hind paw diameter by 1.78%, 4.05%, 5.38% and 4.9% respectively (Figure 2; Table 3). The anti-inflammatory activity of the extract at the dosage of 100 mg/kg and 150 mg/kg showed no significant difference and were comparable to aspirin (reference drug) (p>0.05; Table 3). However, the anti-inflammatory activity of the extract at the dose level of 50 mg/kg body weight was significantly different from 100 mg/kg and 150 mg/kg body weight (p<0.05; Table 3) and comparable to normal control (p>0.05; Table 3).

### Comparison between the anti-inflammatory activities of *Kigelia africana* and *Acacia hockii*

In comparison, the anti-inflammatory activity of the two extracts at the dose level of 50 mg/kg body weight in the 1, 2, 3 and 4 hours of the test period was not significantly different with p values of 0.12, 0.96, 0.70 and 0.64 respectively ( $p > 0.05$ ). The anti-inflammatory activity of the two extracts at the dose level of 100 mg/kg body weight was significantly different in the first hour after treatment with p values of 0.04 ( $p < 0.05$ ). However, the anti-inflammatory activity of the two extracts at the dosage of 100 mg/kg body weight in the 2, 3 and 4 hours was not significantly different with p values of 0.32, 0.40 and 0.20 respectively ( $p > 0.05$ ). The anti-inflammatory activity of the two extracts at the dose level of 150 mg/kg body weight in the 1, 3 and 4 hours of the test period showed no significant difference with p values of 0.27, 0.16 and 0.42 respectively ( $p > 0.05$ ). However, the anti-inflammatory activity of the two extracts the dose level of 150 mg/kg body weight in the 2 hour of the treatment period showed significant difference with p values of 0.004 ( $p < 0.05$ ). Both extracts were more efficient in the fourth hour at the dose level of 150 mg/kg body weight (Figure 3).



**Figure 3:** Comparison of percentage reduction in paw diameter of methanolic leaf extract of *Kigelia africana* and stem bark *Acacia hockii* on carrageenan-induced inflammation in mice.

### Qualitative phytochemical screening

The qualitative phytochemical screening of the methanolic leaf extract of *K. africana* showed the presence of flavonoids, phenolics, cardiac glycosides, steroids, and terpenoids (Table 4). However, the methanolic stem bark extract of *A. acacia* demonstrated the presence of alkaloids, cardiac glycosides, flavonoids, saponins, phenolics, steroids, and terpenoids (Table 4).

Phytochemical	Leaf <i>Kigelia africana</i>	Stem bark <i>Acacia hockii</i>
Alkaloids	-	+
Flavonoids	+	+
Steroids	+	+
Saponins	-	+
Terpenoids	+	+
Cardiac glycosides	+	+
Phenolics	+	+

**Table 4:** Qualitative phytochemical composition of methanolic leaf extract of *Kigelia africana* and stem bark extract of *Acacia hockii*. Present phytochemical is denoted by (+) sign; absent phytochemical is indicated by (-) sign.

### Discussion

The present study evaluated for the anti-inflammatory (antiphlogistic) activity of methanolic leaf extract of *Kigelia africana* (Lam.) Benth and methanolic stem bark extract of *Acacia hockii* De Wild on carrageenan-induced paw edema in mice. Carrageenan, dextran, arachidonic acid, dextran, histamine, serotonin and formalin-induced paw edema; cotton pellet induced granuloma; Freund's adjuvants are the standard agents for causing acute, sub-acute and chronic inflammation respectively in animal models [22-25].

Carrageenan is a natural carbohydrate obtained from edible red seaweeds [26]. It is widely used to induce acute inflammation in experimental animals [27] and hence the choice in the present study. A freshly prepared solution of 1% carrageenan in normal saline as an intraplantar injection at a dose of 50-150  $\mu$ l is commonly used to induce inflammation [28].

The carrageenan-induced inflammation is described as a biphasic event in which various mediators operates to produce an inflammatory response [29]. The first mediators detectable in the early phase (1 hour) include histamine, serotonin, and cyclooxygenase. On the other hand, the late phase (over 1 hour) is sustained by the production of PGE2 and it is mediated by bradykinin and leukotrienes [6, 30]. Inducible nitric oxide synthase (iNOS) and COX-2 enzyme are responsible for the production of an enormous amount of inflammatory mediators [26, 31]. Carrageenan-induced inflammation is also associated with enhanced levels of the endogenous pyrogenic cytokines such as tumor necrosis factor and interleukins (IL-1 and IL-6) which act as pro-inflammatory mediators [32].

