

Synergistic Bioefficacy of Botanical Insecticides against *Zabrotes subfasciatus* (Coleoptera: Bruchidae) a Major Storage Pest of Common Bean

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Abstract

This experiment was conducted to determine possibilities of synergism among insecticidal plants against *Z. subfasciatus* with a view of augmenting potency and reducing dosage rates. Leaf and seed powders of five insecticidal plants, namely *Jatropha curcas* (L.), *Datura stramonium* (L.), *Chenopodium ambrosioides* (L.), *Schinus molle* (L.) and *Azadirachta indica* (A. Juss) were mixed to 1% and 2%w/w unitary and binary formulations. The synthetic insecticide primiphos methyl at the rate of 0.1/100 gm grain dust and untreated grains were used as positive and negative controls, respectively. Most binary formulation had better efficacy than their constituent unitary formulation especially at lower dosage rates. Synergistic combination of botanical powders resulted in highest adult mortality, F₁ progeny reduction and lowest weevil perforation index and weight loss comparable to chemical standard primiphos methyl. Among the botanical combinations, bean seeds treated with binary formulation of *C. ambrosioides* with *D. stramonium*, *J. curcas* and *S. molle* gave the best efficacy in controlling *Z. subfasciatus*.

Keywords: *Z. subfasciatus*; Binary formulation; Botanical insecticides; *P. vulgaris*; Synergism

Introduction

The common bean (*Phaseolus vulgaris* L.) is one of the important food and cash crops in eastern and southern Africa [1,2]. Pre-harvest and post-harvest damage by insect pests, inter alia, is a major limiting factor of bean production, especially in smallholder farming conditions, under which most beans are grown in the region. Stored beans suffer heavy losses in terms of both quality and quantity mostly by bean bruchids [1]. Common bean weevil, *Acanthoscelides obtectus* and Mexican bean weevil *Zabrotes subfasciatus* are the most important species of bruchids attacking stored beans, causing yield losses reaching up to 38% [3,4]. Bruchids infestation damage quantity, quality and viability of bean seed [3]. The degree of loss depends on the storage period and storage conditions. Ref. [5] reported an average grain loss of 60% within 3-6 months of storage period due to bean bruchids.

To reduce storage losses due to insect pests, synthetic insecticides have been recommended. However, their use is limited under small scale farming condition due to high costs and infrequent supply [6,7]. Besides, indiscriminate use of insecticides may result in undesirable consequences such as resistance development by the pest, secondary pest outbreaks, wide spread environmental hazards and risk to spray operators [8,9]. For these reasons, development of other alternative control methods such as botanical insecticides have gained significant importance in bruchid management [4,10,11]. Use of botanical insecticides not only confers effective pesticidal effect against bruchids but also serves as ecologically sound and economically feasible control option with low health risks to consumers [8,12]. Different plant extracts may act synergistically to effectively inhibit pest growth and developments compared with a single constituent extract and development of pest resistance is less likely when used over time [13-15].

Even though encouraging efforts that have been made in the last 2-3 decades, to identify botanicals with better insecticidal potential for bruchid management [16-18] limited information is available in their synergistic potential, toxicology, optimal application and species specificity. Moreover, recommended rates were often high which created inconvenience in practical application of botanical.

Hence, the current study was undertaken to examine the prospect of synergism among combinations of crude botanical formulations with the objective of enhancing effectiveness of constituent botanical in mixtures and reducing dosage rates. The botanical plants were chosen based on their local availability and their potential for bean bruchids control [17,19]. The insecticidal plants and parts used in this study are shown in Table 1.

Materials and Methods

Insect rearing

Adult bean bruchids (*Z. subfasciatus*) were obtained from laboratory culture reared on disinfested common bean variety, Awash-1. The experimental insects were maintained under laboratory condition (27 ± 3°C, 60 ± 10% RH, 12L:12D) at Melkassa Agricultural Research Center (8°24'N; 39°21'E). The food medium (bean seeds) used for insect rearing was first disinfested by keeping the grains in the oven at 40°C for 4 hours and allowed to cool for 2 hrs before use [20]. Infestation was done by introducing 100 parental adults (1:1 sex ratio) in 1 L volume of glass jars containing 250 g of bean grains. The parental adults were sieved off 13 days after oviposition period and the grains were kept under laboratory condition until the emergence of F₁ progeny. New generations of adult bean bruchids (*Z. subfasciatus*) obtained from this culture were used in the experiment.

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No.	Scientific name	Common name	Parts used
1	<i>Azadirachta indica</i>	Neem tree	Seed
2	<i>Chenopodium ambrosioides</i>	Mexican tea	Leaf
3	<i>Datura stramonium</i>	Thorn-apple	Leaf
4	<i>Jatropha carcus</i>	Physic nut	seed
5	<i>Schinus molle</i>	Pepper tree	Seed

Table 1: List of botanical plants and parts to be used against *Z. subfasciatus*.

Plant materials and treatment formulations

Fresh plant parts (leaves and seeds) of the botanical plants *J. carcus*, *S. molle* and *Datura stramonium* were collected from MARC and the surroundings. Whereas, plant materials from the other two insecticidal plants *C. ambrosioides* and *A. indica* were collected from natural habitat in Addis Ababa (9°1'48"N 38°44'24"E) and Worer Agricultural Research Center (9°20' 27" N and 40°10' 53"E), respectively. The plant materials were air dried and crushed separately into fine powder using a pestle and mortar. The resultant powder was further sieved through a 0.25 mm mesh to obtain a fine dust. The powders were weighed into 0.5 and 1 gm samples and then mixed appropriately to constitute binary formulation at either 1 or 2% w/w admixture on 100 gm bean samples. The unitary formulations were weighed into 1 and 2 gm samples and admixed with 100 gm bean samples to represent dosage rates of 1 or 2% w/w respectively.

