

## Role of Smad3 and S1P Signaling in Mandibular Condylar Cartilage Homeostasis

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### Abstract

Osteoarthritis (OA), the most common degenerative joint disease, results from an imbalance between chondrocyte-controlled anabolic and catabolic processes. OA is characterized by progressive degradation of components of the Extracellular Matrix (ECM) within the articular cartilage, correlated with secondary inflammation. Several studies had investigated the morphological and biochemical changes during OA progression. However, a comprehensive study of the OA pathogenesis still remains to be elucidated to find the best therapy for OA. In this review, recent advances in our understanding of the mechanisms of action of Sphingosine 1-phosphate (S1P) and Smad3 independently and in relation to Temporomandibular Joint Osteoarthritis (TMJ-OA) will be discussed. S1P receptors are expressed on the cell surface and are internalized upon binding of the bioactive lipid, S1P, as part of the migratory response. Meanwhile, Smad3 is an intracellular signaling molecule that mediates signaling from transforming growth factor- $\beta$  (TGF- $\beta$ ) and activin receptors. Crosstalk between the TGF- $\beta$ /Smad3 and S1P/S1P<sub>3</sub> signaling pathways regulates cell motility and apoptosis in chondrocyte cells. Thus, Smad3/S1P<sub>3</sub> signaling in chondrocytes may be responsible for the development of TMJ-OA, and the potential for these proteins to represent targets for the treatment of TMJ-OA warrants further study.

**Keywords:** Smad3; S1P; Chondrocyte; Migration; TMJ-OA

### Introduction

Mandible, a part of the human masticatory system, through contractions of the neuromuscular controls direction of joint loads as dictated by dental eruption and growth of the Temporomandibular Joint (TMJ) eminence [1,2]. The motion of the mandible is relative to the cranial base and distributes the normal stresses of function (speaking and chewing) and parafunction (bruxism and clenching) [3]. A number of clinical orofacial conditions that involve the masticatory musculature, the TMJ, and associated structures are referred to as Temporomandibular Disorders (TMD). A severe TMD is osteoarthritis (OA) which often affects the TMJ of patients and involves changes in the subchondral bone and progressive cartilage degradation [4].

In the mandibular condyle, endochondral ossification is the primary process by which subchondral bone is formed and this process is regulated by endogenously expressed factors in chondrocyte. Loss of cartilage integrity caused by (bio)mechanical, biochemical, inflammatory, or immunologic in character disturbs the chondrocyte-controlled balance between synthesis and degradation of the ECM components [5,6]. Increased synthesis and activity of proteases, resulting in an initially degradation of articular cartilage [6,7]. During late stage of OA, severe fibrillated and eroded tissue is may appear and neovascularization of TMJ articular cartilage may be present. Denudation of subchondral bone is frequently seen. Synovial membrane may appear hypervascularization and hyperthropic, or fibrotic and disc displacement and perforation may develop [6].

To date, the relationship between subchondral bone abnormalities and the onset of TMJ-OA has not been determined. It is hypothesized

that the accumulation of chondroprogenitor cells at injury sites is due to the migration of these cells from the surrounding matrix [8-12]. Migratory chondroprogenitor cells that are present in cartilage represent a valuable resource for improving cell recruitment into cartilage defects without the need for perforation of the subchondral bone plate. In addition, migratory chondroprogenitor cells have the potential to support the endogenous repair of blunt injured cartilage when traumatic chondrocyte loss has occurred. However, the potential physiologic and/or pathologic functions of chondroprogenitor cells and their migratory effects on healing in TMJ-OA joints remain unknown.

Several studies have reported that a subset of the effects elicited by the TGF- $\beta$ /Smad3 signaling pathway are transmitted via a pathway that is initiated by activation of Sphingosine Kinase (Sphk), followed by intracellular generation of the bioactive lipid, Sphingosine 1-phosphate (S1P) [13]. Here, we highlight that TGF- $\beta$ /Smad3 signaling influences cartilage homeostasis by influencing S1P/S1P receptor signaling and chondrocyte migration.

### Pathogenesis of TMJ-OA

In elderly adults, chronic disability is most often caused by OA. In the early stages of OA, it has recently been demonstrated that low bone mineral density and increased bone turnover are observed in the knee joint [14-16]. Efficacies of bone resorption inhibitors for the rescue of OA have also been reported [15,17,18]. Taken together, these results suggest that abnormal subchondral bone remodeling is important in the pathogenesis of knee OA.

