

Xia et al., Chemo Open Access 2017, 6:4 DOI: 10.4172/2167-7700.1000246

# The Progress of Diallyl Disulfide in Anti-Cancer

#### Xia LZ<sup>#</sup>, Liao Q<sup>#</sup>, Wang H, Nie S, Liu Q, Oyang L, Chen X, Tan S, Tian Yo, Su M, Lin, Xia Luo J, Wang H and Zhou Y\*

Hunan Key Laboratory of Translational Radiation Oncology, Hunan Cancer Hospital and The Affiliated Cancer Hospital of Xiangya School of Medicine, Central South University, 283 Tongzipo Road, Changsha 410013, Hunan, PR China

#### #Authors contributed equally.

\*Corresponding author: Yujuan Zhou, Hunan Key Laboratory of Translational Radiation Oncology, Hunan Cancer Hospital and The Affiliated Cancer Hospital of Xiangya School of Medicine, Central South University, 283 Tongzipo Road, Changsha 410013, Hunan, PR China, Tel: 86-731-88651681; Fax: 86-731-88651999; Email: yujany\_zhou@163.com

Received date: November 15th, 2017; Accepted date: November 21st, 2017; Published date: November 24th, 2017

**Copyright:** ©2017 Xia LZ, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

#### Abstract

Considerable evidence in recent years suggests that garlic has anti-proliferative effects on various types of cancer. Garlic contains water-soluble and oil-soluble sulfur compounds. Oil-soluble compounds (OSCs), such as diallyl sulfide (DAS), diallyl disulfide (DADS), diallyl trisulfide (DATS) and ajoene are more effective than water-soluble compounds in protection against cancer. DADS, a major organosulfur compound derived from garlic, can reduce carcinogen-induced cancers in experimental animals and inhibit the proliferation of various types of cancer cells. The mechanisms of these action of DADS include activation of metabolic enzymes that detoxify carcinogens, suppression of the formation of DNA adducts, antioxidant effects, regulation of cell-cycle progression, induction of apoptosis, and inhibition of angiogenesis and metastasis. These topics are discussed in depth in this review.

**Keywords:** DADS; Cell-cycle; Apoptosis; Angiogenesis; Invasion; Organosulfur compounds; iNOS; COX-2

#### Introduction

Cancer is one of the most severe health problems that human beings are facing now. Epidemiological and experimental studies have shown that natural or synthetic ingredients in diet may affect the development of tumors. The daily diets made up of resveratrol, curcumin, genistein, garlic, and capsaicin can greatly reduce the risk of cancer or slower down the tumor progression [1]. As far back as ancient times, Egyptians realized that eating garlic could improve human health. In recent years, the anti-tumor effects of garlic and its active constituents have attracted extensive attention [2]. Garlic, commonly used as a food ingredient in cook, has been found to be associated with lower risk of certain cancers, such as lung, colorectal, and prostate cancers [3-5]. The organosulfur compounds in garlic can inhibit the development of cancer cell through targeting multiple signaling pathways that mediate the cell cycle arrest, apoptosis and antioxidants of cancer cells and the metabolism of carcinogens [6-9]. The intake of garlic is negatively associated with the occurrence and development of tumors, including oropharyngeal, esophageal, colorectal, laryngeal, breast, ovarian and prostate cancer and renal cell carcinoma [5,10].

As the main component of garlic oil-soluble compounds (OSCs), the antitumor and immunomodulatory functions of diallyl disulfide (DADS) have attracted attention of researchers [3]. More and more studies showed that DADS inhibits the development and progression of tumors, and the mechanism of action may be closely related to induction of cell cycle arrest and apoptosis, inhibition of oncogenic pathways and elimination of free radicals and oxidants [11]. This article updates the progression in DADS studies.

#### The anti-tumor effects of DADS

DADS, as one of the OSCs of garlic, has the potential of inhibiting tumor cell proliferation, promoting tumor cell apoptosis and inhibiting tumor cell metastasis in addition to the anti-inflammatory [12] and antibacterial activity [13]. Importantly, as a food ingredient, DADS has low toxicity and could be used as alternative chemotherapeutic agents for carcinomas [14]. DADS exerts anti-cancer effects via activating antioxidant enzymes to degrade carcinogens, inhibiting formation of carcinogen-DNA adducts [15,16], inducing cell cycle arrest and apoptosis [17-19], and inhibiting differentiation [20], angiogenesis, migration and invasion [21-23], etc. For instance, DADS can reduce 35% to 60% of DNA damage induced by N-nitrosodimethylamine in liver as assessed by the comet assay [24]. In liver cancer cells exposed to cytotoxic chemicals, DADS can accumulate intracellular ROS and induce the dysregulation of mitochondrial membrane potential, triggering DNA damage-induced G2/M phase arrest and mitochondrial apoptotic pathway [19]. Similarly, DADS can suppress apoptosis resistance induced by DCA through inhibiting the DCAinduced ROS production, inflammatory factors, IkBa phosphorylation, and expression of p50 in the nucleus in a dose-dependent manner in human barrett's epithelial cells [25,26]. On the other hand, an in vitro study reported that the oil-soluble compound diallyl disulfide were more potent inhibitor of human esophageal squamous cell carcinoma by inducing p53/p21-mediated G2/M phase arrest and apoptosis via p53/p21 and MEK-ERK pathway [27]. Recently, DADS was reported to inhibit the invasion and metastasis of MCF-7 cells in vitro by downregulating p38 activity [28]. The anti-tumor effect of DADS is achieved by regulating many complex signaling pathways. Additional studies on DADS are still needed.

#### The anti-tumor mechanisms of DADS

Garlic organosulfur compounds were tested for their anticancer potential in many isolated cell systems. However, the mechanisms underlying the anticancer effects of this class of compounds are not fully understood [29]. Several mechanisms have been proposed, including antioxidant activity, cell growth inhibition, effective stimulation of the immune response, and induction of apoptosis, which coincides with an increase in the percentage of cells blocked in the G2/M phase of the cell cycle [30]. In recent years, the studies have shown that DADS can suppress inflammation and apoptosis by inhibiting the ROS and NF- $\kappa$ B signaling pathways [10].

# Modulation of cytochrome p450-dependent monooxygenase by DADS

Cytochrome p450-dependent monooxygenase plays an important role in the metabolism of drugs, steroids, fat soluble vitamins, carcinogens, pesticides and other chemicals [31]. Carcinogenesis is a multistage and complex process that requires binding of a chemical carcinogen to DNA and formation of DNA adduct. Blockage of DNA adducts is an essential and important early step in prevention of carcinogenesis [32]. It's well-known that chemical carcinogens are often required to modulate metabolic activity through cytochrome p450-dependent monooxygenase and acceleration of carcinogen detoxification via induction of phase-I enzymes [33,34]. Animal experiments have also confirmed that DADS can promote the degradation of carcinogens by regulating the activity of cytochrome p450-dependent monooxygenase or activating the metabolism of phase-II enzymes [35]. For instance, Nkrumah-Elie and his colleagues had found that DADS can inhibit the carcinogenesis of BP which required metabolic activation to display its full carcinogenic potential by the cytochrome p450 pathway [36], via modulating the activation of multiple phases I and II enzymes, including CYP1A1, CYP1A2, CYP1B1, and epoxide enzymes in HepG2 cells [37-39]. Many studies have indicated that inhibition of CYP2E1 (cytochrome p450 2E1) with DADS leads to significant decrease in the single strand DNA damage, which can result in significant reduction in the DNA adduct level [16,40,41]. Not all of the subunits of cytochrome p450-dependent monooxygenase are inhibited by DADS, but DADS can degrade the substance closely associated with cyclophosphamide-induced progressive toxicity and oxidative stress by significantly up-regulation of CYP2B1/2 and CYP3A1 [42,43]. Therefore, the role of DADS in cytochrome p450 is also needed to be confirmed by a large number of studies.