Toxicity assessment

Healthy disinfected common bean seeds (100 gm) treated with different unitary and binary formulations of botanical insecticide powders were placed in the 1 L volume glass jar. The glass jars tops were covered with nylon mesh to allow aeration and held in place with rubber bands. The effectiveness of the different treatments was assessed by introducing 8 pairs of 2-4 days old bruchids obtained from laboratory culture to the treated and untreated grains. The synthetic insecticide primiphos methyl at the rate of 0.1/100 gm grain dust and untreated grains were used as positive and negative controls, respectively. Percent insect mortality was calculated using Abbott's formula by counting number of dead insects in each jar 24 hrs, 48 hrs, 72 hrs, 96 hrs and 120 hrs after treatment application/ adult introduction. Adults were considered dead when no response was observed after probing them with forceps. At the end of each assessment, dead insects were removed. The experiment was arranged in completely randomized design (CRD) with three replications.

$$\text{Abbot's formula: } Pt = \frac{Po - Pc}{100 - Pc} \times 100$$

Where P_t =percent (%) mortality; P_o =observed mortality; P_c =control mortality

Effect of powders on F_1 progeny

After toxicity assessment of plant powders, remaining *Z. subfasciatus* adults on treated and untreated jars were kept for additional 10 days and were sieved and discarded (both live and dead). The infested jars were further maintained under laboratory condition ($7 \pm 3^\circ\text{C}$, $60 \pm 10\%$ RH, 12L: 12D) until adult emergence and effect of treatments on the F_1 progeny were assessed. To avoid overlapping generation, the number of F_1 progeny was counted upon emergence for a period of 45 days since the initial date of adult introduction. Percentage reduction in adult emergence or inhibition rate (% IR) was calculated using the following formula:

$$\% \text{ IR} = \frac{(Cn - Tn)}{Cn} \times 100$$

Where C_n =number of newly emerged insects in the untreated (control) jar

T_n =number of insects in the treated jar

Grain damage assessment

To determine grain damage level, samples of 100 grains were taken randomly from the treated and control jars. Both treated and untreated grains were assessed for extent of bruchids damage using exit-holes as a measure of damage to the grain. The number of damaged grains (with characteristic hole) and undamaged grains were counted and weighed. Percentage grains weight loss was calculated using the following formula.

$$\text{Weight loss (\%)} = \frac{(UNd) - (DNu)}{U(Nd + Nu)} \times 100$$

Where U=weight of undamaged grain; Nd=number of damaged grains; D=weight of damaged grain; N_u =number of undamaged grains.

Moreover, grains that are riddled with exit-holes were counted and the percentage damage (PD) and weevil perforation index (WPI) was calculated according to methods in Ref. [21,22] respectively.

$PD = (\text{total number of treated grains perforated} / \text{total number of grains}) \times 100$

$WPI = (\% \text{ of treated grains perforated} / \% \text{ of control grains perforated} + \% \text{ of treated grains perforated}) \times 100$

Germination test

Germination test was carried out by randomly picking 80 undamaged grains from each treatment jar. Then 20 grains from each treated and control groups were placed separately on a moistened filter paper in Petri dishes and kept at room temperature. Each treatment was replicated four times where healthy grains without botanical insecticide powder application were used as a control. The numbers of germinated grains were recorded starting from the first date of germination. Percent germination was computed using the following formula:

$$\text{Viability index (\%)} = \frac{NG}{TG} \times 100$$

Where NG=number of grains germinated and TG=total number of grains tested in each Petri dish.

Data analysis

All data were checked for normality before they were subjected to analysis. Data which lacked normality were transformed using appropriate transformations method. Data were analyzed with analysis of variance (ANOVA) using General Linear Model (GLM) in SAS software. Significant means were separated using Student-Newma Keuls (SNK) test.

Results

Effects of different botanical powder combinations on bruchids mortality

Results on adult mortality of *Z. subfasciatus* 24-120 hrs after application of unitary and binary formulations of different botanical powders at 1% w/w and 2% w/w dosage rates on common beans grain are shown in Table 2 and 3 and Figure 1. Significant difference ($P < 0.001$) in adult mortality was observed among different treatments depending on type of botanicals and their combinations, dosage rates

Treatments	Adult mortality (% mean \pm SE)					F-value	P-value
	24 hrs*	48 hrs	72 hrs	96 hrs	120 hrs		
<i>A. indica</i>	16.67 \pm 2.08Ec [~]	47.92 \pm 4.17Cb	64.58 \pm 5.51Ca	70.83 \pm 4.17Fa	80.28 \pm 2.41Da	34.73	P<0.0001
<i>C. ambrosioides</i>	27.08 \pm 4.17Dd	60.42 \pm 2.08Bc	72.92 \pm 2.08Bb	87.5 \pm 3.61Ca	93.22 \pm 3.85Ba	58.01	P<0.0001
<i>D. stramonium</i>	4.17 \pm 2.08Gc	27.08 \pm 2.08Eb	50 \pm 3.61Ea	64.58 \pm 4.17Ha	71.53 \pm 7.22Fa	35.73	P<0.0001
<i>J. curcas</i>	22.92 \pm 5.51Dc	47.92 \pm 5.51Cb	68.75 \pm 7.2Ca	79.17 \pm 7.51Fa	87.83 \pm 6.47Ca	14.21	P = 0.0004
<i>S. molle</i>	14.58 \pm 2.08Fc	35.42 \pm 5.51Db	54.17 \pm 4.17Da	62.5 \pm 0.00Ia	71.81 \pm 0.69Fa	40.13	P<0.0001
<i>A. indica</i> + <i>C. ambrosioides</i>	33.33 \pm 2.08Cd	60.42 \pm 4.17Bc	70.83 \pm 2.08Cb	81.25 \pm 0.00Ea	90.97 \pm 0.28Ca	75.44	P<0.0001
<i>A. indica</i> + <i>D. stramonium</i>	29.17 \pm 4.17Db	56.25 \pm 3.61Ba	62.5 \pm 3.61Ca	72.92 \pm 5.51Fa	80.42 \pm 3.97Da	17.18	P=0.0002
<i>A. indica</i> + <i>J. curcas</i>	31.25 \pm 3.61Dc	54.17 \pm 5.51Cb	70.83 \pm 2.08Ca	81.25 \pm 3.61Ea	88.75 \pm 4.39Da	26.99	P<0.0001
<i>A. indica</i> + <i>S. molle</i>	33.33 \pm 2.08Cd	60.42 \pm 8.33Bc	72.92 \pm 5.51Bb	83.33 \pm 2.08Da	88.92 \pm 4.58Ca	16.71	P=0.0002
<i>C. ambrosioides</i> + <i>D. stramonium</i>	35.42 \pm 4.17Cd	64.58 \pm 2.08Bc	75.0 \pm 3.61Bb	89.58 \pm 2.08Ba	93.44 \pm 2.16Ba	57.44	P<0.0001
<i>C. ambrosioides</i> + <i>J. curcas</i>	22.92 \pm 2.08Dd	47.92 \pm 2.08Cc	75.0 \pm 3.61Bb	87.5 \pm 3.61Ca	94.36 \pm 3.61Ba	82.84	P<0.0001
<i>C. ambrosioides</i> + <i>S. molle</i>	35.42 \pm 5.51Cd	58.33 \pm 2.08Bc	70.83 \pm 5.51Cb	85.42 \pm 5.51Ca	90.36 \pm 3.61Ba	21.19	P<0.0001
<i>D. stramonium</i> + <i>J. curcas</i>	31.25 \pm 3.61Dc	47.92 \pm 5.51Cb	68.75 \pm 6.25Ca	77.08 \pm 2.08Fa	86.81 \pm 1.81Da	22.52	P<0.0001
<i>D. stramonium</i> + <i>S. molle</i>	37.5 \pm 3.61Bb	54.17 \pm 2.08Ca	60.42 \pm 5.51Ca	66.67 \pm 5.51Ga	75.83 \pm 6.31Ea	6.06	P=0.0097
<i>J. curcas</i> + <i>S. molle</i>	22.92 \pm 4.17Dd	47.92 \pm 4.17Cc	64.58 \pm 5.51Cb	70.83 \pm 8.33Fa	74.72 \pm 3.37Da	15.45	P=0.0003
<i>Primiphose methyl</i>	87.5 \pm 7.22Aa	91.67 \pm 4.17Aa	100 \pm 0.0Aa	100 \pm 0.0Aa	97.92 \pm 2.08Aa	2.12	P=0.1532
Control (untreated)	0.00 \pm 0.00Ha	0.00 \pm 0.00Fa	0.00 \pm 0.00Fa	0.00 \pm 0.00Ja	2.08 \pm 2.08Ga	1.00	P=0.4516
F-value	23.85	20.11	20.48	25.93	28.53		
P-value	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001		