However, the TMJ is one of the most common sites of OA and TMJ-OA may be part of generalized OA [19-21]. TMJ-OA is present in 70% persons of 73-75-year age group and 89% of patients with or without

reduction of disc displacement [20,22]. TMJ-OA can affect all TMJ tissues to induce anatomical changes and severe pain [4].

It is hypothesized that the subchondral bone has an etiological role in TMJ-OA pathology based on recent observations that increased remodeling of mandibular condylar subchondral bone occurs in the early stages of TMJ-OA [23-25]. Moreover, the relationship between the development of TMJ-OA and the abnormalities in subchondral bone remains to be determined, and this is an area of active study.

The key mediators of cartilage degradation *in vivo* and *in vitro* include the Matrix Metalloproteinase (MMPs) and members of the closely related family of a disintegrin and Metalloproteases (ADAMs) with Thrombospondin motifs (ADAM-TS) [26,27]. Roles for matrix MMP-13 and ADAM-TS5 in this degeneration process have been demonstrated [28-33]. Subsequently, key roles have been identified for complement component 5 (C5) [34] and hypoxia-inducible factor-2 $\alpha$  (HIF-2 $\alpha$ ) [35,36]. As OA progresses, articular chondrocytes express interleukin-1 (IL-1), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), runt-related transcription factor 2 (RUNX2), alkaline phosphatase, MMP-13, and type X collagen. Concomitantly, articular cartilage exhibits expanded calcified cartilage zones and low levels of proteoglycans [37-42].

Nitric Oxide (NO) as a free radical play a role in the apoptosis of chondrocytes and inhibits proteoglycan synthesis. These may contribute to the abnormal chondral calcification and osteophyte formation [43,44]. Mitogen-inducible gene-6 (Mig-6), immediate early response gene encoding via threonine kinase receptors, plays an important role in maintaining joint homeostasis. Thus, the involvement of other cells may engage in the pathogenesis of OA [45].

In OA, a primary concern is the degeneration of articular cartilage [33]. The initial and repair stage of OA is characterized biochemically by an increased synthesis of ECM components, DNA, and metabolic activity of the chondrocytes. Accounting for the proliferation, mitoses, and clustering observed histologically [6,42,46]. The repair response is mediated by growth factors (e.g., insulin-like growth factor-1 (IGF-1) and transforming growth factor- $\beta$  (TGF- $\beta$ )), and is partially determined by the diffusibility of these growth factor through the cartilage matrix to the chondrocytes [47-49]. Moreover, growth factors that are normally bound to the ECM components will be released by cartilage degradation and thereby stimulate the chondrocytes in their repair responses. Balance between repair and degradation established, an increased synthesis of the ECM components equals their degradation due to an increased protease activity [6].

To date, the molecular mechanism responsible for mediating the progression from defective subchondral bone to degeneration of articular cartilage in OA remains largely uncharacterized. Acknowledgment of the imbalance anabolic and inflammatory/catabolic pathways has led to explored interest in treatment of OA with limited side effect that may be able to encourage maintenance of bone turnover and chondral homeostasis [39,44].

### The TGF- $\beta$ /Smad3 signaling system and OA

There are three subfamilies of TGF- $\beta$  that closely related to the mammalian isoforms, TGF- $\beta$ 1, - $\beta$ 2, and - $\beta$ 3 [50,51]. Proteins of the TGF- $\beta$  family mediate signaling pathways via serine/ threonine kinase receptors [52]. Specifically, type II serine/threonine kinase receptors (TGF $\beta$ R II) are activated following their binding of type I serine/threonine kinase receptors (TGF $\beta$ R I) [53].

Intracellular Smad proteins (50-70 kDa), particularly Smad2 and Smad3, then transmit this activation signal and that of activin to the nucleus. In addition, when a Smad protein is activated by a receptor, its phosphorylated form is able to heterodimerize with Smad4 and translocate to the nucleus where the complex mediates the transactivation of specific target genes. Meanwhile, Smad1, Smad5, and Smad8 transduce signaling from BMP. Conversely, Smad6 and Smad7 provide an inhibitory function whereby phosphorylation of pathway-specific Smads is inhibited and signal transduction is disrupted [53].

