

**Review Article** 

Gupta, Immunome Res 2013, 9:1 DOI: 10.4172/1745-7580.1000067

# Role of Dendritic Cells in Autoimmunity in Aged Humans

# Sudhir Gupta\*

Programs in Primary Immunodeficiency and Aging, Division of Basic and Clinical Immunology, University of California, Irvine, California, USA

#### Abstract

Immunological tolerance to self-antigen is mediated by deletion of self-reactive lymphocytes by apoptosis, unresponsiveness to self-antigens (anergy), regulation by T cells (Treg), and efficient removal of apoptotic bodies by phagocytic cells. Dendritic cells (DCs) play an important role in both tolerance (predominantly via induction of Treg) and inthe induction of immune response to non-self antigens. Therefore, an impairment of DCs functions may result in the loss of tolerance and induction of immune response to self-antigens, resulting in autoimmunity. Aging represents a paradox of impaired response to non-self antigens, and an increased response to self-antigens, resulting in an increased susceptibility to infections, and development of autoimmunity in aged humans. I have reviewed the role of DCs and mechanisms involved in autoimmunity, including epigenetic changes in aged DNA, and histone modifications in chromatin in human aging.

**Keywords:** Epigenetic changes; IFN-I; IFN-III; mDC; pDC; Chromatin modification; Cytokines; Treg

## Introduction

Aging represents a unique paradox of autoimmunity and impaired response to exogenous antigens resulting in an increased susceptibility to infections. This is of particular significance since patients with autoimmune diseases are not at risk for increased susceptibility to infections, unless on immunosuppressive/biological therapy. Aged humans develop a variety of autoantibodies, and display poor response to vaccines and increased susceptibility to both viral and bacterial infections. The progressive impairment in immune functions include progressive T cell deficiency, which is contributed by thymic involution [1], increased apoptosis of T cells and T cell subsets [2-4], and impaired priming of T cells by DCs [5-7]; B cell dysfunctions are revealed by impaired specific antibody response to vaccines, and development of autoimmunity [8-12].

Dendritic cells are a heterogeneous population of hematopoietic antigen-presenting cells. They play a major role in initiating and shaping both innate and adaptive immune responses, and in the maintenance of immunological tolerance [13-20]. Recent studies in humans and experimental models suggest that DCs are involved in the pathogenesis of autoimmune diseases. There are two major subpopulations of DCs, namely "conventional" (cDCs), which are also known as myeloid DCs, and plasmacytoid DCs (pDCs). The cDCs are known to differentiate from the common myeloid hematopoietic precursors. It appears there is a significant plasticity in the DC lineage. pDCs could be derived from either myeloid or lymphoid precursors [21]. Furthermore, pDCs and cDCs maintain plasticity even after their differentiation [22]. Therefore, a numbers of functions are shared between two subtypes of DCs; for example, production of IFN-I and IFN-III, albeit their relative concentrations may be different among two DC subtypes, and priming of naïve T cells. The cDCs exist in peripheral tissues, secondary lymphoid organs, and in the circulating blood. pDCs circulate in the blood and enter lymphoid organs through high endothelial venules. Compare to cDCs, pDCs express different sets of Toll like receptors (TLRs) [23]. pDCs express TLR7 and TLR9 and upon stimulation secrete large amounts of IFN-a [24]. In response to microbial infection, monocytes migrate into inflammatory sites and differentiate into DCs [24]. In humans, in vitro activation of monocytes with GM-CSF and IL-4 induces differentiation of monocytes into monocyte-derived DCs, which serves as a model for cDCs. In this review I will refer them as mDCs.

# The Role of DCs in Immune Tolerance and Autoimmunity

Knight and colleague [25] provided initial evidence for the role of DCs in autoimmunity. They demonstrated that DCs from animals with EAE could transfer the disease in naïve recipients.

DCs play a role in both central and peripheral tolerance. In the central tolerance, thymic DCs have shown to cross present selfantigens, which have been acquired from medullary thymic epithelial cells (mTECs) [26,27]. In addition, thymic DCs may also facilitate generation of natural Treg (nTreg) [28]. There is also an evidence that peripheral DCs might migrate into the thymus, and present peripheral self-antigens to induce clonal deletion or to generate Treg [29,30]. In general, DCs appear to play only a minor role in central tolerance; mTECs appear to play the major role in inducing central tolerance.