*Hours after treatment application; [~]Means followed by the same letter (s) within a column (upper case letters) and within a row (lowercase letters) are not significantly different using Student-Newman-Keuls (SNK) test (P<0.05). Effectiveness of botanicals and their combinations was determined by computing percent insect mortality (Abbotts 1925) and comparing the mortality data by ANOVA using GLM procedure.

Table 2: Mortality (% mean \pm SE) of adult *Z. subfasciatus* on common bean seeds admixed with unitary and binary formulations (2% w/w) of different botanical insecticide powders.

Treatments	F1 progeny	% Inhibition Rate (IR)	Weevil Perforation Index(WPI)	% Weight Loss
<i>A. indica</i>	18.33 \pm 0.88c [~]	58.89 \pm 7.56d	37.36 \pm 3.23b	1.33 \pm 0.45a
<i>C. ambrosioides</i>	1.00 \pm 1.0f	98.33 \pm 1.67a	3.51 \pm 3.51e	0.04 \pm 0.04c
<i>D. stramonium</i>	22.33 \pm 3.18b	48.52 \pm 13.31e	32.77 \pm 11.56b	1.00 \pm 0.50b
<i>J. curcas</i>	14.00 \pm 2.08d	69.26 \pm 5.29c	18.80 \pm 1.02d	0.95 \pm 0.46b
<i>S. molle</i>	10.00 \pm 1.73e	78.33 \pm 3.53b	19.58 \pm 2.50c	0.19 \pm 0.18c
<i>A. indica</i> + <i>C. ambrosioides</i>	3.00 \pm 1.73f	92.78 \pm 3.89a	10.50 \pm 5.44e	0.05 \pm 0.05c
<i>A. indica</i> + <i>D. stramonium</i>	4.33 \pm 2.19e	91.67 \pm 4.19a	7.00 \pm 3.76e	0.17 \pm 0.13c
<i>A. indica</i> + <i>J. curcas</i>	3.33 \pm 0.33f	92.59 \pm 1.21a	10.45 \pm 1.38e	0.18 \pm 0.12c
<i>A. indica</i> + <i>S. molle</i>	7.33 \pm 0.88e	83.15 \pm 4.38b	18.90 \pm 2.49d	1.13 \pm 0.11b
<i>C. ambrosioides</i> + <i>D. stramonium</i>	1.33 \pm 0.88f	97.59 \pm 1.45a	3.04 \pm 1.63e	0.00 \pm 0.00c
<i>C. ambrosioides</i> + <i>J. curcas</i>	1.67 \pm 0.33f	96.48 \pm 0.49a	4.49 \pm 1.23e	0.00 \pm 0.00c
<i>C. ambrosioides</i> + <i>S. molle</i>	0.00 \pm 0.00f	100.00 \pm 0.00a	0.00 \pm 0.00g	0.00 \pm 0.00c
<i>D. stramonium</i> + <i>J. curcas</i>	7.67 \pm 1.20e	83.15 \pm 3.05b	8.63 \pm 1.54e	0.18 \pm 0.09c
<i>D. stramonium</i> + <i>S. molle</i>	8.00 \pm 1.15e	82.59 \pm 2.59b	12.17 \pm 1.73e	0.35 \pm 0.10c
<i>J. curcas</i> + <i>S. molle</i>	4.67 \pm 1.20e	90.37 \pm 1.03a	12.02 \pm 0.84e	0.22 \pm 0.09c
<i>Primiphos methyl</i>	0.33 \pm 0.33f	99.26 \pm 0.74a	1.19 \pm 1.19f	0.04 \pm 0.04c
Control (untreated)	47.00 \pm 7.00a	0.00 \pm 0.00f	50.00 \pm 0.00a	1.5 \pm 0.41a
F-value	28.23	31.00	14.29	4.89
P-value	<0.0001	<0.0001	<0.0001	<0.0001

Table 3: Mean number of F₁ progeny produced (mean \pm SE), % inhibition rate (IR), weevil perforation index (WPI) and % weight loss caused by *Z. subfasciatus* on common bean seeds admixed with different unitary and binary botanical powder formulations at 1% w/w dosage rate.

and time after treatment application. Significantly higher *Z. subfasciatus* mortality was recorded under binary botanical formulations compared to unitary formulation at both dosage rates (1% w/w and 2% w/w). For example, mean bruchid mortality recorded 120 hrs after application of unitary formulation was 55% while for binary formulation it was 75%.

Adult *Z. subfasciatus* mortality due to botanical insecticide application was directly related to exposure time. Longer duration of exposure after treatment application resulted in significantly higher adult mortality and vice versa for both unitary and binary botanical formulations (P<0.05). For instance, mean adult mortality due to *C.*

ambrosioides+*D. stramonium* treatment was only 35.42% after 24 hrs while the same botanical formulation caused 89.58% mortality after 96 hrs (4 days) (Table 3). Overall, lowest *Z. subfasciatus* mortality was recorded 24 hrs after treatment application where as the highest mortality was recorded 96 hrs after treatment application. In the present study, no significant difference in percent adult mortality was observed between 96 hrs and 120 hrs after treatment application except for binary formulation of *C. ambrosioides*+*J. curcas* at 1% w/w dosage rate.