When chondrocyte-specific deletion of Smad3 was achieved in mice, OA in the knee joint was induced [54]. Correspondingly, in humans, mutations in Smad3 have been found in the MH2 domain of Smad3 protein, a region that is extremely well conserved among other species and among other Smad proteins that are associated with early onset of OA [55]. More recently, when overexpression of TGF- $\beta$ 1 was achieved in murine subchondral bone, mandibular condyle degradation was observed [56]. In our own study of mandibular condylar subchondral bone, spontaneous abnormalities were found to induce progressive cartilage degradation in Smad3-/- mice [57].

Chondrocyte death is commonly accepted as a hallmark of OA. It has been observed that the extent of chondrocyte death that occurs positively correlates with the severity of osteoarthritic cartilage depletion and destruction [58,59]. In our recent study, cell death in the condylar cartilage of Smad3-/- mice appeared to be progressive since the numbers of both TUNEL+ and active caspase-3+ and caspase-9+ cells did not significantly differ from those detected in 1-month-old Smad3-/- mice, yet they were markedly higher in the 4-month-old Smad3-/- mice [57].

### S1P/S1PR system

In both healthy and disease states, the bioactive Sphingolipid metabolite, sphingosine 1-phosphate (S1P), contributes to regulating many cellular processes [60]. For example, the ability of S1P to act via a family of cell surface receptors and to play a critical role in the migration of immune cells throughout the body has been well studied. In addition, control of cell trafficking is a well characterized aspect of the involvement of S1P in disease [61]. To date, there are five G-protein-coupled receptors at the cell surface that have been found to be specific for S1P. They include S1PR<sub>1-5</sub>, and activation of S1P<sub>1</sub> is critical for immune cell trafficking [62]. However, in the glomeruli of rats with diabetic nephropathy, it was recently observed that S1P signals are preferentially transmitted through S1P<sub>2</sub>, rather than S1P<sub>1</sub> [63]. It is possible that this biased delivery of S1P signals may mediate the pathogenesis of endothelial injuries in diabetic nephropathy [63]. S1P<sub>2</sub> was also recently shown to be expressed in enteric neurons and migrating cranial crest cells, while expression of S1P<sub>1</sub> is significant in the neuroepithelium [64]. Meanwhile, S1P<sub>4</sub> and S1P<sub>5</sub> are expressed at later stages in neurons [64]. S1P<sub>3</sub> primarily localizes to the cell surface on the plasma membrane, and high expression levels of S1P<sub>3</sub> have been detected in lung, heart, kidney, spleen, diaphragm, and intestine tissues [65]. Moreover, for neurogenesis and for expression of smooth muscle alpha-actin following arterial injury [66], S1P<sub>3</sub> has been found to be essential [67]. S1P<sub>3</sub> also contributes to the migration of thyroid cancer cells [68] and VEGF-A secretion induced by S1P [69]. Furthermore, compared with other S1P receptor subtypes, S1P<sub>3</sub> receptor antagonists have no effect on S1P-induced Mitogen-Activated Protein Kinase (MAPK) activation [70]. Thus, a role for S1P<sub>3</sub> in MAPK signaling has been excluded.

Challenges for regenerative therapy approaches currently include modulation of MMPs, recruitment of chondroprogenitor cells to affected cartilage, and the impact of the extracellular matrix on cell migration [8-10]. Regarding the latter, the regulatory functions of bioactive lysophospholipids, primarily S1P, in cell migration have led to the identification of these proteins as potent mediators of wound healing and tissue repair. Moreover, S1P is released from most cells after they are stimulated by growth factors such as TGF- $\beta$ . Therefore, S1P receptors should be considered in chondrocyte cell migration, despite a role of S1P receptors in OA being largely uncharacterized. Renal mesangial cells express several S1P receptors (e.g., S1P<sub>1-5</sub>), and these receptors potentially mediate mobilization of intracellular calcium, cell proliferation, and activation of the classic MAPK signaling cascade [71]. In the present study, wild type and Smad3<sup>-/-</sup> chondrocyte cells derived from condylar cartilage were analyzed [57]. The former expressed higher levels of S1P<sub>3</sub> compared with the other S1P receptors assayed. Conversely, expression of S1P<sub>3</sub> by the Smad3<sup>-/-</sup> primary chondrocytes was significantly weaker. This difference in S1P<sub>3</sub> expression was further enhanced following stimulation with TGF- $\beta$  [57]. These results are consistent with the observation that signaling via the Sphk1/S1P<sub>3</sub> axis is enhanced during the transdifferentiation of myoblasts into myofibroblasts in response to TGF [72,73]. However, it is important to note that knee hyaline articular cartilage is distinct from mandibular condylar cartilage.

