

Proteomics in the Light of Integral Value Transformations

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Abstract

In this paper, proteomics have been studied in the light of Integral Value Transformations (IVTs) which was introduced in. For case study, a Human olfactory receptor OR1D2 protein sequence has been considered as the initial sequence and then different IVTs have been used to evolve OR1D2 into some other proteomic sequences. As ensued, it has been found that some of the generated sequences have been mapped to another olfactory receptor in Human or in some other species. Also it has been corroborated through fractal dimension that some of the fundamental protein properties have been nearly intact. Thus, we propose a methodology via which proteins having the same properties can be connected or grouped. This study will help to comprehend proteomic evolutionary network through IVTs.

Keywords: Olfactory receptors (ORs); Box-counting dimension; Proteomics

Introduction

The study of proteins such as structures, functions and evolutions is universally known to as *Proteomics*, was first coined in 1997 to make an analogy with *Genomics*, the study of the genes [1]. After genomics, proteomics is considered to be the next step in the study of biological systems. While we humans probably have only some 21 thousand genes, we possess at least 10 times that number of different proteins. The study of proteomics is important because proteins are responsible for both the structure and the functions of all living things. Genes are simply the instructions for making proteins. Therefore, a proper quantitative understanding of proteins characteristics and their inter-network are required. In this paper, an olfactory receptor OR1D2 has been considered for our analysis. Interestingly, on applying IVT systematically, we have been able to show that each DNA sequence at various discrete time instances in IVT evolutions can be directly mapped to another specific proteomic sequences existing in different species. A number of the fundamental properties namely percentage of accessible residues, alpha helix (Chou & Fasman), amino acid composition (%), beta sheet (Chou & Fasman), beta turn (Chou & Fasman), coil (Deleage & Roux), hydrophobicity (Aboderin) and total beta strand have been considered for the protein properties of the IVT generated sequences. All protein plots for all the IVT generated sequences including OR1D2 (the primitive sequence) have been generated using MATLAB (bioinformatics toolbox). Then box-counting dimension for each of the protein plots have been calculated through BENOITTM. This study will help us to ascertain potential new drugs for the treatment of various diseases.

Some Reviews and Fundamentals

In this section, we describe very briefly about IVTs, fractal and proteins.

Notion of integral value transformation (IVT)

Let us define the Integral Value Transformations (IVTs) in N_0^K as the following [2-5]:

$$IVT^{(p,k)}_j : N_0^K \longrightarrow N_0$$

$$IVT^{(p,k)}_j((n_1, n_2, \dots, n_k) = (f_j(a_0^{(n_1)}, a_0^{(n_2)}, \dots, a_0^{(n_k)}) \\ f_j(a_1^{(n_1)}, a_1^{(n_2)}, \dots, a_1^{(n_k)}) \dots \dots \dots f_j(a_{(l-1)}^{(n_1)}, a_{(l-1)}^{(n_2)}, \dots, a_{(l-1)}^{(n_k)}))_p = m$$

where

$$n_1 = (a_0^{(n_1)} a_1^{(n_1)} \dots a_{(l-1)}^{(n_1)})_p, n_2 = \\ (a_0^{(n_2)} a_1^{(n_2)} \dots a_{(l-1)}^{(n_2)})_p, \dots, n_k = (a_0^{(n_k)} a_1^{(n_k)} \dots a_{(l-1)}^{(n_k)})_p \\ f_j : \{0, 1, 2, \dots, p-1\}^k \longrightarrow \{0, 1, 2, \dots, p-1\}$$

m is the decimal conversion from the p adic number.

Obviously for $IVT^{(p,k)}_j$ system there are p^{p^k} number of possible rules and out of them the function number that we select is indicated by j.

