

The Relationship between Mitochondria Ca²⁺ Intake Mediated by Mitochondria-associated Endoplasmic Reticulum Membranes and Tumor Genesis

Zijian Zhu^{1#}, Qingzhi Ma^{2#}, Qian Wang³, Xiacheng Sun³, Zhanhua Zhang¹, Lele Ji^{3*} and Qichao Huang^{3*}

¹The Fourth Brigade, School of Basic Medical Sciences, The Fourth Military Medical University, Xi'an 710032, China

²The Third Brigade, School of Basic Medical Sciences, The Fourth Military Medical University, Xi'an 710032, China

³State Key Laboratory of Cancer Biology and Experimental Teaching Center of Basic Medicine, Fourth Military Medical University, Xi'an, China

#Contributed equally to this work

*Corresponding author: Qichao Huang, State Key Laboratory of Cancer Biology and Experimental Teaching Center of Basic Medicine, Fourth Military Medical University, 169 West Changle Street, Xi'an 710032, China, Tel: +86-29-84774518; E-mail: huangqichao1@163.com

Lele Ji, Experimental Teaching Center of Basic Medicine, Fourth Military Medical University, 169 West Changle Street, Xi'an 710032, China, Tel: +86-29-84774766; E-mail: seasonglad@126.com

Received date: January 17, 2019; Accepted date: February 14, 2019; Published date: February 22, 2019

Copyright: © 2019 Zhu Z, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Mitochondria-associated endoplasmic reticulum membranes (MAMs) are regions of the endoplasmic reticulum (ER) tethered to mitochondria, which play a key role in mediating material transfer and signal transduction between the two organelles. The findings from recent studies on MAMs contributed to deeper understanding of the complexities associated with the structure, the important proteins involved and the intricacies in the related biological pathways. A large number of Ca²⁺ transporter proteins and their regulatory proteins are located on MAMs, which finely regulate a series of important cellular activities such as mitochondrial Ca²⁺ homeostasis, ATP production and cell apoptosis. MAMs are also enriched with many oncogenic proteins and tumor suppressor proteins, which are closely related to the regulation of Ca²⁺ transport. Therefore, the role of MAMs in tumorigenesis has received extensive attention. In this review, we focused on the regulatory mechanisms of Ca²⁺ transport mediated by MAMs and their role in tumorigenesis, aiming to acquire the new insight to further understanding the pathogenesis of tumors.

Keywords: Mitochondria; Mitochondria-associated endoplasmic reticulum membranes; Ca²⁺ signal; Tumor

Introduction

Mitochondria and the endoplasmic reticulum (ER) regulate numerous cellular processes, and are critical contributors to cellular and whole-body homeostasis. Interestingly, about 5-20% of the mitochondrial membranes are directly in contact with ER [1]. Therefore, the mitochondria and ER cannot be considered as static structures, they intimately communicate, forming very dynamic platforms termed Mitochondria-associated endoplasmic reticulum membranes (MAMs). With the development of super-resolution fluorescence imaging, electron tomography and proteomics, MAMs have been found in various eukaryotes [2,3]. In particular, the MAMs accommodate flux of Ca²⁺ from the ER to mitochondria, which decode them into specific inputs to regulate essential functions, including metabolism, energy production and apoptosis [4-6]. Furthermore, previous studies have suggested that many human diseases are closely linked to the mitochondria abnormal Ca²⁺ intake mediated by MAMs, such as tumor genesis [7] and neurodegeneration[8]. Hence, MAMs are not simply be considered as a static bridge between the ER and mitochondria, but also as dynamic organelles that play a variety of roles both in physiological and pathological processes that are crucial in maintaining the health or establishing a disease due to functional disturbances. Recently, there is an increased focus on MAMs because numerous oncogenic proteins

and tumor suppressors were found on the MAMs, which exert an important influence on cell fate and the emerging picture of MAMs seems to indicate that deregulated MAMs-mediated mitochondrial Ca²⁺ intake play an important role in tumor genesis. This review has focused on the findings from publications from the past ten years.