The evaluation of anti-inflammatory activity of methanolic leaf extracts of *K. africana* and stem bark extract of *A. hockii* demonstrated a significant anti-inflammatory activity on carrageenan-induced paw edema in Swiss albino mice (Figure 1 and 2; Tables 2 and 3). These findings were consistent with other studies carried out on herbal medicines using animal models. A similar study carried out by [33], demonstrated a significant anti-inflammatory activity of dichloromethane: methanolic leaf extracts of *Caesalpinia volkensii* and *Maytemus obscura* on carrageenan-induced paw edema in mice. Similarly, methanolic stem bark extract of *Tinospora cordifolia* Wild, fruits of *Emblca officinalis* and rhizomes of *Cyperus rotundus* Linn demonstrated anti-inflammatory effect in rodents [34].

The NSAIDs such as diclofenac, ibuprofen, indomethacin, naproxen and acetaminophen are commonly prescribed in the medication of inflammation [35]. The NSAIDs block the enzyme cyclooxygenase 2 (COX-2) which stimulate biosynthesis PGE<sub>2</sub>. There are two types COX enzymes; COX-1, and COX-2. The COX-2 produces prostaglandins that promote inflammation while the COX-1 produces prostaglandins that support platelets and protect the stomach [36]. The NSAIDs less inhibit the initial phase of carrageenan-induced paw edema, and this is attributed to the release of histamine, serotonin and bradykinin. However, the second phase is attributed to the induction of inducible COX-2 and can be blocked using NSAIDs [37]. It is therefore believed that the anti-inflammatory activities of leaf extract of *K. africana* and stem bark extract of *A. hockii*, inhibited the synthesis and release of prostaglandins to manage edema.

The present study used a dose range of 50 mg/kg, 100 mg/kg, and 150 mg/kg body weight to test for the anti-inflammatory activity of the extracts in mice. A similar study carried out by [38], evaluated for the anti-inflammatory activity of *Cissus quadrangularis* using a dose range of 50 mg/kg, 100 mg/kg, and 150 mg/kg body weight in rats. Similarly, a study carried out on the anti-inflammatory activity of leaf extract of *Musanga cecropioides* used a dose range of 50 mg/kg, 100 mg/kg, 150 mg/kg, and 200 mg/kg [39].

The methanolic leaf extract of *K. africana* and stem bark extract of *A. hockii* demonstrated dose-dependent response on carrageenan-induced paw edema in mice (Figure 1 and 2; Table 2 and 3). These findings were in agreement with a study carried out on the anti-inflammatory properties of dichloromethane: methanolic extracts of *Caesalpiina volkensii* Harms and *Maytemus obscura* in mice [33]. Similarly, another study carried out on the evaluation of anti-inflammatory activity of *Strophanthus hispidus* in experimental animals, showed a dose-dependent manner [40]. Furthermore, another study carried out by [41], on anti-inflammatory properties of *Terbium chamaedrys* showed a dose-dependent response in animal models.

The methanolic leaf extract of *K. africana* and stem bark extract of *A. hockii* showed minimal anti-inflammatory activities at lower dose levels of 50 and 100 mg/kg body weight compared to 150 mg/kg body weight (Figure 1 and 2; Table 2 and 3). The reference drug (diclofenac) achieved its maximum anti-inflammatory activity in the third hour (Figure 1 and 2; Table 2 and 3); its activity decreased subsequently probably due to metabolism and excretion of the drugs. The maximum anti-inflammatory activity of methanolic leaf extracts of *K. africana* and stem bark extract *A. hockii* was achieved at the dosage of 150 mg/kg body weight in the fourth hour (Figure 1 and 2; Table 2 and 3), indicating slow but steady passive diffusion of the bioactive constituent's across the cell membrane in the peritoneal cavity [42].

The methanolic leaf extract of *K. africana* and stem bark extract of *A. hockii* at different dose levels did not reduce paw diameter in the first and second hours compared to the third and fourth hours (Figure 1 and 2; Table 2 and 3). However, the extract of *K. africana* and stem bark extract of *A. hockii* at the dose of 150 mg/kg body weight was more effective in the fourth hour of treatment compared to diclofenac (reference drug) in the same hour (Figure 1 and 2; Table 2 and Table 3). These findings showed that the leaf extract of *K. africana* and stem bark extract *A. hockii* were able to inhibit the synthesis of prostaglandins more than the conventional drug diclofenac (Figure 1 and 2; Table 2 and Table 3).

