

# Abundance of Monocyte Subsets Determines Susceptibility to Perinatal Hepatic Inflammation

Sarah Mohamedaly, Anas Alkhani, Amar Nijagal\*

Department of Surgery and the Liver Center, University of California, San Francisco, CA 94143-0570, USA

## ABSTRACT

The devastating consequences of perinatal liver inflammation contribute to a pressing need to develop therapeutics for the diseases that underly this condition. Biliary atresia (BA) is a perinatal inflammatory disease of the liver that results in obliterative cholangiopathy and rapidly progresses to liver failure, requiring transplantation. The ability to develop targeted therapies requires an understanding of the immune mechanisms that mitigate perinatal liver inflammation. This article reviews our recent findings demonstrating that in a murine model of perinatal hepatic inflammation, Ly6c<sup>Lo</sup> non-classical monocytes express a pro-reparative transcriptomic profile and that the relative abundance of Ly6c<sup>Lo</sup> monocytes promotes resolution of perinatal liver inflammation, rendering neonatal pups resistant to disease. We also examine the lineage relationship between monocyte subsets, reviewing data that suggests classical monocytes are a precursor for non-classical monocytes, and the alternative possibility that separate progenitors exist for each subset. Although a precursor-product relationship between classical and non-classical monocytes might exist in certain environments, we argue that they may also arise from separate progenitors, which is evident by sustained Ly6c<sup>Lo</sup> non-classical monocyte expansion when Ly6c<sup>Hi</sup> monocytes are absent. An improved understanding of monocyte subsets and their developmental trajectories during perinatal hepatic inflammation will provide insight into how therapies directed at controlling monocyte function may help alleviate the devastating consequences of diseases like BA.

**Keywords:** Biliary atresia (BA); Inflammation; Monocyte

## INTRODUCTION

Perinatal hepatic inflammation can have devastating, life-threatening consequences. Biliary atresia (BA) is an example of a progressive inflammatory disease that occurs in the liver of neonates and infants. BA results in rapid obliteration of the biliary tree, leading to liver failure and cirrhosis. Accounting for approximately 75% of liver transplants performed in children under the age of 2 years, BA is the most common indication for pediatric liver transplant [1,2]. The primary treatment of BA relies on the Kasai portoenterostomy, a palliative procedure in most cases as 70% of children still develop progressive liver failure [2]. The etiology of BA is not well understood, but is thought to be attributed to multiple factors including genetic predisposition, immune dysregulation, and toxic and infectious causes [2]. The timing of injury is also not well understood. Compelling evidence supports the idea that the pathogenesis of BA occurs in utero

[3]. The result of this perinatal insult triggers an inflammatory cascade in the liver. A better understanding of the mechanisms by which perinatal hepatic inflammation is initiated and resolved is an important step in identifying therapeutic targets that can halt the development of progressive liver injury and the need for liver transplantation in diseases like BA.

The uniform observation of periportal inflammation in patients with BA indicates that immune-mediated mechanisms are central to the pathogenesis and resolution of hepatic inflammation. Attempts to decrease periportal inflammation in patients with BA have included corticosteroids and intravenous immunoglobulin after portoenterostomy, but these treatments have not improved post-Kasai bile drainage or changed the overall survival of patients with their native liver [4,5]. The lack of clear efficacy using these treatments suggests that immunomodulatory agents that have broad effects may lack the specificity necessary to mitigate disease

**Correspondence to:** Amar Nijagal, Department of Pediatric Surgery, University of CA, San Francisco, CA 94143-0570, USA, Tel: 415-476-4086; E-mail: Amar.Nijagal@ucsf.edu

**Received:** January 04, 2021; **Accepted:** January 18, 2021 **Published:** January 25, 2021

**Citation:** Mohamedaly S, Alkhani A, Nijagal A (2021) Abundance of Monocyte Subsets Determines Susceptibility to Perinatal Hepatic Inflammation. *Immunotherapy (Los Angel)*, Vol.7 Iss.1 No:169.

**Copyright:** © 2021 Mohamedaly S, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

progression.