Structure Basis of MAMs

The association between the ER and mitochondria was first visualized in the 1970s with electron microscopy by Morre et al. [9]. It was not until 1990s, however, the Vance group made great progress in the MAMs field by presenting a detailed protocol describing the isolation of pure MAMs fractions by differential ultracentrifugation [1]. In recent years, multiple methods have been developed to dissect MAMs' specific properties and the protein composition, either using biochemical or fluorescent microscopy-based strategies. We have enormously extended our comprehension on MAMs that MAMs contain several crucial proteins involved in many biological pathways. In addition, some researches show that MAMs are closely related to cellular lipid metabolism [1] and energy metabolism [10].

Molecular Components of the MAMs in Ca²⁺ Transfer

It has recently become clear that MAMs are crucial for highly efficient transmission of Ca²⁺ from the ER to mitochondria, thus controlling fundamental processes involved in energy production and also determining the cell fate by triggering or preventing apoptosis.

Therefore, many Ca²⁺ transporter proteins, such as Inositol 1,4,5-trisphosphate receptors (IP3Rs), voltage-dependent anion channel (VDAC)[11], mitochondrial calcium uniporter (MCU) and their regulatory proteins were identified on MAMs [12](Figure 1).

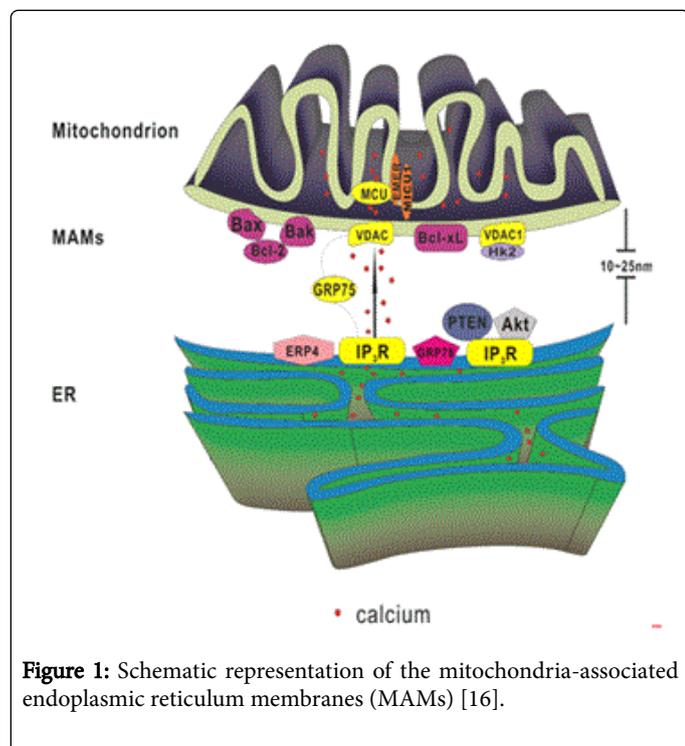


Figure 1: Schematic representation of the mitochondria-associated endoplasmic reticulum membranes (MAMs) [16].

IP3Rs

A key role in the control of Ca²⁺ signals is attributed to the inositol 1,4,5-trisphosphate (IP3) receptors (IP3Rs), the main Ca²⁺-release channels in the ER. As expected, all three IP3R isoforms (IP3R1, IP3R2 and IP3R3) are also enriched in MAMs and precisely control Ca²⁺ transfer into mitochondria [13]. When the G protein-coupled receptors are activated, intracellular IP3 binding with IP3Rs can lead to the release of Ca²⁺. This creates a micro-domain in which the Ca²⁺ concentrations are manifold higher than in the cytosol, allowing for rapid mitochondrial Ca²⁺ uptake [14]. In addition, some metabolites can also affect Ca²⁺ transfer efficient in MAMs by regulating the activity of IP3Rs. For example, glucose-regulated protein 78 (GRP78) promotes the activity of IP3Rs and increases the Ca²⁺ intake of mitochondria [15]. A fraction of endoplasmic reticulum protein 44 (ERp44) also localizes to the MAM, where it interacts with IP3R1 and competes with BiP/Grp78 for the same binding site on the IP3R1. Cells over-expressing ERp44 show reduced IP3R1 Ca²⁺-release [16,17]. In addition, ATP can promote the release of Ca²⁺ mediated by IP3R, but heparin is the specific inhibitor [13].