The antiphlogistic activity of methanolic leaf extract of *K. africana* and stem bark extract of *A. hockii*, could be due to the presence of

bioactive constituents that exhibit anti-inflammatory action. This could be through inhibition of inflammatory mediators such as prostaglandins, histamine, serotonin and lysosome [43]. The qualitative phytochemical screening of methanolic leaf extract of *K. africana* indicated the presence of flavonoids, steroids, terpenoids, cardiac glycosides and phenolics while the stem bark extract of *A. hockii* indicated the presence of flavonoids, alkaloids, steroids, saponins, terpenoids, cardiac glycosides and phenolics (Table 4). The presence of some these bioactive compounds such as alkaloids, flavonoids, terpenoids, and steroids have shown to exhibit antiphlogistic activity in experimental animals [44, 45].

Flavonoids have been reported to inhibit pro-inflammatory mediators such as TNF- $\alpha$  and phospholipase A<sub>2</sub> [44]. Furthermore, some flavonoids respond by blocking both the cyclooxygenase and lipoxygenase pathways of the arachidonate cascade at relatively high concentration while at the lower level only the lipoxygenase pathway is blocked [46]. Research findings have revealed that triterpenoids suppresses some function of macrophages, neutrophils and also inhibit nitric oxide (NO), NF- $\kappa$ B signaling and PGE<sub>2</sub> production responsible for inflammation induction [47]. The NF- $\kappa$ B can detect noxious stimuli, such as infectious agents, cellular injuries and free radicals, and then directs DNA to synthesize inflammatory cytokines. Thus, their inhibition leads to management of edema [48]. Steroids also attenuate inflammation by inhibiting phospholipase A<sub>2</sub>, which hydrolyzes arachidonic acid from membrane phospholipids and subsequent formation of prostanoids and leukotrienes via the cyclooxygenase and lipoxygenase pathways.

## Conclusions

The methanolic leaf extract of *K. africana* and stem bark extract of *A. hockii* showed potent anti-inflammatory activity on carrageenan-induced paw edema in mice. The anti-inflammatory activity of leaf extract of *K. africana* and stem bark extract of *A. hockii* demonstrated a dose-dependent response and were comparable to diclofenac (reference drug). The extracts were most active at the dose level of 150 mg/kg body weight in the fourth hour of treatment.

The extracts of *K. africana* and *A. hockii* could, therefore, be an alternative bio-resource for generating anti-inflammatory agents. However, further studies are necessary to elucidate the mechanism behind this effect and their active compounds. The present study, therefore, scientifically confirms and supports the traditional use of *K. africana* and *A. hockii* in the management of inflammation.

## References

1. Calixto JB, Campos MM, Otuki MF, Santos ARS (2004) Anti-inflammatory compounds from plant origin. Part II. Modulation of pro-inflammatory cytokines, chemokines and adhesion molecules. *Planta Medica* 70: 93-103.
2. Hurley JV (1972) *Acute inflammation*. Edinburgh, London: Churchill Livingstone.
3. Shah BN, Patel NP, Pandya P (2008) Role of leukotriene in inflammation and antileukotriene therapy. *Journal of Pharmacy Research* 1: 113-123.
4. Williams JG, Maier RV (1992) The inflammatory response. *Journal of Intensive Care Medicine* 7: 53-66.
5. Posadas I, Bucci M, Roviezzo F, Rossi A, Parente L, et al. (2004) Carrageenan-induced mouse paw edema is biphasic, age-weight dependent and displays differential nitric oxide cyclooxygenase-2 expression. *British Journal of Pharmacology* 142: 331-338.