DCs appear to play a role in peripheral tolerance by supporting the homeostasis of peripheral Treg cells. DCs can polarize naïve CD4+ T cells to Treg cells (iTreg) in the presence of TGF- $\beta$  [31,32]. Recently it has been suggested that migratory cDCs and not the resident DCs in the lymph nodes induce the development of Treg specific to a particular self-antigen [33]. It appears that DCs facilitate the induction and/or maintenance of Treg cells; however; they are not indispensible for both the induction and maintenance of Treg. This is further supported by lack of significant autoimmunity in patients with primary immunodeficiency of DCs [34]. In peripheral tolerance, nTreg generated in the thymus by mTECs may more important than iTreg generated by DCs in the periphery. In general, DCs may induce tolerance by inducing Treg and induction of energy.

A role of DCs have been demonstrated in a number of autoimmune

\*Corresponding author: Sudhir Gupta, MD, Ph.D., Medical Sciences I, C-240, University of California, Irvine, Irvine, CA 92697, USA, Tel: (949) 824-5818; Fax: (949) 824-4362; E-mail: squpta@uci.edu

Received September 24, 2013; Accepted October 21, 2013; Published October 26, 2013

Citation: Gupta S (2013) Role of Dendritic Cells in Autoimmunity in Aged Humans. Immunome Res 9: 067. doi: 10.4172/1745-7580.1000067

**Copyright:** © 2013 Gupta S. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

diseases and animal models of autoimmune diseases, including systemic lupus erythematosus (SLE), multiple sclerosis (MS), experimental autoimmune encephalitis (EAE), and autoimmune myocarditis [35].

### Role of Apoptosis in Tolerance and Autoimmunity

Apoptosis (programmed cell death) also plays an important role in maintaining tolerance and immune homeostasis [2]. Almost every cell is programmed and equipped to die only to be replaced by new cell to main homeostasis. These apoptotic cells contains a number of selfantigens, and therefore, must be removed by neighboring phagocytic cells and self-antigens degraded to avoid their uptake by DCs and presentation to self-reactive lymphocytes. Apoptosis is induced by death receptor pathway, mitochondrial pathway, and endoplasmic reticulum stress pathway [2,36-38]. Apoptotic cells express a number of surface antigens to provide a 'eat me' signals, which are recognized by receptors on phagocytic cells. Apoptosis also provide a major mechanism for deletion of self-reactive T cells (central tolerance). Therefore, disorders of apoptosis are associated with autoimmunity and autoimmune diseases [39]. An impaired apoptosis, resulting in failure to remove self-reactive lymphocytes, and increased apoptosis with excessive load for the phagocytic cells, and defects in uptake of phagocytic cells may lead to late necrosis of apoptotic blebs resulting in increased release and therefore, exposure of self-antigens to DCs resulting in autoimmunity and autoimmune diseases.

## Role of Interferons in Autoimmunity

Secretion of interferons (IFNs) from virus-infected cells is a hallmark of host antiviral immunity. In addition, interferons modulate both innate and adaptive immune responses. DCs are major producers of both IFN-I (IFN- $\alpha$ , IFN- $\beta$ , IFN- $\kappa$ , IFN- $\omega$ ) and IFN-III [IFN- $\lambda$ 1 (IL-29),  $\lambda$ 2 (IL-28a),  $\lambda$ 3 (IL-28b)]. mDCs produce greater amount of IFN-III, whereas pDCs are major source of IFN-I [13,14,17,40,41].

IFN-I promote the differentiation of naïve CD4+ T cells to TH1 cells, and survival of activated T cells, and enhance the cytotoxicity of CD8+ T cells and NK Cells [42-44]. In addition, IFN-I enhance antibody production, and induces isotype class switch in B cells [45-47]. Furthermore, IFN-I enhances antigen cross presentation and maturation by DCs by up-regulating co-stimulatory molecules and TLRs, and inducing secretion of pro-inflammatory cytokines [48,49]. IFN-I and IFN-I regulated genes are increased in systemic [50,51], and organ-specific autoimmune diseases [52], suggesting its role in autoimmunity. Autoantigens from apoptotic cells stimulate pDCs to prduce large amounts of IFN-I, which induces B cell differentiation and Ig class switch, including autoreactive B cells resulting in autoantibody production.

