

Review

Schwann Cell Supplementation in Neurosurgical Procedures after Neurotrauma

Santiago R. Unda*

Instituto de Biotecnología, Centro de Investigación e Innovación Tecnológica, Universidad Nacional de La Rioja, Argentina

*Corresponding author: Santiago R. Unda, Instituto de Biotecnología, Centro de Investigación e Innovación Tecnológica, Universidad Nacional de La Rioja, Argentina, Tel: 3804277348; E-mail: santiagounda94@gmail.com

Rec Date: January 12, 2018, Acc Date: March 13, 2018, Pub Date: March 16, 2018

Copyright: © 2018 Santiago R. Unda. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Nerve trauma is a common cause of quality of life decline, especially in young people. Causing a high impact in personal, psychological and economic issues. The Peripheral Nerve Injury (PNI) with a several grade of axonotmesis and neurotmesis represents a real challenge for neurosurgeons. However, the basic science has greatly contribute to axonal degeneration and regeneration knowledge, making possible to implement in new protocols with molecular and cellular techniques for improve nerve re-growth and to restore motor and sensitive function. The Schwann cell transplantation from different stem cells origins is one of the potential tools for new therapies. In this briefly review is included the recent results of animal and human neurosurgery protocols of Schwann cells transplantation for nerve recovery after a PNI.

Keywords: Schwann cells; Neurotrauma; Peripheral nerve injury; Cell therapy

Introduction

With an annual incidence of approximating 13 to 23 per 100,000 persons per year [1] the Peripheral Nerve Injuries (PNI), have a great impact in the people's quality of life. The expected recovery from this injuries depends on critical factors as the type of injury, the patient's age and the delay before intervention [2]. However, it's well known that after a PNI the Peripheral Nervous System (PNS) has an innate capacity to regenerate but is often insufficient and functional recovery is incomplete [3].

In 1943, Seddon introduced a classification system to describe nerve injury [4] that included neurapraxia, axonotmesis, and neurotmesis [5]. A few years later, Sunderland expanded the axonotmesis grade into three different levels, for a total of five grades [6]. In general, the several axonotmesis injuries and neurotmesis requires complicated microsurgery techniques to restore nerve continuity and functional recovery.

Nerve transfer and nerve grafting are some of most helpful techniques for PNI repair. This procedures have been enhanced with cellular and molecular protocols. The combination of neurosurgery interventions and transplanted Schwann cells improves an auspicious support for peripheral nerve repair and has been investigated in preclinical and clinical studies as a strategy to circumvent the limitations of surgical repair [7].

In this briefly review is included the recent results of animal and human neurosurgery protocols of Schwann cells transplantation for nerve recovery after a PNI.

Sources of Schwann Cells

Schwann cells are embryologically derived from the neural crest [8]. The capacity of Schwann cells for proliferation, growth factor secretion, immune modulation, remyelination and migration make them potential targets in neural repair [9]. Some of the sources of Schwann cells are a population of Neural Crest pluripotent and Stem Cells (NCSCs) in sciatic nerve and Dorsal Root Ganglia (DRG), the NCSCs placed in a specified differentiation medium and the most favorable nanofiber matrix [10] are influenced to convert into a Schwann cell lineage.

Mesenchymal Stromal Cells (MSCs) or bone marrow stromal cells are multipotent somatic stem cells, from a non-hematopoietic precursor found in the bone marrow capable to differentiate into neural cell types including Schwann-like cells [11]. In vitro using an indirect co-culture system, the DRG cells extend their neurites significantly higher when they are expose to differentiate MSCs (dMSCs) compared to undifferentiated MSCs (uMSCs) [12] and in animal models the transplantation of this Schwann-like cells derived from MCSs were able to differentiate into myelinating cells, capable of supporting nerve fibre re-growth and functional recovery after nerve injury [13].