## Promotion of the antioxidant response by DADS

The term ROS encompasses a wide range of molecules, including superoxide (O2-), hydrogen peroxide (H2O2), hydroxyl radical, and peroxynitrite [44]. Redox control is achieved through the cooperative and coordinated action of antioxidant enzymes, such as catalase and low-molecular-weight antioxidant compounds, such as glutathione (GSH) [45]. Apart from being directly implicated in carcinogenesis, ROS are strongly involved in tumor proliferation, survival, and resistance to chemotherapeutic agents [46]. On the one hand, the constitutive high level of ROS in some cancer cells appears to promote their proliferation. Additional amounts of ROS above a certain threshold may further cause cell-cycle arrest and/or apoptosis [47]. Recent studies have shown that DADS achieves anti-tumor effects by activating antioxidant responses via inhibiting the expression of ROS [15]. The progressive toxicity and oxidative stress induced by DADS play an important role in the pathogenesis of cancer through changing the structure of chromosomes [48,49]. Accordingly, it was observed that NADPH oxidase and ROS play a pivotal role in DADS-induced apoptosis [50]. DADS caused pronounced intracellular ROS

accumulation, phosphorylation, and expression of Bcl-2 to cause apoptosis through the activation of JNK and inactivation of ERK1/2 and Akt.

The effects of DADS on IL-1β-induced intracellular ROS production and lipid peroxidation were detected and the proteins expression of Nrf2, Bax, Bcl-2, caspase-3, total and phosphorylated JNK, and p38 MAPKs were evaluated [26]. Nuclear transcriptional factor 2 (Nrf2) is an activator of the antioxidant responsive element (ARE) [51]. In DMBA/TPA-induced cancer cells, the nuclear localization of Nrf2 is reduced, leading to the inhibition of the activation of ARE, resulting in decreased activity of the antioxidant enzymes which activity is essential to antioxidant responses [52]. However, DADS is capable to inhibit the tumorigenesis and progression of TPA-induced tumor via promoting the localization of Nrf2 by up-regulating the expression of p21, or inhibiting the degradation of Nrf2 [15]. Therefore, Nrf2 is critical for the inhibition of mouse skin cancer by DADS and can become a promisinglytherapeutic target. However, the mechanisms that DADS inhibit the activation of oncogenic factors and the expression of protooncogenes activated by carcinogenic factors are still not very clear, which requires a large number of studies to clarify.

#### Induction of G2/M phase arrest by DADS

Studies have confirmed that the anti-proliferative activity of DADS is associated with a decrease in the percentage of cells in the G1 phase and a phase G2/M arrest in many human cancer cells [53], but the mechanisms are not very clear. Previous studies indicated that DADS induces G2/M phase arrest in MGC803 cells [54]. High phosphorylation of checkpoint kinase 1(Chk1), low phosphorylation of Chk2 and the low expression of Cdc25c, 14-3-3 and cyclin B1 are closely associated with the inactivation of G2/M checkpoint [55]. However, some researchers found that the overexpression of Chk1, but not Chk2, exhibited increased accumulation in G2/M phase [56]. In addition, DADS can cause high phosphorylation of Cdc2 and decrease the expression of Cdc25c protein that is critical in G2/M arrest [57]. Actually, Cdc2 activation depends on the dephosphorylation of Tyr15 by Cdc25c [27]. However, p53 and the p53-responsive gene, p21, can decrease cyclin B1 and Cdc2 expression by inhibiting either Cdc2 kinase activity or blocking the interaction of cyclin B1-Cdc2 complexes with their substrates, leading to G2/M-phase cell cycle arrest, which indicated that p53/p21 signaling pathway may be the mechanism underlying G2/M phase arrest [58-60]. In addition, DADS can induce G2/M phase cell cycle arrest by activating p38 [57], and suppressing the ATR/Chk1/Cdc25C/cyclinB1 signaling pathway by specifically activating Chk1 [61].

DNA damage has been reported to induce activation of cell cycle arrest to enhance cellular DNA repair capacity [62]. Yang and other researchers found that DADS could induce the DNA damage through up-regulating ROS protein levels in skin cancer cells, and then activate p21, one of the downstream genes of p53, to affect G2/M regulators, including Wee l kinase, Cdc25c and Cdc2 (p34), etc. Simultaneously, p53 can also activate the mitochondrial apoptotic pathway by activating caspase-9, caspase-3 and PARP, thereby promoting apoptosis in skin cancer cells [62]. Recent studies have verified that DADS-induced DNA damage can cause specific phosphorylation of the Ser345 site of the Chk1 protein and interact with the downstream effectors that induce G2/M phase arrest in PaCa-2 MIA cancer cells [19]. The phosphorylation of Chk1 is able to induce the phosphorylation of Ser216 site of the Cdc25c [63]. The degradation of

Cdc25c protein significantly inhibits the formation of the complex of cell protein dependent kinase, including Cdc2/cyclin B1 (a crucial driver of phase G2/M transitions), etc [64]. Ling et al. found that DADS could increase the rate of phase G0/G1 cells and accordingly reduce the rate of phase S cells in leukemiacells HL-60 treated with different concentrations of DADS [20]. But the mechanisms need to be further elucidated.

#### Activation of the apoptotic pathways by DADS

Apoptosis is a phenomenon of programmed cell death in normal cells. Apoptosis pathways contain the death receptor-mediated extrinsic pathway [65], the intrinsicmitochondrial pathway [65], granzyme B-mediated pathway [66], and the endoplasmic reticulum stress-mediated pathway [67]. The protein closely associated with apoptosis mainly contains four important protein molecules involved in apoptotic pathways: caspases (the proteases which execute cell death), adapter proteins (caspase activators), Bcl-2 family proteins and inhibitors of apoptosis proteins (IAPs) [68]. In the experiment of apoptotic cell models established by pretreatment of 10 µM DADS in HeLa cells with 2Gy high-LET carbon beams, some researchers found that DADS can obviously promote the activation of apoptosis signal via up-regulating the balance between Tap73 pathway (transcriptionally active p73 isoforms) and  $\Delta Np73$  (N-terminally truncated anti-apoptotic p73 isoforms). At the same time, DADS can deregulate important molecules involved in internal and external apoptotic pathways, such as apoptotic protease-activating factor-1(APAF1), IL10 and FASLG [69]. Among them, APAF1 plays a key role in the mechanism of apoptosis and can activate the caspase cascade reaction responsible for the apoptotic effector phase [70]. In addition, IL-10 in serum induces apoptosis of T cells by activating the caspase-8 pathway by Fas signaling [71]. New research confirms that receptor for advanced glycation endproduct (RAGE) can specifically deliver DADS into three negative breast cancer cells and significantly enhance the cytotoxicity of DADS by reducing anti-apoptotic proteins and upregulation of pro-apoptotic protein [72]. The researchers have found that DADS-RAGE-SLN can be obviously absorbed by MDA-MB231 and upregulate the expression of pro-apoptotic proteins (Bax, caspase-3, and caspase-9) and downregulate the expression of antiapoptotic proteins, such as Bcl-2 family (Bcl-2, Bcl-xl, Mcl-1) and survivin through the experiment of establishing the model of solid lipid nanoparticles with DADS-loaded solid lipid location nanoparticles (DADS-SLN), and DADS-loaded solid lipid location nanoparticles with the RAGE positioning (DADS-RAGE-SLN) [72]. In fact, caspase-3 plays an important role as the central effector for initiation of apoptosis [73]. DADS promoted the apoptosis of ECA109 cells by downregulating Bcl-2 mRNA expression and upregulating the Bax expression in a dose-dependent manner, thus increasing the Bax/ Bcl-2 ratio, leading to a pro-apoptotic process via caspase-3 pathway. In one word, DADS controls cell apoptosis by activating caspase-3, upregulating Bax/Bcl-2 ratio and downregulating the MEK-ERK signaling pathway [19,27,41,74].