## IMMUNE SYSTEM MITIGATION OF PERINATAL LIVER INFLAMMATION

Current research supports the involvement of macrophages [6] and dendritic cells [7] in activating natural killer (NK) cells and neutrophils, respectively, to induce a proinflammatory response. Immune signals from NK cells [7], CD4<sup>+</sup> T-helper cells [8-11], and gamma-delta T-cells [12] have also been implicated in the pathogenesis of perinatal inflammation in mice. Additionally, monocytes play an important role in mitigating hepatic inflammation and are among the initial wave of leukocytes recruited to hepatic parenchyma at the time of insult [13]. However, their role in perinatal hepatic inflammatory diseases like BA is poorly understood.

Our current understanding of monocyte function during inflammation is largely based on experiments in adult animals [14-17]. Previous data support the idea that classical monocytes (defined by cell surface expression of Ly6c<sup>Hi</sup>Ccr2<sup>Hi</sup>Cx3cr1<sup>int</sup>) perform pro-inflammatory functions, whereas non-classical monocytes (Ly6c<sup>Lo</sup>Ccr2<sup>Lo</sup>Cx3cr1<sup>Hi</sup>) aid in pro-reparative functions [13,17-20]. In the physiologic state, Ly6c<sup>Lo</sup>Ccr2<sup>Lo</sup>Cx3cr1<sup>Hi</sup> non-classical monocytes act as vascular scavengers, surveilling the endothelium and eliminating luminal microparticles [21,22], whereas circulating Ly6c<sup>Hi</sup>Ccr2<sup>Hi</sup>Cx3cr1<sup>int</sup> classical monocytes are believed to be selectively recruited to inflamed tissues, where they participate in a pro-inflammatory response and give rise to monocyte-derived macrophages [23-25].

We investigated the role of classical and non-classical monocyte subsets in the pathogenesis and resolution of perinatal liver inflammation using a mouse model of perinatal hepatic inflammation. Rhesus rotavirus (RRV) infection in neonatal BALBc mice results in progressive hepatic inflammation that resembles the histologic findings of human BA [26]. Our findings demonstrate that (i) Ly6c<sup>Lo</sup> non-classical monocytes express a pro-reparative transcriptomic signature during the perinatal period, (ii) the abundance of Ly6c<sup>Lo</sup> non-classical monocytes inversely correlates with susceptibility to RRV-mediated perinatal liver injury, and (iii) experimental manipulation of Ly6c<sup>Lo</sup> non-classical monocytes can render neonatal pups resistant to perinatal liver injury [27].

The transcriptomic profile of Ly6c<sup>Lo</sup> non-classical monocytes in the perinatal liver had higher expression of anti-inflammatory genes such as Il4ra and Tgfb1 than did Ly6c<sup>Hi</sup> classical monocytes. Furthermore, we observed a physiologic abundance of Ly6c<sup>Lo</sup> non-classical monocytes compared to Ly6c<sup>Hi</sup> classical monocytes in the late-gestation liver. Specifically, Ly6c<sup>Lo</sup> non-classical monocytes were the predominant monocyte population in the late-gestation liver, outnumbering classical monocytes by 2.5-fold, before equalizing to the known 1:1 relationship that exists in circulation after birth [20].

Given the known anti-inflammatory profile of Ly6c<sup>Lo</sup> non-classical monocytes, we questioned whether this relative abundance of Ly6c<sup>Lo</sup> non-classical monocytes confers resistance to perinatal liver inflammation. Using fetal surgery techniques that we previously described [28,29], we infected late-gestation fetuses with RRV and confirmed inflammatory changes in the fetal liver. We observed that 75% of live-born mice infected with RRV in utero lived beyond 21 days, which was significantly higher than the proportion surviving after neonatal injection (21%). Furthermore, unlike mice injected postnatally, the mice infected in utero did not demonstrate neonatal growth restriction or weight loss. These findings suggested

that the observed physiologic differences in monocyte subsets may account for the resistance to disease observed in late-gestation fetal mice.