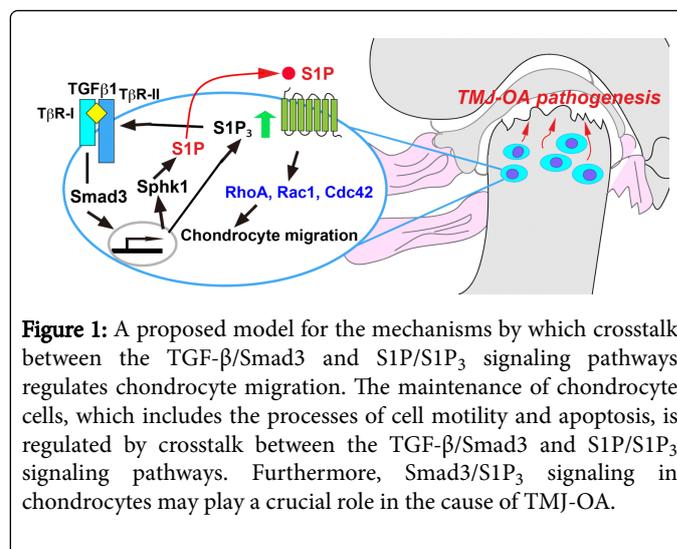
### S1P and TGF- $\beta$ /Smad3 crosstalk in wound healing

It has been observed that TGF- $\beta$  increases Sphk1 activity and up-regulates mRNA and protein levels of Sphk1 in dermal fibroblasts [74]. Thus, it is hypothesized that crosstalk between TGF- $\beta$  and S1P regulates MMP expression. S1P utilizes signaling by its receptors to stimulate phosphorylation and activation of TGF $\beta$ R I kinase, thereby leading to phosphorylation of Smad2 and Smad3 independent of TGF- $\beta$  ligand, as well as an induction of both proliferation and migration in keratinocytes [75]. Abrogation of Smad3 appears to prevent S1P-mediated effects [57,72,75,76], and this suggests a surprising, and yet essential, role for Smad3 in the signaling cascade of the lysophospholipid, S1P. A role for S1P<sub>3</sub> in Smad3 activation was confirmed with the use of small interfering RNA (siRNA) targeting S1P<sub>3</sub> and suramin [57,72]. Correspondingly, suramin was reported to be a selective agonist of the S1P<sub>3</sub> receptor in vitro [77]. Abrogation of S1P-stimulated Smad3 activation by siRNA targeting TGF $\beta$ R II further supports the hypothesis that TGF $\beta$ R II is a component of the S1P signaling cascade [57].

For cell migration, Rho GTPases are critical for coordinating the cellular responses involved [78,79]. In the present study, the Rho GTPases that were assayed exhibited increased levels of activity following stimulation by TGF- $\beta$ . However, when primary chondrocyte cells were transfected with siRNA targeting S1P<sub>3</sub> and then were stimulated with TGF- $\beta$ , the activity levels of GTP-Rac1, GTP-RhoA, and GTP-Cdc42 decreased [57].

### Conclusion

Overall, these findings suggest a model in which chondrocyte cells are maintained via crosstalk between the TGF- $\beta$ /Smad3 and S1P/S1P<sub>3</sub> signaling pathways. The crosstalk between these pathways also regulates cell motility and apoptosis in these cells. Thus, Smad3/S1P<sub>3</sub> signaling in chondrocytes may be responsible for the development of TMJ-OA, and the potential for these proteins to represent targets for the treatment of TMJ-OA warrants further study (Figure 1).



**Figure 1:** A proposed model for the mechanisms by which crosstalk between the TGF- $\beta$ /Smad3 and S1P/S1P<sub>3</sub> signaling pathways regulates chondrocyte migration. The maintenance of chondrocyte cells, which includes the processes of cell motility and apoptosis, is regulated by crosstalk between the TGF- $\beta$ /Smad3 and S1P/S1P<sub>3</sub> signaling pathways. Furthermore, Smad3/S1P<sub>3</sub> signaling in chondrocytes may play a crucial role in the cause of TMJ-OA.

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### Conflicts of Interest

The authors declare no conflict of interest.