Let us fix the domain of IVTs as N_0 (k=1) and thus the above definition boils down to the following:

$$IVT^{p,1}_j(x) = (f_j(x_n) f_j(x_{n-1}) \dots \dots \dots f_j(x_1))_p = m$$

where m is the decimal conversion from the p adic number, and

$$x = (x_n x_{n-1} \dots \dots x_1)_p$$

Now, let us denote the set of $IVT^{p,1}_j$ as

$$T^{p,1} = \left\{ \begin{array}{l} IVT^{p,1}_j : N_0 \rightarrow N_0 \\ 0 \leq j < p^p, IVT^{p,1}_j(x) = (f_j(x_n) f_j(x_{n-1}) \dots \dots \dots f_j(x_1))_p = m \\ \text{where m is the decimal conversion} \\ \text{from the p adic number and } x = (x_n x_{n-1} \dots \dots x_1)_p \end{array} \right\}$$

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$$T^{p,1} = \left\{ \begin{array}{l} IVT^{p,1}_j : N_0 \rightarrow N_0 \mid 0 \leq j < P^p, IVT^{p,1}_j(x) \\ = (f_j(x_n) f_j(x_{n-1}) \dots f_j(x_1))_p = m \end{array} \right\}$$

Where m is the decimal conversion from the P adic number and $x = (x_n x_{n-1} \dots x_1)_p$

Let us define the IVT in N_0 in 4-adic number systems. There are 256 ($4^{(4^1)}$) one variable four state CA rules. Corresponding to each of those CA rules there are 256 IVTs in 4 adic system in one dimension.

$IVT^{4,1}_\#$ is mapping a non-negative integer to a non-negative integer.

$$IVT^{4,1}_\#(a) = ((f_\#(a_n) f_\#(a_{n-1}) \dots f_\#(a_1))_4 = b$$

Where 'a' is a non-negative integer and $a = (a_n a_{n-1} \dots a_1)_4$ and 'b' is the decimal value corresponding to the 4-adic number.

For an example, let us consider $a = 225 = (3201)_4$ and $\# = 120$ so $f_\#(0) = 0; f_\#(1) = 2; f_\#(2) = 3$ and $f_\#(3) = 1$

Therefore,

$$IVT^{(4,1)}_{120}(225) = (f_{120}(3) f_{120}(2) f_{120}(0) f_{120}(1))_4 = (1302)_4 = 114.$$

Consequently, $IVT^{4,1}_{120}(225) = 114.$

Let us denote $T^{(4,1)}_\#$ as set of all $IVT^{p,k}_\#$ transformations. It is worth noting that there are $4! = 24$ number of bijective functions in $\mathfrak{T}^{4,1}_\#$. So out of the 256 ($4^{(4^1)}$) transformations in $\mathfrak{T}^{4,1}_\#$ four are linear and rest is nonlinear [6].

Fractal and fractal dimension

Our artificial world can be described easily through Euclidean geometric shapes but there are many things in nature such as shape of cloud, geometry of lightning etc. could not be described through Euclidean geometry. Many mathematicians descended the challenge for a fair enough description of natural objects but after a long period in 1975, B. Mandelbrot took up the challenge and gave the birth of a new geometry to describe nature which is known to us as 'Fractal Geometry' (in short 'Fractal'). The precise definition of "Fractal" according to Benoit Mandelbrot is a set for which the Hausdorff Besicovitch dimension strictly exceeds the topological dimension. To gain a quantitative insight of Fractal, some fractal parameters namely Fractal dimension, Hurst exponent, succolarity, lacunarity etc. are also introduced in the literature. A brief discussion follows about one of the well-known methods of calculating fractal dimension namely 'Box-Counting method'.

Box-Counting Method: This method computes the number of cells required to entirely cover an object, with grids of cells of varying size. Practically, this is performed by superimposing regular grids over an object and by counting the number of occupied cells. The logarithm of $N(r)$, the number of occupied cells, versus the logarithm of $1/r$, where r is the size of one cell, gives a line whose gradient corresponds to the box dimension [7].

Problem in protein structures

Proteins are an important class of biological macromolecules present

in all organisms. After the structure of DNA was discovered by James Watson and Francis Crick, who used the experimental evidence of Maurice Wilkins and Rosalind Franklin (among others), serious efforts to understand the nature of the encoding of proteins began. George postulated that a three-letter code must be employed to encode the 20 standard amino acids used by living cells to encode proteins, because 3 is the smallest integer n such that 4^n is at least 20 [8]. The three-dimensional structures of proteins were first determined by X-ray diffraction analysis; Perutz and Kendrew shared the 1962 Nobel Prize in Chemistry for these discoveries. At present, more than ten thousand protein structures were found with their atomic details. The structure of the protein is ultimately defined by its primary structure, or amino acid sequence. There are no theories or computational techniques at the moment which will allow us to predict the new protein folding by its sequence. Even, how protein sequences and their tertiary structures are evolved during evolution remains unclear. Therefore proper understanding is required at the primary structure level i.e. in the amino acids sequence level of proteins.