We then hypothesized that experimental expansion of Ly6c<sup>Lo</sup> non-classical monocytes in the neonatal liver would spare postnatal mice from hepatic inflammation. Indeed, after depleting both Ly6c<sup>Hi</sup> classical monocytes and neutrophil populations in a postnatal pup we found a significant increase in Ly6c<sup>Lo</sup> non-classical monocytes and a high Ly6c<sup>Lo</sup>:Ly6c<sup>Hi</sup> ratio similar to that of the late-gestation fetus. We found that this environment of Ly6c<sup>Lo</sup> non-classical monocyte expansion conferred protection against RRV-mediated perinatal inflammation. Our findings were confirmed histologically, demonstrating less severe inflammatory changes in the liver parenchyma with near resolution of inflammation in most pups. This result suggested that resistance to perinatal inflammation was either due to diminished levels of pro-inflammatory classical monocytes and neutrophils, or due to the abundance of non-classical monocytes. To distinguish these possibilities, we inhibited Ly6c<sup>Lo</sup> non-classical monocytes in the setting of Ly6c<sup>Hi</sup> classical monocyte and neutrophil depletion and found that susceptibility to RRV-mediated inflammation was restored. These findings demonstrate that the abundance of Ly6c<sup>Lo</sup> non-classical monocytes is indeed associated with resolution of RRV-mediated hepatic inflammation [27]. Collectively, the results of these experiments indicate that non-classical monocytes are crucial for resolution of perinatal hepatic inflammation and suggest that these cells should serve as targets for therapies designed to mitigate the effects of perinatal liver inflammation.

## LINEAGE RELATIONSHIP BETWEEN LY6C<sup>Hi</sup> CLASSICAL MONOCYTES AND LY6C<sup>Lo</sup> NON-CLASSICAL MONOCYTES

Our data also informs the lineage relationship between non-classical monocytes and classical monocytes. The expansion of Ly6c<sup>Lo</sup> non-classical monocytes that occurs in the absence of Ly6c<sup>Hi</sup> classical monocytes suggests that non-classical monocytes may not originate from classical monocytes during RRV-mediated perinatal hepatic inflammation.

There are currently two hypotheses that address the development of classical and non-classical monocyte subsets. The most accepted hypothesis supports the idea that classical monocytes serve as a precursor for non-classical monocytes [17,24,30-32]. An alternative hypothesis that has been proposed is that separate progenitors exist for each subset [23,30]. Studies exploring these hypotheses have focused on bone marrow monocytes in the adult mouse. To the best of our knowledge, studies have not been replicated in the neonatal mouse or specifically in the setting of perinatal hepatic inflammation.

The first hypothesis, originally described by Sunderkötter et al., demonstrated that non-classical monocytes are generated from classical monocytes in peripheral circulation [24]. In the original experiments, the authors ablated all monocyte populations by intravenous administration of clodronate-loaded liposomes and then used fluorescence-activated cell sorter analysis to monitor re-emergence of the monocyte subsets in peripheral blood [24]. Ly6c<sup>Hi</sup> classical monocytes re-emerged on day 2 after ablation, whereas Ly6c<sup>Lo</sup> non-classical monocytes re-emerged on days 3 and 5 [24]. These findings suggest that Ly6c<sup>Hi</sup> classical monocytes serve as a

precursor for Ly6c<sup>Lo</sup> non-classical monocytes.

Subsequent studies utilizing similar methods to Sunderkötter et al. also observed this precursor-product relationship between the two monocyte subsets [30,33,34]. Using pulse chase technology, Yona et al. supported this linear developmental relationship [31]. The authors mapped BrdU (5-bromo-2'-deoxyuridine) incorporation in monocyte subsets, showing that Ly6c<sup>Hi</sup> classical monocytes were rapidly present in the peripheral blood, whereas Ly6c<sup>Lo</sup> non-classical monocytes were observed in the peripheral blood after 3 days despite incorporation of BrdU in the bone marrow at 6 hours [31]. The appearance of classical monocytes first, followed by non-classical monocytes, support the idea that Ly6c<sup>Hi</sup> monocytes give rise to Ly6c<sup>Lo</sup> monocytes.

Del Sacco et al. investigated the relationship between monocyte subsets in the liver using spinning-disk fluorescent confocal intravital microscopy (SD-IVM) [17]. Monocytes subsets were identified by the cell surface markers using Ccr2 and Cx3cr1, with classical monocytes being Ccr2<sup>Hi</sup> Cx3cr1<sup>Lo</sup> and non-classical monocytes defined as Ccr2<sup>Lo</sup> Cx3cr1<sup>Hi</sup>. At baseline, ~5% of patrolling lymphocytes in the liver vasculature were Ccr2<sup>Hi</sup> classical monocytes and no Cx3cr1<sup>Hi</sup> non-classical monocytes were detected. Upon injury, Ccr2<sup>Hi</sup> classical monocytes began to accumulate around the injured area between 8 and 72 hours later, whereas Cx3cr1<sup>Hi</sup> non-classical monocytes became prevalent between 48 and 72 hours with identical localization to the Ccr2<sup>Hi</sup> cells [17]. There was also a transition from Ccr2<sup>Hi</sup> Cx3cr1<sup>Lo</sup> to Ccr2<sup>Lo</sup> Cx3cr1<sup>Hi</sup> within the tissue microenvironment surrounding the focal site of injury that was dependent on a local milieu of cytokines and independent of the recruitment of additional cells to the injury site [17]. This phenotypic transition was correlated with tissue repair and thus suggestive of a local, cytokine-driven in situ reprogramming of pro-inflammatory classical monocytes into reparative non-classical monocytes during liver injury.