### References

1. Nickel JC, McLachlan KR, Smith DM (1988) Eminent development of the postnatal human temporomandibular joint. J Dent Res 67: 896-902.
2. Kuroda S, Tanimoto K, Izawa T, Fujihara S, Koolstra JH, et al. (2009) Biomechanical and biochemical characteristics of the mandibular condylar cartilage. Osteoarthritis Cartilage 17: 1408-1415.
3. Aryaei A, Vapniarsky N, Hu JC, Athanasiou KA (2016) Recent Tissue Engineering Advances for the Treatment of Temporomandibular Joint Disorders. Curr Osteoporos Rep 14: 269-279.
4. Scrivani SJ, Keith DA, Kaban LB (2008) Temporomandibular disorders. N Engl J Med 359: 2693-2705.
5. Maldonado M, Nam J (2013) The role of changes in extracellular matrix of cartilage in the presence of inflammation on the pathology of osteoarthritis. Biomed Res Int 2013: 284873.
6. Dijkgraaf LC, de Bont LG, Boering G, Liem RS (1995) The structure, biochemistry, and metabolism of osteoarthritic cartilage: a review of the literature. J Oral Maxillofac Surg 53: 1182-1192.
7. Mort JS, Billington CJ (2001) Articular cartilage and changes in arthritis: matrix degradation. Arthritis Res 3: 337-341.
8. Gerter R, Kruegel J, Miosge N (2012) New insights into cartilage repair - the role of migratory progenitor cells in osteoarthritis. Matrix Biol 31: 206-213.
9. Khan IM, Williams R, Archer CW (2009) One flew over the progenitor's nest: migratory cells find a home in osteoarthritic cartilage. Cell Stem Cell 4: 282-284.
10. Koelling S, Kruegel J, Irmer M, Path JR, Sadowski B, et al. (2009) Migratory chondrogenic progenitor cells from repair tissue during the later stages of human osteoarthritis. Cell Stem Cell 4: 324-335.