Methods and Results

Method of sequence generation through IVTs

The domain of action of IVTs is a set of non-negative positive integers. So it is required to have a numeric sequence corresponding to each of the proteomic sequence. A simple mapping f is defined below:

Let $\mathcal{P} = \{A, C, D, E, F, G, H, I, K, L, Q, N, P, Q, R, S, T, V, W, Y\}$ be the set of amino acid codes and

$$\mathcal{N} = \{0, 1, 2, 3, \dots, 19\}, f : \mathcal{P} \rightarrow \mathcal{N} \text{ as } f(A) = 0; f(C) = 1, f(D) = 2, f(E) = 3, \dots, f(W) = 18 \text{ and } f(Y) = 19$$

Therefore, a protein sequence is now simply a string of twenty variables namely 0, 1, 2...19 as per coding scheme f .

Starting from a protein sequence to generate another proteomic like sequences, it is required to have all the IVTs in a particular $T^{(p,1)}_\#$, which maps \mathcal{N} to itself (bijective rules).

The list of some such IVTs in $T^{(p,1)}_\#$ is given below in Table 1.

Now we apply Integral Value Transformations ($IVT^{(p,1)}_\#$) systematically [3-10]:-

Firstly, divide the whole one dimensional initial sheet of proteomic sequence (numeric sequence) of length n into multiple blocks. We designate the initial sequence as $S(t_0)$.

Secondly, we apply bijective domain preservative transformations (need not to be all distinct) taken from $\mathfrak{T}^{p,1}_\#$ for different p starting from 2 to 19) over each of the r different blocks in t_0 . We call such application of different rules to different blocks as *Hybrid Application of IVTs*. In other words, we are getting $S(t_1)$ from $S(t_0)$ through hybrid application of IVTs. Next, we follow this step successively as long as we wish to iterate. The results, on applying the proposed systematic technique of application of IVTs on OR1D2 are enumerated in the following subsections.

Results

Here we discuss the results on applying different IVTs in two following cases.

On applying $IVT_{\#}^{p,1}$: The proteomic sequence of OR1D2 is of length 312 (sequence shown below in Text-1). Choose $r=50$, so there are 7 blocks are there. The following two IVTs are used to generate $S(t_i)$ as shown below in Table 2.

MDGGNQSEGSEFLLGMSESPEQQRILFWMFLSMYLVTVVGNVLIILAI

SDSRLHTPVYFFLANLSFTDLFFVTNTIPKMLVN-LQSHNKAISYAGCLTQ

LYFLVSLVALDNLILAVMAYDRYVAICCPHLYTTAMSPKLCILLSLCVW

LSVLYGLIHTLLMTRVTFGSRKIHIFCEMYVLLRMACS-NIQINHTVLI

P-adic	$\mathfrak{T}^{p,1}_{\#}$	$\mathfrak{T}^{2,1}_{\#}$	$\mathfrak{T}^{3,1}_{\#}$	$\mathfrak{T}^{4,1}_{\#}$	$\mathfrak{T}^{5,1}_{\#}$	$\mathfrak{T}^{6,1}_{\#}$	$\mathfrak{T}^{7,1}_{\#}$	$\mathfrak{T}^{8,1}_{\#}$	$\mathfrak{T}^{9,1}_{\#}$
#	1 2	5 11 21	99 114 147 177 180 210 414 434 694 714	28805 28955 28985 297393	28565 28595 28745 297051	297093 5135375 5135431	102907844 102907916	297435 5135886 5135942	102908572 102908644
			225 228 894	29860 29890 30040 299109	299151 5138959 5139015	102913676 102913748	299835 5139981	102915132	

Table.1: IVTs in $\mathfrak{T}^{p,1}_{\#}$.

