

## Sustained Release of Benomyl Fungicide from Psyllium and Acrylic Acid-Based Hydrogels for its Use in Agricultural Sector

Kiran K<sup>1\*</sup> and Balbir SK<sup>2</sup><sup>1</sup>Department of Chemistry, Rajkiya Kanya Maha Vidyalaya, Shimla, India<sup>2</sup>Department of Chemistry, NIT, Jalandhar, India

\*Corresponding author: Kiran K, Department of Chemistry, Rajkiya Kanya Maha Vidyalaya, Shimla, India, Tel: 01772807959; E-mail: kgrewalcg@gmail.com

Rec date: March 03, 2018; Acc date: April 20, 2018; Pub date: April 29, 2018

Copyright: © 2018 Kiran K, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### Abstract

In this current investigation functionalization of psyllium has been done with acrylic acid in the presence of potassium per-sulphate (KPS)-hexamethylene tetramine (HMTA) as an initiator-crosslinker system. After the initial optimization of different parameters for the synthesis of the hydrogel, the candidate polymer, i.e., (Psy-cl-poly(AA)-IA), has been studied for its sustained release behavior of 'benomyl' fungicide for its further use in agricultural sector. The diffusion exponent 'n' for Psy-cl-poly(AA)-IA has been found to be 1.16. The 'n' value indicated Case-II type diffusion.

**Keywords:** Biopolymer; Psyllium; Acrylic acid; Sustained fungicide release

### Introduction

Recent years have witnessed tremendous applications of pesticides, herbicides, fungicides and fertilizers on agricultural land. These chemicals were exploited without considering the disadvantages of the same. After being used for at-least two to three decades, the world is now observing the hazardous effects of these chemicals. The properties of pesticide such as volatility, leaching and photo-degradation which led to increased dosage of pesticides. Excess pesticides in the soil can be harmful to us in many ways. Out of the total amount of applied pesticides, it is only the little amount (5-10%), which is effectively used, rest being lost in many ways. These lost pesticides are one of the major components of water pollution and can harm living species by intruding the food chain. These chemicals act on the nervous system, circulatory system and genetic system, hence, creating disorders which may be carried over to generations together.

Controlled release of pesticides can thus be a major remedial factor. The release mechanism follows the processes such as diffusion, degradation or a combination of both. Controlled pesticide delivery using polymeric materials has not been much explored by the scientists worldwide. Use of polymeric matrix such as alginate [1,2], ethyl cellulose [3], lignin [4,5] and starch [6-10] for the controlled and sustained release of pesticides has been earlier reported in the literature. Dogan [11] studied the *in vitro* effect of pesticides such as lambda-cyhalothrin, deltamethrin, diazinon, dorzolamide and brinzolamide on carbonic anhydrase enzyme obtained from the blood of fish *Oncorhynchus mykiss* and *Cyprinus carpio* and compared it with carbonic anhydrase inhibitors. Fernandez-Perez et al. [12] studied the use of polymeric matrix to reduce the leaching of herbicide (diuron) into the soil and hence the ground water contamination. The use of polymeric matrix loaded with pesticides help in attaining sustained activity [13], reducing evaporation, photo-degradation [14] and leaching [15].

The present study dealt with the controlled and sustained release of pesticide (Benomyl) using bio-polymer i.e., psyllium-based hydrogels, crosslinked with acrylic acid as a monomer and using potassium persulphate- hexamethylene tetramine as an initiator-crosslinker system.

### Materials and Methods

Plantago Psyllium (Sidhpur Sat-Isabgol Factory), acrylic acid (Merck-Schuchardt), potassium per sulphate (S. D. Fine), hexamethylene tetramine (S. D. Fine) and Benomyl (IFFCO) were used as received. Electronic balance (Libror AEG-220, Shimadzu) was used for weighing purpose.

FTIR spectra of the samples were taken on Perkin Elmer RXI Spectrophotometer using KBr pellets. Scanning electronic microscopic measurements of the samples were taken on Jeol Steroscan 150 Microscope. Thermo-gravimetric Analysis/Differential Thermal Analysis (TGA/DTA) studies were carried-out on Linseis, L-81 11 thermal analyzer in air at a heating rate of 10°C/min. X-ray diffraction studies were carried-out on X-ray diffractometer (Bruker AXS D8 Advance). Crystalline index (C.I.), which measures the orientation of the crystals in a polysaccharide to the polysaccharide axis, was determined by using the wide-angle X-ray diffraction counts at 2θ angle close to 21.297° and 34.744°. The counter reading at peak intensity of 21.297° is said to represent the crystalline material and the peak intensities at 34.744° corresponds to the amorphous material in Psyllium [16]. Percentage crystallinity (%Cr) [17] and the crystalline index [18] were calculated as follows:

