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Silver Nanoparticles Biosynthesis by *Fusarium oxysporum* and Determination of Its Antimicrobial Potency

Nida Tabassum Khan*, Mahum Jameel and Jibran Jameel

Department of Biotechnology, Balochistan University of Information Technology Engineering and Management Sciences (BUITEMS), Quetta, Pakistan

Abstract

Nanotechnology encompasses the engineering of nanoparticles with enhanced functionality and improved stability which is quite different from the bulk form of the same material. Fungi as bionanofactories for the production of diverse metallic nanocrystals is the right choice because it not only offers an ecofriendly and cost effective procedure but also provide easy and simple down streaming for product recovery. Fungal filtrate of *Fusarium oxysporum* isolated from banana fruit was used for the amalgamation of silver nanoparticles. Appearance of light brown colour of fungal filtrate upon incubation with AgNO₃ indicates silver nanoparticle formation with strong absorption in the visible region at 440 nm as determined by ultra violet visible spectroscopy. Disc diffusion assay showed enhanced antibacterial and antifungal activity of silver nanoparticles against pathogens like *Candida albicans*, *Escherichia coli*, *Candida krusei*, *Staphylococcus aureus* and *Aspergillus flavus*.

Keywords: Green synthesis; *Fusarium oxysporum*; Silver nanoparticles; Ultra violet spectroscopy

Introduction

Now a days, silver nanoparticles have gained significant consideration due to their unique characteristics and diverse application like electrical conduction [1], surface-enhanced Raman scattering [2], nucleotide sequencing [3], antibacterial and antifungal activities etc. [4,5]. Numerous methods were available for the fabrication of silver nanoparticles such as heat decomposition in organic solvents [6], reduction with or without the presence of stabilizers [7], photoreduction [8,9], and electromagnetic reduction [10-12] etc. But these above mentioned approaches are costly and involve the use of compounds which may pose potential environmental risks. Therefore there is a need to develop an eco-friendly procedure that applies biological principles in nanoparticle formation i.e., biomimetic approach [13-15] such as the use of fungus [16], proteins [17], plants or plant extracts [18-20] for the production of metallic nanoparticles that do not employ harmful compounds. Silver nanoparticle can be produced by a variety of biological systems such as bacteria, plants, fungi but among these, eukaryotic fungi is the suitable candidate with unique features like increased growth and rapid reproduction by virtue of mycelial branching, capability to produce number of enzymes with the ability to bio-accumulate different metal nanoparticles by bioreduction, ability to grow under extreme conditions etc. make them suitable for such purposes [21-23]. In current study silver nanoparticles were produced from Fusarium oxysporum under controlled in vitro conditions and the mycosynthesized silver nanoparticles were than tested for their antimicrobial activity against different disease causing pathogens.

Materials and Methods

Fusarium oxysporum was isolated from banana fruit on PDA (potato dextrose agar) at 28°C for three to four days. The fungus was identified using morphological characteristics such as colony colour, texture of mycelia etc. *F. oxysporum* was grown on CD (cezapex Dox) broth (glucose (10 g), magnesium sulphate (0.5 g), yeast extract (1 g), Potassium dihydrogen phosphate (1 g), calcium chloride (0.5 g), zinc sulphate (0.01 g), ferrous sulphate (0.01 g) and Sodium nitrate (2 g) dissolved in one liter of distilled water and autoclaved for 15 min at 121°C at 15 psi (pound/square inches). The flasks containing

CD medium inoculated with F. oxysporum was kept for 5 days on rotatory shaker at 25°C at 150 rpm. The mycelia were harvested using Whatman's filter paper no. 42 followed by extensive washing twice to remove media components. 50 ml of obtained fungal filtrate was then incubated with 10 ml of AgNO, (10 mM) at 28°C for 24 hrs in dark to avoid any photochemical reactions. Freshly prepared CD broth with silver nitrate solution was run as the control. Confirmation of silver nitrate formation is indicated by a colour change in the experimental flask. Bioreduction of Ag metal ions in the aqueous solution was monitored by means of ultra violet spectroscopy (JENWAY 6305) from 400 to 500 to determine optimum wavelength. Antimicrobial activity of the silver nanoparticles were determine using dis diffusion assay carried on Yeast Malt extract agar (YM) against Candida albicans, Escherichia coli, Candida krusei, Staphylococcus aureus and Aspergillus flavus by placing one cm sterile filter paper disc impregnated with 60 µL of colloidal siver nanoparticle. Ruler scale was used to estimate the diameter of inhibition zone.