BLOCK	Sequence-1 in 2 adic IVT	Sequence-1 in 3 adic IVT	Sequence-1 in 4 adic IVT
Block-1	$IVT^{2,1}_1$	$IVT^{3,1}_5$	$IVT^{4,1}_{99}$
Block-2	$IVT^{2,1}_1$	$IVT^{3,1}_5$	$IVT^{4,1}_{114}$
Block-3	$IVT^{2,1}_2$	$IVT^{3,1}_{11}$	$IVT^{4,1}_{147}$
Block-4	$IVT^{2,1}_1$	$IVT^{3,1}_{11}$	$IVT^{4,1}_{177}$
Block-5	$IVT^{2,1}_2$	$IVT^{3,1}_{21}$	$IVT^{4,1}_{180}$
Block-6	$IVT^{2,1}_2$	$IVT^{3,1}_{21}$	$IVT^{4,1}_{210}$
Block-7	$IVT^{2,1}_2$	$IVT^{3,1}_{21}$	$IVT^{4,1}_{225}$
BLOCK	Sequence-1 in 5 adic IVT	Sequence-1 in 6 adic IVT	Sequence-1 in 7 adic IVT
Block-1	$IVT^{5,1}_{194}$	$IVT^{6,1}_{28565}$	$IVT^{7,1}_{297051}$
Block-2	$IVT^{5,1}_{214}$	$IVT^{6,1}_{28595}$	$IVT^{7,1}_{297093}$
Block-3	$IVT^{5,1}_{294}$	$IVT^{6,1}_{28745}$	$IVT^{7,1}_{297393}$
Block-4	$IVT^{5,1}_{334}$	$IVT^{6,1}_{28805}$	$IVT^{7,1}_{297435}$
Block-5	$IVT^{5,1}_{414}$	$IVT^{6,1}_{28985}$	$IVT^{7,1}_{299109}$
Block-6	$IVT^{5,1}_{434}$	$IVT^{6,1}_{28955}$	$IVT^{7,1}_{299151}$
Block-7	$IVT^{5,1}_{694}$	$IVT^{6,1}_{29860}$	$IVT^{7,1}_{299793}$
BLOCK	Sequence-1 in 8 adic IVT	Sequence-1 in 9 adic IVT	
Block-1	$IVT^{8,1}_{5135375}$	$IVT^{9,1}_{102907844}$	
Block-2	$IVT^{8,1}_{5135431}$	$IVT^{9,1}_{102907916}$	
Block-3	$IVT^{8,1}_{5135886}$	$IVT^{9,1}_{102908572}$	
Block-4	$IVT^{8,1}_{5135942}$	$IVT^{9,1}_{102908644}$	
Block-5	$IVT^{8,1}_{5138959}$	$IVT^{9,1}_{102913676}$	
Block-6	$IVT^{8,1}_{5139015}$	$IVT^{9,1}_{102913748}$	
Block-7	$IVT^{8,1}_{5139981}$	$IVT^{9,1}_{102915132}$	

Table 2: IVTs from $\mathfrak{T}^{p,1}_{\#}$ used for generation of $\mathfrak{T}^{p,1}_{\#}$.

ATGCFILIPFGFVIISYVLIIRAILRIPSVSKKYKAFSTCASH-LGAVSL

FYGTLCMVYVKPLHTYSVKDSVATVMYAVVTPMMN-PFIYSLRNKDMHGAL

GRLLDKHFKRLT

Text 1: Protein Sequence of OR1D2.

Similarly, other $S(t_i)$ can be generated applying the IVTs in different blocks of the $S(t_{i-1})$ as tabulated in *supl.met-I*. We have generated 90 such $S(t_i)$ s corresponding to OR1D2 in each $\mathfrak{T}^{p,1}_{\#}$ system (for $p=2, 3 \dots 20$) (available in *supl.met.-II*).

All these generated sequences have been blast in the NCBI database for significant similarity. The blast result is shown in *supl.met.-III*.

Most of the generated sequences are mapped to olfactory receptors (specifically close to OR1D2) in different organisms like *homo sapiens*, *pan troglodytes*, *lagothrixlagotricha* etc. Some of the sequences are not mapped due to the fact that they are more conserved sequence than OR1D2.

Also we have been observed that some of the protein primary structural properties (listed below) are intact with respect to the two dimensional protein plot graphs (using bioinformatics toolbox of Matlab-R2010b) for each of the generated sequences.