$$\%Cr = \frac{I_{21.297}}{I_{21.297} + I_{34.744}} \times 100$$

$$C.I. = \frac{I_{21.297} - I_{34.744}}{I_{21.297}}$$

where  $I_{21.297}$  and  $I_{34.744}$  are the crystalline and amorphous intensities at 2θ scale close to 21.297° and 34.744°, respectively. The concentration of the fungicide at definite time interval was studied

using UV-VIS spectrophotometer (UV-VIS Spectrophotometer-Systronics 118), the fungicide loaded polymers were dried at 40°C using thermostatic incubator (Scientific Instruments Ltd.) and the weighing was performed using electronic weighing balance (Libror-220, Shimadzu).

### Sample preparation

Synthesis of different crosslinked graft copolymers of Psyllium with acrylic acid was carried-out in-air (IA). Psyllium (1.0 g) was taken in a flask having a known amount of solvent. A known amount of monomer was added to this reaction mixture followed by the addition of an initiator and crosslinker dissolved previously in minimum amount of the solvent. Mixture was made uniform by thorough stirring. The reaction was carried-out under required temperature. After completion of reaction, the homopolymer was removed with water. The synthesized polymers were dried to constant weight in hot air oven at 50°C. Optimization was carried-out as a function of percent grafting ( $P_g$ ) and percent swelling ( $P_s$ ) which were calculated as per the equations [19,20]:

$$P_g = \frac{W_f - W_b}{W_b} \times 100 \dots \dots \dots (a)$$

where  $W_f$ =weight of functionalized polymer,  $W_b$ =weight of backbone polymer.

$$P_s = \frac{W_s - W_d}{W_d} \times 100 \dots \dots \dots (b)$$

where,  $W_s$  and  $W_d$  are the weights of swollen polymer and dry polymer, respectively.

### Swelling studies of candidate polymer

The purpose of swelling studies is to optimize the different conditions best suited for maximum swelling. These studies in distilled water have been carried-out using gravimetric method as a function of time (2, 4, 8, 16 and 24 h) and temperature (25, 37, 45 and 50°C). 100 mg of the polymer sample was immersed in 250 ml of water. Sample was taken-out after a definite interval of time, wiped and weighed. The process was repeated till a constant weight was reached. The percent swelling was calculated as per the following equation.

$$P_s = \frac{W_s - W_d}{W_d} \times 100 \dots \dots \dots (b)$$

After optimization of time as a function of  $P_s$ , the temperature for maximum percent swelling was optimized.

### Loading of fungicides onto polymeric matrices

Saturated aqueous solution of fungicide was prepared and  $\lambda_{max}$  was noted down. 750 mg of polymeric sample was immersed in 100 ml of fungicide solution for 24 h. The polymer was then taken-out from the solution, wiped off with tissue paper and was kept for drying in incubator at 40C. Dried fungicide-loaded polymer was washed with distilled water to remove any surface adhered molecules and was studied for fungicide release kinetics as a function of release time.

**Mathematical analysis:** Mathematical modeling of fungicide release with hydrogel was studied as per the methods reported earlier in the literature [21-25]. The empirical equation used to describe the water uptake which is the weight gain ( $M_s$ ) can be presented as [26].

$$M_s = Kt^n \dots \dots \dots (1)$$

where, 'k' and 'n' are constant. 'n'=0.5 reveals the normal Fickian diffusion whereas 'n'=1.0 signifies Case II diffusion. Non-Fickian or anomalous diffusion is characterized with value of n between 0.5 and 1.0 [26]. Evaluation of fungicide release from the swellable polymers can be assessed from the above power law expression. Here,  $M_s$  is replaced with  $M_t/M_\infty$  and the expression is modified as [27]:

$$\frac{M_t}{M_\infty} = Kt^n \dots \dots \dots (2)$$

where,  $M_t/M_\infty$  is the fractional release of fungicide in time t. 'k' is the constant which is characteristic of polymer-fungicide system. 'n' is the diffusion exponent characteristic of the release mechanism. Value of 'n' and 'k' can be evaluated from the slope and intercept of the plot between  $\ln M_t/M_\infty$  versus  $\ln t$ , respectively. This equation can be applied until 60% of the total fungicide is released.