Mortality effect of botanicals was dose dependent especially for

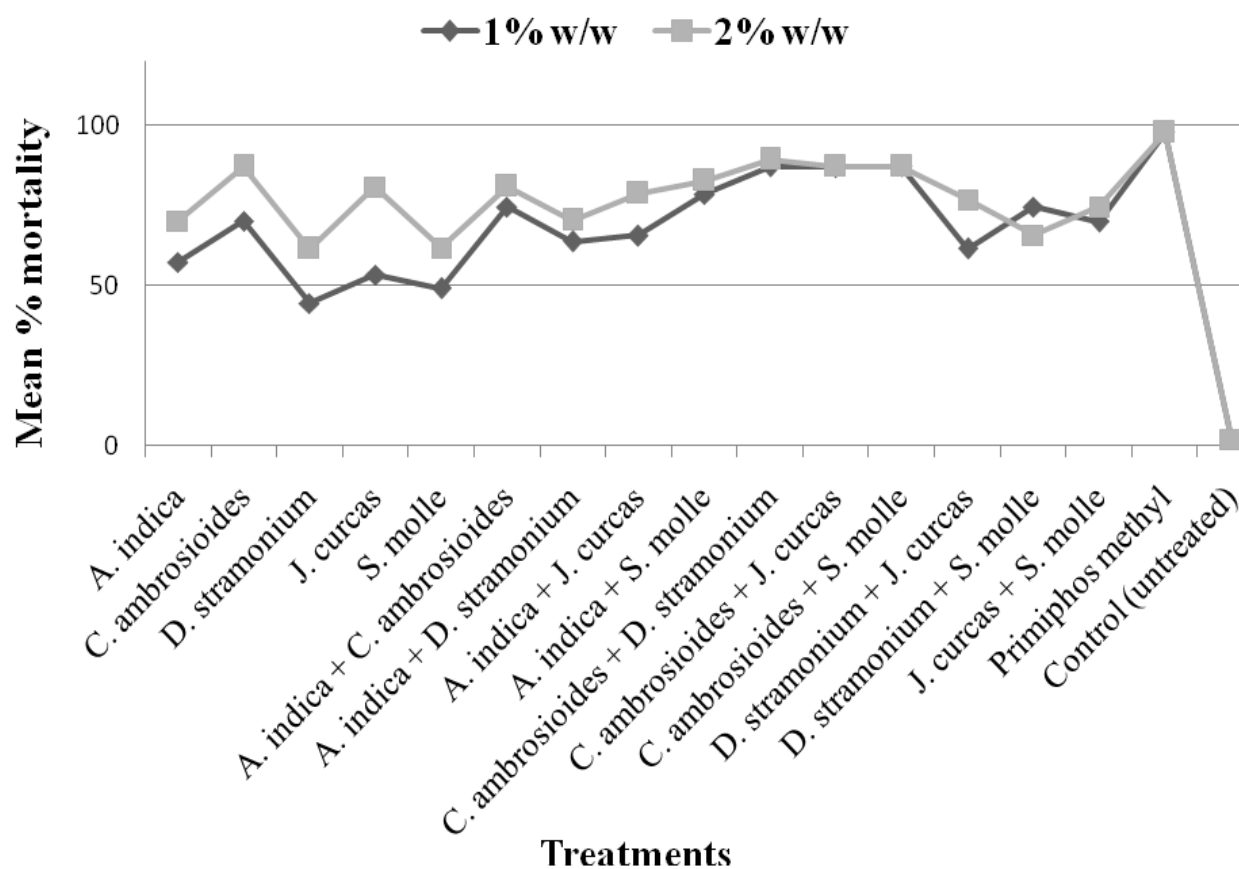


Figure 1: Cumulative mortality effect of unitary and binary formulations of botanical insecticide powders on adult *Z. subfasciatus*, applied at dosage rates of 1% w/w and 2% w/w.

Treatments	F1 progeny	% Inhibition Rate (IR)	Weevil Perforation Index(WPI*)	% Weight Loss
<i>A. indica</i>	8.67 ± 3.71b ^{***}	82.96 ± 4.86e	17.35 ± 4.63d	0.33 ± 0.18b
<i>C. ambrosioides</i>	0.33 ± 0.33b	99.26 ± 0.74a	2.42 ± 1.21i	0.14 ± 0.07b
<i>D. stramonium</i>	11.33 ± 2.33b	76.11 ± 2.00g	21.06 ± 1.31c	0.17 ± 0.05b
<i>J. curcas</i>	6.33 ± 3.33b	87.78 ± 4.75d	13.29 ± 4.79f	0.17 ± 0.17b
<i>S. molle</i>	9.33 ± 1.76b	78.89 ± 6.19f	31.41 ± 2.79b	0.77 ± 0.31b
<i>A. indica</i> + <i>C. ambrosioides</i>	2.33 ± 1.20b	95.37 ± 2.57b	4.29 ± 2.97i	0.18 ± 0.09b
<i>A. indica</i> + <i>D. stramonium</i>	5.33 ± 0.88b	87.96 ± 2.73d	19.35 ± 1.95d	0.46 ± 0.17b
<i>A. indica</i> + <i>J. curcas</i>	2.33 ± 0.88b	94.26 ± 2.80b	6.7 ± 2.16h	0.03 ± 0.03b
<i>A. indica</i> + <i>S. molle</i>	5.67 ± 1.45b	87.96 ± 2.91d	13.78 ± 2.31f	0.17 ± 0.09b
<i>C. ambrosioides</i> + <i>D. stramonium</i>	0.67 ± 0.33b	98.70 ± 0.67a	2.14 ± 1.09i	0.04 ± 0.04b
<i>C. ambrosioides</i> + <i>J. curcas</i>	0.00 ± 0.00b	100.00 ± 0.00a	0.00 ± 0.00k	0.00 ± 0.00b
<i>C. ambrosioides</i> + <i>S. molle</i>	0.00 ± 0.00b	100.00 ± 0.00a	0.00 ± 0.00k	0.00 ± 0.00b
<i>D. stramonium</i> + <i>J. curcas</i>	7.00 ± 1.00b	84.44 ± 2.94e	14.94 ± 3.65e	0.1 ± 0.09b
<i>D. stramonium</i> + <i>S. molle</i>	6.33 ± 0.88b	86.48 ± 0.19d	14.07 ± 3.83f	0.55 ± 0.23b
<i>J. curcas</i> + <i>S. molle</i>	4.33 ± 0.33b	90.56 ± 0.85c	11.61 ± 4.91g	0.22 ± 0.22b
Primiphos methyl	0.33 ± 0.33b	99.26 ± 0.74a	1.19 ± 1.19j	0.04 ± 0.04b
Control (untreated)	47.00 ± 7.00a	0.00 ± 0.00h	50.00 ± 0.00a	1.51 ± 0.41a
F-value	22.60	70.75	21.32	4.91
P-value	<0.0001	<0.0001	<0.0001	<0.0001

*WPI value above 50 indicate negative protectant ability; ^{***}Means followed by the same letter within a column are not significantly different using Student-Newman-Keuls (SNK) test (P<0.05). The data was analyzed by ANOVA using GLM procedure (SAS2002-2008)

Table 4: Mean number of F₁ progeny produced (mean ± SE), % inhibition rate (IR), weevil perforation index (WPI) and % weight loss caused by *Z. subfasciatus* on common bean grain admixed with different unitary and binary botanical powder formulations at 2% w/w dosage rate.