IFN-III (IFN- $\lambda$ s) is family of interferons encoded by 3 different genes that were discovered in 2003 [53,54]. They play a major role in viral defense; however, provides much better protection at mucosal surfaces than IFN-I [40,55].

Though IFN-I and IFN-III signal through different receptors, they share a common downstream signaling pathway, a common set of interferon-stimulated genes (ISGs), and share many biological properties, including anti-viral and anti-proliferative activities [55,56].

Since a role of IFN-I in autoimmunity and autoimmune diseases has been demonstrated [50-52], it is expected that IFN-III may also play a role in autoimmunity and autoimmune diseases. Increased serum IFN- $\lambda$ 1 levels have been observed in patients with rheumatoid arthritis as compared to healthy controls and patients with osteoarthritis [57].

Mannechet et al. [59] have shown that *in vitro*, IFN- $\lambda$ -treated DCs induce proliferation of FoxP3-expressing regulatory T cells. More recently, Rynda et al. [60] have demonstrated that endogenous IFN-III (IL-28) protects against EAE in the absence of Treg cells, and treatment of animals with neutralizing antibodies against IL-28 render mice susceptible to EAE, suggesting a role of IFN-III in tolerance.

#### Autoimmunity in Aging

#### Role of apoptosis in autoimmunity in aging

In contrast to progressive decline in immune functions with advancing age, there is an increased reactivity to self and endogenous antigens as evidenced by the presence and increased titers of a variety of autoantibodies [8-12], which suggest a loss of peripheral tolerance in aging. The information regarding mechanisms of impaired tolerance in human aging is limited. Apoptosis plays an important role in the effector functions and immune homeostasis. One of the critical steps in apoptosis is a rapid uptake of apoptotic cells and apoptotic bodies by neighboring phagocytic cells, resulting in intracellular degradation of self-antigens, and induction of anti-inflammatory response and generation of Treg. We have shown that in aging, apoptosis of T cells and T cell subsets is increased [2-4,61,62], whereas DCs are impaired in their capacity to uptake apoptotic cells [63]. As a consequence apoptotic cells would undergo secondary necrosis with additional proteolytic degradation of specific autoantigens, leading to the release endogenous danger signals like nuclear antigens clustered in apoptotic blebs and bodies (e.g. chromatic, DNA, RNA, histones etc), resulting in maturation of DCs, presentation of self-antigens to lymphocytes and induction of T cell immunity to self antigens, and stimulation of autoreactive B cells and production of autoantibodies.

# Role of DCs in Autoimmunity in Aging

#### Increased reactivity of aged DCs to self-DNA

DCs are unique antigen-presenting cells because of their capacity to prime naïve T cells [14,16-18]. DCs therefore function as initiators of T cell immunity. DCs can prime or tolarize T cells. Under physiological conditions, DCs play a role in unresponsiveness to self-antigens. DCs are essential for both central and peripheral tolerance. Impaired clearance of apoptotic cells has been implicated in autoimmune diseases like lupus [64,65]. Therefore, we examined the priming capacity of young and aged DCs to self-DNA. DNA was purified from young humans and delivered intracellularly to mDCs from young and aged subjects and examined for the activation and cytokine production by mDCs and their capacity to induce T cell proliferation [66]. DNA-primed mDCs from aged subjects upregulated co-stimulatory molecules, and secreted increase levels of IL-6 and IFN-α as compared to young mDCs. Similar increased in cytokine secretion was observed by aged mDCs in response to late apoptotic cells. Furthermore, young DNA-primed aged mDCs induced autologous T cell proliferation, whereas young DNA-primed young mDCs did not induce T cell proliferation, suggesting a role of DCs in increased reactivity to self-DNA and a loss of tolerance in aged humans. This increased reactivity to DNA is independent of TLR-9. Furthermore, expression of the cytosolic DNA sensor DAM1 was comparable between young and aged, suggesting steps downstream of cytosolic sensor may be involved in self-reactivity to DNA by aged DCs. Since there are several DNA censors, a possibility of involvement of one of the other DNA sensors cannot be excluded. One of the steps

downstream of DNA censors is the interferon-responsive factor-3 (IRF-3). We observed an increased activation of IRF-3 transcription factor in mDCs from aged in response to Intracellular self-DNA [61]. Furthermore, mDCs from aged display higher basal levels of NF-kB activation, suggesting that DCs from aged are in an activated state. Panda et al. [67] also observed increased basal levels of cytokines in aged DCs.