#### Regulation of the miRNAs by DADS

MicroRNAs (miRNAs) are a class of post-transcriptional regulators that negatively regulate plant development, signal transduction, and response to abiotic stresses and pathogen invasions [75-77]. Recently, numerous studies found that DADS could activate/or inhibit the expression of some miRNAs to regulate cell growth, cell cycle progression and apoptosis of human cancer cells [32,78]. For example, miR-200b and miR-22 synergistically induces apoptosis of human GCs (gastric cancer cells) by enhancing theeffects of DADS in vitro and in vivo via the Wnt-1 signaling pathway [79]. However, different miRNAs in different contexts are aberrantly expressed in various cancers, such as prostatic cancer, esophageal squamous cell carcinoma, breast cancer, and gastric cancer. For example, the miR-22 shows different effects in these cancers [80-83]. DADS can silence HIF-1a (hypoxia inducible factor 1a) via increasing expression of miR-22 to repress VEGF (vascular endothelial growth factor) expression to block angiogenesis, leading to the disruption of cancer progression [79,84]. The upregulation of miR-22 induced by DADS may post-transcriptionally target cyclin A2 and CDKN1A (cyclin-dependent kinase inhibitor 1A) to arrest the cell cycle in G0/G1 phage in CRC and liver cancer cells, respectively [85,86]. The recent researches have confirmed that DADS induces apoptosis of SGC-7901 cells by upregulation of miR-34a, via inhibition of the PI3K-Akt signaling pathway [87]. In addition, DADS induced neovasculogenesis through the modulation of crucial signaling pathways and the suppression of miR 221 via mediating the upregulation of c-kit expression and the phosphorylation (activation) of the Akt signaling cascade in human EPCs (endothelial progenitor cells) [32]. However, the role of DADS on miRNAs is still in infancy and needs to be studied by researchers.

#### Inhibition of tumor metastasis by DADS

Epithelial-mesenchymal transition (EMT) is an important phenomenon in the development of tumorigenesis, and is also an important mechanism for the invasion of tumor cells and secondary metastasis [88]. The cells with EMT are present certain anti-apoptotic activity and drug resistance [89]. The signaling pathways involved in EMT include TGF- $\beta$ , Wnt, Hedgehog, Notch, ILK, and uPAR (urokinase-type plasminogen activator receptor) [90-95]. A recent study has revealed that DADS induces the reversal of the EMT and inhibits cell growth by inactivating the  $\beta$ -catenin signaling pathway in breast cancer cells [96]. Interestingly, DADS inhibits the occurrence of EMT in tumor cells by inhibiting multiple EMT-related signaling pathways.

Previous researches demonstrated that the DADS-induced downregulation of uPAR results in the inhibition of the ERK/Fra-1 pathway in addition to gastric cancer cell migration and invasion [97]. DADS downregulates LIM kinase-1 (LIMK1), vimentin and EMTrelated proteins (Slug, Snail), and upregulates E-cadherin and TIMP-3 (tissue inhibitor of metalloproteinase-3) expression which are the signs of the occurrence of EMT, suggesting that DADS may reverse EMT by downregulating LIMK1 [97,98]. DADS suppresses actin cytoskeletal remodeling and cell pseudopodia formation, and the occurrence of EMT via down regulating LIMK1 expression to decrease the expression of vimentin, CD34, and Ki-67; increase E-cadherin expression; and impede cancer cell proliferation, angiogenesis, and EMT alteration, thereby inhibiting invasion and metastasis [98-100]. Actually, the effects of DADS on E-cadherin and vimentin expression may result in blockage of the ERK/Fra-1 pathway by inhibition of LIMK1 expression and activity.

MMPs (matrix metalloprotease) have many physiological functions in tumor metastasis, *via* degrading extracellular matrix and basement membrane, which then caused the integrity of basement membrane to be localized for infiltration and distant metastasis [101]. Thus inhibition of MMPs activity could prevent or suppress the tumor metastasis [102,103]. Park et al. have found that DADS significantly inhibited cell motility and invasive activity by decreasing MMP activity

#### Page 3 of 8

and tightening TJs (tight junctions) [104]. The anti-invasive activity of DADS was associated with inhibition of MMP-2 and MMP-9 activities through elevation of TIMPs expression, and increased TIMPs/MMPs ratio is a key factor in regulation of the anti-metastatic process [104]. Recently, the DADS was confirmed that it could inhibit the invasion and migration of MCF-7 cells in a dose-dependent manner by downregulated the expression of vimentin and MMP-9 and up-regulated Ecadherin expression in the cells via down-regulating the activity of p38 which activates the TGF- $\beta$  signaling pathway [15]. The recent research found that DADS dose-dependently inhibited HIF-1a transcriptional activity and hypoxia-induced hematogenous metastasis of MDA-MB-231 cells rather than inhibition of HIF-1a mRNA expression or ubiquitin proteasome degradation [23]. In addition, DADS can alter cell morphology and cell motility via inhibiting the activation of PI3K/Akt signal pathway, and then inhibited the tumor migration and invasion [105]. However, the inhibition by DADS on tumor metastasis is a complex process, and it is not limited to affecting one key molecule related to tumor metastasis, more studies are needed.

#### The inhibition of angiogenesis by DADS

Angiogenesis is essential for proper tissue development, but neovascularization can facilitate the progression of pathological conditions, such as tumor growth and survival. Aberrant neovascularization is an essential pathogenic mechanism in many human diseases, including tumor growth and survival [106,107]. Tumor angiogenesis results in abnormally formed, tortuous, and poorly organized vessels that exhibit altered permeability [108,109]. Previous researches have confirmed that the aberrant activated TNF-a receptor signaling is associated with coordination of tumor angiogenesis and metastasis [110]. DADS can inhibit the CCL2 (CC chemokine-2) releasing induced by TNF-a to play an antiangiogenesis role in MDA-MB-231 cells. The decreasing level of CCL2 attenuates the phosphorylation of ERK which is a key molecule of the MAPK signaling pathway, leading to the weakening of angiogenesis in tumor cells [111]. Activation of ERK and Akt signaling can promote the angiogenesis of HCC (hepatocellular carcinoma) [112]. Recently, DADS has been shown to reduce migration, invasion, and angiogenesis of human colon cancer in part mediated by NF-KB, ERK1/2, JNK1/2, and p38 signaling [113]. Xiao et al. found that the pexpression decreased Akt protein with the increasing DADS concentration[114], and led to attenuating the proangiogenesis effect of VEGF. The effect of DADS on angiogenesis may be related to the regulation of metalloproteinase activation which is essential for the pro-angiogenic growth factors available to their receptors [115].