**Diffusion coefficients:** Analysis of fungicide release from various hydrogels can be performed by calculating the diffusion coefficients. Diffusion process can be adequately described through Fick's first and second law. Integral diffusion for the cylindrical hydrogel can be given as [28]:

$$\frac{M_t}{M_\infty} = 4 \times \frac{Dt}{\pi l^2} \dots \dots \dots (3)$$

where,  $M_t/M_\infty$  is the fractional release,  $M_t$  is the fungicide released at time 't', and  $M_\infty$  is the fungicide release at equilibrium, D is the diffusion coefficient and l is the thickness of the sample.

The average diffusion coefficient (DA) for the 50% release of the fungicide can be calculated by putting  $M_t/M_\infty=0.5$  in Eq. 3, and can be presented as:

$$D_A = \frac{0.049l^2}{t^{1/2}} \dots \dots \dots (4)$$

where,  $t^{1/2}$  is the time required for the 50% release of fungicide.

Eq. 5 gives the value of late diffusion coefficient and can be calculated as [27,28]:

$$\frac{M_t}{M_\infty} = 1 - \left(\frac{8}{\pi^2}\right) \exp\left(\frac{-\pi^2 Dt}{l^2}\right) \dots \dots \dots (5)$$

$$D_L = \frac{(slope)l^2}{8} \dots \dots \dots (6)$$

The slope of plot between  $\ln(1 - M_t/M_\infty)$  and time 't' has been used for the evaluation of DL [27].

## Results and Discussion

### Optimization of different reaction parameters

The different reaction parameters such as initiator concentration, amount of solvent, reaction time, reaction temperature, pH, monomer concentration and crosslinker concentration were optimized and the scheme of results for optimization of the reaction parameters is illustrated in Table 1 [29].

S. No.	Initiator Conc.	Amount of solvent (ml)	Reaction time (min.)	Reaction temperature(°C)	pH	Monomer conc.mol/L	Crosslink-er conc. mol/L	Percent grafting	Percent swelling
1.	0	25	120	65	7	0.725	0.02853	0	-
2.	$9.24 \times 10^{-3}$	25	120	65	7	0.725	0.02853	73	-
3.	$18.49 \times 10^{-3}$	25	120	65	7	0.725	0.02853	125	-
4.	$27.74 \times 10^{-3}$	25	120	65	7	0.725	0.02853	84	-
5.	$36.99 \times 10^{-3}$	25	120	65	7	0.725	0.02853	81	-
6.	$18.49 \times 10^{-3}$	10	120	65	7	0.725	0.02853	32	-
7.	$18.49 \times 10^{-3}$	15	120	65	7	0.725	0.02853	61	-
8.	$18.49 \times 10^{-3}$	20	120	65	7	0.725	0.02853	118	-
9.	$18.49 \times 10^{-3}$	25	120	65	7	0.725	0.02853	124	-
10.	$18.49 \times 10^{-3}$	30	120	65	7	0.725	0.02853	80	-
11.	$18.49 \times 10^{-3}$	25	30	65	7	0.725	0.02853	0	-
12.	$18.49 \times 10^{-3}$	25	60	65	7	0.725	0.02853	116	-
13.	$18.49 \times 10^{-3}$	25	90	65	7	0.725	0.02853	118	-
14.	$18.49 \times 10^{-3}$	25	120	65	7	0.725	0.02853	125	-
15.	$18.49 \times 10^{-3}$	25	150	65	7	0.725	0.02853	90	-
16.	$18.49 \times 10^{-3}$	25	120	55	7	0.725	0.02853	97	-
17.	$18.49 \times 10^{-3}$	25	120	60	7	0.725	0.02853	118	-
18.	$18.49 \times 10^{-3}$	25	120	65	7	0.725	0.02853	125	-
19.	$18.49 \times 10^{-3}$	25	120	70	7	0.725	0.02853	110	-
20.	$18.49 \times 10^{-3}$	25	120	75	7	0.725	0.02853	0	-
21.	$18.49 \times 10^{-3}$	25	120	65	4	0.725	0.02853	5	-
22.	$18.49 \times 10^{-3}$	25	120	65	7	0.725	0.02853	125	-
23.	$18.49 \times 10^{-3}$	25	120	65	9	0.725	0.02853	9	-
24.	$18.49 \times 10^{-3}$	25	120	65	7	0.29	0.02853	-	2309
25.	$18.49 \times 10^{-3}$	25	120	65	7	0.58	0.02853	-	3244
26.	$18.49 \times 10^{-3}$	25	120	65	7	0.807	0.02853	-	3090
27.	$18.49 \times 10^{-3}$	25	120	65	7	1.16	0.02853	-	2466
28.	$18.49 \times 10^{-3}$	25	120	65	7	1.45	0.02853	-	2192
29.	$18.49 \times 10^{-3}$	25	120	65	7	0.58	0.01426	-	2000
30.	$18.49 \times 10^{-3}$	25	120	65	7	0.58	0.02853	-	3200
31.	$18.49 \times 10^{-3}$	25	120	65	7	0.58	0.04279	-	2987
32.	$18.49 \times 10^{-3}$	25	120	65	7	0.58	0.05706	-	2875
33.	$18.49 \times 10^{-3}$	25	120	65	7	0.58	0.07133	-	2600