11. Seol D, McCabe DJ, Choe H, Zheng H, Yu Y, et al. (2012) Chondrogenic progenitor cells respond to cartilage injury. *Arthritis Rheum* 64: 3626-3637.
12. Joos H, Wildner A, Hogrefe C, Reichel H, Brenner RE (2013) Interleukin-1 beta and tumor necrosis factor alpha inhibit migration activity of chondrogenic progenitor cells from non-fibrillated osteoarthritic cartilage. *Arthritis Res Ther* 15: R119.
13. Watterson KR, Lanning DA, Diegelmann RF, Spiegel S (2007) Regulation of fibroblast functions by lysophospholipid mediators: potential roles in wound healing. *Wound Repair Regen* 15: 607-616.
14. Bouchgua M, Alexander K, Carmel EN, d'Anjou MA, Beauchamp G, et al. (2009) Use of routine clinical multimodality imaging in a rabbit model of osteoarthritis--part II: bone mineral density assessment. *Osteoarthritis Cartilage* 17: 197-204.
15. Pelletier JP, Boileau C, Brunet J, Boily M, Lajeunesse D, et al. (2004) The inhibition of subchondral bone resorption in the early phase of experimental dog osteoarthritis by licofelone is associated with a reduction in the synthesis of MMP-13 and cathepsin K. *Bone* 34: 527-538.
16. Xie L, Lin AS, Kundu K, Levenston ME, Murthy N, et al. (2012) Quantitative imaging of cartilage and bone morphology, reactive oxygen species, and vascularization in a rodent model of osteoarthritis. *Arthritis Rheum* 64: 1899-1908.
17. Kadri A, Funck-Brentano T, Lin H, Ea HK, Hannouche D, et al. (2010) Inhibition of bone resorption blunts osteoarthritis in mice with high bone remodelling. *Ann Rheum Dis* 69: 1533-1538.
18. Zhu S, Chen K, Lan Y, Zhang N, Jiang R, et al. (2013) Alendronate protects against articular cartilage erosion by inhibiting subchondral bone loss in ovariectomized rats. *Bone* 53: 340-349.
19. Griffin CJ, Powers R, Kruszynski R (1979) The incidence of osteo-arthritis of the temporomandibular joint in various cultures. *Aust Dent J* 24: 94-106.
20. Schmitter M, Essig M, Seneadza V, Balke Z, Schroder J, et al. (2010) Prevalence of clinical and radiographic signs of osteoarthrosis of the temporomandibular joint in an older persons community. *Dentomaxillofac Radiol* 39: 231-234.
21. Abrahamsson AK, Kristensen M, Arvidsson LZ, Kvien TK, Larheim TA, et al. (2017) Frequency of temporomandibular joint osteoarthritis and related symptoms in a hand osteoarthritis cohort. *Osteoarthritis Cartilage* 25: 654-657.
22. Das SK (2013) TMJ osteoarthritis and early diagnosis. *J Oral Biol Craniofac Res* 3: 109-110.
23. Embree M, Ono M, Kilts T, Walker D, Langguth J, et al. (2011) Role of subchondral bone during early-stage experimental TMJ osteoarthritis. *J Dent Res* 90: 1331-1338.
24. Jiao K, Niu LN, Wang MQ, Dai J, Yu SB, et al. (2011) Subchondral bone loss following orthodontically induced cartilage degradation in the mandibular condyles of rats. *Bone* 48: 362-371.
25. Zhang J, Jiao K, Zhang M, Zhou T, Liu XD, et al. (2013) Occlusal effects on longitudinal bone alterations of the temporomandibular joint. *J Dent Res* 92: 253-259.
26. Parks WC, Wilson CL, Lopez-Boado YS (2004) Matrix metalloproteinases as modulators of inflammation and innate immunity. *Nat Rev Immunol* 4: 617-629.
27. Tang BL (2001) ADAMTS: a novel family of extracellular matrix proteases. *Int J Biochem Cell Biol* 33: 33-44.
28. Glasson SS, Askew R, Sheppard B, Carito B, Blanchet T, et al. (2005) Deletion of active ADAMTS5 prevents cartilage degradation in a murine model of osteoarthritis. *Nature* 434: 644-648.
29. Kozaci LD, Buttle DJ, Hollander AP (1997) Degradation of type II collagen, but not proteoglycan, correlates with matrix metalloproteinase activity in cartilage explant cultures. *Arthritis Rheum* 40: 164-174.
30. Porter S, Clark IM, Kevorkian L, Edwards DR (2005) The ADAMTS metalloproteinases. *Biochem J* 386: 15-27.