Besides bone marrow stromal cells and NCSCs, skin, adipose, and olfactory tissues provide other potential cell sources [14]. The Skin Precursor Cells SKPCs that can differentiate into neural crest cell types that display similar features to Schwann cells and peripheral neurons [15]. After peripheral and spinal cord injury the SKPCs provided a bridge across the lesion site, increased the size of the spared tissue rim, myelinated spared axons within the tissue rim, reduced reactive gliosis, and an environment that was highly conducive to axonal growth making possible to enhance the locomotor activity [16].

The differentiated adipose-derived Schwann cells expressed a range of intrinsic neurotrophic factors [17]. The addition of this cells to the site of repair promote a neuroprotective effect evidenced by the expression of anti-apoptotic m-RNA of Bcl-2 in the dorsal root ganglia. Other studies showed that adipose-derived Schwann cells enhanced regeneration distance in a similar manner to differentiated bone marrow mesenchymal stem cells without limitations of the donor-site morbidity associated with isolation of Schwann cells [18].

Finally, the induced Pluripotent Stem Cells (iPSCs) since their discovery in 2006 [19] have been propose as a new tool for cell replacement therapy in nerve injury. NCSCs derived from iPSCs can be immune compatible with a potential expansion of these cells, making them a better source for the regeneration of peripheral nerve [20]. However, aberrant differentiation into unwanted lineages, might complicate their therapeutic utility.

In summary the three-principal mechanism of action in stem cell transplantation is represented in Figure 1. Their potential depends basically in differentiate into Schwann-like cells, increase neurotrophic production and myelin formation [21].



Figure 1: Mechanism of stem cell transplantation to enhance a proper nerve restoration. Differentiation of Schwann-like cells, usually the stem cells are maintained in differentiation medium for 2 weeks in most protocols that includes chemical induction, biological treatment, gene transfection, or co-culture with neural cells. Neurotrophic production, secretion of bioactive neurotrophic molecules provides a proper microenvironment for neural cell survival and neurogenesis. Myelin formation, determines the regeneration quality and functional recovery.

Schwann cells transplantation after PNI in Humans

The mechanisms of traumatic nerve degeneration depend on the underlying trauma, but eventually with severe enough injuries Wallerian degeneration ensues [22]. Schwann cells provide bioactive substrates on which axons migrate and have been shown to release factors that regulate axonal outgrowth [23]. Recently, clinical protocols of neurosurgery interventions with Schwann cells transplantation have been applied.

The steps to produce a safety protocol to apply in humans combined a large list of researches that include: the isolation of the SCs from adult, human nerve tissue [24], the ability to induce division and growth of the cells once isolated with the use of particular mitogens (heregulin b1/forskolin), and confirming the safety of these cells and making sure that they did not produce tumorous growths in vivo [25].

In 2017, the first human experience of Schwann cells transplantation for sciatic nerve restore [26]. The study evidence that

autologous Schwann cells may be a viable option for long-gap nerve defect repairs. Both patients respond correctly with an improvement in their motor and sensitive function. This preliminary results shows the potential use of the cell therapy for improve patient's quality of life after neurotrauma. In Figure 2 is represented the schematic protocol for future neurosurgery interventions with microsurgery combined with Schwann cell transplantation.



Figure 2: Schematic representation of autologous Schwann cells transplantation in patients after a peripheral nerve injury. This protocol included a biopsy from sural nerve of about 5cm, a dissociation to get human Schwann cells and expansion with especific mitogens (heregulin b1/forskolin). Finally, transplantation by a microinjection of 29 million autologous Schwann cells within a collagen matrix after a nerve graft.

