

# **Cloning & Transgenesis**

# Recent Research Progress on MicroRNA's in Medicago Truncatula

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#### Abstract

MicroRNAs (miRNAs) are small, endogenous RNAs that play important regulatory roles in development and stress response by down-regulating gene expression post-transcriptionally. MiRNA study in *Medicago truncatula* is advancing quickly. This paper will introduce this field to readers from five aspects: nodulation, development of roots, response to drought, metal ions and phytohormone.

Keywords MicroRNA; Medicago truncatula; Research progress

#### Introduction

In plants, there are two kinds of small RNAs, endogenous small interfering RNAs (siRNAs) and microRNAs (miRNAs). Of them, miRNAs have an approximate length of 21 nt, and play key roles in negatively regulating gene expression by targeting corresponding messenger RNAs (mRNAs) [1,2]. It has been reported that miRNAs play a regulatory role in many processes in plants. In the field of growth and development, they are involved in seed germination [3], leaf morphogenesis [4,5], floral organ and root development [6-8]. In addition, they participated in responses of plants to various abiotic stresses, such as drought [9,10], cold [11], salinity [12], oxidative stress [13], metal ion toxicity [14,15], and nutrition starvation [16,17].

As the third model plant after Arabidopsis and rice, the miRNA study in *Medicago truncatula* is not so much detailed in contrast to Arabidopsis and rice [18-20]. However, the research development is so rapid. Here, the recent research progress on microRNAs will be introduced from several areas in *M. truncatula*.

#### Nodulation

As leguminous species, the roots of *M. truncatula* have the ability to interact with rhizobia to develop nitrogen-fixing root nodules. miR164 may be the first miRNA found to be involved in nodule formation, which can target *MtHAP2-1* encoding a transcription factor of the CCAAT-binding family with an ability of controlling nodule meristem function. Overexpression of miR164 leads to block of nodule development by down-regulating the expression of *MtHAP2-1* [21]. mR166 is another miRNA which affects nodule formation by negatively regulating several class-III HD-ZIP genes, and overexpression it makes number of symbiotic nodules decrease [8]. Lelandais-Briere et al. have identified many miRNAs in root nodules of *M. truncatula* by the method of high-throughput sequencing technology, which supplies abundant message for researchers about miRNA in nodules development [22].

#### **Development of Roots**

The root system is crucial for resource acquisition from soil. miRNAs involved in root development in *M. truncatula* have been identified [23]. Of them, overexpression of miR166 can reduce the number of lateral roots and induce ectopic development of vascular bundles in roots [8]. In addition, miR396 can regulate the architecture of the root system by targeting *MtGRF5* [24].

#### Drought

Thirty-two known members of 10 miRNA families and 8 new miRNAs/new members of known miRNA families were identified to be responsive to drought stress by high-throughput sequencing of sRNA libraries from *M. truncatula*. The targets of these miRNAs were involved in development, transcription, protein degradation, detoxification, nutrient status and cross adaptation [25]. An interesting discovery is miR398 targeting two Cu/Zn superoxide dismutases was found to be down-regulated transcriptionally in response to drought, different from the results reported by Trindale et al. [10]. This difference may results from the different extent and duration of drought stress, and it is needed to be studied sequentially.

#### **Metal Ions**

The expression response of miRNAs to aluminum and mercury toxicity was also been analyzed. Twenty-three miRNAs were responsive to  $Al^{3+}$ . By analysis the expression pattern of these miRNAs, it's found that the majority of these mRNAs was down-regulated and responded rapidly to  $Al^{3+}$  [15]. miRNAs and their targets response to  $Hg^{2+}$  were identified by high-throughput sequencing, too [14]. From these two articles, we found miR390 and miR2199 were all down-regulated by  $Al^{3+}$  and  $Hg^{2+}$ .

#### Phytohormone

The target of miR164, *MtNAC1* can be down-regulated by auxin homologous naphthaleneacetic acid (NAA), whose homolog *AtNAC1* plays a key role in auxin signal transport in Arabidopsis, but the function of *MtNAC1* is different from *AtNAC1* [7,26]. In addition, ethylene-responsive miRNAs in roots of *M. truncatula* have been identified by high-throughput sequencing. Eight miRNAs were down-

regulated after exposure to ethylene, and the potential role of these miRNAs in the ethylene-induced inhibition of root elongation was discussed [27].