Arguably, the most compelling evidence supporting the idea that Ly6c<sup>Hi</sup> classical monocytes may give rise to Ly6c<sup>Lo</sup> non-classical monocytes was demonstrated by adoptive transfer methods. Varol et al. demonstrated that in the absence of inflammation, transferred bone marrow Ly6c<sup>Hi</sup> classical monocytes home back to the bone marrow of recipient mice in which myeloid dendritic cell progenitors (MDPs) are depleted and participate in Ly6c<sup>Lo</sup> non-classical monocyte differentiation [35]. The authors excluded MDPs on the basis that they are Ly6c negative; however, recent evidence supports the presence of common monocyte progenitors (cMoPs) that are Ly6c positive, confounding the results of this study [36]. The recent discovery of common monocyte progenitors (cMoPs) certainly raises the question of whether other precursors exist in the lineage relationship between MDPs and final monocyte subsets that have yet to be discovered.

Challenging this precursor-product relationship between monocytes subsets, Nahrendorf et al. used adoptive transfer methods to demonstrate sequential recruitment of different monocyte subsets to the myocardium after a myocardial ischemic injury in a biphasic manner [37]. Findings demonstrated that Ly6c<sup>Hi</sup> classical monocytes were present at the site of injury during the first 3 days (phase I) and Ly6c<sup>Lo</sup> non-classical monocytes increased between days 4-7 (phase II) [37-39]. The adoptive transfer methods in CCR2 knockout mice demonstrated the following (i) efficient accumulation of adoptively transferred Ly6c<sup>Lo</sup> monocytes in infarcts in phase II; (b) the absence of Ly6c<sup>Hi</sup> monocytes in phase I, but presence of Ly6c<sup>Lo</sup> monocytes in phase II, in infarcts of CCR2<sup>-/-</sup> mice; and (c) presence of Ly6c<sup>Hi</sup>

monocytes in phase I, but absence of Ly6c<sup>Lo</sup> monocytes in phase II, in infarcts of CX3CR1<sup>-/-</sup> mice [37]. The expansion of Ly6c<sup>Lo</sup> non-classical monocytes when Ly6c<sup>Hi</sup> classical monocytes are absent argue against in situ conversion of Ly6c<sup>Hi</sup> classical monocytes to Ly6c<sup>Lo</sup> non-classical monocytes during tissue repair.

## CONCLUSION

While our findings demonstrate the importance of Ly6c<sup>Lo</sup> non-classical monocytes in resolution of perinatal liver inflammation and raise the possibility of a separate progenitor relationship between the monocyte subsets, the evidence supporting an alternate hypothesis of a precursor-product relationship demonstrates the need for continued investigation and further understanding of this developmental relationship. This is especially relevant in the setting of perinatal liver inflammation, given its morbid consequences. Deeper understanding of the role of monocyte subsets and particularly their developmental lineage during inflammation can provide insight for future targeted therapy that could halt inflammation and alleviate the burden of devastating inflammatory conditions, such as BA.