The protein properties which we have considered here are as follows:

- Prop-1: Accessible residues (%)
- Prop-2: Alpha helix (Chou & Fasman)
- Prop-3: Amino acid composition (%)
- Prop-4: Beta sheet (Chou & Fasman)
- Prop-5: Beta turn (Chou & Fasman)
- Prop-6: Coil (Deleage & Roux)
- Prop-7: Hydrophobicity (Aboderin)
- Prop-8: Total beta strand

Corresponding to each property of the $S(t_i)$, we have had eight protein plot graphs from which we have calculated box counting dimensions using BENOITTM.

The data for OR1D2 sequence are stated below in the Table 3. The rest of the data are available in the *supl.met-IV*. We have observed that the box-counting dimensions for all the eight protein plots corresponding to each of the protein property for all the generated sequences $S(t_i)$ s are almost same to the same of OR1D2. The data for all the box counting dimension of protein plots for the $S(t_i)$ generated through the $\mathfrak{T}^{2,1}_{\#}$ system is shown below. Hereby we can come to a conclusion that these IVTs preserve the protein properties of the strings. It is to be noted that all these IVTs are bijective; therefore one can switch from one protein to another protein through the IVTs without encumbering the protein properties. Most of the $S(t_i)$ (IVT generated sequences) preserve all the eight protein properties. It is to be noted that in the case $\mathfrak{T}^{2,1}_{\#}$ system, $S(t_1)$ and $S(t_2)$ are both mapped to G-protein-coupled receptor in OR1D2 in human. Also they follow all the protein properties as in OR1D2 (Table 4).

But interestingly, there are many $S(t_i)$ in different $\mathfrak{T}^{p,1}_{\#}$ systems, do not map significantly in any organisms but they possess the protein properties as in OR1D2. One of the main reasons for this is that most of the sequences are conserved whereas OR1D2 is not so. Some of the

Sequence	Property	Box-counting dimension
OR1D2	Prop1	1.91092
	Prop2	1.91103
	Prop3	1.90855
	Prop4	1.91141
Sequence	Property	Box-counting dimension
	Prop5	1.91095
	Prop6	1.91348
	Prop7	1.90989
	Prop8	1.91071

Table 3: Box-counting dimension for protein plots of OR1D2.

Sequence	Property	Box-counting dimension
$S(t_1)$	Prop1	1.92694
	Prop2	1.91117
	Prop3	1.90976
	Prop4	1.91111
	Prop5	1.91113
	Prop6	1.93038
	Prop7	1.91021
	Prop8	1.91144
$S(t_2)$	Prop1	1.91124
	Prop2	1.91099
	Prop3	1.91389
	Prop4	1.90948
	Prop5	1.91064
	Prop6	1.93051
	Prop7	1.91398
	Prop8	1.90983
$S(t_3)$	Prop1	1.91045
	Prop2	1.91049
	Prop3	1.90994
	Prop4	1.91299
	Prop5	1.92765
	Prop6	1.91648
	Prop7	1.92813
	Prop8	1.91448
$S(t_4)$	Prop1	1.91294
	Prop2	1.91495
	Prop3	1.91084
	Prop4	1.9108
	Prop5	1.91155
	Prop6	1.91577
	Prop7	1.9281
	Prop8	1.93043
$S(t_5)$	Prop1	1.91443
	Prop2	1.91431
	Prop3	1.91259

	Prop4	1.93055
	Prop5	1.92909
	Prop6	1.91638
	Prop7	1.92901
	Prop8	1.91676
$S(t_6)$	Prop1	1.92863
	Prop2	1.928
	Prop3	1.91431
	Prop4	1.9295
	Prop5	1.91133
	Prop6	1.91751
	Prop7	1.91379
	Prop8	1.91292
$S(t_7)$	Prop1	1.91421
	Prop2	1.928
	Prop3	1.9142
	Prop4	1.91614
	Prop5	1.9101
	Prop6	1.91402
	Prop7	1.9108
	Prop8	1.91314
$S(t_8)$	Prop1	1.9104
	Prop2	1.91378
	Prop3	1.91039
	Prop4	1.91287
	Prop5	1.91177
	Prop6	1.91392
	Prop7	1.90987
	Prop8	1.91378
$S(t_9)$	Prop1	1.91428
	Prop2	1.91129
	Prop3	1.91367
	Prop4	1.91337
	Prop5	1.91263
	Prop6	1.91431
	Prop7	1.91084
	Prop8	1.91413
$S(t_{10})$	Prop1	1.91082
	Prop2	1.9108
	Prop3	1.91081
	Prop4	1.91337
	Prop5	1.91263
	Prop6	1.91514
	Prop7	1.91084
	Prop8	1.9176

Table 4: Box-counting dimension for all protein plots of $\mathfrak{T}^{p,1}_{\#}$ in $\mathfrak{T}^{p,1}_{\#}$.