# Role of Epigenetic modifications in aging DNA in autoimmunity

Epigenetic regulation of gene expression occurs via chemical modification such as histone acetylation and methylation, without alteration in the nucleotide sequence in the genome [68]. Human DNA undergoes age-associated genetic and epigenetic changes [69,70]. During aging, cells and tissues become hypomethylated while selected genes become progressively hypermethylated [71]. There is a relationship between genomic instability, DNA damage, and DNA repair mechanisms, which are in aging resulting in DNA lesions with single and double-stranded breaks [72]. Furthermore, oxidative damage to DNA has been implicated in aging and age-related degenerative disorders [73].

Human DNA is generally inert and does not stimulate DCs. We demonstrated that DNA from aged mononuclear cells when introduced into young mDCs resulted in upregulation of co-stimulatory molecules CD80 and CD86, and increased secretion of IFN- $\alpha$ , as compared to young DNA, suggesting an increased immunogenicity of aged DNA [74]. We also showed that DNA from aged subjects is hypomethylated, and when aged DNA was hypermethylated comparable to methylation of young DNA, aged DCs could no longer induced increased secretion of IFN-a, demonstrating that immunogenicity of mammalian DNA correlates inversely with DNA methylation. Finally, we observed that intracellular delivery of oxidative-damaged DNA did not result in the activation of mDCs, which suggest that DNA damage per se does not increase immunogenicity of aged DNA, and hypomethylation of DNA is responsible for its increased immunogenicity in aging. It remains to be determined which site-specific hypomethylation confer increased immunogenicity to self-DNA.

#### Interferons in aging

The role of interfrons in defense against viruses is well established. IFN-I are known to have anti-proliferative and antitumor activities [44,49,56]. IFN-III also display anti-viral activity; however, predominantly at mucosal surfaces because of more restricted expression of IFN-III receptors as compared to IFN-I receptors, which are more widely expressed [75,76]. IFN-II has predominantly immunoregulatory role. A role of IFN-Ia in autoimmunity and autoimmunity is well documented. IFN-Ia expression is increased in autoimmune diseases [40,50-52] and IFN-Ia treatment has been associated with exacerbation or development of certain autoimmune diseases [77-79]. This is in contrast to use of IFN- $\beta$  in the treatment of multiple sclerosis, an autoimmune disease. A role of IFN-III in immune regulation and autoimmunity has not yet established; however, abnormal and dysregulated expression of IFN-III and IFN-III receptors in certain autoimmune diseases, including rheumatoid arthritis and systemic lupus erythematosus has been reported [57,58].

Because of increased susceptibility of aged humans to viral infection, especially respiratory tract infections, and development of autoimmunity, others and we have investigated the production of IFN-I and IFN-III by DCs from healthy young and aged subjects. pDC from aged are impaired in secreting both IFN-I and IFN-III in response to Influenza virus (TLR signal) and CpG (TLR9 signaling) [7,67,80-83]. However, the expression of TLR7 and TLR9 on aged pDCs is comparable to young [7]. Furthermore, we demonstrated that the protein expression of downstream signaling molecules IRAK-1, Myd88, and IRF-7 in aging pDCs is comparable to young; however, CpG and influenza virus-induced IRF-7 phosphorylation in aged pDCs is impaired [7]. pDCs from aged are also impaired in priming CD8+ T cells as determined by proliferation, perforin and granzyme production, and IFN- $\gamma$  secretion [7]. pDCs are also impaired in priming of CD4+ T cells as determined by CD4+ T cell proliferation.

We also examined a role of mDCs in responses to influenza virus. The phenotype of aged and young mDcs was comparable following activation with heat-inactivated influenza virus. However, mDCs from aged were impaired in their ability to produce Influenza virusinduced both IFN-I and IFN-III. Panda et al. [67] also demonstrated decreased IFN-I production in aged mDCs. We examine whether histone modification (epigenetic changes) play a role in impaired interferon secretion by mDCs in aged humans [84]. Association of IFN-A2 and IFN-l1 (IL-29) promoter to H3K4me3 and H3K9me3 is altered in aged DCs. This impaired association was specific to IFN-A2 and IFN-l1 since no such impairment of association was observed between histones and TNF- $\alpha$  promoter, and no association of IFN-2A, and IL-29 (IFN- $\lambda$ ) non-promoter to H3K4me3 and H3K9me3 was observed. Additionally association of IFN-A2, IFN-l, TNF-a promoter to H3K4me and HeK9me in aged mDCs is unstable. Both IFN-α and IFN-β have shown to mediate their effects though a common IFNAR1 and IFNAR2 [85,86]. However, recently it has been reported that IFN-β may not use IFNR2 and uses a different partner with IFNAR1 [85]. The expression of IFNAR1/2 and response to IFN- $\beta$  remains to be studied.