## REFERENCES

1. Verkade HJ, Bezerra JA, Davenport M, Schreiber RA, Vergani GM, Hulscher JB, et al. Biliary atresia and other cholestatic childhood diseases: Advances and future challenges. *J Hepatol.* 2016;65(3):631-642.
2. Hartley JL, Davenport M, Kelly DA. Biliary atresia. *The Lancet.* 2009;374:1704-1713.
3. Harpavat S, Garcia-Prats JA, Shneider BL. Newborn bilirubin screening for biliary atresia. <https://doi.org/10.1056/NEJMc1601230>. 2016.
4. Nijagal A, Perito ER. Treating biliary atresia: The challenge continues. *J Pediatr Gastroenterol Nutr.* 2019;68(4):464-465.
5. Mack CL, Spino C, Alonso EM, Bezerra JA, Moore J, Goodhue C, et al. A phase I/IIa trial of intravenous immunoglobulin following portoenterostomy in biliary atresia. *J Pediatr Gastroenterol Nutr.* 2019;68(4):495-501.
6. Mohanty SK, Ivantes CAP, Mourya R, Pacheco C, Bezerra JA. Macrophages are targeted by rotavirus in experimental biliary atresia and induce neutrophil chemotaxis by Mip2/Cxcl2. *Pediatr Res.* 2010;67(4):345-351.
7. Saxena V, Shivakumar P, Sabla G, Mourya R, Chougnet C, Bezerra JA, et al. Dendritic cells regulate natural killer cell activation and epithelial injury in experimental biliary atresia. *Sci Transl Med.* 2011;3(102):102ra94.
8. Lages CS, Simmons J, Maddox A, Jones K, Karns R, Sheridan R, et al. The dendritic cell-Th17-macrophage axis controls cholangiocyte injury and disease progression in murine and human biliary atresia. *Hepatology Baltim Md.* 2017;65(1):174-188.
9. Shivakumar P, Campbell KM, Sabla GE, Miethke A, Tiao G, McNeal MM, et al. Obstruction of extrahepatic bile ducts by lymphocytes is regulated by IFN- $\gamma$  in experimental biliary atresia. *J Clin Invest.* 2004;114(3):322-329.
10. Feldman AG, Tucker RM, Fenner EK, Pelanda R, Mack CL. B cell deficient mice are protected from biliary obstruction in the rotavirus-induced mouse model of biliary atresia. *Plos One.* 2013;8(8):e73644.
11. Li J, Bessho K, Shivakumar P, Mourya R, Mohanthy SK, Santos JLD, et al. Th2 signals induce epithelial injury in mice and are compatible with the biliary atresia phenotype. *J Clin Invest.* 2011;121(11).