VDAC

VDAC is a large, high-conductance, weakly anion-selective channel that represents the primary permeability pathway through which solutes enter the mitochondria. VDAC is associated with MAMs and controls metabolic cross-talk between mitochondria and the rest of the cell by allowing the influx and efflux of metabolites, ions, nucleotides, Ca²⁺ and more [18]. Human cells have three distinct VDAC genes (VDAC1, VDAC2 and VDAC3), with VDAC1 representing the best

characterized one [19]. For example, VDAC1 selectively interacts with IP3R3, thereby potentiating the transfer of low-amplitude apoptotic Ca²⁺ signals to mitochondria. Therefore, VDAC1 mediates Ca²⁺ released by ER transmits into the mitochondrial intermembrane space [19]. Arbel N found that the over expression of VDAC1 led mitochondrial Ca²⁺ increase in skeletal muscle cells and Hela cells [20]. Consistently, VDAC1 knockdown decreased mitochondrial Ca²⁺ [21]. In addition, it has been reported that, glucose-regulated protein 75 (GRP75) promoted the connection between VDAC1 and IP3Rs with the subsequent mitochondrial Ca²⁺ intake [22].

MCU

The mitochondrial Ca²⁺ uniporter is a complex of proteins including the Ca²⁺ selective pore-forming subunit MCU and accessory proteins including MICU1, MICU2, MCUR1 and EMRE located in the mitochondrial inner membrane (IMM) and also enriched in MAMs [23-25]. Ca²⁺ crosses the IMM through the MCU depending on the considerable driving force represented by the negative Trans membrane potential. Several lines of evidence indicate that MICU1 and MICU2 operate together with MCU [24]. MICU1 acts as a gatekeeper of the uniporter complex, preventing Ca²⁺ entry under resting conditions and activating the channel at high cytosolic Ca²⁺ concentrations [26]. While, MICU2, a paralog of MICU1, can inhibit MICU1-mediated Ca²⁺ uptake [27]. Moreover, EMRE was required for the interaction of MCU with MICU1 and MICU2. Thus, EMRE bridges the calcium-sensing role of MICU1 and MICU2 with the calcium-conducting role of MCU [28]. In addition, the expression of MCU appears to be controlled by microRNA-25, which can efficiently reduce MCU levels and subsequent mitochondrial Ca²⁺ transfer [29]. Therefore, Ca²⁺ transferred by MAMs is an intricate and tightly regulated process.

Influence of Ca²⁺ Intake on Mitochondrial Function

The main physiological role of mitochondrial Ca²⁺ uptake was assessed to be the control of metabolic activity of the mitochondria. It has been reported that various enzymes directly involving in the Krebs cycle are modulated by mitochondrial matrix Ca²⁺ [30]. For example, pyruvate dehydrogenase (PDH), which converts pyruvate into acetyl-CoA, is phosphorylated and activated when the level of mitochondrial Ca²⁺ is elevated [30,31]. While mitochondrial Ca²⁺ was also demonstrated to increase the affinity of dehydrogenase (ICDH) or oxoglutarate dehydrogenase (OGDH) with their substrates [31]. In addition, mitochondrial Ca²⁺ also directly activates the electron transport chain and the activity of F0F1ATP synthase [32,33]. On the contrary, the mitochondrial Ca²⁺ overload results in dramatic alterations in mitochondrial functions, including increased reactive oxygen species (ROS) production and “mitochondrial permeability transition pore”(mPTP) activity[34]. The mPTP opening can induce mitochondrial swelling, and these large-scale alterations of organelle morphology allow the release of caspase cofactors into the cytosol, which can lead to cell death finally [35]. Therefore, Mitochondria are not only the energy powerhouse of the cell but also a major hub for cellular Ca²⁺ signaling crucial for cell life and death.

Impact of Ca²⁺ Transfer Regulated by MAMs on Tumor Genesis

Recently, increasing evidence is beginning to reveal that the abnormal remodeling of mitochondrial Ca²⁺ homeostasis has

important roles in tumor initiation and progression [36]. For example, decrease of Ca²⁺ in mitochondria was reported to shift the cancer cells toward glycolysis, providing chemo resistance but leading to a poor overall survival [37]. Notably, a wide range of tumor suppressors and oncogenic proteins were identified to be located on MAMs and play important roles in mitochondrial Ca²⁺ transfer [37]. Generally, tumor suppressors were believed to promote the mitochondrial Ca²⁺ uptake, while oncogenic proteins exert opposite roles [38]. Therefore, enhancing the mitochondrial Ca²⁺ uptake through MAMs might be a potential strategy for cancer treatment.