31. Stanton H, Rogerson FM, East CJ, Golub SB, Lawlor KE, et al. (2005) ADAMTS5 is the major aggrecanase in mouse cartilage in vivo and in vitro. *Nature* 434: 648-652.
32. Tortorella MD, Malfait AM, Deccico C, Arner E (2001) The role of ADAM-TS4 (aggrecanase-1) and ADAM-TS5 (aggrecanase-2) in a model of cartilage degradation. *Osteoarthritis Cartilage* 9: 539-552.
33. Neuhold LA, Killar L, Zhao W, Sung ML, Warner L, et al. (2001) Postnatal expression in hyaline cartilage of constitutively active human collagenase-3 (MMP-13) induces osteoarthritis in mice. *J Clin Invest* 107: 35-44.
34. Wang Q, Rozelle AL, Lepus CM, Scanzello CR, Song JJ, et al. (2011) Identification of a central role for complement in osteoarthritis. *Nat Med* 17: 1674-1679.
35. Saito T, Fukai A, Mabuchi A, Ikeda T, Yano F, et al. (2010) Transcriptional regulation of endochondral ossification by HIF-2alpha during skeletal growth and osteoarthritis development. *Nat Med* 16: 678-686.
36. Yang S, Kim J, Ryu JH, Oh H, Chun CH, et al. (2010) Hypoxia-inducible factor-2alpha is a catabolic regulator of osteoarthritic cartilage destruction. *Nat Med* 16: 687-693.
37. Amano K, Densmore M, Nishimura R, Lanske B (2014) Indian hedgehog signaling regulates transcription and expression of collagen type X via Runx2/Smads interactions. *J Biol Chem* 289: 24898-24910.
38. He Y, Siebuhr AS, Brandt-Hansen NU, Wang J, Su D, et al. (2014) Type X collagen levels are elevated in serum from human osteoarthritis patients and associated with biomarkers of cartilage degradation and inflammation. *BMC Musculoskelet Disord* 15: 309.
39. Zhen G, Wen C, Jia X, Li Y, Crane JL, et al. (2013) Inhibition of TGF-beta signaling in mesenchymal stem cells of subchondral bone attenuates osteoarthritis. *Nat Med* 19: 704-712.
40. Zheng Q, Zhou G, Morello R, Chen Y, Garcia-Rojas X, et al. (2003) Type X collagen gene regulation by Runx2 contributes directly to its hypertrophic chondrocyte-specific expression in vivo. *J Cell Biol* 162: 833-842.
41. Mitchell PG, Magna HA, Reeves LM, Lopresti-Morrow LL, Yocum SA, et al. (1996) Cloning, expression, and type II collagenolytic activity of matrix metalloproteinase-13 from human osteoarthritic cartilage. *J Clin Invest* 97: 761-768.
42. Goldring MB (2000) Osteoarthritis and cartilage: the role of cytokines. *Curr Rheumatol Rep* 2: 459-465.
43. Hashimoto S, Ochs RL, Rosen F, Quach J, McCabe G, et al. (1998) Chondrocyte-derived apoptotic bodies and calcification of articular cartilage. *Proc Natl Acad Sci U S A* 95: 3094-3099.
44. Freitag J, Bates D, Boyd R, Shah K, Barnard A, et al. (2016) Mesenchymal stem cell therapy in the treatment of osteoarthritis: reparative pathways, safety and efficacy - a review. *BMC Musculoskelet Disord* 17: 230.
45. Staal B, Williams BO, Beier F, Vande Woude GF, Zhang YW (2014) Cartilage-specific deletion of Mig-6 results in osteoarthritis-like disorder with excessive articular chondrocyte proliferation. *Proc Natl Acad Sci U S A* 111: 2590-2595.
46. Goldring MB, Otero M, Plumb DA, Dragomir C, Favero M, et al. (2011) Roles of inflammatory and anabolic cytokines in cartilage metabolism: signals and multiple effectors converge upon MMP-13 regulation in osteoarthritis. *Eur Cell Mater* 21: 202-220.
47. Fortier LA, Barker JU, Strauss EJ, McCarrel TM, Cole BJ (2011) The role of growth factors in cartilage repair. *Clin Orthop Relat Res* 469: 2706-2715.
48. Civinini R, Nistri L, Martini C, Redl B, Ristori G, et al. (2013) Growth factors in the treatment of early osteoarthritis. *Clin Cases Miner Bone Metab* 10: 26-29.
49. Wang L, Lazebnik M, Detamore MS (2009) Hyaline cartilage cells outperform mandibular condylar cartilage cells in a TMJ fibrocartilage tissue engineering application. *Osteoarthritis Cartilage* 17: 346-353.
50. Roberts AB, Sporn MB (1988) Transforming growth factor beta. *Adv Cancer Res* 51: 107-145.