#### Forecast

From the Web of Science, it's known that there are many papers about genome-wide identification of microRNAs from different treatments of Medicago truncatula. According to the newest miRNA database (miRBase 20, released in Jun. 2013), M. truncatula has the most amount of precursors and mature miRNAs in plants, even more than Arabidopsis and rice. But the functional study level of miRNAs in M. truncatula is not as high as Arabidopsis and rice. For example, two Cu/Zn superoxide dismutase genes (CSD1 and CSD2), a copper chaperone gene (CCS1) and a subunit gene of cytochrome c oxidase (COX5b-1) were all identified as target genes of miR398 in Arabidopsis. These genes were reported to be involved in response to diverse abiotic and biotic stresses, such as oxidative stress, salt stress and heat stress, Cu and phosphate deficiency, and addition of sucrose or plant pathogens, forming a detailed regulatory net of miR398 in Arabidopsis [13,28-32]. Because most of miRNAs have been identified and to find novel miRNAs from M. truncatula is more and more difficult, researchers are beginning to pay more attention to the functional study of important miRNAs in diverse biochemical and physiological processes, which will become the trend of miRNAs study in *M. truncatula*.

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## References

- 1. Jones-Rhoades MW, Bartel DP, Bartel B (2006) MicroRNAS and their regulatory roles in plants. Annu Rev Plant Biol 57: 19-53.
- Khraiwesh B, Arif MA, Seumel GI, Ossowski S, Weigel D, et al. (2010) Transcriptional control of gene expression by microRNAs. Cell 140: 111-122.
- Reyes JL, Chua NH (2007) ABA induction of miR159 controls transcript levels of two MYB factors during Arabidopsis seed germination. Plant J 49: 592-606.
- 4. Palatnik JF, Allen E, Wu X, Schommer C, Schwab R, et al. (2003) Control of leaf morphogenesis by microRNAs. Nature 425: 257-263.
- 5. Liu Z, Jia L, Wang H, He Y (2011) HYL1 regulates the balance between adaxial and abaxial identity for leaf flattening via miRNA-mediated pathways. J Exp Bot 62: 4367-4381.
- 6. Chen X (2004) A microRNA as a translational repressor of APETALA2 in Arabidopsis flower development. Science 303: 2022-2025.
- Guo HS, Xie Q, Fei JF, Chua NH (2005) MicroRNA directs mRNA cleavage of the transcription factor NAC1 to downregulate auxin signals for arabidopsis lateral root development. Plant Cell 17: 1376-1386.
- Boualem A, Laporte P, Jovanovic M, Laffont C, Plet J, et al. (2008) MicroRNA166 controls root and nodule development in Medicago truncatula. Plant J 54: 876-887.
- Li WX, Oono Y, Zhu J, He XJ, Wu JM, et al. (2008) The Arabidopsis NFYA5 transcription factor is regulated transcriptionally and posttranscriptionally to promote drought resistance. Plant Cell 20: 2238-2251.
- Trindade I, Capitão C, Dalmay T, Fevereiro MP, Santos DM (2010) miR398 and miR408 are up-regulated in response to water deficit in Medicago truncatula. Planta 231: 705-716.