$S(t_i)$ are not mapped to any of the ORs in any organism although the box-counting dimension for all the protein plots are intact as it is in OR1D2. It is our strong conviction that these $S(t_i)$ serve the purpose for replacement of OR1D2 in the genetic evolutionary future. In the next section we are about to discuss the case on applying the bijective IVTs from $\mathfrak{T}^{20,1}_{\#}$ (Table 5).

On Applying $IVT^{(20,1)}_{\#}$: We have chosen a few bijective IVTs (available in *supl. met.-I*) from $\mathfrak{T}^{20,1}_{\#}$ system to generate $S(t_i)$ from

the protein code for OR1D2 (methodology is discussed in 3.1). Here all the $S(t_i)$ have been blasted in NCBI and they all are mapped to G protein-coupled receptor, or MOR30-1, hypothetical protein and conserved hypothetical protein in different organisms ranging from human to plasmodium species (data shown in *supl. met.-III*). The box counting dimension is still intact for all the protein plots for all the IVT generated sequence in $\mathfrak{T}^{20,1}_{\#}$ system as shown in (Figure 1) (raw data shown in *supl. met.-IV*). It is noted that the number of bijective, domain preservative IVTs is increased as p increased in $T^{(p,1)}_{\#}$. Consequently the sequential conservation is inversely proportional to p.

Summary and Conclusion

In summary, we have seen that IVTs steer a given OR sequence of a species to another of the same or different (most likely) species, preserving the protein properties of the original sequence. This

Sequence	Property	Box-counting dimension
$S(t_1)$	Prop1	1.90836
	Prop2	1.91371
	Prop3	1.92937
	Prop4	1.91313
	Prop5	1.92746
	Prop6	1.9128
	Prop7	1.91234
	Prop8	1.91291
$S(t_2)$	Prop1	1.91418
	Prop2	1.91204
	Prop3	1.91182
	Prop4	1.91205
	Prop5	1.91418
	Prop6	1.92998
	Prop7	1.9099
	Prop8	1.91351
$S(t_3)$	Prop1	1.91459
	Prop2	1.91308
	Prop3	1.91151
	Prop4	1.91464
	Prop5	1.91434
	Prop6	1.91216
	Prop7	1.91306
	Prop8	1.91321
$S(t_4)$	Prop1	1.91087
	Prop2	1.91468
	Prop3	1.90957
	Prop4	1.90991
	Prop5	1.92755
	Prop6	1.9159
	Prop7	1.9104
	Prop8	1.91369
$S(t_5)$	Prop1	1.91448
	Prop2	1.91485
	Prop3	1.92691
	Prop4	1.914
	Prop5	1.9123
	Prop6	1.91203
	Prop7	1.92751