Since functions of mDCs and pDCs and production of both IFN-I and IFN-III are impaired, it is unlikely that altered IFN production in aging plays an significant role in autoimmunity in aging; perhaps impaired uptake of apoptotic bodies, and increased reactivity to selfantigen and production of pro-inflammatory cytokines by aged DCs may predominantly contribute to autoimmunity associated with aging. An impaired interferon production may be responsible for increased susceptibility to viral infections in aged humans.

#### References

- 1. Rezzani R, Nardo L, Favero, G, Peroni M, Rodella LF (2013) Thymus and aging: morphological, radiological, and functional overview. Age (Dordr).
- Gupta S (2000) Molecular steps of cell suicide: An insight into immune senescence. J Clin Immunol 20: 229-239.
- Gupta S (2005) Molecular mechanisms of apoptosis in the cells of the immune system in human aging. Immunol Rev 205: 114-129.
- Gupta S, Agrawal S, Agrawal A, Su H, Gollapudi S (2006) A paradox of Immunodeficiency and inflammation: Lessons learned from apoptosis. Immun Ageing 5: 1-8.
- Yu J, Dong H, Mann ER, Knight SC, Yaqoob P (2013) Ageing impairs the T cell response to dendritic cells Immunobiology 218: 1077-1084.
- Gupta S (2013) Role of dendritic cells in innate and adaptive immune responses in aged humans. Exp Gerontol, In press.
- Sridharan A, Esposo M, Kaushal K, Tay J, Osann K, et al. (2010) Ageassociated impaired plasmacytoid dendritic cell functions lead to decreased CD4 and CD8 T cell immunity. AGE 33: 363-376.
- Rowley MJ, Buchanan H, Mackay IR (1968) Reciprocal change with age in antibody to extrinsic and intrinsic antigens. Lancet 2: 24-26.
- 9. Manavalan JS, Kirman I, Zhao K, Weksler ME (1998) Aging and autoimmunity

In: Rose NR, Mackay IR, editors The autoimmune diseases. Academic Press, San Diego, 783–94.