**Table 1:** Scheme for optimization of various reaction parameters.

### FTIR spectroscopy

The IR spectrum of Psyllium [29] showed broad peaks at 3780.9  $\text{cm}^{-1}$  and 3427.6  $\text{cm}^{-1}$  (O-H stretching bonded absorption of carbohydrates), 2925.8  $\text{cm}^{-1}$  ( $\text{CH}_2$  asymmetric stretching), 1378.8  $\text{cm}^{-1}$  ( $\text{CH}$ ,  $\text{CH}_2$  and OH in-plane bending in carbohydrates), 1039.5  $\text{cm}^{-1}$  (C-O stretching region as complex bands, resulting from C-O and C-O-C stretching vibrations), 897  $\text{cm}^{-1}$  and 533  $\text{cm}^{-1}$  (pyranose rings). Whereas, Psy-cl-poly(AA)-IA showed peaks at 2857.1  $\text{cm}^{-1}$  (carboxylic acid O-H stretching), 1737.9  $\text{cm}^{-1}$  (C=O stretching in carboxylic acid), 1636.4  $\text{cm}^{-1}$  (strong C.....O asymmetric stretching) and 1400  $\text{cm}^{-1}$  (coupled OH in-plane bending and C-O stretching) besides peaks obtained with that of Psyllium.

### Scanning electron microscopic studies (SEM)

SEM of psyllium and Psy-cl-poly(AA)-IA clearly indicated the morphological changes which are brought about as a result of crosslinking onto the backbone [29].

### Thermogravimetric analysis and differential thermal analysis studies (TGA/DTA)

Thermo-gravimetric analysis of Psyllium and Psy-cl-poly(AA)-IA was carried-out as a function of percent weight loss versus temperature at a rate of 10 $^{\circ}\text{C}/\text{min}$  in air. The analysis was performed in order to examine the changes in thermal properties of the Psyllium brought about by graft copolymerization with acrylic acid in air [29]. Psyllium exhibited initial decomposition temperature (IDT) at 229.3 $^{\circ}\text{C}$  and final decomposition temperature (FDT) at 601.9 $^{\circ}\text{C}$ . Two stage decomposition has been found between 229.3 $^{\circ}\text{C}$  to 316.4 $^{\circ}\text{C}$  (wt. loss=47.16%) and 316.4 $^{\circ}\text{C}$  to 601.9 $^{\circ}\text{C}$  (wt. loss=38.0%). DTA curve of Psyllium indicated one endothermic peak at 67.1 $^{\circ}\text{C}$  (-2.3  $\mu\text{V}$ ) and two exothermic peaks at 298.9  $^{\circ}\text{C}$  (12.7  $\mu\text{V}$ ) and 487.9 $^{\circ}\text{C}$  (16.0  $\mu\text{V}$ ). It has been found that in case of Psyllium, IDT (229.3 $^{\circ}\text{C}$ ) is higher as compared to that of Psy-cl-poly(AA)-IA (172.2 $^{\circ}\text{C}$ ). However, FDT of Psy-cl-poly(AA)-IA has been found to be higher (620.5 $^{\circ}\text{C}$ ) than that of Psyllium (601.9 $^{\circ}\text{C}$ ). Two-stage decomposition from 172.2 $^{\circ}\text{C}$  to 496.1 $^{\circ}\text{C}$  (wt. loss=40.8%) and 496.1 $^{\circ}\text{C}$  to 620.5 $^{\circ}\text{C}$  (wt. loss=31.0%) was observed. DTA studies of this crosslinked sample showed one endothermic peak at 138.7 $^{\circ}\text{C}$  (-3.1  $\mu\text{V}$ ) and one exothermic peak at 534.3 $^{\circ}\text{C}$  (22.4  $\mu\text{V}$ ). It shows that exothermic combustion of Psy-cl-poly(AA)-IA persists at higher temperature as compared to that of Psyllium [30].