Future trends and challenges

One of the main challenges in this topic is to minimize the nonfunctional and erroneous connections [27]. A combination of slow axonal regeneration, structural changes in muscle targets, and an increasingly less supportive stromal environment for regeneration all contribute to incomplete functional recovery [28]. So, we need to take some considerations in future protocols for treat patients with acute, severe and long length peripheral nerve injuries. The autologous transplantation of SC is the gold standard, they enhance the regeneration process through their secretion of neurotrophic and tropic cues however, in the clinical practice with certain patients it may not result very convenient for the several weeks that takes to culture a reasonable amount of SC. In the other hand, the allogenic SC transplantation could be culture very fast but this protocols should include immunosuppression treatments. Carrying to possibility to infections and side effects. Moreover, there ethical commitments on this topic haven't been totally discussed.

Another dimension on this topic is in the engineering process, nerve conduits need to be a perfect scaffolds with enough permeability to provide sufficient diffusion of oxygen and metabolites for supporting Schwann cells proliferation but should also prevent fibroblast infiltration [29]. In this context, the main considerations for approach new protocols should consider, an early and appropriate diagnose of nerve injury, including a mechanism of injury and length of nerve defect. If a patient is candidate to a SC transplant, then we should consider which kind of transplant will bring better outcomes, the autologous or the allogenic with the pros and cons mentioned. Furthermore, in future clinical trials are also needed to standardize rehabilitation strategies in order to restore nerve function after injury [30].

Conclusion

The implementation of cell therapy protocols like the Schwann cells or Schwann-like cells transplantation represents a potential tool to improve the quality of life of patients with peripheral nerve injuries after a neurotrauma. However, there's still a need for more clinical trials with different origins of Schwann-like cells and neurosurgery techniques to identify the proper protocols for distinct kinds of nerve injuries and patients.

References

- 1. Taylor CA, Braza D, Rice JB, Dillingham T (2008) The incidence of peripheral nerve injury in extremity trauma. Am J Phys Med Rehabil Assoc Acad Physiatr 87: 381-385.
- Riley DC, Boyer RB, Deister CA, Pollins AC, Cardwell NL (2017) Immediate Enhancement of Nerve Function Using a Novel Axonal Fusion Device After Neurotmesis. Ann Plast Surg 79: 590-599.
- 3. Kim TH, Yoon SJ, Lee WC, Kim JK, Shin J et al. (2011) Protective effect of GCSB-5, an herbal preparation, against peripheral nerve injury in rats. J ethnopharmacol 136: 297-304.
- Sullivan R, Dailey T, Duncan K, Abel N, Borlongan CV (2016). Peripheral Nerve Injury: Stem Cell Therapy and Peripheral Nerve Transfer. Int J Mol Sci 17: 2101.
- 5. Seddon HJ (1943) Three types of nerve injury. Brain 66: 237-288.
- 6. Sunderland S (1951) A classification of peripheral nerve injuries producing loss of function. Brain 74: 491-516.
- Udina E, Rodríguez FJ, Verdú E, Espejo M, Gold BG et al. (2004) FK506 enhances regeneration of axons across long peripheral nerve gaps repaired with collagen guides seeded with allogeneic Schwann cells. Glia 47: 120-129.
- 8. Woodhoo A, Sommer L (2008) Development of the Schwann cell lineage: from the neural crest to the myelinated nerve. Glia 56: 1481-1490.
- Armati PJ, Mathey EK (2013) An update on Schwann cell biology-Immunomodulation, neural regulation and other surprises. J Neurol Sci 333: 68-72.
- Ren YJ, Zhang S, Mi R, Liu Q, Zeng X et al. (2013) Enhanced differentiation of human neural crest stem cells towards the Schwann cell lineage by aligned electrospun fiber matrix. Acta Biomater 9: 7727-7736.
- Wakao S, Hayashi T, Kitada M, Kohama M, Matsue D et al. (2010) Longterm observation of auto-cell transplantation in non-human primate reveals safety and efficiency of bone marrow stromal cell-derived Schwann cells in peripheral nerve regeneration. Exp Neurol 223:537-547.
- 12. Ladak A, Olson J, Tredget EE, Gordon T (2011) Differentiation of mesenchymal stem cells to support peripheral nerve regeneration in a rat model. Exp Neurol 228: 242-252.
- Dezawa M, Takahashi I, Esaki M, Takano M, Sawada H (2001) Sciatic nerve regeneration in rats induced by transplantation of in vitro differentiated bone-marrow stromal cells. Eur J Neurosci 14: 1771-1776.