- 10. Tomer Y, Shoenfeld Y (1988) Aging and autoantibodies. Autoimmunity 1: 145-149.
- Weksler ME, Goodhardt M (2002) Do age-associated changes in 'physiologic' autoantibodies contribute to infection, atherosclerosis, and Alzheimer's disease? Exp Gerontol 37: 971-979.
- Franceschi C, Monti D, Sansoni P, Cossarizza A (1995) The immunology of exceptional individuals The lessons of Centenarians Immunology Today 16: 12-16.
- 13. Merad M, Manz MG (2009) Dendritic cell homeostasis. Blood 113: 3418-3427.
- Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, et al. (2000) Immunobiology of dendritic cells. Ann Rev Immunol 18: 767-811.
- Gallo PM, Gallucci S (2013) The dendritic cell response to classic, emerging, and homeostatic danger signals Implications for autoimmunity. Front Immunol 4:138.
- Hubo M, Trinschek B, Kryczanowsky F, Tuettenberg A, Steinbrink K, et al. (2013) Costimulatory molecules on immunogenic versus tolerogenic human dendritic cells. Frontiers in Immuno I 4: 1-14.
- 17. Collin M, McGovern N, Haniffa M (2013) Human dendritic cell subsets. Immunology 29.
- Ueno H, Schmitt N, Klechevsky E, Pedroza-Goanzales A, Matsui T, et al. (2010) Harnessing human dendritic cell subsets for medicine. Immunol Rev 234: 199-212.
- Steinman RM (2012) Decision about dendritic cells: past, present, and future. Ann Rev 30: 1-22.
- Lewis KL, Reizis B (2012) Dendritic cells: arbiters of immunity and immunological tolerance. Cold Spring Harb Perspect Biol 4: a007401.
- Shigemetssu H, Reizis B, Iwasaki H, Mizuno S, Hu D, et al. (2004) Plasmacytoid dendritic cells activate lymphoid-specific genetic programs irrespective of their cellular origin. Immunity 21: 43-53.
- Diebold SS, Montoya M, Unger H, Alexopoulos L, Roy P, et al. (2003) Viral infection switches non-plasmacytoid dendritic cells into high interferon producers. Nature 424: 324-328.
- Akira S, Uematsu S, Takeuchi O (2006) Pathogen recognition and innate immunity. Cell 124: 783-801.
- 24. Siegal FP, Kadowaki N, Shodell M, Fitzgerald-Bocarsly PA, Shah et al. (1999) The nature of principal type 1 interferon-producing cells in human blood. Science 284: 1835-1837.
- Knight SC, Mertin J, Stackpoole A, Clark J (1983) Induction of immune responses in vivo with small numbers of veiled (dendritic) cells. Proc Natl Acad Sci 80: 6032-6035.
- Gallegos AM, Beavan MJ (2004) Central tolerance to tissue-specific antigens mediated by direct and indirect antigen presentation. J Exp Med 200: 1039-1049.
- 27. Hubert FX (2011) AIRE regulates the transfer of antigens from mTECs to dendritic cells for induction of thymic intolerance. Blood 118: 2462-2472.
- Lei Y, Ripen AM, Ishimaru N, Ohigashi I, Nagasawa T, et al. (2011) Airedependent production of XCL1 mediates medullary accumulation of thymic dendritic cells and contributes to regulatory T cell development. J Exp Med 208: 383-394.
- Bonasio R, Scimone ML, Schaerli P, Grabie N, Lichtman AH, et al. (2006) Clonal deletion of thymocytes by circulating dendritic cells homing to the thymus. Nature Immunol 7: 1092-1100.
- Proietto A, van Dommelen S, Zhou P, Rizzitelli A, D'Amico A, et al. (2008) Dendritic cells in the thymus contribute to T regulatory cell induction. Proc Natl Acad Sci 105: 19869-19874.
- Yamazaki S, Iyoda T, Tarbell K, Olson K, Velinzon K, et al. (2003) Direct expansion functional CD25+ CD4+ regulatory T cells by antigen-processing dendritic cells. J Exp Med 198: 235-247.
- Sela U, Olds P, Park A, Schlesinger SJ, Steinman RM (2011) Dendritic cells induce antigen-specific regulatory T cells that prevent graft versus host disease and persist in mice. J Exp Med 208: 2489-2496.
- 33. Vitali C, Mingozzi F, Broggi A, Barresi S, Zolezzi F, et al. (2012) Migratory,

and not lymphoid resident dendritic cells maintain peripheral self tolerance and prevent autoimmunity via induction of iTreg cells. Blood 120: 1237-1245.