### X-Ray diffraction (XRD) studies

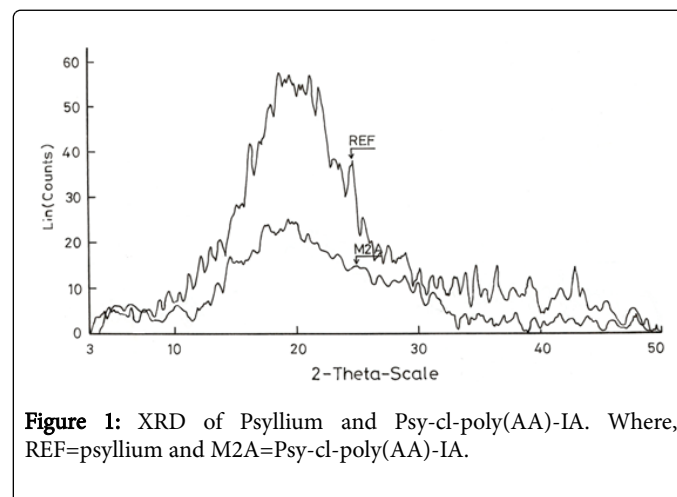
It has been observed that Psyllium exhibited 75.76% percentage crystallinity and crystalline index was found to be 0.6800. Whereas, Psy-cl-poly(AA)-IA exhibited 64.91% percentage crystallinity and the crystalline index of 0.4592 (Table 2).

S. No.	Sample	Pg SE $\pm$	at 2 $\theta$ -Scale		%Cr	C.I.
			121.297	134.744		
1	Psyllium	-	100	32	75.76	0.68

2	Psy-cl-poly(AA)-IA	125.0 $\pm$ 1.00	98	53	64.91	0.4592
---	--------------------	------------------	----	----	-------	--------

**Table 2:** Percentage crystallinity (%Cr) and Crystalline Index (C. I.) of Psyllium and Psy-cl-poly(AA). Where, I=relative intensity; IA=in air; P<sub>g</sub>=percent grafting.

It is evident from Figure 1 that spectrum of Psyllium is more convex than that of Psy-cl-poly(AA)-IA. The incorporation of monomer moiety onto the Psyllium had impaired its crystallinity, thereby decreasing the %Cr [31]. The 2  $\theta$ , I and d-values of graft copolymer has been found to be quite different than that of Psyllium. Psy-cl-poly(AA)-IA showed the relative intensity of 49 (d=2.2175) at 42.9 $^{\circ}$  2  $\theta$ -scale. This behaviour is due to the involvement of primary bonding like covalent bonding between Psyllium and acrylic acid during graft copolymerization [16].

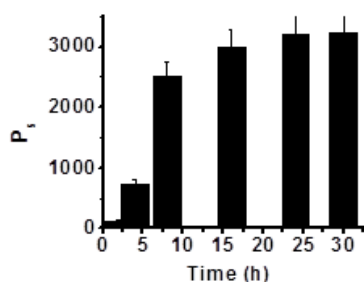


**Figure 1:** XRD of Psyllium and Psy-cl-poly(AA)-IA. Where, REF=psyllium and M2A=Psy-cl-poly(AA)-IA.

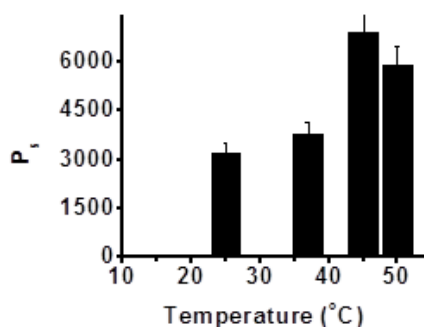
### Swelling studies of the candidate hydrogel

**Effect of time on percent swelling:** Figure 2a reveals the effect of swelling time onto Ps. It was observed that the Ps increased with increase in swelling time upto 24 h at 25 $^{\circ}\text{C}$  and thereafter, attained a constant value. This can be explained based on the fact that after 24 h the porous network of the polymer got saturated with the solvent molecules with no more accommodation for further solvent molecules.

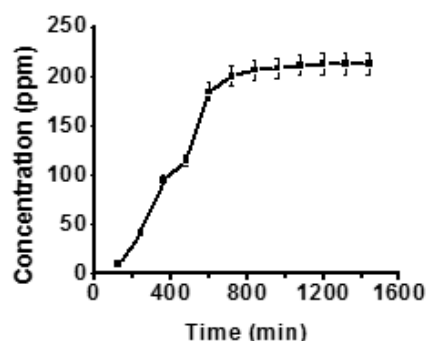
**Effect of temperature on percent swelling:** It was observed that maximum Ps (6900  $\pm$  15.27) was found at 45 $^{\circ}\text{C}$  which decreased with further increase in temperature (Figure 2b). It was since beyond optimum temperature, the polymer matrix starts crumbling, thereby, leading to desorption and decreased Ps.