However, pervious study found that DADS and DATS dosedependently enhance the neovasculogenesis of human EPCs (endothelial progenitor cells), in part through the up-regulation of ckit protein as well as the activation of the PI3K/Akt/NF- $\kappa$ B cascades and MAPK/ERK pathway [32]. DADS significantly induces the phosphorylation of the Akt, and I- $\kappa$ Ba proteins in human EPCs thus activating the PI3K/Akt and NF- $\kappa$ B signaling pathways. Meanwhile, DADS also augment the phosphorylation of the GSK-3 $\beta$  protein and stabilize its downstream target  $\beta$ -catenin protein through the suppression of the phosphorylated  $\beta$ -catenin level, resulting in the inactivation of  $\beta$ -catenin signaling pathway [40,114]. Based on the proangiogenic effects of DADS in human EPCs, whether or not the DADS has the same effect in tumor still needs further researches. Page 4 of 8

#### The inhibition of inflammation by DADS

Inflammation is a main component of the host immune response to infection, and can have both protective and pathogenic roles. Some pro-inflammatory cytokines play pathogenic roles during tumorigenesis [116]. Particularly, chronic inflammation creates and maintains a tumor microenvironment where there is migration and a relative abundance of immune inflammatory cells and cytokines [117]. For example, studies have shown that tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), IL-8 and IL-10 levels are higher in patients with inflammation-associated cancer [118]. DADS has been shown to have both immunomodulatory and anti-inflammatory effects in several types of cancer [7,119].

Although a recent research paper reported that the antiinflammatory activity of garlic OSCs help reduce TNF-a, IL-6, and iNOS [120], whether DADS can suppress inflammatory bowel disease and the molecular mechanisms haven't been investigated yet [116]. Many studies has been shown that DADS could suppress nitric oxide production through inhibiting the activation of NF-kB (nuclear factor kappa B), and attenuating expression of inducible nitric oxide synthase (iNOS) and blocking the nuclear translocation of NF-KB [121]. Interestingly, ROS are recognized as an important link between inflammation and cancer, who can induce tumor cell proliferation and mutagenesis and inhibits apoptosis via activating various signaling pathways, such as NF-KB signaling pathway. Constitutive NF-KB activation in inflammatory bowel disease is associated with an increased risk of developing colorectal cancer [122]. The recent research indicated that DADS exerted its protective effects against colorectal tumors by suppressing inflammation in the colitis-induced colorectal cancer AOM/DSS mouse model [123]. The suppressive effect of DADS on NF-KB activity is likely due to its ability to inhibit the nuclear translocation of the phosphorylated NF-kB heterodimeric Rel-A (P65S536)-p50 complex. Meanwhile, DADS can induce the inactivation of GSK-3( $\alpha/\beta$ ) that can activate the NF- $\kappa$ B signaling cascade by enhancing the transcriptional activity of NF-KB in the nucleus by blocking its phosphorylation of serine residue 21/9 [124,125] . Previous studies have confirmed that DADS can obviously decrease ROS production as a natural antioxidant both in vivo and in vitro and pro-inflammatory cytokines levels by inhibiting the phosphorylation of IkBa and the nuclear translocation of p65 and p50 to inactivate the NF-κB signaling pathways [126]. Subsequent researches have verified that DADS can inhibit the expression of iNOS and cyclooxygenase (COX-2) in activated RAW 264.7 cells. Due to the inhibition of signal transduction for the iNOS and COX-2 gene expression and the direct inhibition of iNOS and COX-2, the production of NO and PGE2 (prostaglandin E2) is significantly inhibited. NO, PGE2, and related enzymes have been implicated as important mediators in the inflammation process. DADS has antiinflammatory activity, including the reduction of pro-inflammatory cytokines, such as TNF- $\alpha$ , a key player in the development of tumors. The concrete anti-inflammatory mechanism of DADS does not elucidate very clearly, which need to further study.

#### Conclusions

As a major component of garlic, more and more evidence indicates that DADS could have broad-spectrum anti-tumor effects, but has no toxic effects in healthy cells. The mechanism of action of DADS is closely related with antioxidants, regulation of cell-cycle arrest, induction of apoptosis and inhibition of angiogenesis and invasion (Figure 1).



**Figure 1:** Schema pattern of molecular mechanisms of DADS on anti-cancer effects; DADS inhibits the development and progression of tumors, and the mechanism of action may be closely related to induction of cell cycle arrest and apoptosis, inhibition of oncogenic pathways and elimination of free radicals and oxidants, which achieved by regulating many complex signaling pathways, such as inhibiting the ROS, PI3K/Akt, MAPK and NF- $\kappa$ B signaling pathways.

However, the applications of DADS are only demonstrated *in vitro* and *in vivo* experiments on cancer cell lines and animals. It is implemented into clinical practice thus far because the potential use of DADS in the clinical treatment of cancer has not been excavated. The high concentrations of DADS used in animal studies are unlikely to be physiologically achievable in humans if such compounds are considered for clinical applications, and how to modify the DADS construction to get the clinical effects of cancer treatment is a challenge. Nevertheless, base on the existing studies, it indicates that DADS may be a prospective agent for multi-targeted prevention and/or treatment against human cancer.

## Acknowledgement

This work was supported in part by grants from the National Natural Science Foundation of China (81402006).

## References

- 1. Walczak K, Marciniak S, Rajtar G (2017) Cancer chemoprevention selected molecular mechanisms. Postepy Hig Med Dosw 71: 149-61.
- Xie X, Huang X, Tang H, Ye F, Yang L, et al. (2017) Diallyl Disulfide Inhibits Breast Cancer Stem Cell Progression and Glucose Metabolism by Targeting CD44/PKM2/AMPK Signaling. Curr Cancer Drug Targets.
- Horn N, Miller G, Ajuwon KM, Adeola O (2017) Garlic diallyl disulfide and diallyl trisulfide mitigates effects of pro-oxidant induced cellular stress and has immune modulatory function in LPS-stimulated porcine epithelial cells. J Anim Sci 95: 4045-4051.
- 4. Talib WH (2017) Consumption of garlic and lemon aqueous extracts combination reduces tumor burden by angiogenesis inhibition, apoptosis induction, and immune system modulation. Nutrition 44: 89-97.
- Reiter J, Levina N, van der Linden M, Gruhlke M (2017) Diallylthiosulfinate (Allicin), a Volatile Antimicrobial from Garlic (Allium sativum), Kills Human Lung Pathogenic Bacteria, Including MDR Strains, as a Vapor. Molecules.
- Iciek M, Kwiecien I, Wlodek L (2009) Biological properties of garlic and garlic-derived organosulfur compounds. Environ Mol Mutagen 50: 247-265.
- Wallace GC, Haar CP, Vandergrift WA, Giglio P, Dixon-Mah YN, et al. (2013) Multi-targeted DATS prevents tumor progression and promotes apoptosis in ectopic glioblastoma xenografts in SCID mice via HDAC inhibition. J Neurooncol 114: 43-50.