- Collin M, Bigley V, Haniffa M, Hambleton S (2011) Human dendritic cell deficiency: the missing ID. Nat Rev Immunol 11: 575-583.
- Ganguly D, Haak S, Sisirak V, Reizis B (2013) The role of dendritic cells in autoimmunity. Nature Rev Immunol 13: 566-577.
- Wilson NS, Dixit VM, Ashkanazi A (2009) Death receptor signal transducers: Nodes of coordination in immune signaling networks Nature Immunol 10: 348-355.
- Martinou JC, Green DR (2001) Breaking the mitochondrial barrier. Nat Rev Mol Cell Biol 2: 63-67.
- Orrenius S, zhivotocsky B, Nicotera N (2003) Regulation of cell death: the calcium apoptosis link. Nat Rev Mol Cell Biol 4: 552-565.
- Oliveira JB, Gupta S (2008) Disorders of apoptosis: mechanisms for autoimmunity in primary immunodeficiency diseases. J Clin Immunol 28: S20-S28.
- Ank N, Paludan SR (2009) Type III IFNs: New layer of complexity in innate antiviral immunity. Biofactors 35: 82-87.
- Fitzgerald-Bocarsly P, Dai J, Singh S (2008) Plasmacytoid dendritic cells and Type I IFN: 50 years of convergent history. Cytokine Growth Factor Rev 19: 3-19.
- Rogge D, Biffi M, Penna G, Minetti LJ, Preski DH, et al. (1998) The role of Stat4 in species specific regulation of Th development by type I IFNs. J Immunol 161: 6567-6574.
- Marrack P, Kappler J, Mitchell T (1999) Type I interferons keep activated T cells alive. J Exp Med 189: 521-530.
- 44. de Weerd NA, Samarajiwa SA, Hertzog PJ (2007) Type I interferon receptors: biochemistry and biological functions. J Biol Chem 282: 20053-20057.
- Braun D, Caramalho I, Damengeot J (2000) IFN-alpha/beta enhances BCRdependent B cell responses. Int Immunol 14: 411-419.
- 46. Jego G, Paliucka AK, Blanck JP, Chalouni C, Pascual V, et al. (2003) Plasmacytoid dendritic cells induced plasma cell differentiation through type I interferon and interleukin-6. Immunity 19: 225-234.
- 47. Le Bon A, Thompson C, Kamphuis E, Durand V, Rossman C et al. (2006) Enhancement of antibody responses through direct stimulation of B and T cells by type I IFN. J Immunol 176, 2074-2078.
- Beignon AS, Skoberne M, Bhardwaj N (2003) Type I interferons promote crosspriming: more functions for old cytokines. Nat Immunol 4: 939-941.
- Tough DF (2004) Type I interferon as a link between innate and adaptive immunity through dendritic cell stimulation. Leuk Lymphoma 45: 257-264.
- Kirou KA, Lee C, George S, Louca K, Papagiannis IG, et al. (2004) Coordinate overexpression of interferon-alpha-induced genes in systemic lupus erythematosus. Arthritis Rheum 50: 3958-3967.
- Gottenberg JE, Cagnard N, Lucchesi C, Letourneur F, Mistou S, et al. (2006) Activation of IFN pathways and plasmacytoid dendritic cell recruitment in target organs of primary Sjögren's syndrome. Proc Natl Acad Sci (USA) 103: 2770-2775.
- Mavragni CP, Niewwold TB, Chatzigeorgiou A, Danielides S, Thomas D, et al. (2013) Increased serum type I interferon activity in organ-specific autoimmune disorders: clinical, imaging, and serological associations. Front Immunol 4: 1-9.
- Sheppard P, Kindsvogel W, Xu W, Henderson K, Schlutsmeyer S, et al. (2003) IL-28, IL-29 and their class II cytokine receptor IL-28R. Nat Immunol 4: 63-68.
- Kotenk SV, Gallagher G, Baurin VV, Lewis-Antes A, Shen M, et al. (2003) IFN-lambdas mediate antiviral protection through a distinct class II cytokines receptor complex. Nat Immunol 4: 69-77.
- Donnelly RP, Kotenko SV (2010) Interferon-lambda: A new addition to an old family. J Interfer Cytok Res 30: 55-564.
- George PM, Badiger R, Alazawi T, Foster GR, Michell JA (2012) Pharmacology and therapeutic potential of interferons. Pharmacol Ther135: 44-53.
- 57. Wu Q, Yang Q, Sun H, Li M, Zhang Y, et al. (2013) Seurm IFN-λ1 is abnormally elevated in rheumatoid arthritis patients. Autoimmunity 46: 40-43.
- 58. Lin SC, Kuo CC, Tsao JT, Lin LJ (2012) Profiling the expression of interleukin (IL)-28 and IL-28 receptor α in systemic lupus erythematosus patients. Eur J Clin Invest 42: 61-69.