- 11. Lv DK, Bai X, Li Y, Ding XD, Ge Y, et al. (2010) Profiling of cold-stressresponsive miRNAs in rice by microarrays. Gene 459: 39-47.
- 12. Zhao B, Ge L, Liang R, Li W, Ruan K, et al. (2009) Members of miR-169 family are induced by high salinity and transiently inhibit the NF-YA transcription factor. BMC Mol Biol 10: 29.
- Sunkar R, Kapoor A, Zhu JK (2006) Posttranscriptional induction of two Cu/Zn superoxide dismutase genes in Arabidopsis is mediated by downregulation of miR398 and important for oxidative stress tolerance. Plant Cell 18: 2051-2065.
- Zhou ZS, Zeng HQ, Liu ZP, Yang ZM (2012) Genome-wide identification of Medicago truncatula microRNAs and their targets reveals their differential regulation by heavy metal. Plant Cell Environ 35: 86-99.
- Chen L, Wang T, Zhao M, Tian Q, Zhang WH (2012) Identification of aluminum-responsive microRNAs in Medicago truncatula by genomewide high-throughput sequencing. Planta 235: 375-386.
- Bari R, Datt Pant B, Stitt M, Scheible WR (2006) PHO2, microRNA399, and PHR1 define a phosphate-signaling pathway in plants. Plant Physiol 141: 988-999.
- 17. Chiou TJ, Aung K, Lin SI, Wu CC, Chiang SF, et al. (2006) Regulation of phosphate homeostasis by MicroRNA in Arabidopsis. Plant Cell 18: 412-421.
- Rajagopalan R, Vaucheret H, Trejo J, Bartel DP (2006) A diverse and evolutionarily fluid set of microRNAs in Arabidopsis thaliana. Genes Dev 20: 3407-3425.
- Fahlgren N, Howell MD, Kasschau KD, Chapman EJ, Sullivan CM, et al. (2007) High-throughput sequencing of Arabidopsis microRNAs: evidence for frequent birth and death of MIRNA genes. PLoS One 2: e219.
- Sunkar R, Zhou X, Zheng Y, Zhang W, Zhu JK (2008) Identification of novel and candidate miRNAs in rice by high throughput sequencing. BMC Plant Biol 8: 25.
- Combier JP, Frugier F, de Billy F, Boualem A, El-Yahyaoui F, et al. (2006) MtHAP2-1 is a key transcriptional regulator of symbiotic nodule development regulated by microRNA169 in Medicago truncatula. Genes Dev 20: 3084-3088.
- 22. Lelandais-Brière C, Naya L, Sallet E, Calenge F, Frugier F, et al. (2009) Genome-wide Medicago truncatula small RNA analysis revealed novel microRNAs and isoforms differentially regulated in roots and nodules. Plant Cell 21: 2780-2796.
- Eyles RP, Williams PH, Ohms SJ, Weiller GF, Ogilvie HA, et al. (2013) microRNA profiling of root tissues and root forming explant cultures in Medicago truncatula. Planta 238: 91-105.
- 24. Bazin J, Khan GA, Combier JP, Bustos-Sanmamed P, Debernardi JM, et al. (2013) miR396 affects mycorrhization and root meristem activity in the legume Medicago truncatula. Plant J 74: 920-934.
- Wang T, Chen L, Zhao M, Tian Q, Zhang WH (2011) Identification of drought-responsive microRNAs in Medicago truncatula by genome-wide high-throughput sequencing. BMC Genomics 12: 367.
- 26. D'haeseleer K, Den Herder G, Laffont C, Plet J, Mortier V, et al. (2011) Transcriptional and post-transcriptional regulation of a NAC1 transcription factor in Medicago truncatula roots. New Phytol 191: 647-661.
- 27. Chen L, Wang T, Zhao M, Zhang W (2012) Ethylene-responsive miRNAs in roots of Medicago truncatula identified by high-throughput sequencing at whole genome level. Plant Sci 184: 14-19.
- Abdel-Ghany SE, Pilon M (2008) MicroRNA-mediated systemic downregulation of copper protein expression in response to low copper availability in Arabidopsis. J Biol Chem 283: 15932-15945.
- Jagadeeswaran G, Saini A, Sunkar R (2009) Biotic and abiotic stress down-regulate miR398 expression in Arabidopsis. Planta 229: 1009-1014.
- 30. Jia X, Wang WX, Ren L, Chen QJ, Mendu V, et al. (2009) Differential and dynamic regulation of miR398 in response to ABA and salt stress in Populus tremula and Arabidopsis thaliana. Plant Mol Biol 71: 51-59.

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- 31. Pant BD, Musialak-Lange M, Nuc P, May P, Buhtz A, et al. (2009) Identification of nutrient-responsive Arabidopsis and rapeseed microRNAs by comprehensive real-time polymerase chain reaction profiling and small RNA sequencing. Plant Physiol 150: 1541–1555.
- 32. Guan Q, Lu X, Zeng H, Zhang Y, Zhu J (2013) Heat stress induction of miR398 triggers a regulatory loop that is critical for thermotolerance in Arabidopsis. Plant J 74: 840-851.