- Li W, Tian H, Li L, Li S, Yue W, et al.(2012) Diallyl trisulfide induces apoptosis and inhibits proliferation of A549 cells in vitro and in vivo. Acta Biochim Biophys Sin 44: 577-583.
- Lai KC, Hsu SC, Yang JS, Yu CC, Lein JC, et al.(2015) Diallyl trisulfide inhibits migration, invasion and angiogenesis of human colon cancer HT-29 cells and umbilical vein endothelial cells, and suppresses murine xenograft tumour growth. J Cell Mol Med 19: 474-484.
- 10. Galeone C, Pelucchi C, Levi F, Negri E, Franceschi S, et al. (2006) Onion and garlic use and human cancer. Am J Clin Nutr 84: 1027-1032.
- 11. Antony ML, Singh SV (2011) Molecular mechanisms and targets of cancer chemoprevention by garlic-derived bioactive compound diallyl trisulfide. Indian J Exp Biol 49: 805-816.
- 12. Park HY, Kim ND, Kim GY, Hwang HJ, Kim BW, et al.(2012) Inhibitory effects of diallyl disulfide on the production of inflammatory mediators and cytokines in lipopolysaccharide-activated BV2 microglia. Toxicol Appl Pharmacol 262: 177-184.
- Busquet M, Calsamiglia S, Ferret A, Carro MD, Kamel C (2005) Effect of garlic oil and four of its compounds on rumen microbial fermentation. J Dairy Sci 88: 4393-4404.
- 14. Yeruva L, Elegbede JA, Carper SW (2008) Methyl jasmonate decreases membrane fluidity and induces apoptosis through tumor necrosis factor receptor 1 in breast cancer cells. Anticancer Drugs 19: 766-776.
- Shan Y, Wei Z, Tao L, Wang S, Zhang F, et al. (2016) Prophylaxis of Diallyl Disulfide on Skin Carcinogenic Model via p21-dependent Nrf2 stabilization. Sci Rep 6: 35676.
- Sapkota M, Hottor TK, DeVasure JM, Wyatt TA, McCaskill ML (2014) Protective role of CYP2E1 inhibitor diallyl disulfide (DADS) on alcoholinduced malondialdehyde-deoxyguanosine (M1dG) adduct formation. Alcohol Clin Exp Res 38: 1550-1558.
- 17. Puccinelli MT, Stan SD (2017) Dietary Bioactive Diallyl Trisulfide in Cancer Prevention and Treatment. Int J Mol Sci 18.
- Choi YH (2017) Diallyl trisulfide induces apoptosis and mitotic arrest in AGS human gastric carcinoma cells through reactive oxygen speciesmediated activation of AMP-activated protein kinase. Biomed Pharmacother 94: 63-71.
- 19. Saini V, Manral A, Arora R, Meena P, Gusain S, et al. (2017) Novel synthetic analogs of diallyl disulfide triggers cell cycle arrest and apoptosis via ROS generation in MIA PaCa-2 cells. Pharmacol Rep 69: 813-821.
- Ling H, He J, Tan H, Yi L, Liu F, et al. (2017) Identification of potential targets for differentiation in human leukemia cells induced by diallyl disulfide. Int J Oncol 50: 697-707.
- 21. Shin SS, Song JH, Hwang B, Park SL, Kim WT, et al. (2017) Angiopoietin-like protein 4 potentiates DATS-induced inhibition of proliferation, migration, and invasion of bladder cancer EJ cells; involvement of G2/M-phase cell cycle arrest, signaling pathways, and transcription factors-mediated MMP-9 expression. Food Nutr Res.
- 22. Hayashida R, Kondo K, Morita S, Unno K, Shintani S, et al. (2017) Diallyl Trisulfide Augments Ischemia-Induced Angiogenesis via an Endothelial Nitric Oxide Synthase-Dependent Mechanism. Circ J 81: 70-78.
- 23. Wei Z, Shan Y, Tao L, Liu Y, Zhu Z, et al. (2017) Diallyl trisulfides, a natural histone deacetylase inhibitor, attenuate HIF-1alpha synthesis, and decreases breast cancer metastasis. Mol Carcinog 56: 2317-2331.
- 24. Singh V, Belloir C, Siess MH, Le Bon AM (2006) Inhibition of carcinogen-induced DNA damage in rat liver and colon by garlic powders with varying alliin content. Nutr Cancer 55: 178-184.
- 25. Feng C, Luo Y, Nian Y, Liu D, Yin X, et al. (2017) Diallyl Disulfide Suppresses the Inflammation and Apoptosis Resistance Induced by DCA Through ROS and the NF-kappaB Signaling Pathway in Human Barrett's Epithelial Cells. Inflammation 240: 818-831.
- 26. Hosseinzadeh A, Jafari D, Kamarul T, Bagheri A, Sharifi AM (2017) Evaluating the Protective Effects and Mechanisms of Diallyl Disulfide on Interlukin-1beta-Induced Oxidative Stress and Mitochondrial Apoptotic Signaling Pathways in Cultured Chondrocytes. J Cell Biochem 118: 1879-1888.