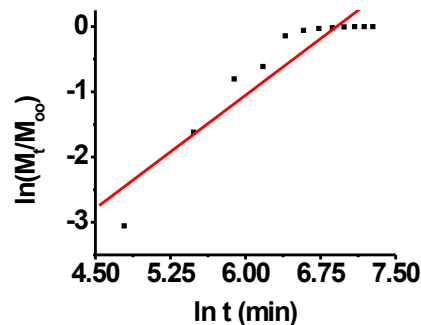
**Figure 2a:** Effect of swelling time onto Ps of Psy-cl-poly(AA)-IA in distilled water.



**Figure 2b:** Effect of swelling temperature onto Ps of Psy-cl-poly(AA)-IA in distilled water.



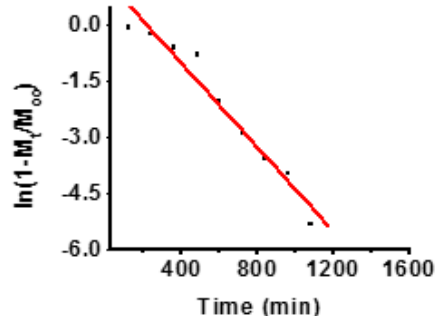
**Figure 3:** Sustained release of Benomyl with Psy-cl-poly(AA)-IA as a function of time at 6.0 pH.



**Figure 4:** Plot of  $\ln M_t/M_\infty$  versus  $\ln t$  for the Benomyl release behaviour of Psy-cl-poly(AA)-IA at 6.0 pH.

### Sustained release of benomyl

**Fungicide (Benomyl) release kinetics of Psy-cl-poly(AA)-IA:** Psy-cl-poly(AA)-IA exhibited  $10 \pm 1.15$  ppm initial release of fungicide in 2 h and maximum release at the time interval of 20 h was found to be  $212 \pm 4.93$  ppm (Figure 3). The diffusion exponent 'n' for Psy-cl-poly(AA)-IA has been found to be 1.16. The 'n' value indicated Case-II type diffusion in which the rate of diffusion is very rapid as compared with the relaxation process [32]. The Case-II type diffusion has been confirmed from the characteristic sigmoid curve obtained in the plot between  $M_t/M_\infty$  versus  $t^{1/2}$ . Gel characteristic constant 'k' was found to be  $6.51 \times 10^{-4}$  (Figure 4). Moreover, the values of initial diffusion coefficient have been found to be greater than late diffusion coefficient indicating more fungicide release in the early stages (Tables 3 and 4). Huang et al. [33] also reported Case-II diffusion at higher pH values for the release of ketoprofen using hydrogels based on cationic guar gum and acrylic acid monomer (Figures 4-6).



**Figure 5:** Plot of  $\ln(1-M_t/M_\infty)$  versus time for the Benomyl release behaviour of Psy-cl-poly(AA)-IA at 6.0 pH.

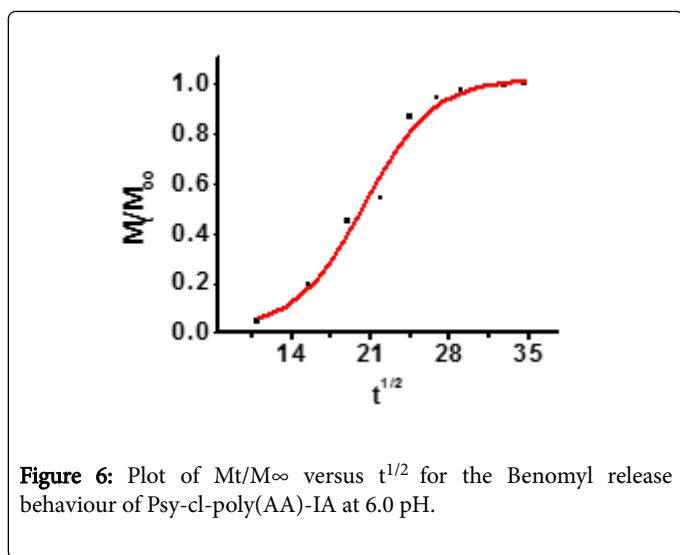


Figure 6: Plot of  $M_t/M_\infty$  versus  $t^{1/2}$  for the Benomyl release behaviour of Psy-cl-poly(AA)-IA at 6.0 pH.