Results and Discussion

Sufficient biomass of *Fusarium oxysporum* was obtained in CD broth because of utilization of the sugar source glucose and all the other essential nutrients required by the fungal mycelia to grow and continuous agitation helps in uniform distribution of all the nutrients available in the CD medium (Figure 1) [20].

Colour change of the filtrate incubated with $AgNO_3$ to brown depicted silver nanoparticle formation (Figure 2). Silver nanoparticles were found to be quite stable in the fungus supernatant [24]. Silver

*Corresponding author : Nida Tabassum Khan, Department of Biotechnology, Faculty of Life Sciences and Informatics, Balochistan University of Information Technology Engineering and Management Sciences (BUITEMS), Quetta, Pakistan, Tel: +92-3368164903; E-mail: nidatabassumkhan@yahoo.com

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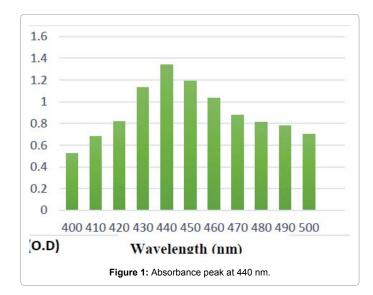
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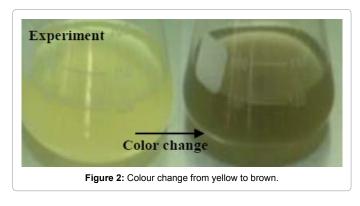
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nanoparticle exhibits strong absorption in the visible region. Ultraviolet visible spectrum of the sample revealed absorbance peak at 440 nm which is specific for silver nanoparticles [10]. A single peak indicates the formation of spherical nanoparticles with wide spread distribution (Figure 3). However no change in colour was observed in freshly prepared CD media incubated with silver nitrate indicating absence of silver nanoparticles.

Antimicrobial potency of silver nanoparticles were tested against various pathogens and compared with the controls. The diameter of inhibition zone is shown in Table 1.







S.no	Test pathogens	Zone of inhibition (cm) Silver nanoparticles (60 µL)
1	Staphylococcus aureus	0.9
2	Candida albicans	0.8
3	Candida krusei	0.8
4	Aspergillus flavus	0.8
5	Escherichia coli	1.0

 Table 1: Silver nanoparticle antimicrobial activity against various pathogens.

Silver nanoparticles could inhibit two different pathogenic bacteria including *Escherichia coli* and *Staphylococcus aureus*. Thus displayed broad antibacterial spectrum as increased activity was seen against these two bacteria Staphylococcus aureus and Escherichia coli with a wide clear inhibition zone of 0.9 cm and 1 cm around the sterile discs was observed. Enhanced antifungal potency of these mycosynthesized silver nanoparticles was seen against the pathogenic *Candida* species such as *Candida krusei* and *Candida albicans*. Similar results were reported while using silver nanoparticles synthesized at different physioculture conditions by Nayak et al. [25]. Silver nanoparticles produce from *F. oxysporum* were also found to be effective against *Aspergillus flavus*. Therefore because of their effective antimicrobial properties silver nanoparticles could have potential clinical applications.

Conclusion

Green mycofabrication of Silver nanoparticles using *Fusarium* oxysporum is an ecofriendly and cost effective process which can be easily achieved in lab under standard conditions forming stable colloidal silver nanoparticles of spherical morphology with a potential to be used as anti-bacterial and antifungal agents.