- Rodrigues MCO, Rodrigues AA, Glover LE, Voltarelli J, Borlongan CV (2012) Peripheral Nerve Repair with Cultured Schwann Cells: Getting Closer to the Clinics. Scientific World J 413091.
- 15. McKenzie IA, Biernaskie J, Toma JG, Midha R, Miller FD (2006) Skinderived precursors generate myelinating Schwann cells for the injured and dysmyelinated nervous system. J Neurosci 26: 6651-6660.
- Biernaskie J, Sparling JS, Liu J, Shannon CP, Plemel JR, et al. (2007) Skinderived precursors generate myelinating Schwann cells that promote remyelination and functional recovery after contusion spinal cord injury. J Neurosci 27: 9545-9559.
- 17. Reid AJ, Sun M, Wiberg M, Downes S, Terenghi G, et al. (2011) Nerve repair with adipose-derived stem cells protects dorsal root ganglia neurons from apoptosis. Neurosci 199: 515-522.
- Di Summa PG, Kingham PJ, Raffoul W, Wiberg M, Terenghi G, et al. (2010) Adipose-derived stem cells enhance peripheral nerve regeneration. J Plast Reconstruct Aesthet Surg 63: 1544-1552.
- 19. Xu W, Cox CS, Li Y (2011) Induced pluripotent stem cells for peripheral nerve regeneration. J Stem Cells 6: 39.
- Wang A, Tang Z, Park IH, Zhu Y, Patel S, et al. (2011) Induced Pluripotent Stem Cells for Neural Tissue Engineering. Biomaterials 32: 5023–5032.
- Jiang L, Jones S, Jia X (2017) Stem Cell Transplantation for Peripheral Nerve Regeneration: Current Options and Opportunities. Int J Mol Sci 18: 94.
- 22. Lehmann HC, Höke A (2015) Use of engineered Schwann cells in peripheral neuropathy: Hopes and hazards. Brain Res 1638: 97-104.
- 23. Fowler JR, Lavasani M, Huard J, Goitz RJ (2015) Biologic strategies to improve nerve regeneration after peripheral nerve repair. J Reconstr Microsurg 31: 243-248.
- Morrissey TK, Kleitman N, Bunge RP (1991) Isolation and functional characterization of Schwann cells derived from adult peripheral nerve. J Neurosci 11: 2433-2442.
- 25. Levi AD, Burks SS, Anderson KD, Dididze M, Khan A et al. (2016) The Use of Autologous Schwann Cells to Supplement Sciatic Nerve Repair With a Large Gap: First in Human Experience. Cell Transplant 25: 1395-1403.
- 26. Gersey ZC, Burks SS, Anderson KD, Dididze M, Khan A et al. (2017) First human experience with autologous Schwann cells to supplement sciatic nerve repair: report of 2 cases with long-term follow-up. Neurosurg Focus 42: E2.
- 27. Hood B, Levene HB, Levi AD (2009) Transplantation of autologous Schwann cells for the repair of segmental peripheral nerve defects. Neurosurg Focus. 26: E4.
- Grinsell D, Keating CP (2014) Peripheral Nerve Reconstruction after Injury: A Review of Clinical and Experimental Therapies. BioMed Res Int 698256.
- 29. Chang C, Hsu S, Yen H, Chang H (2007) Effects of unidirectional permeability in asymmetric poly(DL-lactic acid-co-glycolic acid) conduits on peripheral nerve regeneration: an in vitro and in vivo study. J Biomed Mater Res B Appl Biomater 83: 206-215.
- 30. Pallua N, Suscheck CV (2011) Tissue Engineering: From Lab to Clinic. Berlin: Springer-Verlag.