18	M	211
	SE	4.04
	SD	7
20	M	212
	SE	4.93
	SD	8.54
22	M	--
	SE	--
	SD	--

Table 3: Benomyl release behaviour of Psy-cl-poly(AA)-IA. Where No. of samples used in each case=3; M=mean; SE=Standard error of mean; SD=Standard deviation; --=constant release.

Time	Sample a	Psy-cl-poly(AA)-IA
2	M	10
	SE	1.52
	SD	2.64
4	M	42
	SE	1.15
	SD	2
6	M	95
	SE	2.08
	SD	3.6
8	M	115
	SE	3
	SD	5.19
10	M	184
	SE	2.08
	SD	3.6
12	M	200
	SE	5.5
	SD	9.53
14	M	206
	SE	3.78
	SD	6.55
16	M	208
	SE	4.72
	SD	8.18

Sample Code	Structure of benomyl	Diffusion exponent 'n'	Gel characteristic constant 'k' × 10 <sup>-4</sup>	Diffusion coefficients (cm <sup>2</sup> /min)		
				Di × 10 <sup>-5</sup>	DA × 10 <sup>-5</sup>	DL × 10 <sup>-5</sup>
Psy-cl-poly(AA)-IA		1.16	6.51	0.0144	0.46	0.9005

Table 4: Diffusion exponent, Gel characteristic constant and Diffusion coefficients for the release of Benomyl through Psy-cl-poly(AA)-IA at pH 6.0. Where Di=Initial diffusion coefficient, DL=late diffusion coefficient, DA=average diffusion coefficient.

## Conclusion

The current study dealt with the synthesis of psyllium and acrylic acid-based hydrogels and their utilization in the sustained delivery of a fungicide for use in agriculture sector. It has been found that the polymer synthesized can sustain delivery of benomyl up to 20 h with release of 212 ppm. The rate of thermal decomposition of the synthesized polymer is greater as compared to that of psyllium. The diffusion exponent 'n' for Psy-cl-poly(AA)-IA has been found to be 1.16. The 'n' value indicated Case-II type diffusion. Hence the synthesized polymer can be quite important from the technological point of view.

## Acknowledgements

We are thankful to Ministry of Human Resource for financing the above research and thankful to Department of Chemistry, Himachal Pradesh University, for their constructive advice during research.

## References

- Johnson RM, Pepperman AB (1995) Soil column mobility of metribuzin from alginate-encapsulated controlled release formulations. Journal of Agricultural and Food Chemistry 43: 241-246.
- Kulkarni AR, Soppimath KS, Aminabhavi TM, Dave AM, Mehta MH (2000) Glutaraldehyde crosslinked sodium alginate beads containing liquid pesticide for soil application. Journal of Controlled Release 63: 97-105.