- 27. Yin X, Zhang R, Feng C, Zhang J, Liu D, et al. (2014) Diallyl disulfide induces G2/M arrest and promotes apoptosis through the p53/p21 and MEK-ERK pathways in human esophageal squamous cell carcinoma. Oncol Rep 32: 1748-1756.
- 28. Chen XX, Liu XW, Zhou ZG, Chen XY, Li LD, et al. (2016) Diallyl disulfide inhibits invasion and metastasis of MCF-7 breast cancer cells in vitro by down-regulating p38 activity. Nan Fang Yi Ke Da Xue Xue Bao 36: 814-818.
- 29. Diretto G, Rubio-Moraga A, Argandona J, Castillo P, Gomez-Gomez L, et al. (2017) Tissue-Specific Accumulation of Sulfur Compounds and Saponins in Different Parts of Garlic Cloves from Purple and White Ecotypes. Molecules.
- 30. Wahid S, Versiani MA, Jahangir S, Jawaid K, Shafique M, et al. (2017) Phytochemical and Biological Activities of Pseudocalymma elegans: A False Garlic. Chem Biodivers.
- Yamazaki H (2017) Differences in Toxicological and Pharmacological Responses Mediated by Polymorphic Cytochromes P450 and Related Drug-Metabolizing Enzymes. Chem Res Toxicol 30: 53-60.
- 32. Chiang EP, Chiu SC, Pai MH, Wang YC, Wang FY, et al. (2013) Organosulfur garlic compounds induce neovasculogenesis in human endothelial progenitor cells through a modulation of MicroRNA 221 and the PI3-K/Akt signaling pathways. J Agric Food Chem 61: 4839-4849.
- Powolny AA, Singh SV (2008) Multitargeted prevention and therapy of cancer by diallyl trisulfide and related Allium vegetable-derived organosulfur compounds. Cancer Lett 269: 305-314.
- 34. Guo Z, Sevrioukova IF, Denisov IG, Zhang X, Chiu TL, et al. (2017) Heme Binding Biguanides Target Cytochrome P450-Dependent Cancer Cell Mitochondria. Cell Chem Biol 24: 1259-1275.
- Yi L, Su Q (2013) Molecular mechanisms for the anti-cancer effects of diallyl disulfide. Food Chem Toxicol 57: 362-370.
- 36. Jiang H, Shen YM, Quinn AM, Penning TM (2005) Competing roles of cytochrome P450 1A1/1B1 and aldo-keto reductase 1A1 in the metabolic activation of (+/-)-7,8-dihydroxy-7,8-dihydro-benzo[a]pyrene in human bronchoalveolar cell extracts. Chem Res Toxicol 18: 365-374.
- Belloir C, Singh V, Daurat C, Siess MH, Le Bon AM (2006) Protective effects of garlic sulfur compounds against DNA damage induced by direct- and indirect-acting genotoxic agents in HepG2 cells. Food Chem Toxicol 44: 827-834.
- 38. Arranz N, Haza AI, Garcia A, Moller L, Rafter J, et al. (2007) Protective effects of organosulfur compounds towards N-nitrosamine-induced DNA damage in the single-cell gel electrophoresis (SCGE)/HepG2 assay. Food Chem Toxicol 45: 1662-1669.
- 39. Garcia A, Haza AI, Arranz N, Delgado ME, Rafter J, et al. (2008) Organosulfur compounds alone or in combination with vitamin C protect towards N-nitrosopiperidine- and N-nitrosodibutylamineinduced oxidative DNA damage in HepG2 cells. Chem Biol Interact 173: 9-18.
- 40. Ko JW, Shin JY, Kim JW, Park SH, Shin NR, et al. (2017) Protective effects of diallyl disulfide against acetaminophen-induced nephrotoxicity: A possible role of CYP2E1 and NF-kappaB. Food Chem Toxicol 102: 156-165.
- 41. Ko JW, Park SH, Shin NR, Shin JY, Kim JW, et al. (2017) Protective effect and mechanism of action of diallyl disulfide against acetaminopheninduced acute hepatotoxicity. Food Chem Toxicol 109: 28-37.
- 42. Kim SH, Lee IC, Baek HS, Moon C, Kim SH, et al. (2014) Induction of cytochrome P450 3A1 expression by diallyl disulfide: protective effects against cyclophosphamide-induced embryo-fetal developmental toxicity. Food Chem Toxicol 69: 312-319.
- **43.** Kim SH, Lee IC, Baek HS, Shin IS, Moon C, et al. (2014) Mechanism for the protective effect of diallyl disulfide against cyclophosphamide acute urotoxicity in rats. Food Chem Toxicol 64: 110-118.
- 44. Smith RL, Tan JME, Jonker MJ, Jongejan A, Buissink T, et al. (2017) Beyond the polymerase-gamma theory: Production of ROS as a mode of NRTI-induced mitochondrial toxicity. PLoS One.
- 45. Williams CF, Vacca AR, Dunham L, Lloyd D, Coogan MP, et al. (2016) The redox-active drug metronidazole and thiol-depleting garlic

compounds act synergistically in the protist parasite Spironucleus vortens. Mol Biochem Parasitol 206: 20-28.

- De Gianni E, Fimognari C (2015) Anticancer Mechanism of Sulfur-Containing Compounds. Enzymes 37: 167-192.
- 47. Eleftheriadis T, Pissas G, Antoniadi G, Liakopoulos V, Stefanidis I (2017) Allopurinol protects human glomerular endothelial cells from high glucose-induced reactive oxygen species generation, p53 overexpression and endothelial dysfunction. Int Urol Nephrol.
- Wang DD, Liu Y, Li N, Zhang Y, Jin Q, et al. (2017) Induction of CYP1A1 increases gefitinib-induced oxidative stress and apoptosis in A549 cells. Toxicol In Vitro.
- Schmitt G, Barrow P (2016) Regulatory Approaches to Non-clinical Reproductive Toxicity Testing of Anti-Cancer Drugs. Anticancer Agents Med Chem.
- 50. Wang W, Yao GD, Shang XY, Gao JC, Zhang Y, et al. (2017) Eclalbasaponin I from Aralia elata (Miq.) Seem. reduces oxidative stressinduced neural cell death by autophagy activation. Biomed Pharmacother 97: 152-161.
- 51. Lee IC, Kim SH, Baek HS, Moon C, Kang SS, et al. (2014) The involvement of Nrf2 in the protective effects of diallyl disulfide on carbon tetrachloride-induced hepatic oxidative damage and inflammatory response in rats. Food Chem Toxicol 63: 174-185.
- 52. Kwei KA, Finch JS, Thompson EJ, Bowden GT (2004) Transcriptional repression of catalase in mouse skin tumor progression. Neoplasia 6: 440-448.
- Liao QJ, Su J, He J, Song Y, Tang HL, et al. (2009) Effect of diallyl disulfide on cell cycle arrest of human colon cancer SW480 cells. Ai Zheng 28: 138-141.
- 54. Ling H, Zhang LY, Su Q, Song Y, Luo ZY, et al. (2006) Erk is involved in the differentiation induced by diallyl disulfide in the human gastric cancer cell line MGC803. Cell Mol Biol Lett 11: 408-423.
- 55. Ashra H, Rao KV (2006) Elevated phosphorylation of Chk1 and decreased phosphorylation of Chk2 are associated with abrogation of G2/M checkpoint control during transformation of Syrian hamster embryo (SHE) cells by Malachite green. Cancer Lett 237: 188-198.
- 56. Bo S, Hui H, Li W, Hui L, Hong X, et al. (2014) Chk1, but not Chk2, is responsible for G2/M phase arrest induced by diallyl disulfide in human gastric cancer BGC823 cells. Food Chem Toxicol 68: 61-70.
- 57. Paul P, Rajendran SK, Peuhu E, Alshatwi AA, Akbarsha MA, et al. (2014) Novel action modality of the diterpenoid anisomelic acid causes depletion of E6 and E7 viral oncoproteins in HPV-transformed cervical carcinoma cells. Biochem Pharmacol 89: 171-184.
- Zhang Z, Wang CZ, Du GJ, Qi LW, Calway T, et al. (2013) Genistein induces G2/M cell cycle arrest and apoptosis via ATM/p53-dependent pathway in human colon cancer cells. Int J Oncol 43: 289-296.
- 59. Ding L, Huang Y, Du Q, Dong F, Zhao X, et al. (2014) TGEV nucleocapsid protein induces cell cycle arrest and apoptosis through activation of p53 signaling. Biochem Biophys Res Commun 445: 497-503.
- 60. Ling H, Wen L, Ji XX, Tang YL, He J, et al. (2010) Growth inhibitory effect and Chk1-dependent signaling involved in G2/M arrest on human gastric cancer cells induced by diallyl disulfide. Braz J Med Biol Res 43: 271-278.
- 61. Wang HC, Yang JH, Hsieh SC, Sheen LY (2010) Allyl sulfides inhibit cell growth of skin cancer cells through induction of DNA damage mediated G2/M arrest and apoptosis. J Agric Food Chem 58: 7096-8103.
- 62. Daugas E, Susin SA, Zamzami N, Ferri KF, Irinopoulou T, et al. (2000) Mitochondrio-nuclear translocation of AIF in apoptosis and necrosis. FASEB J 14: 729-739.
- 63. Reinhardt HC, Yaffe MB (2009) Kinases that control the cell cycle in response to DNA damage: Chk1, Chk2, and MK2. Curr Opin Cell Biol 21: 245-255.
- 64. Fulda S, Debatin KM (2006) Extrinsic versus intrinsic apoptosis pathways in anticancer chemotherapy. Oncogene 25: 4798-4811.