Treatments	Percent Germination (mean \pm SE)	
	1% w/w	2% w/w
<i>A. indica</i>	96.67 \pm 1.12a*	95.00 \pm 2.46a
<i>C. ambrosioides</i>	98.75 \pm 0.65a	97.92 \pm 0.96a
<i>D. stramonium</i>	97.08 \pm 1.14a	97.50 \pm 0.75a
<i>J. curcas</i>	95.42 \pm 1.14a	97.92 \pm 0.74a
<i>S. molle</i>	98.75 \pm 0.65a	97.08 \pm 1.68a
<i>A. indica</i> + <i>C. ambrosioides</i>	96.67 \pm 1.98a	96.67 \pm 1.55a
<i>A. indica</i> + <i>D. stramonium</i>	95.00 \pm 1.51a	96.67 \pm 1.67a
<i>A. indica</i> + <i>J. curcas</i>	92.50 \pm 1.90a	97.08 \pm 1.79a
<i>A. indica</i> + <i>S. molle</i>	97.92 \pm 0.94a	96.67 \pm 1.12a
<i>C. ambrosioides</i> + <i>D. stramonium</i>	92.92 \pm 1.99a	94.58 \pm 2.42a
<i>C. ambrosioides</i> + <i>J. curcas</i>	97.50 \pm 1.44a	99.17 \pm 0.56a
<i>C. ambrosioides</i> + <i>S. molle</i>	96.25 \pm 1.52a	97.50 \pm 1.44a
<i>D. stramonium</i> + <i>J. curcas</i>	94.58 \pm 1.68a	98.33 \pm 0.94a
<i>D. stramonium</i> + <i>S. molle</i>	95.42 \pm 1.44a	97.92 \pm 1.14a
<i>J. curcas</i> + <i>S. molle</i>	93.33 \pm 1.88a	95.83 \pm 1.72a
Primiphos methyl	98.33 \pm 0.71a	98.33 \pm 0.71a
Control (untreated)	97.92 \pm 0.96a	97.92 \pm 0.96a
F-value	2.03	0.69
P-value	P=0.5308	P=0.7992

*Means followed by the same letter within a column are not significantly different using Student-Newman-Keuls (SNK) test ($P < 0.05$). The data was analyzed by ANOVA using GLM procedure (SAS 2002-2008)

Table 5: Effect of unitary and binary botanical formulations treatment on percent germination (mean \pm SE) of common bean seeds.

Treatments	Z. subfasciatus mortality (% mean \pm SE)					F-value	P-value
	24 hrs*	48 hrs	72 hrs	96 hrs	120 hrs		
<i>A. indica</i>	20.83 \pm 5.51Ec**	33.33 \pm 9.08Eb	45.83 \pm 9.08Hb	60.42 \pm 2.08Fa	67.36 \pm 5.14Ga	6.09	P=0.0095
<i>C. ambrosioides</i>	27.08 \pm 9.08Dc	41.67 \pm 7.51Dc	50.0 \pm 7.22Fb	70.83 \pm 4.16Da	79.28 \pm 2.41Ea	8.29	P=0.0032
<i>D. stramonium</i>	12.5 \pm 0.0Fc	31.25 \pm 0.0Eb	43.75 \pm 0.0Ha	47.92 \pm 2.08Ha	54.58 \pm 5.42Ja	31.57	P<0.0001
<i>J. curcas</i>	6.25 \pm 3.61Gd	22.92 \pm 4.17Fc	37.5 \pm 3.61Ib	52.08 \pm 4.17Ga	63.33 \pm 1.82Ha	31.32	P<0.0001
<i>S. molle</i>	10.42 \pm 2.08Gb	20.83 \pm 5.51Fb	35.42 \pm 2.08Ja	47.92 \pm 4.17Ha	59.17 \pm 2.92Ia	22.09	P<0.0001
<i>A. indica</i> + <i>C. ambrosioides</i>	29.17 \pm 7.51Db	56.25 \pm 9.55Ca	70.83 \pm 9.08Ca	72.92 \pm 7.51Da	79.72 \pm 3.37Da	6.14	P=0.0092
<i>A. indica</i> + <i>D. stramonium</i>	12.5 \pm 3.61Fc	39.58 \pm 2.08Db	52.08 \pm 4.17Fa	62.5 \pm 3.61Fa	70.89 \pm 2.50Fa	41.49	P<0.0001
<i>A. indica</i> + <i>J. curcas</i>	22.92 \pm 2.08Db	43.75 \pm 0.0Da	47.92 \pm 2.08Ga	66.67 \pm 7.51Ea	75.69 \pm 9.98Fa	9.86	P=0.0017
<i>A. indica</i> + <i>S. molle</i>	25 \pm 3.61Dd	43.75 \pm 3.61Dc	60.42 \pm 7.51Db	77.08 \pm 9.08Ca	88.61 \pm 7.91Ca	11.44	P=0.0009
<i>C. ambrosioides</i> + <i>D. stramonium</i>	27.08 \pm 4.17Dd	56.25 \pm 3.61Cc	72.92 \pm 2.08Cb	85.42 \pm 2.08Ba	87.22 \pm 3.85Ba	57.63	P<0.0001
<i>C. ambrosioides</i> + <i>J. curcas</i>	37.5 \pm 3.61Bb	45.83 \pm 4.17Db	56.25 \pm 6.25Eb	58.33 \pm 4.17Fb	87.08 \pm 6.67Ba	13.45	P=0.0005
<i>C. ambrosioides</i> + <i>S. molle</i>	35.42 \pm 5.51Cc	62.5 \pm 0.0Bb	77.08 \pm 2.08Ba	87.5 \pm 0.0Ba	92.22 \pm 3.85Ba	48.07	P<0.0001
<i>D. stramonium</i> + <i>J. curcas</i>	20.83 \pm 2.08Ee	39.58 \pm 2.08Dd	50.0 \pm 3.61Fc	58.33 \pm 2.08Fb	61.67 \pm 4.35Ga	30.25	P<0.0001
<i>D. stramonium</i> + <i>S. molle</i>	14.58 \pm 4.17Fb	29.17 \pm 9.08Fb	56.25 \pm 3.61Ea	70.83 \pm 2.08Da	74.58 \pm 3.63Da	26.48	P<0.0001
<i>J. curcas</i> + <i>S. molle</i>	20.83 \pm 4.17Ed	39.58 \pm 7.51Dc	56.25 \pm 6.25Eb	66.67 \pm 7.51Ea	70 \pm 8.32Ea	8.77	P=0.0026
Primiphos methyl	87.5 \pm 7.22 Aa	91.67 \pm 4.17Aa	100 \pm 0.0Aa	100 \pm 0.0Aa	97.92 \pm 2.08Aa	2.12	P=0.1532
Control (untreated)	0.0 \pm 0.0Ha	0.0 \pm 0.0Ga	0.0 \pm 0.0Ka	0.0 \pm 0.0Ia	2.08 \pm 0.08Ka	1.00	P=0.4516
F-value	16.85	13.66	17.79	22.39	18.93		
P-value	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001		