Oncogenic proteins on MAMs

There are some oncogenic proteins on MAMs (Table 1), they can interact with different molecules and inhibit the apoptosis of tumor cells. Among oncogenic proteins, Akt is an important sensor of the bioenergetics of the cell and therefore it is linked to the function of the mitochondria. Recently, several studies have proved that Akt could phosphorylate all IP3R isoforms, thus inhibits Ca²⁺ release from ER and protects cells from apoptosis. Moreover, Akt also was demonstrated to promote the interaction between VDAC1 and hexokinase 2 (HK2) on MAMs through phosphorylation events. Therefore, this association inhibits apoptosis mediated by mitochondrial Ca²⁺. In addition, it has been also found that PTEN could dephosphorylate PIP3 and reverse PI3K/Akt signaling, which further promotes the apoptosis of tumor cell. Bcl-2 protein family also contains numerous anti-apoptotic and pro-apoptotic members. Therefore, Bcl-2 protein family plays an important role in mitochondria dependent apoptosis [39]. Recently, researches have demonstrated that Bcl-2 protein family members interact with different functional domains of IP3Rs and promote or inhibit Ca²⁺ signals and the apoptosis of tumor cells [39, 40]. For example, Williams A. et al. have found that numerous Bcl-2 are rich on MAMs. Moreover, Bcl-2 interacts directly with IP3Rs to inhibit channel opening and ER Ca²⁺-release, thus inhibit tumor cells apoptosis [41]. Monaco G has proved that anti-apoptotic protein Bcl-XL, which is deregulated in several cancer types, exerts its anti-apoptotic functions by inhibiting the activity of Ca²⁺ channels, including IP3Rs and VDAC isoforms [42, 43]. In addition, Bcl-XL also blocks the apoptosis pathway by neutralizing pro-apoptotic Bcl-2 members, such as Bak, Bax, Bid and Bim [44]. Sig1-R is a Ca²⁺-sensitive and ligand-operated receptor chaperone and localizes at MAMs, stabilizes the conformation of IP3R3 and the ER stress sensor IRE1. Normally, Sig1-R forms a complex at MAMs with the chaperone BiP/GRP78 to regulate Ca²⁺ homeostasis between the ER and the mitochondria, but upon Ca²⁺ depletion or via ligand stimulation, Sig1-R dissociates from BiP leading to a prolonged Ca²⁺ signaling into mitochondria via IP3R3. [4-14]

Protein	Impact on ER-mitochondrial Ca ²⁺ transfer	References
AKT	Inhibition of Ca ²⁺ release from ER	[4-43]
Bcl-2	Induction of Ca ²⁺ leakage from ER	[4-42],[4-43]
Bcl-XL	Induction of Ca ²⁺ leakage from ER	[4-44]
Sig-1R	Regulation of Ca ²⁺ homeostasis on MAMs	[4-14]

Table 1: Summary of the ant apoptotic proteins on MAMs.

Tumor suppressors on MAMs

Many tumor suppressors are located on MAMs and here we list several most important tumor suppressors on MAMs (Table 2). The tumor suppressor PTEN is among the most commonly lost or mutated tumor suppressors implicated in human cancers, and it is a key regulator of a wide range of biological functions other than tumor suppression. Recent findings have shown that it localizes at MAMs where it interacts with the IP3R3 and regulates Ca²⁺ release from the ER in a protein phosphatase-dependent manner that counteracts AKT activation; thus, it can inhibit AKT-mediated phosphorylation of IP3R3, which protects from Ca²⁺-mediated apoptosis [4]. In addition, the tumor suppressor PML also localizes at the MAMs where it modulates IP3R3 activity and the ER-mitochondria Ca²⁺ fluxes by promoting the formation of a multi protein complex containing IP3R3, AKT and the protein phosphatase 2A (PP2a) [4-49]. The tumor suppressor p53 regulates tumor genesis also in a Ca²⁺ dependent pathway. P53 physically interacts with SERCA and this increases the efficiency of the transfer of Ca²⁺ ions between the ER and mitochondria, augmenting the propensity of (pre)malignant cells exposed to oncogenic or chemotherapeutic stress to succumb to apoptosis. The interplay between p53 and Ca²⁺ signaling is not limited to chemotherapy but is also relevant for cellular response following the photodynamic therapy (PDT) [4-49].