- 65. Munoz-Pinedo C (2012) Signaling pathways that regulate life and cell death: evolution of apoptosis in the context of self-defense. Adv Exp Med Biol 738: 124-143.
- 66. Logue SE, Cleary P, Saveljeva S, Samali A (2013) New directions in ER stress-induced cell death. Apoptosis 18: 537-546.
- 67. Zhou S, Wang Y, Zhu JJ (2016) Simultaneous Detection of Tumor Cell Apoptosis Regulators Bcl-2 and Bax through a Dual-Signal-Marked Electrochemical Immunosensor. ACS Appl Mater Interfaces 8: 7674-7682.
- Di C, Sun C, Li H, Si J, Zhang H, et al. (2015) Diallyl disulfide enhances carbon ion beams-induced apoptotic cell death in cervical cancer cells through regulating Tap73 /DeltaNp73. Cell Cycle 14: 3725-3733.
- 69. Cecconi F (1999) Apaf1 and the apoptotic machinery. Cell Death Differ 6: 1087-1098.
- 70. Yang X, Sun B, Wang H, Yin C, Wang X, et al. (2015) Increased serum IL-10 in lupus patients promotes apoptosis of T cell subsets via the caspase 8 pathway initiated by Fas signaling. J Biomed Res 29: 232-240.
- Siddhartha VT, Pindiprolu S, Chintamaneni PK, Tummala S, Nandha Kumar S (2017) RAGE receptor targeted bioconjuguate lipid nanoparticles of diallyl disulfide for improved apoptotic activity in triple negative breast cancer: in vitro studies. Artif Cells Nanomed Biotechnol 1-11.
- 72. Mohamed MS, Bishr MK, Almutairi FM, Ali AG (2017) Inhibitors of apoptosis: clinical implications in cancer. Apoptosis.
- 73. Curti V, Di Lorenzo A, Dacrema M, Xiao J, Nabavi SM, et al. (2017) In vitro polyphenol effects on apoptosis: An update of literature data. Semin Cancer Biol 46: 119-131.
- 74. Yang L, Mu X, Liu C, Cai J, Shi K, et al. (2015) Overexpression of potato miR482e enhanced plant sensitivity to Verticillium dahliae infection. J Integr Plant Biol 57: 1078-1088.
- 75. Shukla LI, Chinnusamy V, Sunkar R (2008)The role of microRNAs and other endogenous small RNAs in plant stress responses. Biochim Biophys Acta 1779: 743-748.
- 76. Liu Q, Chen YQ (2009) Insights into the mechanism of plant development: interactions of miRNAs pathway with phytohormone response. Biochem Biophys Res Commun 384: 1-5.
- 77. Kurashige J, Mima K, Sawada G, Takahashi Y, Eguchi H, et al. (2015) Epigenetic modulation and repression of miR-200b by cancer-associated fibroblasts contribute to cancer invasion and peritoneal dissemination in gastric cancer. Carcinogenesis 36: 133-141.
- 78. Tang H, Kong Y, Guo J, Tang Y, Xie X, et al. (2013) Diallyl disulfide suppresses proliferation and induces apoptosis in human gastric cancer through Wnt-1 signaling pathway by up-regulation of miR-200b and miR-22. Cancer Lett 340: 72-81.
- 79. Yang C, Ning S, Li Z, Qin X, Xu W (2014) miR-22 is down-regulated in esophageal squamous cell carcinoma and inhibits cell migration and invasion. Cancer Cell Int 14: 138.
- Knyazev EN, Samatov TR, Fomicheva KA, Nyushko KM, Alekseev BY, et al. (2016) MicroRNA hsa-miR-4674 in Hemolysis-Free Blood Plasma Is Associated with Distant Metastases of Prostatic Cancer. Bull Exp Biol Med 161: 112-115.
- Damavandi Z, Torkashvand S, Vasei M, Soltani BM, Tavallaei M, et al. (2016) Aberrant Expression of Breast Development-Related MicroRNAs, miR-22, miR-132, and miR-212, in Breast Tumor Tissues. J Breast Cancer 19: 148-155.
- Yang M, Jiang N, Cao QW, Sun Q (2016) EDD1 predicts prognosis and regulates gastric cancer growth in vitro and in vivo via miR-22. Biol Chem.
- 83. Yamakuchi M, Yagi S, Ito T, Lowenstein CJ (2011) MicroRNA-22 regulates hypoxia signaling in colon cancer cells. PLoS One 6: e20291.
- Yang F, Hu Y, Liu HX, Wan YJ (2015) MiR-22-silenced cyclin A expression in colon and liver cancer cells is regulated by bile acid receptor. J Biol Chem 290: 6507-6515.
- 85. Shi C, Xu X (2013) MicroRNA-22 is down-regulated in hepatitis B virusrelated hepatocellular carcinoma. Biomed Pharmacother 67: 375-380.

- 86. Wang G, Liu G, Ye Y, Fu Y, Zhang X (2016) Upregulation of miR-34a by diallyl disulfide suppresses invasion and induces apoptosis in SGC-7901 cells through inhibition of the PI3K/Akt signaling pathway. Oncol Lett 11: 2661-2667.
- Su B, Su J, Zeng Y, Liu F, Xia H, et al. (2016) Diallyl disulfide suppresses epithelial-mesenchymal transition, invasion and proliferation by downregulation of LIMK1 in gastric cancer. Oncotarget 7: 10498-10512.
- Du B, Shim JS (2016) Targeting Epithelial-Mesenchymal Transition (EMT) to Overcome Drug Resistance in Cancer. Molecules 21.
- 89. Qi J, Yu Y, Akilli Ozturk O, Holland JD, Besser D, et al. (2016) New Wnt/ beta-catenin target genes promote experimental metastasis and migration of colorectal cancer cells through different signals. Gut 65: 1690-1701.
- 90. Ebbing EA, Steins A, Fessler E, Stathi P, Lesterhuis WJ, et al. (2017) Esophageal Adenocarcinoma Cells and Xenograft Tumors Exposed to Erb-b2 Receptor Tyrosine Kinase 2 and 3 Inhibitors Activate Transforming Growth Factor Beta Signaling, Which Induces Epithelial to Mesenchymal Transition. Gastroenterology 15: 63-76 e14.
- 91. Ding J, Zhou XT, Zou HY, Wu J (2017) Hedgehog signaling pathway affects the sensitivity of hepatoma cells to drug therapy through the ABCC1 transporter. Lab Invest 97: 819-832.
- 92. Xiao S, Chang RM, Yang MY, Lei X, Liu X, et al. (2016) Actin-like 6A predicts poor prognosis of hepatocellular carcinoma and promotes metastasis and epithelial-mesenchymal transition. Hepatology 63: 1256-1271.
- 93. Shibue T, Brooks MW, Weinberg RA (2013) An integrin-linked machinery of cytoskeletal regulation that enables experimental tumor initiation and metastatic colonization. Cancer Cell 24: 481-498.
- 94. Kim H, Choi GH, Na DC, Ahn EY, Kim GI, et al. (2011) Human hepatocellular carcinomas with "Stemness"-related marker expression: keratin 19 expression and a poor prognosis.
- 95. Huang J, Yang B, Xiang T, Peng W, Qiu Z, et al. (2015) Diallyl disulfide inhibits growth and metastatic potential of human triple-negative breast cancer cells through inactivation of the beta-catenin signaling pathway. Mol Nutr Food Res 59: 1063-1075.
- 96. Su B, Su J, He H, Wu Y, Xia H, et al. (2015) Identification of potential targets for diallyl disulfide in human gastric cancer MGC-803 cells using proteomics approaches. Oncol Rep 33: 2484-2494.
- Su J, Zhou Y, Pan Z, Shi L, Yang J, et al. (2017) Downregulation of LIMK1-ADF/cofilin by DADS inhibits the migration and invasion of colon cancer. Sci Rep 7: 45624.
- Hong KO, Lee JI, Hong SP, Hong SD (2016) Thymosin beta4 induces proliferation, invasion, and epithelial-to-mesenchymal transition of oral squamous cell carcinoma. Amino Acids 48: 117-127.
- Liu A, Zhou Z, Dang R, Zhu Y, Qi J, et al. (2016) Neuroligin 1 regulates spines and synaptic plasticity via LIMK1/cofilin-mediated actin reorganization. J Cell Biol 212: 449-463.
- 100. Fan SH, Wang YY, Lu J, Zheng YL, Wu DM, et al. (2015) CERS2 suppresses tumor cell invasion and is associated with decreased V-ATPase and MMP-2/MMP-9 activities in breast cancer. J Cell Biochem 116: 502-513.
- 101. Suzuki S, Honda K, Nanjo H, Iikawa N, Tsuji T, et al. (2017) CD147 expression correlates with lymph node metastasis in T1-T2 squamous cell carcinoma of the tongue. Oncol Lett 14: 4670-4676.
- 102. Thapa N, Anderson RA (2017) PLD and PA Take MT1-MMP for a Metastatic Ride. Dev Cell 43: 117-119.
- 103. Park HS, Kim GY, Choi IW, Kim ND, Hwang HJ, et al. (2011) Inhibition of matrix metalloproteinase activities and tightening of tight junctions by diallyl disulfide in AGS human gastric carcinoma cells. J Food Sci 76: T105-111.
- 104. Xie K, Nian J, Zhu X, Geng X, Liu F (2015) Modulatory role of garlicin in migration and invasion of intrahepatic cholangiocarcinoma via PI3K/AKT pathway. Int J Clin Exp Pathol 8: 14028-14033.
- 105. Lee S, Rho SS, Park H, Park JA, Kim J, et al. (2017) Carbohydrate-binding protein CLEC14A regulates VEGFR-2- and VEGFR-3-dependent signals during angiogenesis and lymphangiogenesis. J Clin Invest 12: 457-471.