12. Klemann C, Schröder A, Dreier A. Interleukin 17, produced by  $\gamma\delta$  T cells, contributes to hepatic inflammation in a mouse model of biliary atresia and is increased in livers of patients. *Gastroenterology*. 2016;150(1):229-241.e5.
13. Brempelis KJ, Crispe IN. Infiltrating monocytes in liver injury and repair. *Clin Transl Immunol*. 2016;5(11):e113.
14. Misharin AV, Cuda CM, Saber R, Tuner JD, Gierut AK, Haines GK, et al. Nonclassical Ly6C<sup>-</sup> monocytes drive the development of inflammatory arthritis in mice. *Cell Rep*. 2014;9(2):591-604.
15. Zigmund E, Varol C, Farache J, Elmaliach E, Sathpathy AT, Friedlander G, et al. Ly6Chi monocytes in the inflamed colon give rise to proinflammatory effector cells and migratory antigen-presenting cells. *Immunity*. 2012;37(6):1076-1090.
16. Grainger JR, Wohlfert EA, Fuss IJ, Bouladoux N, Askenase MH, Legrand F, et al. Inflammatory monocytes regulate pathologic responses to commensals during acute gastrointestinal infection. *Nat Med*. 2013;19(6):713-721.
17. Dal-Secco D, Wang J, Zeng Z, Kolaczowska E, Wong CHY, Petri B, et al. A dynamic spectrum of monocytes arising from the in situ reprogramming of CCR2<sup>+</sup> monocytes at a site of sterile injury. *J Exp Med*. 2015;212(4):447-456.
18. Lee PY, Nelson-Maney N, Huang Y, Levescot A, Wang Q, Wei K, et al. High-dimensional analysis reveals a pathogenic role of inflammatory monocytes in experimental diffuse alveolar hemorrhage. *JCI Insight*. 4(15).
19. Olingy CE, San Emeterio CL, Ogle ME, Krieger JR, Bruce AC, Pfau DD, et al. Non-classical monocytes are biased progenitors of wound healing macrophages during soft tissue injury. *Sci Rep*. 2017;7.
20. Williams M, Mildner A, Yona S. Developmental and functional heterogeneity of monocytes. *Immunity*. 2018;49(4):595-613.
21. Imhof BA, Jemelin S, Ballet R, Vesin C, Schapira M, Karaca M, et al. CCN1/CYR61-mediated meticulous patrolling by Ly6Clow monocytes fuels vascular inflammation. *Proc Natl Acad Sci U S A*. 2016;113(33):E4847-E4856.
22. Finsterbusch M, Hall P, Li A, Devi S, Westthrope CLV, Kitching AR, et al. Patrolling monocytes promote intravascular neutrophil activation and glomerular injury in the acutely inflamed glomerulus. *Proc Natl Acad Sci U S A*. 2016;113(35):E5172-E5181.
23. Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M, Ley K. Development of monocytes, macrophages, and dendritic cells. *Science*. 2010;327(5966):656-661.
24. Sunderkötter C, Nikolic T, Dillon MJ, Rooijen NV, Stehling M, Drevets DA, et al. Subpopulations of mouse blood monocytes differ in maturation stage and inflammatory response. *J Immunol*. 2004;172(7):4410-4417.
25. Palframan RT, Jung S, Cheng G, Weninger W, Luo Y, Dorf M, et al. Inflammatory chemokine transport and presentation in HEV. *J Exp Med*. 2001;194(9):1361-1374.
26. Riepenhoff-Talty M, Schaekel K, Clark HF, Mueller W, Uhnou I, Rossi T, et al. Group A rotaviruses produce extrahepatic biliary obstruction in orally inoculated newborn mice. *Pediatr Res*. 1993;33:394-399.
27. Alkhani A, Levy CS, Tsui M, Rosenberg KA, Polovina K, Mattis AN, Mack M, et al. Ly6c<sup>Lo</sup> non-classical monocytes promote resolution of rhesus rotavirus-mediated perinatal hepatic inflammation. *Sci Rep*. 2020;10(1):7165.
28. Nijagal A, Le T, Wegorzewska M, Mackenzie TC. A mouse model of in utero transplantation. *J Vis Exp JoVE*. 2011;(47).
29. Nijagal A, Wegorzewska M, Jarvis E, Le T, Tang Q, MacKenzie TC. Maternal T cells limit engraftment after in utero hematopoietic cell transplantation in mice. *J Clin Invest*. 2011;121(2):582-592.
30. Peipei ZYF, Graham TD, Catherine HC. Transcriptional control of monocyte development 2014 jeffrey m. hoeg award lecture. *Arterioscler Thromb Vasc Biol*. 2016;36(9):1722-1733.
31. Yona S, Kim KW, Wolf Y, Mildner A, Varol D, Breker M, et al. Fate mapping reveals origins and dynamics of monocytes and tissue macrophages under homeostasis. *Immunity*. 2013;38(1):79-91.
32. Ginhoux F, Jung S. Monocytes and macrophages: Developmental pathways and tissue homeostasis. *Nat Rev Immunol*. 2014;14(6):392-404.
33. Tacke F, Randolph GJ. Migratory fate and differentiation of blood monocyte subsets. *Immunobiology*. 2006;211(6):609-618.
34. Tacke F, Ginhoux F, Jakubzick C, van Rooijen N, Merad M, Randolph GJ. Immature monocytes acquire antigens from other cells in the bone marrow and present them to T cells after maturing in the periphery. *J Exp Med*. 2006;203(3):583-597.
35. Varol C, Landsman L, Fogg DK, Greenshtein L, Gildor B, Margalit R, et al. Monocytes give rise to mucosal, but not splenic, conventional dendritic cells. *J Exp Med*. 2007;204(1):171-180.
36. Hettinger J, Richards DM, Hansson J, Barra MM, Joschko AC, Krijgsveld J, et al. Origin of monocytes and macrophages in a committed progenitor. *Nat Immunol*. 2013;14(8):821-830.
37. Nahrendorf M, Swirski FK, Aikawa E, Stangenberg L, Wurdinger T, Figureiredo JL, et al. The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. *J Exp Med*. 2007;204(12):3037-3047.
38. Satoh T, Nakagawa K, Sugihara F, Kuwahara R, Ashihara M, Yamane F, et al. Identification of an atypical monocyte and committed progenitor involved in fibrosis. *Nature*. 2017;541(7635):96-101.
39. Hanna RN, Carlin LM, Hubbeling HG, Nackiewicz D, Green AM, Punt JA, et al. The transcription factor NR4A1 (Nur77) controls bone marrow differentiation and the survival of Ly6C<sup>-</sup> monocytes. *Nat Immunol*. 2011;12(8):778-785.