References

- Chang LT, Yen CC (1995) Studies on the Preparation and Properties of Conductive Polymers. VIII. Use of Heat Treatment to Prepare Metalized Films from Silver Chelate of PVA and PAN. J Appl Polym Sci 55: 371-374.
- Matejka P, Vlckova B, Vohlidal J, Pancoska P, Baumuruk V, et al. (1992) The Role of Triton X-100 as an Adsorbate and a Molecular Spacer on the Surface of Silver Colloid: A Surface-Enhanced Raman Scattering Study. J Phys Chem 96: 1361-1366.
- Cao YW, Jin R, Mirkin CA (2001) DNA-Modified Core-Shell Ag/Au Nanoparticles. J Am Chem Soc 123: 7961-7962.
- Shahverdi AR, Mianaeian S, Shahverdi HR, Jamalifar H, Nohi AA, et al. (2007) Rapid Synthesis of Silver Nanoparticles Using Culture Supernatants of Enterobacteria: A Novel Biological Approach. Process Biochem 42: 919-923.
- Baker C, Pradhan A, Pakstis L, Pochan DJ, Shah SI, et al. (2005) Synthesis and Antibacterial Properties of Silver Nanoparticles. J Nanosci Nanotechnol 5: 224-249
- Esumi K, Tano T, Torigoe K, Meguro K (1990) Preparation and Characterization of Biometallic Pd-Cu Colloids by Thermal Decomposition of Their Acetate Compounds in Organic Solvents. J Chem Mater 2: 564-567.
- Liz-Marzan LM, Lado-Tourino I (1996) Reduction and Stabilization of Silver Nanoparticles in Ethanol by Nonionic Surfactants. Langmuir 12: 3585-3589.
- Pileni MP (2000) Fabrication and Physical Properties of Self-Organized Silver Nanocrystals. Pure Appl Chem 72: 53-65.
- Sun YP, Atorngitjawat P, Meziani MJ (2001) Preparation of Silver Nanoparticles via Rapid Expansion of Water in Carbon Dioxide Microemulsion into Reductant Solution. Langmuir 17: 5707-5710.
- Henglein A (1993) Physicochemical Properties of Small Metal Particles in Solution: 'Microelectrode' Reactions, Chemisorption, Composite Metal Particles, and the Atom-to-Metal Transition. Phys Chem B 97: 5457-5471.
- Henglein A (1998) Colloidal Silver Nanoparticles: Photochemical Preparation and Interaction with O₂, CCl₄, and Some Metal Ions. Chem Mater 10: 444-446.

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- Henglein A (2001) Reduction of Ag (CN)⁻² on Silver and Platinum Colloidal Nanoparticles. Langmuir 17: 2329-2333.
- Nair B, Pradeep T (2002) Coalescence of Nanoclusters and Formation of Submicron Crystallites Assisted by *Lactobacillus* Strains. Cryst. Growth Des 2: 293-298.
- Klaus T, Joerger R, Olsson E, Granqvist CG (1999) Silver-Based Crystalline Nanoparticles, Microbially Fabricated. Proc Natl Acad Sci USA 96: 13611-13614.
- Konishi Y, Uruga T (2007) Bioreductive Deposition of Platinum Nanoparticles on the Bacterium Shewanella algae. J Biotechnol 128: 648-653.
- Vigneshwaran N, Kathe A (2007) Silver-protein (core-shell) nanoparticle production using spent mushroom substrate. Langmuir 23: 7113-7117.
- 17. Willner I, Baron R, Willner B (2006) Growing Metal Nanoparticles by Enzymes. J Adv Mater 18: 1109-1120.
- Chandran SP, Chaudhary M, Pasricha R, Ahmad A, Sastry M, et al. (2006) Synthesis of gold nanotriangles and silver nanoparticles using Aloe Vera plant extract. Biotechnol Prog 22: 577-583.

- Shankar SS, Rai A, Ahmad A, Sastry M (2004) Rapid synthesis of Au, Ag and bimetallic Au core–Ag shell nanoparticles using Neem (*Azadirachta indica*) leaf broth. J Colloid Interface Sci 275: 496-502.
- Jae YS, Beom SK (2009) Rapid Biological Synthesis of Silver Nanoparticles Using Plant Leaf Extracts. Bioprocess Biosyst Eng 32: 79-84.
- Khambhaty Y, Mody K, Basha S, Jha B (2009) Kinetics, equilibrium and thermodynamic studies on biosorption of hexavalent chromium by dead fungal biomass of marine *Aspergillus niger*. Chem Eng J 145: 489-495.
- Vala AK, Upadhyay RV (2008) on the tolerance and accumulation of arsenic by facultative marine Aspergillus sp. Res J Biotechnol, pp: 366-368.
- Vala AK (2010) Tolerance and removal of arsenic by a facultative marine fungus Aspergillus candidus. Bioresour Technol 101: 2565 -2567.
- Balaji DS, Basavaraja S, Deshpande R, Bedre D, Prabhakar BK, et al. (2009) Extracellular biosynthesis of functionalized silver nanoparticles by strains of *Cladosporium cladosporioides* fungus. Colloids and Surfaces B: Biointerfaces 68: 88-92.
- Nayak RR, Pradhan N, Behera D (2011) Green synthesis of silver nanoparticle by Penicillium purpurogenum NPMF: the process and optimization. J Nanopart Res 13: 3129-3137.