6. Ravi V, Saleem TM, Patel SS, Raamamurthy J, Gauthaman K (2009) Anti-inflammatory effect of methanolic extract of *Solanum nigrum* Linn berries. *International Journal of Applied Research in Natural Products* 2: 33-36.
7. Recio MC, Andujar I, Rios JL (2012) Anti-inflammatory agents from plants: progress and potential. *Curr Med Chem* 19: 2088-2103.
8. Nordqvist C (2015) Inflammation: Causes, Symptoms and Treatment. *Medical News Today*. Tilley L, Coffman TM, Koller BH (2001) Mixed messages: modulation of inflammation and immune responses by prostaglandins and thromboxanes. *Journal of Clinical Investigation* 108: 15-24.
9. Tilley SL, Coffman TM, Koller BH (2001) Mixed messages: modulation of inflammation and immune responses by prostaglandins and thromboxanes. *Journal of Clinical Investigation* 108: 15-24.
10. Warden SJ (2010) Prophylactic use of NSAIDs by athletes: a risk/benefit assessment. *Phys Sportsmed* 38: 132-138.
11. Vane J, Botting R (1987) Inflammation and the mechanism of action of anti-inflammatory drugs. *FASEB J* 1: 89-96.
12. Traversa G, Walker AM, Ippolito FM, Caffari B, Capurso L, et al. (1995) Gastroduodenal toxicity of different nonsteroidal anti-inflammatory drugs. *Epidemiology* 6: 49-54.
13. Kamboj VP (2000) Herbal medicine. *Current Science Bangalore* 78: 35-38.
14. Kareru PG, Kenji GM, Gachanja AN, Keriko JM, Mungai G (2006) Traditional medicines among the Embu and Mbeere peoples of Kenya. *Afr J Tradit Complement Altern Med* 4: 75-86.
15. Vogel HG (2002) Drug discovery and evaluation pharmacological assays. Springer-Verlag Berlin Heidelberg New York 1408: 2-716.
16. Winter CA, Risley EA, Nuss GW (1962) Carrageenin-induced edema in hind paw of the rat as an assay for antiinflammatory drugs. *Proc Soc Exp Biol Med* 111: 544-547.
17. Bamgbose SO, Noamesi BK (1981) Studies on cryptolepine. II: Inhibition of carrageenan induced oedema by cryptolepine. *Planta Med* 41: 392-396.
18. Umamageswari A, Kudagi BL (2015) Anti-inflammatory and analgesic properties of *Ocimum sanctum*: a comparative study using animal models. *International Journal of Basic and Clinical Pharmacology* 4: 981-986.
19. Harbone JB (1998) Phytochemical methods: A guide to modern techniques of plant analysis. Chapman and Hal Publishers, London, United Kingdom 3: 60-66.
20. Kotake CK (2000) Practical Pharmacognosy. Vallabh Prakashan, New Delhi 4: 107-111.
21. Bhagyasri Y, Lavakumar V, Divya Sree MS, Ashok Kumar CK (2015) An overview on anti-inflammatory activity of Indian herbal plants. *International Journal of Research and Pharmaceutical Nano Science* 4: 1-9.
22. Ismail TS, Gopalakrishnan S, Begum VH, Elango V (1997) Anti-inflammatory activity of *Salacia oblonga* Wall. and *Azima tetracantha* Lam. *J Ethnopharmacol* 56: 145-152.
23. Mujumdar AM, Misar AV (2004) Anti-inflammatory activity of *Jatropha curcas* roots in mice and rats. *J Ethnopharmacol* 90: 11-15.
24. Lai SC, Peng WH, Huang SC, Ho YL, Huang TH, et al. (2009) Analgesic and anti-inflammatory activities of methanol extract from *Desmodium triflorum* DC in mice. *The American Journal of Chinese Medicine* 37: 573-588.
25. Kolawole OT, Akiibinu MO, Ayankunle AA, Awe EO (2013) Evaluation of anti-inflammatory and antinociceptive potentials of *Khaya senegalensis* A. Juss (Meliaceae) stem bark aqueous extract. *British Journal of Medicine and Medical Research* 3: 216-229.
26. Necas J, Bartosikova L (2013) Carrageenan: a review. *Veterinari Medicina* 58: 187-205.
27. Paschapur MS, Patil MB, Kumar R, Patil SR (2009) Evaluation of anti-inflammatory activity of ethanolic extract of *Borassus flabellifer* L. male flowers (inflorescences) in experimental animals. *Journal of Medicinal Plants Research* 3: 049-054.
28. Estakhr J, Sanchooli N, Najafi SH, Javdan N (2011) Anti-Inflammatory Activity of Ethanolic Extract of *Physalis alkekengi*. *Research Journal of Pharmaceutical, Biological and Chemical Sciences* 2: 421-425.
29. Gupta M, Mazumder UK, Gomathi P, Selvan VT (2006) Antiinflammatory evaluation of leaves of *Plumeria acuminata*. *BMC Complement Altern Med* 6: 36.