51. Cupp AS, Kim G, Skinner MK (1999) Expression and action of transforming growth factor beta (TGFbeta1, TGFbeta2, and TGFbeta3) during embryonic rat testis development. *Biol Reprod* 60: 1304-1313.
52. Massague J (1998) TGF-beta signal transduction. *Annu Rev Biochem* 67: 753-791.
53. Ten Dijke P, Goumans MJ, Itoh F, Itoh S (2002) Regulation of cell proliferation by Smad proteins. *J Cell Physiol* 191: 1-16.
54. Chen CG, Thuillier D, Chin EN, Alliston T (2012) Chondrocyte-intrinsic Smad3 represses Runx2-inducible matrix metalloproteinase 13 expression to maintain articular cartilage and prevent osteoarthritis. *Arthritis Rheum* 64: 3278-3289.
55. van de Laar IM, Oldenburg RA, Pals G, Roos-Hesselink JW, de Graaf BM, et al. (2011) Mutations in SMAD3 cause a syndromic form of aortic aneurysms and dissections with early-onset osteoarthritis. *Nat Genet* 43: 121-126.
56. Jiao K, Zhang M, Niu L, Yu S, Zhen G, et al. (2014) Overexpressed TGF-beta in subchondral bone leads to mandibular condyle degradation. *J Dent Res* 93: 140-147.
57. Mori H, Izawa T, Tanaka E (2015) Smad3 deficiency leads to mandibular condyle degradation via the sphingosine 1-phosphate (S1P)/S1P3 signaling axis. *Am J Pathol* 185: 2742-2756.
58. Jiao K, Wang MQ, Niu LN, Dai J, Yu SB, et al. (2009) Death and proliferation of chondrocytes in the degraded mandibular condylar cartilage of rats induced by experimentally created disordered occlusion. *Apoptosis* 14: 22-30.
59. Thomas CM, Fuller CJ, Whittles CE, Sharif M (2007) Chondrocyte death by apoptosis is associated with cartilage matrix degradation. *Osteoarthritis Cartilage* 15: 27-34.
60. Maceyka M, Harikumar KB, Milstien S, Spiegel S (2012) Sphingosine-1-phosphate signaling and its role in disease. *Trends Cell Biol* 22: 50-60.
61. Spiegel S, Milstien S (2011) The outs and the ins of sphingosine-1-phosphate in immunity. *Nat Rev Immunol* 11: 403-415.
62. Schwab SR, Cyster JG (2007) Finding a way out: lymphocyte egress from lymphoid organs. *Nat Immunol* 8: 1295-1301.
63. Imasawa T, Kitamura H, Ohkawa R, Satoh Y, Miyashita A, et al. (2010) Unbalanced expression of sphingosine 1-phosphate receptors in diabetic nephropathy. *Exp Toxicol Pathol* 62: 53-60.
64. Meng H, Lee VM (2009) Differential expression of sphingosine-1-phosphate receptors 1-5 in the developing nervous system. *Dev Dyn* 238: 487-500.
65. Takabe K, Paugh SW, Milstien S, Spiegel S (2008) "Inside-out" signaling of sphingosine-1-phosphate: therapeutic targets. *Pharmacol Rev* 60: 181-195.
66. Grabski AD, Shimizu T, Deou J, Mahoney WM Jr, Reidy MA, et al. (2009) Sphingosine-1-phosphate receptor-2 regulates expression of smooth muscle alpha-actin after arterial injury. *Arterioscler Thromb Vasc Biol* 29: 1644-1650.
67. Harada J, Foley M, Moskowitz MA, Waeber C (2004) Sphingosine-1-phosphate induces proliferation and morphological changes of neural progenitor cells. *J Neurochem* 88: 1026-1039.
68. Balthasar S, Samulin J, Ahlgren H, Bergelin N, Lundqvist M, et al. (2006) Sphingosine 1-phosphate receptor expression profile and regulation of migration in human thyroid cancer cells. *Biochem J* 398: 547-556.
69. Balthasar S, Bergelin N, Lof C, Vainio M, Andersson S, et al. (2008) Interactions between sphingosine-1-phosphate and vascular endothelial growth factor signalling in ML-1 follicular thyroid carcinoma cells. *Endocr Relat Cancer* 15: 521-534.
70. Xin C, Ren S, Pfeilschifter J, Huwiler A (2004) Heterologous desensitization of the sphingosine-1-phosphate receptors by purinoceptor activation in renal mesangial cells. *Br J Pharmacol* 143: 581-589.
71. Katsuma S, Hada Y, Ueda T, Shiojima S, Hirasawa A, et al. (2002) Signalling mechanisms in sphingosine 1-phosphate-promoted mesangial cell proliferation. *Genes Cells* 7: 1217-1230.
72. Cencetti F, Bernacchioni C, Nincheri P, Donati C, Bruni P (2010) Transforming growth factor-beta1 induces transdifferentiation of myoblasts into myofibroblasts via up-regulation of sphingosine kinase-1/S1P3 axis. *Mol Biol Cell* 21: 1111-1124.
73. Keller CD, Rivera Gil P, Tolle M, van der Giet M, Chun J, et al. (2007) Immunomodulator FTY720 induces myofibroblast differentiation via the lysophospholipid receptor S1P3 and Smad3 signaling. *Am J Pathol* 170: 281-292.
74. Yamanaka M, Shegogue D, Pei H, Bu S, Bielawska A, et al. (2004) Sphingosine kinase 1 (Sphk1) is induced by transforming growth factor-beta and mediates TIMP-1 up-regulation. *J Biol Chem* 279: 53994-54001.
75. Sauer B, Vogler R, von Wenckstern H, Fujii M, Anzano MB, et al. (2004) Involvement of Smad signaling in sphingosine 1-phosphate-mediated biological responses of keratinocytes. *J Biol Chem* 279: 38471-38479.
76. Radeke HH, von Wenckstern H, Stoldtner K, Sauer B, Hammer S, et al. (2005) Overlapping signaling pathways of sphingosine 1-phosphate and TGF-beta in the murine Langerhans cell line XS52. *J Immunol* (Baltimore, Md. : 1950) 174: 2778-2786.
77. Ancellin N, Hla T (1999) Differential pharmacological properties and signal transduction of the sphingosine 1-phosphate receptors EDG-1, EDG-3, and EDG-5. *J Biol Chem* 274: 18997-19002.
78. Raftopoulou M, Hall A (2004) Cell migration: Rho GTPases lead the way. *Dev Biol* 265: 23-32.
79. Ridley AJ (2001) Rho GTPases and cell migration. *J Cell Sci* 114: 2713-2722.