3. Fernandez-Urrusuno R, Gines JM, Morillo E (2000) Development of controlled release formulations of alachlor in ethylcellulose. *Journal of Microencapsulation* 17: 331-342.
4. Kok FN, Wilkins RM, Cain RB, Arica MY, Alaeddinoglu G, et al. (1999) Controlled release of aldicarb from lignin loaded ionotropic hydrogel microspheres. *Journal of Microencapsulation* 16: 613-623.
5. Cotterill JV, Wilkins RM, Da Silva FT (1996) Controlled release of diuron from granules based on a lignin matrix system. *Journal of Controlled Release* 40: 133-142.
6. Doane WM (1992) Encapsulation of pesticides in starch by extrusion. *Industrial Crops and Products* 1: 83-87.
7. Trimnell D, Shasha BS (1988) Autoencapsulation: A new method for entrapping pesticides within starch. *Journal of Controlled Release* 7: 25-31.
8. Frederiksen HK, Hansen HC (2002) Starch-encapsulated chlorpyrifos: release rate, insecticidal activity and degradation in soil. *Journal of Microencapsulation* 19: 319-331.
9. Shukla PG, Kalidhass B, Shah A, Palaskar DV (2002) Preparation and characterization of microcapsules of water-soluble pesticide monocrotophos using polyurethane as carrier material. *Journal of Microencapsulation* 19: 293-304.
10. Trimnell D, Shasha BS (1990) Controlled release formulations of atrazine in starch for potential reduction of groundwater pollution. *Journal of Controlled Release* 12: 251-256.
11. Dogan S (2006) The in vitro effects of some pesticides on carbonic anhydrase activity of *Oncorhynchus mykiss* and *Cyprinus carpio carpio* fish. *Journal of Hazardous Materials* 132: 171-176.
12. Fernández-Pérez M, Villafranca-Sanchez M, Gonzalez-Pradas E, Flores-Céspedes F (1999) Controlled release of diuron from an alginate-bentonite formulation: water release kinetics and soil mobility study. *Journal of Agricultural and Food Chemistry* 47: 791-798.
13. Coffman CB, Gentner WA (1980) Persistence of several controlled release formulations of trifluralin in greenhouse and field. *Weed Science* 28: 21-23.
14. Schreiber MM, Shasha BS, Ross MA, Orwick PL, Edgecomb DW (1978) Efficacy and rate of release of EPTC and butylate from starch encapsulated formulations under greenhouse conditions. *Weed Science* 26: 679-686.
15. Baur JR (1980) Release Characteristics of Starch Xanthide Herbicide Formulations I. *Journal of Environmental Quality* 9: 379-382.
16. Mishra A, Agarwal M, Bajpai M, Rajani S, Mishra RP (2002) Plantago psyllium mucilage for sewage and tannery effluent treatment. *Iranian Polymer Journal* 11: 381-386.
17. Agrawal AM, Manek RV, Kolling WM, Neau SH (2003) Studies on the interaction of water with ethylcellulose: effect of polymer particle size. *AAPS PharmSciTech* 4: 469-479.
18. Segal LG, Creely JJ, Martin Jr AE, Conrad CM (1959) An empirical method for estimating the degree of crystallinity of native cellulose using the X-ray diffractometer. *Textile Research Journal* 29: 786-794.
19. Kaith BS, Singha AS, Sharma SK (2003) Graft copolymerization of flax fibers with binary vinyl monomer mixtures and evaluation of swelling, moisture absorbance and thermal behavior of the grafted fibers. *Journal of Polymer Materials* 20: 195-200.
20. Kumar K, Kaith BS, Jindal R, Mittal H (2012) Gamma radiation initiated synthesis of Psyllium and acrylic acid based polymeric networks for selective absorption of water from different oil-water emulsions. *Journal of Applied Polymer Science* 124: 4969-4977.
21. Brannon-Peppas L, Peppas NA (1989) Solute and penetrant diffusion in swellable polymers. IX. The mechanisms of drug release from pH-sensitive swelling-controlled systems. *Journal of Controlled Release* 8: 267-274.
22. Bamba M, Puisieux F, Marty JP, Carstensen JT (1979) Physical model for release of drug from gelforming sustained release preparations. *International Journal of Pharmaceutics* 3: 87-92.
23. Korsmeyer RW, Von Meerwall E, Peppas NA (1986) Solute and penetrant diffusion in swellable polymers. II. Verification of theoretical models. *Journal of Polymer Science Part B: Polymer Physics* 24: 409-434.
24. Lee PI (1980) Diffusional release of a solute from a polymeric matrix—approximate analytical solutions. *Journal of Membrane Science* 7: 255-275.
25. Peppas NA, Gurny R, Doelker E, Buri P (1980) Modelling of drug diffusion through swellable polymeric systems. *Journal of Membrane Science* 7: 241-253.
26. Alfrey T, Gurnee EF, Lloyd WG (1966) Diffusion in glassy polymers. In *Journal of Polymer Science: Polymer Symposia*. Wiley Subscription Services Inc, A Wiley Company 12: 249-261.
27. Ritger PL, Peppas NA (1987) A simple equation for description of solute release I. Fickian and non-fickian release from non-swellable devices in the form of slabs, spheres, cylinders or discs. *Journal of Controlled Release* 5: 23-36.
28. Ritger PL, Peppas NA (1987) A simple equation for description of solute release II. Fickian and anomalous release from swellable devices. *Journal of Controlled Release* 5: 37-42.
29. Kaith BS, Kumar K (2008) In vacuum synthesis of psyllium and acrylic acid-based hydrogels for selective water absorption from different oil-water emulsions. *Desalination* 229: 331-341.
30. Kaith BS, Kumar K (2007) Preparation of psyllium mucilage and acrylic acid-based hydrogels and their application in selective absorption of water from different oil/water emulsions, pp: 529-538.
31. Kaith BS, Kalia S (2007) Synthesis and characterization of graft copolymers of flax fiber with binary vinyl monomers. *International Journal of Polymer Analysis and Characterization* 12: 401-412.
32. Ensore DJ, Hopfenberg HB, Stannett VT (1977) Effect of particle size on the mechanism controlling n-hexane sorption in glassy polystyrene microspheres. *Polymer* 18: 793-800.
33. Huang Y, Yu H, Xiao C (2007) pH-sensitive cationic guar gum/poly (acrylic acid) polyelectrolyte hydrogels: swelling and in vitro drug release. *Carbohydrate Polymers* 69: 774-783.