30. Unnisa A, Parven TD (2011) Anti-inflammatory and acute toxicity studies of the extracts from the rhizomes of *Alpinia galanga* Willd. *Der Pharmacia Sinica* 2: 361-367.
31. Handy RL, Moore PK (1998) A comparison of the effects of L-NAME, 7-NI and L-NIL on carrageenan-induced hindpaw oedema and NOS activity. *Br J Pharmacol* 123: 1119-1126.
32. Cuzzocrea S, Sauterin L, De Sarro G, Costantino G, Rombolà L, et al. (1999) Role of IL-6 in the pleurisy and lung injury caused by carrageenan. *J Immunol* 163: 5094-5104.
33. Mwangi BM, Gitahi SM, Njagi JM, Mworio JK, Aliyu U, et al. (2015) Anti-inflammatory Properties of Dichloromethane: Methanolic Leaf Extracts of *Caesalpinia Volkensii* and *Maytenus Obscura* in Animal Models. *International Journal of Current Pharmaceutical Research* 7: 83-87.
34. Mradu G, Dailya B, Arup M (2013) Studies Of Anti Inflammatory, Antipyretic And Analgesic Effects Of Aqueous Extract Of Traditional Herbal Drug Of Rodents. *International Research Journal of Pharmacy* 4: 113-120.
35. Fiorucci S, Santucci L, Cirino G, Mencarelli A, Familiari L, et al. (2000) IL-1 $\beta$  converting enzyme is a target for nitric oxide-releasing aspirin: new insights in the anti-inflammatory mechanism of nitric oxide-releasing non-steroidal anti-inflammatory drugs. *The Journal of Immunology* 165: 5245-5254.
36. Mitchell JA, Akarasereenont P, Thiemermann C, Flower RJ, Vane JR (1993) Selectivity of nonsteroidal antiinflammatory drugs as inhibitors of constitutive and inducible cyclooxygenase. *Proc Natl Acad Sci USA* 90: 11693-11697.
37. Nantel E, Denis D, Gordon R, Northey A, Cirino M, et al. (1999) Distribution and regulation of cyclooxygenase-2 in carrageenan-induced inflammation. *Br J Pharmacol* 128: 853-859.
38. Vijay P, Vijayvergia R (2010) Analgesic, anti-inflammatory and antipyretic activity of *Cissus quadrangularis*. *Journal of Pharmaceutical Science and Technology* 2: 64-71.
39. Sowemimo A, Okwuchuku E, Samuel FM, Ayoola O, Mutiat I (2015) Musanga cecropioides leaf extract exhibits anti-inflammatory and antinociceptive activities in animal models. *Revista Brasileira de Farmacognosia* 25: 506-512.
40. Agbaje EO, Fageyinbo MS (2012) Evaluating Anti-Inflammatory activity of aqueous root extracts of *Strophanthus hispidus* DC. (Apocynaceae). *International Journal of Applied Research in Natural Products* 4: 7-14.
41. Pourmotabbed A, Farshchi A, Ghiasi G, Malek Khatibi P (2010) Analgesic and anti-inflammatory activity of *Teucrium chamaedrys* leaves aqueous extract in male rats. *Iranian Journal of Basic Medical Sciences* 13: 119-125.
42. Hossain E, Mandal SC, Gupta JK (2011) Phytochemical screening and in-vivo antipyretic activity of the methanol leaf-extract of *Bombax malabaricum* DC (Bombacaceae). *Tropical Journal of Pharmaceutical Research* 10: 55-60.
43. Dina TA, Rahman MA, Ahmed NU, Uddin MN (2010) Analgesic and anti-inflammatory properties of *Argyrea argentea* methanol extract in animal model. *Journal of Taibah University for Science* 3: 1-7.
44. Bhaskar VH, Balakrishnan N (2009) Analgesic, anti-inflammatory and antipyretic activities of *Pergularia daemia* and *Carissa carandas*. *Journal of Pharmaceutical Sciences* 17: 168-174.
45. Di Carlo G, Mascolo N, Izzo AA, Capasso F (1999) Flavonoids: old and new aspects of a class of natural therapeutic drugs. *Life Sci* 65: 337-353.
46. Salminen A, Lehtonen M, Suuronen T, Kaarniranta K, Huuskonen J (2008) Terpenoids: natural inhibitors of NF-kappaB signaling with anti-inflammatory and anticancer potential. *Cell Mol Life Sci* 65: 2979-2999.

- 
47. Frantz B, Nordby EC, Bren G, Steffan N, Paya CV, et al. (1994) Calcineurin acts in synergy with PMA to inactivate I kappa B/MAD3, an inhibitor of NF-kappa B. *EMBO J* 13: 861-870.
48. Mencarelli A, Renga B, Palladino G, Distrutti E, Fiorucci S (2009) The plant sterol attenuates inflammation and immune dysfunction in murine models of inflammatory bowel disease. *Biochemical pharmacology* 78: 1214-1223.