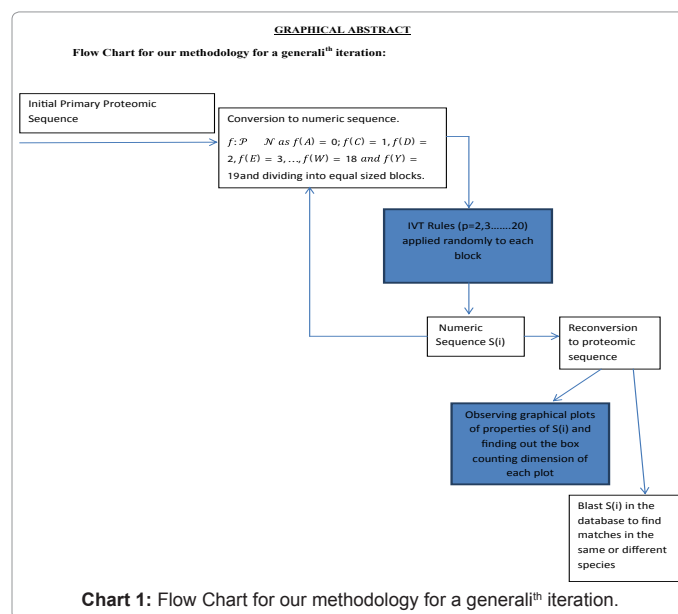
$\tau^{8,1}_{\#}$	Prop8	1.92845
	Prop1	1.91315
	Prop2	1.91176
	Prop3	1.91169
	Prop4	1.91317
	Prop5	1.91348
	Prop6	1.91507
	Prop7	1.91141
$S(t_7)$	Prop8	1.92879
	Prop1	1.91258
	Prop2	1.91057
	Prop3	1.91388
	Prop4	1.91508
	Prop5	1.92907
	Prop6	1.91605
	Prop7	1.91244
$S(t_8)$	Prop8	1.91098
	Prop1	1.92725
	Prop2	1.92767
	Prop3	1.91331
	Prop4	1.91074
	Prop5	1.91459
	Prop6	1.91608
	Prop7	1.90883
$S(t_9)$	Prop8	1.91143
	Prop1	1.90984
	Prop2	1.92917
	Prop3	1.9154
	Prop4	1.91098
	Prop5	1.91336
	Prop6	1.91545
	Prop7	1.91013
$S(t_{10})$	Prop8	1.92845
	Prop1	1.91286
	Prop2	1.91425
	Prop3	1.91506
	Prop4	1.91402
	Prop5	1.92938
	Prop6	1.91632
	Prop7	1.91337
Prop8	1.9125	

Table 5: Box-counting dimension for all protein plots of $\tau^{p,1}_{\#}$ in $\tau^{p,1}_{\#}$.

methodology will be helpful to mimic the genomic evolution procedure artificially, which is required for genetic replacement therapy. IVTs may also be considered to be a platform to comprehend the morphological connections among the various species. A naïve question to the biologists, which rose amongst us:

Suppose, we are given an olfactory receptor *or1* of a species *s1* which help it to identify the odors *x1*, *x2*,...

Now, we apply the proposed methodology to *or1* and obtain a new olfactory receptor *or2* (supposedly) of species *s2*.



So, does *or2* help *s2* in identifying the same odors *x1*, *x2*,...?

In the near future, we are really interested to explore the underlying biological methodology that governs the entire process.

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References

1. Crick F (1988) Chapter 8: The genetic code. What mad pursuit: a personal view of scientific discovery. New York: Basic Books 89-101.
2. Hassan SS, Choudhury PP, Singh R, Das S, Nayak BK (2010) Collatz Function like Integral Value Transformations, Alexandria Journal of Mathematics 1: 30-35.
3. Hassan SS, Choudhury PP, Guha R, Chakraborty S, Goswami A, et al. (2011) DNA Sequence Evolution through Integral Value Transformations. Interdisciplinary Sciences: Computational Life Sciences 4: 128-132.
4. Hassan SKS, Choudhury PP, Goswami A (2010) Underlying Mathematics in Diversification of Human Olfactory Receptors in Different Loci. Interdisciplinary Sciences: Computational Life Sciences, Springer.
5. Hassan SS, Choudhury PP, Goswami A, De Sarkar N, Fangal V (2011) Quantification of miRNAs and Their Networks in the light of Integral Value Transformations. Nature Precedings.
6. Hassan SK.S, Roy A, Choudhury PP, Nayak B.K. (2011) One Dimensional p-adic Integral Value Transformations, arXiv.
7. Mandelbrot BB (1982) The Fractal Geometry of Nature. W.H. Freeman and Company, New York.
8. Yu Xia, Michael Levitt (2004) Simulating protein evolution in sequence and structure space. Current Opinion in Structural Biology 14: 202-207
9. Cattani C (2010) Fractals and hidden symmetries in DNA. Mathematical Problems in Engineering.
10. Zu-Guo Y, Anh V, Zhi-Min G, Shun-Chao L (2002) Fractals in DNA sequence analysis, Chinese Phys11: 1313.