- Mennechet FJ, Uze G (2006) Interferom lambda treated dendritic cells specifically induce proliferation of FoxP3-expressing suppressor T cells. Blood 107: 4417-4423.
- Rynda A, Maddaloni M, Ochoa-Reparaz J, Callis G, Pascual DW (2013) IL-28 supplants requirement of Treg cells in protein 1-mediated protection against murine experimental autoimmune encephalitis (EAE). PLOS One.
- Aggarwal S, Gupta S (1998) Increased apoptosis of T cell subsets in aging humans: Altered expression of Fas (CD95), Fas ligand, Bcl-2, and Bax. J Immunol 160: 1627-1637.
- 62. Aggarwal S, Gollapudi S, Gupta S (1999) Increased TNF-α-induced apoptosis in lymphocytes from aged humans: changes in TNF-α receptor expression and activation of caspases. J Immunol 162, 2154-2161.
- Agrawal A, Agrawal S, Cao JN, Su H, Osann K, et al. (2007) Altered innate immune functioning of dendritic cells in aging humans: Role of PI3Kinase signaling pathway. J Immunol 178, 6912-6922.
- 64. Hardin JA (2003) Directing autoimmunity to nucleoprotein particles, the impact of dendritic cells and interferon α in lupus. J Exp Med 197: 681-687.
- 65. Shoshan Y, Mevorach D (2004) Accelerated autoimmune disease in MRL/Mpj-Faslpr but not in MRL/Mpj following immunization with high load of syngeneic late apoptotic cells. Autoimmunity 37: 103-109.
- Agrawal A, Tay J, Ton S, Agrawal S, Gupta S (2009) Increased reactivity of dendritic cells from aged subjects to self antigen, the human DNA. J Immunol 182: 1138-1145.
- Panda A, Qian F, Mohanty S, van Duin D, Newman FK, et al. (2010) Ageassociated decrease in TLR function in primary human dendritic cells predicts influenza vaccine response. J Immunol 184: 2518-2527.
- Bernstein E, Meissner A, Lender ES (2007) The mammalian epigenome. Cell 128, 669-681.
- Chen JH, Hales CN, Ozanne SE (2009) DNA damage, cellular senescence, and organismal ageing: causal or correlative? Nucleic Acid Res 35: 7417-7428.
- 70. Richardson BC (2002) Role of DNA methylation in the regulation of cell function: Autoimmunity, aging, and cancer. J Nutr132: 2401S-2405S.
- Issa JP (2003) Age-related epigenetic changes and the immune system. Clin Immunol 109:103-108.
- 72. Lombard DB, Chua KF, Mostoslavsky R, Franco S, Costissa M, et al. (2005) DNA repair, genomic stability, and aging Cell 120: 497-512.

- Martien S, Abbadie C (2007) Acquisition of oxidative DNA damage during senescence: the first step toward carcinogenesis? Ann NY Acad Sci 1119: 51-63.
- 74. Agrawal A, Tay J, Yaang GE, Agrawal S, Gupta S (2010) Age-associated epigenetic modifications in human DNA increase its immunogenicity. Aging 2: 93-100.
- Mordstein M, Kochs G, Dunoutier L, Renauld JC, Paludan SR, et al. (2008) Interferon-lambda contributes to innate immunity in mice against influenza A virus but not against hepatotropic virus. PLos Pathol 4: e1000151.
- Mordstein M, Newgebauer E, Ditt V, Jessen B, Rieger T, et al. (2010) Lambda interferon renders epithelial cells of the respiratory and gastrointestinal tracts resistant to viral infections. J Virol 84: 5670-5677.
- Devendra D, Eisenbarth GS (2004) Interferon alpha- a potential link in the pathogenesis of viral-induced type I diabetes and autoimmunity. Clinical Immunolo 111: 225-233.
- Prummel MF, Laurberg P (2003) Interferon alpha and autoimmune thyroid disease. Thyroid 13: 547-551.
- Ronnblom LE, Elm GV, Oberg KE (1991) Autoimmunity after alpha-interferon therapy for malignant carcinoid tumors. Ann Int Med 115: 178-183.
- Abb J, Abb H, Deinhardt F (1984) Age-related decline of human interferon alpha and interferon gamma. production Blut 48: 285-289.
- Canaday DH, Amponsah NA, Jones I, Tisch DJ, Hornick TR, et al. (2010) Influenza-induced production of interferon-alpha is defective in geriatric individuals. J Clin Immunol 30: 373-383.
- Shodell M, Siegal FP (2002) Circulating, interferon-producing plasmacytoid dendritic cells decline during human ageing. Scand J Immunol 56: 518-521.
- Teig N, Moses D, Gieseler S, Schauer U (2002) Age-related changes in human blood dendritic cell subpopulations. Scand J Immunol 55: 453-457.
- Prakash S, Agrawal S, Cao J, Gupta S, Agrawal A (2012) Impaired secretion of interferons by dendritic cells from aged subjects to influenza Role of histone modification. Age 35: 1785-1797.
- 85. de Weerd NA, Vivian JP, Nguyen TK, Mangan NE, Gould JA, et al. (2013) Structural basis of a unique interferon-β signaling axis mediated via the receptor IFNR1. Nat Immunol 14: 901-907.
- 86. Fensterl V, Sen GC (2008) Interferons and viral infections. Biofactors 35: 14-20.

Page 5 of 5