Protein	Impact on ER-mitochondrial Ca ²⁺ transfer	References
PTEN	Regulation of Ca ²⁺ release via IP3R3	[4-7]
PML	Modulation of the ER-mitochondria Ca ²⁺ flux	[4-48], [4-49]
P53	Modulation of Ca ²⁺ transfer from ER to mitochondria interacting with Serca	[4-49]

Table 2: Summary of the tumor suppressors on MAMs.

Conclusion

We presented clear evidence to indicate that loss of Calcium homeostasis in the mitochondria due to defective transfer between the ER and mitochondria mediated by MAMs has been shown to contribute to tumor genesis. And above all, we can conclude that there are two main ways that proteins on MAMs affect tumor cells fate, one is to interact with Ca²⁺ tunnels, such as IP3R and VDAC, and another way is the mutual effect between oncogenic proteins and tumor suppressors. As a consequence, MAMs dysfunctions have been linked to many types of human cancer. However, several outstanding questions still need to be answered before reaching a complete mechanistic and functional understanding of the MAMs-mediated mitochondrial Ca²⁺ uptake. For example, how does cancer cell regulate the dynamics of the structure of MAMs and the protein located in MAMs? Which proteins located in MAMs is crucial for cancer cell survival, inflammation, and therapy responses? Additionally, it is still not clear how MAMs modulate the mitochondrial unfolded protein response (UPR) and ER stress in cancer cells. All these are outstanding questions that await future studies.

Acknowledgement

This work was supported by the National Natural Science Foundation of China (No. 81772935) and the State Key Laboratory of Cancer Biology Project (CBSKL2017Z02).