Page 7 of 8

- 106. Wang J, Chen J, Guo Y, Wang B, Chu H (2017) Strategies targeting angiogenesis in advanced non-small cell lung cancer. Oncotarget 8: 53854-53872.
- 107. Blagosklonny MV (2001) Hypoxia-inducible factor: Achilles' heel of antiangiogenic cancer therapy. Int J Oncol 19: 257-262.
- 108. Aggarwal C, Somaiah N, Simon G (2012) Antiangiogenic agents in the management of non-small cell lung cancer: where do we stand now and where are we headed? Cancer Biol Ther 13: 247-263.
- Balkwill F (2006) TNF-alpha in promotion and progression of cancer. Cancer Metastasis Rev 25: 409-416.
- 110. Bauer D, Redmon N, Mazzio E, Taka E, Reuben JS, et al. (2015) Diallyl disulfide inhibits TNFalpha induced CCL2 release through MAPK/ERK and NF-Kappa-B signaling. Cytokine 75: 117-126.
- 111. Yan Q, Jiang L, Liu M, Yu D, Zhang Y, et al. (2017) ANGPTL1 interacts with integrin alpha1beta1 to suppress HCC angiogenesis and metastasis by inhibiting JAK2/STAT3 signaling. Cancer Res.
- 112. Lai KC, Hsu SC, Kuo CL, Yang JS, Ma CY, et al. (2013) Diallyl sulfide, diallyl disulfide, and diallyl trisulfide inhibit migration and invasion in human colon cancer colo 205 cells through the inhibition of matrix metalloproteinase-2, -7, and -9 expressions. Environ Toxicol 28: 479-488.
- 113. Xiao L, Yin XC, Cao QQ (2016) The role of the PI3K/AKT signaling pathway in DADS-induced apoptosis of K562 cells. Zhongguo Dang Dai Er Ke Za Zhi 18: 1050-1054.
- 114. Thejass P, Kuttan G (2007) Inhibition of angiogenic differentiation of human umbilical vein endothelial cells by diallyl disulfide (DADS). Life Sci 80: 515-521.
- 115. Morihara N, Hino A, Miki S, Takashima M, Suzuki JI (2017) Aged garlic extract suppresses inflammation in apolipoprotein E-knockout mice. Mol Nutr Food Res 61.
- 116. Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, et al. (1995) Weight-reducing effects of the plasma protein encoded by the obese gene. Science 269: 543-546.
- 117. Netea MG, Balkwill F, Chonchol M, Cominelli F, Donath MY, et al. (2017) A guiding map for inflammation. Nat Immunol 18: 826-831.

- 118. Schafer G, Kaschula CH (2014) The immunomodulation and antiinflammatory effects of garlic organosulfur compounds in cancer chemoprevention. Anticancer Agents Med Chem 14: 233-240.
- 119. Quintero-Fabian S, Ortuno-Sahagun D, Vazquez-Carrera M, Lopez-Roa RI (2013) Alliin, a garlic (Allium sativum) compound, prevents LPSinduced inflammation in 3T3-L1 adipocytes. Mediators Inflamm.
- 120. Ryu JH, Park HJ, Jeong YY, Han S, Shin JH, et al. (2015) Aged red garlic extract suppresses nitric oxide production in lipopolysaccharide-treated RAW 264.7 macrophages through inhibition of NF-kappaB. J Med Food 18: 439-445.
- 121. Atreya I, Atreya R, Neurath MF (2008) NF-kappaB in inflammatory bowel disease. J Intern Med 263: 591-596.
- 122. Saud SM, Li W, Gray Z, Matter MS, Colburn NH, et al. (2016) Diallyl Disulfide (DADS), a Constituent of Garlic, Inactivates NF-kappaB and Prevents Colitis-Induced Colorectal Cancer by Inhibiting GSK-3beta. Cancer Prev Res (Phila) 9: 607-615.
- 123. Hoeflich KP, Luo J, Rubie EA, Tsao MS, Jin O, et al. (2000) Requirement for glycogen synthase kinase-3beta in cell survival and NF-kappaB activation. Nature 406: 86-90.
- 124. Shakoori A, Mai W, Miyashita K, Yasumoto K, Takahashi Y, et al. (2007) Inhibition of GSK-3 beta activity attenuates proliferation of human colon cancer cells in rodents. Cancer Sci 98: 1388-1393.
- 125. Pratheeshkumar P, Thejass P, Kutan G (2010) Diallyl disulfide induces caspase-dependent apoptosis via mitochondria-mediated intrinsic pathway in B16F-10 melanoma cells by up-regulating p53, caspase-3 and down-regulating pro-inflammatory cytokines and nuclear factor-kappabeta-mediated Bcl-2 activation. J Environ Pathol Toxicol Oncol 29: 113-125.
- 126. Chu CC, Wu WS, Shieh JP, Chu HL, Lee CP, et al. (2017) The Anti-Inflammatory and Vasodilating Effects of Three Selected Dietary Organic Sulfur Compounds from Allium Species. J Funct Biomater 8.

Page 8 of 8