* Hours after treatment application; **Means followed by the same letter (s) within a column (upper case letters) and within a row (lower case letters) are not significantly different using student Newman Keuls (SNK) test ($P < 0.05$). Effectiveness of botanicals and their combinations was determined by computing percent insect mortality (Abbotts 1925) and comparing the mortality data by ANOVA using GLM procedure

Table 6: Mortality (% mean \pm SE) of adult *Z. subfasciatus* on common bean seeds admixed with unitary and binary formulations (1%w/w) of different botanical insecticide powders.

unitary formulations. An increased *Z. subfasciatus* mortality was observed at higher doses for unitary formulation. For example, *Z. subfasciatus* mortality due to *J. curcas* application at 1%w/w dosage rate was 53.33% which increased to 80.83% at higher dosage rate (2%w/w). On the other hand, notable increase in adult mortality due to higher dose was not observed in most binary formulations. For instance, binary formulations of *C. ambrosioides*, *D. stramonium*, *J. curcas* and *S. molle* had more or less similar mortality effects at lower and higher application rates. An increase in adult mortality due to higher application rate in binary formulation was observed mainly during *J. curcas* combination with *A. indica* and *D. stramonium*.

Effects of botanical insecticides on weevil perforation index (WPI) and percent grains weight loss

Weevil Perforation Index (WPI) and % weight loss due to *Z. subfasciatus* on common bean seeds admixed with different application rates of unitary and binary botanical powder formulations is presented in Tables 4 and 5. All botanical insecticide formulations resulted in positive protective effect against damage by *Z. subfasciatus*, as the WPI values for all treatments were significantly less than 50. Generally, binary formulations showed better protectant ability compared to unitary formulation. For instance, the mean WPI for binary formulation at 1% w/w dosage rate was 8.72% whereas the mean WPI for unitary formulation was 22.41%. WPI was reduced with an increase dosage rate especially for unitary formulation. Binary formulations *D. stramonium*+*C. ambrosioides*, *C. ambrosioides*+*J. curcas* and *S. molle*+*C. ambrosioides* gave the best protection against *Z. subfasciatus* damage, with WPI value less than 5 at both test doses. This protective effect due to botanical formulation was on par with standard synthetic insecticide, primiphos methyl.

Weight loss due to *Z. subfasciatus* was significantly reduced ($P<0.0001$) after application of both unitary and binary botanical formulations compared to untreated control 7 weeks after infestation. The untreated bean grain had the highest weight loss due to damage by *Z. subfasciatus*. Overall, binary formulations had better effect in reducing weight loss compared to unitary formulation. For instance, mean weight loss after seed treatment with unitary formulation was 41% while that of binary formulation was 15% at 1%w/w dosage rate. Among the botanical treatments, the highest weight loss (65%) was recorded on beans grain treated with *A. indica* while the lowest weight loss due to *S. molle*+*C. ambrosioides* treatment at 1%w/w dosage rate. There was neither seed damage nor weight loss recorded on bean grains treated with binary formulations of *C. ambrosioides*+*J. curcas* and *S. molle*+*C. ambrosioides* at 2% w/w.

Effects of unitary and binary botanical formulations treatment on percent germination

Germination percent of common bean seeds treated with different unitary and binary botanical powder formulation is presented in Table 5. There was no significant ($P>0.05$) difference in the percent germination between disinfected common bean seeds treated with different botanical insecticide formulations and untreated control at both dosage rates. The percent germination of bean seed treated with different botanical powder formulations ranged between 92-99%, which was as good as untreated control, indicating botanical treatment didn't have effect on germination rate.

Discussion

Several studies have been carried on the potential of botanical

insecticides in controlling insect pests including stored grain pests; however, only a few studies considered their synergistic combination. Results from the present study demonstrated synergistic potential of different botanical insecticide powders in controlling of bean bruchids (*Z. subfasciatus*) on stored beans (*P. vulgaris*). Overall, binary formulations had better effect in reducing damage compared to unitary formulation as assessed by different control parameters such as adult mortality (Table 6), F_1 progeny production, percent inhibition, and weevil perforation index (WPI) and weight loss. Moreover, combining more than one botanical insecticide which works synergistically will also make difficult for pests to develop resistance.

Significantly higher adult mortality was recorded in binary formulation compared to unitary formulation. For instance, adult mortality which ranged from 44.58-53.33% due to application unitary formulation of *D. stramonium*, *J. curcas* and *S. molle* increased to over 87% when combined with *C. ambrosioides*, even at lower dosage rate (1% w/w). This indicated combining different botanical insecticides will enhance their potency in controlling *Z. subfasciatus*. The current findings concur with previous reports which showed enhanced potency of botanicals in controlling pests when combined as binary formulations. For example, combination of *A. indica* with a pyrethroid resulted in a more effective management of silver white fly (*Bemisia argentifolii*), a major greenhouse pest of horticultural flowers [23]. Similarly, biological activities of botanical plants, tobacco (*Nicotiana tabacum*), Mexican marigold (*Tagetes minuta*), tephrosia *Tephrosia vogelli*, and *A. indica* were significantly enhanced in their binary formulation against common bean weevil, *A. obtectus* on stored beans reported reduced number of pests on cowpea plants and increased yield of grains as a result of synergistic activity of mixed botanical extract from herbal landraces.