References

- Vance JE (1990) Phospholipid synthesis in a membrane fraction associated with mitochondria. *J Biol Chem* 265: 7248-7256.
- Csordas G, Renken C, Varnai P, Walter L, Weaver D, et al. (2006) Structural and functional features and significance of the physical linkage between ER and mitochondria. *J Cell Biol* 174: 915-921.
- Rizzuto R, Pinton P, Carrington W, Fay FS, Fogarty KE, et al. (1998) Close contacts with the endoplasmic reticulum as determinants of mitochondrial Ca²⁺ responses. *Science* 280: 1763-1766.
- Bonora M, Patergnani S, Rimessi A, De Marchi E, Suski JM, et al. (2012) ATP synthesis and storage. *Purinergic Signal* 8: 343-57.
- Denton RM, Randle PJ, Martin BR (1972) Stimulation by calcium ions of pyruvate dehydrogenase phosphate phosphatase. *Biochem J* 128: 161-163.
- McCormack JG, Halestrap AP, Denton RM (1990) Role of calcium ions in regulation of mammalian intramitochondrial metabolism. *Physiol Rev* 70: 391-425.
- Arruda AP, Pers BM, Parlakgul G, Guney E, Inouye K, et al. (2014) Chronic enrichment of hepatic endoplasmic reticulum-mitochondria contact leads to mitochondrial dysfunction in obesity. *Nat Med* 20: 1427-1435.
- Krols M, van Isterdael G, Asselbergh B, Kremer A, Lippens S, et al. (2016) Mitochondria-associated membranes as hubs for neurodegeneration. *Acta Neuropathol* 131: 505-523.
- Morre DJ, Merritt WD, Lembi CA (1971) Connections between mitochondria and endoplasmic reticulum in rat liver and onion stem. *Protoplasma* 73: 43-49.
- Theurey P, Rieusset J (2017) Mitochondria-Associated Membranes Response to Nutrient Availability and Role in Metabolic Diseases. *Trends Endocrinol Metab* 28: 32-45.
- Naghdi S, Hajnoczky G (2016) VDACC2-specific cellular functions and the underlying structure. *Biochim Biophys Acta* 1863: 2503-2514.
- Rizzuto R, De Stefani D, Raffaello A, Mammucari C (2012) Mitochondria as sensors and regulators of calcium signalling. *Nat Rev Mol Cell Biol* 13: 566-578.
- Ivanova H, Vervliet T, Missiaen L, Parys JB, De Smedt H, et al. (2014) Inositol 1,4,5-trisphosphate receptor-isoform diversity in cell death and survival. *Biochim Biophys Acta* 1843: 2164-2183.
- Carafoli E (2010) The fateful encounter of mitochondria with calcium: how did it happen?. *Biochim Biophys Acta* 1797: 595-606.
- Hayashi T, Su TP (2007) Sigma-1 receptor chaperones at the ER-mitochondrion interface regulate Ca²⁺ signaling and cell survival. *Cell* 131: 596-610.
- Marchi S, Patergnani S, Missiroli S, Morciano G, Rimessi A et al. (2018) Mitochondrial and endoplasmic reticulum calcium homeostasis and cell death. *Cell calcium* 69: 62-72.
- Higo T, Hattori M, Nakamura T, Natsume T, Michikawa T, et al (2005) Subtype-specific and ER luminal environment-dependent regulation of inositol 1,4,5-trisphosphate receptor type 1 by ERp44. *Cell* 120: 85-98.
- Colombini M (2016) The VDACC channel: Molecular basis for selectivity. *Biochim Biophys Acta* 1863: 2498-2502.
- De Stefani D, Bononi A, Romagnoli A, Messina A, De Pinto V, et al. (2012) VDACC1 selectively transfers apoptotic Ca²⁺ signals to mitochondria. *Cell Death Differ* 19: 267-273.
- Arbel N, Ben-Hail D, Shoshan-Barmatz V (2012) Mediation of the antiapoptotic activity of Bcl-xL protein upon interaction with VDACC1 protein. *J Biol Chem* 287: 23152-23161.
- Rapizzi E, Pinton P, Szabadkai G, Wieckowski MR, Vandecasteele G, et al. (2002) Recombinant expression of the voltage-dependent anion channel enhances the transfer of Ca²⁺ microdomains to mitochondria. *J Cell Biol* 159: 613-624.
- Szabadkai G, Bianchi K, Varnai P, De Stefani D, Wieckowski MR, et al. (2006) Chaperone-mediated coupling of endoplasmic reticulum and mitochondrial Ca²⁺ channels. *J Cell Biol* 175: 901-911.
- Kirichok Y, Krapivinsky G, Clapham DE (2004) The mitochondrial calcium uniporter is a highly selective ion channel. *Nature* 427: 360-364.
- Patron M, Checchetto V, Raffaello A, Teardo E, Vecellio Reane D, et al. (2014) MICU1 and MICU2 finely tune the mitochondrial Ca²⁺ uniporter by exerting opposite effects on MCU activity. *Mol Cell* 53: 726-737.
- Raffaello A, De Stefani D, Sabbadin D, Teardo E, Merli G, et al. (2013) The mitochondrial calcium uniporter is a multimer that can include a dominant-negative pore-forming subunit. *EMBO J* 32: 2362-2376.
- Mallilankaraman K, Doonan P, Cardenas C, Chandramoorthy HC, Muller M, et al. (2012) MICU1 is an essential gatekeeper for MCU-mediated mitochondrial Ca²⁺ uptake that regulates cell survival. *Cell* 151: 630-644.
- Kamer KJ, Mootha VK. (2014) MICU1 and MICU2 play nonredundant roles in the regulation of the mitochondrial calcium uniporter. *EMBO Rep* 15: 299-307.
- Sancak Y, Markhard AL, Kitami T, Kovacs-Bogdan E, Kamer KJ, et al. (2013) EMRE is an essential component of the mitochondrial calcium uniporter complex. *Science* 342: 1379-1382.
- Zaglia T, Ceriotti P, Campo A, Borile G, Armani A, et al. (2017) Content of mitochondrial calcium uniporter (MCU) in cardiomyocytes is regulated by microRNA-1 in physiologic and pathologic hypertrophy. *Proc Natl Acad Sci U S A* 114: E9006-E9015.
- Denton RM (2009) Regulation of mitochondrial dehydrogenases by calcium ions. *Biochim Biophys Acta* 1787: 1309-1316.
- Denton RM, Richards DA, Chin JG (1978) Calcium ions and the regulation of NAD⁺-linked isocitrate dehydrogenase from the mitochondria of rat heart and other tissues. *Biochem J* 176: 899-906.
- Territo PR, Mootha VK, French SA, Balaban RS (2000) Ca²⁺ activation of heart mitochondrial oxidative phosphorylation: role of the F(0)/F(1)-ATPase. *American journal of physiology* 278: C423-35.
- Glancy B, Willis WT, Chess DJ, Balaban RS. (2013) Effect of calcium on the oxidative phosphorylation cascade in skeletal muscle mitochondria. *Biochemistry* 52: 2793-2809.
- Bonora M, Wieckowski MR, Chinopoulos C, Kepp O, Kroemer G, et al. (2015) Molecular mechanisms of cell death: central implication of ATP synthase in mitochondrial permeability transition. *Oncogene* 34: 1608.
- Hwang MS, Schwall CT, Pazarentzos E, Datler C, Alder NN, et al. (2014) Mitochondrial Ca²⁺ influx targets cardiolipin to disintegrate respiratory chain complex II for cell death induction. *Cell Death Differ* 21: 1733-1745.
- Ren T, Zhang H, Wang J, Zhu J, Jin M, et al. (2017) MCU-dependent mitochondrial Ca²⁺ inhibits NAD⁺/SIRT3/SOD2 pathway to promote ROS production and metastasis of HCC cells. *Oncogene* 36: 5897-5909.
- Bittremieux M, Parys JB, Pinton P, Bultynck G (2016) ER functions of oncogenes and tumor suppressors: Modulators of intracellular Ca²⁺ signaling. *Biochim Biophys Acta* 1863: 1364-1378.
- Herrera-Cruz MS, Simmen T (2017) Cancer: Untethering Mitochondria from the Endoplasmic Reticulum?. *Front Oncol* 7: 105.
- Pinton P, Ferrari D, Rapizzi E, Di Virgilio F, Pozzan T, et al. (2001) The Ca²⁺ concentration of the endoplasmic reticulum is a key determinant of ceramide-induced apoptosis: significance for the molecular mechanism of Bcl-2 action. *EMBO J* 20: 2690-2701.
- Pinton P, Ferrari D, Magalhaes P, Schulze-Osthoff K, Di Virgilio F, et al. (2000) Reduced loading of intracellular Ca²⁺ stores and downregulation of capacitative Ca²⁺ influx in Bcl-2-overexpressing cells. *J Cell Biol* 148: 857-862.
- Williams A, Hayashi T, Wolozny D, Yin B, Su TC, et al. (2016) The non-apoptotic action of Bcl-xL: regulating Ca²⁺ signaling and bioenergetics at the ER-mitochondrion interface. *J Bioenerg Biomembr* 48: 211-225.

42. Monaco G, Decrock E, Arbel N, van Vliet AR, La Rovere RM, et al. (2015) The BH4 domain of anti-apoptotic Bcl-XL, but not that of the related Bcl-2, limits the voltage-dependent anion channel 1 (VDAC1)-mediated transfer of pro-apoptotic Ca²⁺ signals to mitochondria. *J Biol Chem* 290: 9150-9161.
43. Tsai MF, Jiang D, Zhao L, Clapham D, Miller C (2014) Functional reconstitution of the mitochondrial Ca²⁺/H⁺ antiporter Letm1. *J Gen Physiol* 143: 67-73.
44. Brunelle JK, Letai A (2009) Control of mitochondrial apoptosis by the Bcl-2 family. *J Cell Sci* 122: 437-441.
45. Manning BD, Toker A (2017) AKT/PKB Signaling: Navigating the Network. *Cell* 169: 381-405.
46. Szado T, Vanderheyden V, Parys JB, De Smedt H, Rietdorf K, et al. (2008) Phosphorylation of inositol 1,4,5-trisphosphate receptors by protein kinase B/Akt inhibits Ca²⁺ release and apoptosis. *Proc Natl Acad Sci U S A* 105: 2427-2432.
47. Marchi S, Marinello M, Bononi A, Bonora M, Giorgi C, et al. (2012) Selective modulation of subtype III IP(3)R by Akt regulates ER Ca(2)(+) release and apoptosis. *Cell Death Dis* 3: e304.
48. Manning BD, Cantley LC (2007) AKT/PKB signaling: navigating downstream. *Cell* 129: 1261-1274.
49. Masliab-Planchon J, Pasmant E, Luscan A, Laurendeau I, Ortonne N, et al. (2013) MicroRNAome profiling in benign and malignant neurofibromatosis type 1-associated nerve sheath tumors: evidences of PTEN pathway alterations in early NF1 tumorigenesis. *BMC genomics* 14: 473.