Results from the present study demonstrated reduction in the application rates of constituent individual formulations without compromise in control efficacy due to enhanced potency in combining botanical insecticides. Higher bruchids mortality was achieved after seed treatment with binary formulation of *C. ambrosioides* with *D. stramonium*, *J. curcas* and *S. molle* at lower dosage rate which was on a par with the mortality recorded due to synthetic insecticide primiphos methyl. Besides, low dose binary formulations were more effective in controlling *Z. subfasciatus* than most of their constituent unitary botanical formulations at high dosage rate. In the current study, a binary formulation in which *C. ambrosioides* was included had the best mortality effect, often over 85%. Specifically, combination of *C. ambrosioides* with *D. stramonium*, *J. curcas* and *S. molle* were most potent as highest *Z. subfasciatus* mortality was achieved with very small quantity/proportion of botanicals applied. Besides, there was no significant difference in adult mortality due to these binary formulations at lower (1% w/w) and higher (2% w/w) dosage rates. Previous studies have demonstrated reduced application rates of synthetic insecticides due to increased potency of binary formulations.

Toxic effects of unitary and binary botanical formulations in the current study were directly related to exposure time of the pest to the treatments. Highest *Z. subfasciatus* mortality was recorded at longest exposure periods after botanical treatment and vice versa. It was also found out that mortality effect of botanical insecticides was dose dependent particularly for unitary formulations. An increased *Z. subfasciatus* mortality was observed at higher doses of unitary formulation. Interestingly, some binary formulations had more or less similar effects at both dosage rate, for example, binary formulation of *C. ambrosioides* *D. stramonium*, *J. curcas* and *S. molle*. Among

individual formulations tested, lowest *Z. subfasciatus* mortality was recorded by *D. stramonium* at 1%w/w while the highest mortality was observed by *C. ambrosioides* at higher rate (2%w/w) application. The current findings are in agreement with the previous report by G/selase and Getu where mortality effect of botanicals was shown to be dose and exposure time dependent.

Overall, bean grain treatment with unitary and binary botanical formulations induced significant high reduction in F_1 progeny production by *Z. subfasciatus* compared to the untreated control. Besides, binary formulations showed better reduction in adult emergence compared to their unitary formulation. Bean grain treatment with binary formulations of *C. ambrosioides*+*A. indica*, *D. stramonium*+*C. ambrosioides*, *C. ambrosioides*+*J. curcas*, *S. molle*+*C. ambrosioides*, *A. indica*+*J. curcas* resulted in the highest reduction F_1 progeny produced. Moreover, there was no significant different in F_1 progeny reduction due to synthetic chemical primiphos methyl and the binary formulations. Significantly high reduction in F_1 progeny as a result of binary formulations application, demonstrated by none or below unitary adult emergent number, strongly suggested enhanced potency of different botanical combinations. Pest attack is population dependent where high pest populations build up lead to high infestation and damage, which in turn depends on number of emerging adults [24,25]. The highly toxic effects of the binary formulations against F_1 progeny in this study indicated the potential of synergists as an effective control option against *Z. subfasciatus*.

The synergistic effect of botanical insecticides in suppressing in F_1 progeny could be due to combined factors such as, increased adult mortality, ovicidal and larvicidal properties of botanical formulations and/or presence of chemicals that interfere with insect feeding [26,27]. Previous investigation on wheat treated with *A. indica* and *A. boonei* powder attributed suppression of F_1 generation of *S. zeamais* to high mortality of adult insects which disrupts mating and sexual communication as well as deterring females from laying eggs and affecting developmental stages of insects. Related studies showed botanical powder treatment act as oviposition-deterrent, inhibit oviposition by weakening adult bruchid to lay fewer eggs and kill the hatching larvae afterwards [28,29]. In related study reported neem seed kernel admixed to the groundnuts at the rate of 5% reduced the adult emergence of *C. serratus*. Even though synergistic combination of botanicals in insect suppression has not been widely examined, several studies revealed the potential of botanical insecticides in reducing F_1 progeny production on different insect pests.

Damage by *Z. subfasciatus* infestation was significantly reduced after treating bean grains with unitary and binary botanical formulations compared to untreated control. Binary formulations had better effect in reducing damage compared to their unitary formulations as judged by low to none weevil perforation index and weight loss. This has further confirmed that combining some botanicals as binary formulation will enhance their biological activity to effectively reduce damage by *Z. subfasciatus* on stored beans. Among the different botanical synergists, combination *C. ambrosioides* with *D. stramonium*, *J. curcas* and *S. molle* showed the most effective protecting ability against *Z. subfasciatus*. This was demonstrated by the least weevil perforation index and percent weight loss recorded after their application at lowest dosage rate (1% w/w). In addition to enhanced efficacy, botanical synergist discussed here have favorable toxicological properties such as rapid degradation, low residues and are safe for the consumer which make them preferred biopesticides in storage pest control.

Results from germination test demonstrated that all botanical powder formulations used to treat bean seeds against *Z. subfasciatus* didn't have negative effect in germination percents of the seeds at both dosage rates. Hence, bean seed for planting can be protected and kept viable from storage pest by treating them botanical formulations similar to the grain stored for food purposes. Though this is the first time to test combined effect botanical synergists on germination, previous study on seeds treated with unitary botanical formulation showed no significant effect on the germination rate. Our study results are in agreement with several reports which stated botanical insecticides which provided protection against storage pests didn't affect seed quality and viability [30-32]. In summary, the findings from the current study underscored synergistic combinations of botanicals enhance effective control of storage pests by optimizing potency of constituent botanicals while reducing dosage rates. Toxicity effect of these binary formulations was comparable with the standard chemical pesticide primiphos methyl. Besides, use of botanical insecticides has several comparative advantages over synthetic insecticides, which include low cost, in the context of small holder farmers, availability, reduced environmental pollution and minimal toxicity to humans and livestock [33]. The insecticidal properties of most of botanical plants studied here have been reported against different insect pests [34,35]. However, none of these studies considered synergistic potential of botanicals and similar insecticidal effects were reported at relatively high dosage rates and after longer exposure time. A new dimension of utilizing synergistic combination of different botanical formulations offers an excellent opportunity to increase the efficacy and reduce application rates of biopesticides in effort to successfully control storage pests of agricultural crops [36]. Hence, the authors recommend incorporation the information/ knowledge generated on synergistic combination of botanicals into regular biorational crop protection practice especially by resource limited small scale bean farmers.

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References

1. Abate T, Ampofo JKO (1996) Insect pests of common bean in Africa: Their ecology and management. Ann Rev Entomol 41: 45-75.
2. Schmale I, Wackers FL, Cardona C, Dorn S (2002) Field Infestation of *Phaseolus vulgaris* by *Acanthoscelides obtectus* (Coleoptera: Bruchidae), Parasitoid Abundance, and Consequences for Storage Pest Control. J Environ Entomol 31: 859-863.
3. Schoonhoven AV, Cardona C (1986) Main insect pests of stored beans and their control. Centre International de Agriculture Tropical.
4. Negasi F (1994) Studies on the economic importance and control of bean bruchids in haricot bean. Alemaya University.
5. Getu E, Ibrahim A, Iticha F (2003) Review of lowland pulse insect pest research in Ethiopia. Proceedings of grain legume workshop, Addis Ababa, Ethiopia.
6. Isman BM (2008) Perspective Botanical insecticides: for richer, for poorer. Pest Manag Sci 64: 8-11.
7. Kareru P, Rotich Z K, Maina EW (2013) Use of Botanicals and Safer Insecticides Designed in Controlling Insects: The African Case. Agricultural and Biological Sciences.
8. Williams P, Hammitt J (2001) Chemicals derived from natural sources are generally perceived to pose less environmental risks than synthetic pesticides. J Entomol Sci 45: 45-51.

9. Bruce TJA (2010) Tackling the threat to food security caused by crop pests in the new millennium. J Food Secu 2: 133-141.
10. Songa JM, Rono W (1998) Indigenous methods for bruchid beetle (Coleoptera: Bruchidae) control in stored beans (*Phaseolus vulgaris* L.). Inter J Pest Manag 44: 1-4.
11. Tadesse A (2008) Increasing Crop Production through Improved Plant Protection. Volume I. Plant Protection Society of Ethiopia (PPSE), Addis Ababa, Ethiopia.
12. Ileke KD, Oni MO (2011) Toxicity of some plant powders to maize weevil, *Sitophilus zeamais* (motschulsky) [Coleoptera: Curculionidae] on stored wheat grains (*Triticumaestivum*). Afric J Agric Res 6: 3043-3048.
13. Agona JA, Muyinza H (2003) Synergistic potential of different botanical combinations against bean bruchids in storage. Afric Crop Sci Conf Proc 6: 216-219.
14. Oparaeke AM, Dike MC, Amatoobi CI (2005) Evaluation of botanical mixtures for insect pests management on cowpea plants. J Agric Rural Devel Trop Subtrop 106: 41-48.
15. Rahman A, Talukder FA (2006) Bioefficacy of some plant derivatives that protect grain against the pulse beetle, *Callosobruchus maculatus*. J of Insec Sci 6: 3-4.
16. Hill J, Schoonhoven AV (1981) Effectiveness of vegetable oil fractions in controlling the Mexican bean weevil on stored beans. J Econ Entomol 74: 478-479.
17. Gselase A, Getu E (2009) Evaluation of botanical plants powders against *Zabrotes subfasciatus* (Boheman) (Coleoptera: Bruchidae) in stored haricot beans under laboratory condition. Afric J Agric Res 4: 1073-1079.
18. Ileke KD, Bulus DS (2012) Evaluation of contact toxicity and fumigant effect of some medicinal plant and pirimiphos methyl powders against cowpea bruchid, *Callosobruchus maculatus* (fab.) [Coleoptera: chrysomelidae] in stored cowpea seeds. J Agric Sci 4: 279-284.
19. Abate T, Negasi F, Ayalew G (1992) Botanicals in pest management Presented. CEE/EPC Joint Conference, Addis Ababa.
20. Jembere B (2002) Evaluation of the toxicity potential of *Milletia ferruginea* (Hochest) Baker against *Sitophilus zeamais* (Motsch.). Inter J Pest Manag 48: 29-32.
21. Adedire CO, Ajayi TS (1996) Assessment of insecticidal properties of some plants as Grain protectants against the maize weevil, *Sitophilus zeamais* (Mots). Niger J Entomol 13: 93-101.
22. Fatope MO, Nuhu AM, Mann A, Takeda Y (1995) Cowpea weevil bioassay: a simple pre-screen for plants with grain protectant effect. Inter J Pest Manag 41: 84-86.
23. Greer L (2000) Greenhouse IPM: Sustainable Whitefly Control. Pest Management Technical Note.
24. Southwood TRE (1978) Ecological Methods. New York, USA.
25. Odum EP (1983) Basic Ecology. Holt-Saunders International Edition, Tokyo, Japan.
26. Ofuya TI (1990) Oviposition deterrence and ovicidal property of some plant powders against *Callosobruchus maculatus* in stored cowpea (*Vigna unguiculata*) seeds. J Agric Sci 115: 343-345.
27. Omotoso OT (2005) Insecticidal and insect productivity reduction capacities of aloe vera and *bryophyllumpinnatum* on *tribolium castaneum* (herbst). Afr J Appl Zool Envi Biol 7: 95-100.
28. Mulatu B, Gebremedhin T (2000) Oviposition-deterrent and toxic effects of various botanicals on the Adzuki bean beetle, *Callosobruchus chinensis* L. Insec Sci Appl 20: 33-38.
29. El-Atta HA, Ahmed A (2002) Comparative effects of some botanicals for the control of the seed weevil *Caryedon serratus* Olivier (Col., Bruchidae). J Appl Entomol 126: 577-583.
30. Onu I, Aliyu M (1995) Evaluation of powdered fruits of four peppers (*Capsicum* spp.) for the control of *Callosobruchus maculatus* (F) on stored cowpea seed. Inter J Pest Manag 41: 143-145.
31. Keita SM, Vincent C, Schmit JP, Arnason JT, Belanger A (2001) Efficacy of essential oil of *Ocimum basilicum* L. and *O. gratissimum* L. applied as an insecticidal fumigant and powder to control *Callosobruchus maculatus* (Fab.) (Coleoptera: Bruchidae). J Stor Prod Res 37: 339-349.
32. Dejen A (2002) Evaluation of some botanicals against maize weevil, *Sitophilus zeamais* Motsch. (Coleoptera: Curculionidae) on stored sorghum under laboratory condition at Sirinka. Pest Manag J Ethiop 6: 73-78.
33. Elhag EA (2000) Deterrent effects of some botanical products on oviposition of the cowpea bruchid *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). Inter J Pest Manag 46: 109-113.
34. Asmanizar A, Djamin D, Indris AB (2011) Evaluation of *jatropha carcus* and *annona muricata* seed crude extracts against *Sitophilus zeamais* infesting stored rice. J Entomol 9: 13-22.
35. Abbott WS (1925) A method of computing the effectiveness of insecticides. J Econ Entomol 18: 265-267.
36. Su HCF (1991) Toxicity and repellency of chenopodium oil to four species of stored-product insects. J Entomol Sci 26: 178-182.