

**Review Article** 

# Progress in Understanding the Pathogenesis of Systemic Sclerosis with Lung Involvement: The Contribution of Proteomic Studies

Cinzia Scambi<sup>1\*</sup>, Lucia De Franceschi<sup>2</sup>, Silvia Bosello<sup>3</sup>, Paola Caramaschi<sup>1</sup>, Gianfranco Ferraccioli<sup>3</sup> and Domenico Biasi<sup>1</sup>

<sup>1</sup>Department of Medicine, Section of Rheumatology, University of Verona and AOUI, Verona, Italy <sup>2</sup>Department of Medicine, Section of Internal Medicine, University of Verona and AOUI, Verona, Italy <sup>3</sup>Department of Medicine, Division of Rheumatology, Catholic University, Roma, Italy

### Abstract

Systemic sclerosis (SSc) is a chronic inflammatory disease involving the skin and various internal organs. SSc is characterized by microvascular dysfunction, activation of the immune system and tissue fibrosis. Endothelial cell damage seems to be the initiating factor, but the precise triggering events that underlie the development of the disease remain unclear.

Lung involvement is a frequent complication, which causes increased morbidity and mortality in patients with SSc. In particular, pulmonary arterial hypertension and interstitial lung disease are the two major clinical diseases that affect SSc patients, but the current treatments appear to have no satisfying effects on these pulmonary complications, until yet.

Recently researches using innovative technologies have highlighted several peptides that might contribute to better understand the underlying pathogenic mechanisms of pulmonary injury and could promote more effective therapeutic strategies for SSc patients. Here, we focus on the major proteomic studies on biological fluid from patients with SSc.

**Keywords:** Proteomics; Pulmonary arterial hypertension; Interstitial lung disease; Systemic sclerosis

## Introduction

Systemic sclerosis (SSc) is an autoimmune disease that affects skin and internal organs.

The incidence of SSc is approximately 20 per million and the cause of it remains poorly understood [1].

Current hypotheses in SSc pathogenesis suggest that several exogenous agents activate an abnormal cellular and humoral immunity, in genetically predisposed subjects [2]. In this way, products of the immune activation cause vascular damage possibly through the production of autoantibodies and inflammatory peptides that induce vascular permeability, up-regulation of adhesion molecules and endothelial cell (EC) apoptosis. These vascular dysfunctions enhance the infiltration of mononuclear cells in affected organs and cause tissue fibrosis.

SSc is clinically diverse in terms of skin and organ involvement. Normally, it is distinct in two major clinical subtypes: the diffuse form, that typically leads to a rapid sclerosis of the whole skin and it is associated with important body organs fibrosis, including lungs, heart, digestive tract and kidneys; the limited form, that is characterized by sclerosis of the distal parts of the skin and it is associated with a much slower progression of visceral fibrosis.

In general, two major pulmonary syndromes are associated with SSc: the pulmonary arterial hypertension (PAH) and the interstitial lung disease (ILD) [3,4].

Both PAH and ILD have a great impact on morbidity and mortality of SSc patients, despite the rate progression is variable through SSc patients population [5,6]. As current therapy appears to have modest effects on these conditions, it is necessary better understanding their underlying pathogenic mechanisms.

The basic science studies, integrated with clinical evaluation of SSc patients, are direct to identify either new biological disease marker(s)

suitable for clinical practice or new signaling pathways involved in the pathogenesis of SSc disease [5,7].

Although progresses have been made in the pathogenesis of lung involvement in SSc patients, much still remains to be investigated. The recent development of innovative technologies for massive protein analysis allows the researchers to shed light on SSc disease. In SSc patients with lung involvement, proteomic studies mainly focus on two biological liquid interfaces: the bronchoalveolar lavage fluid (BALF) and the serum.

In the present review, we discuss the more significant proteomic studies on BALF and serum in SSc patients with lung disease.

## Proteomic High-Throughput Approaches in BALF Analysis from SSc Patients with Pulmonary Involvement

BALF is a suitable way to evaluate lower respiratory tract abnormalities in clinical setting and to sample the biological components such as proteins of the epithelial fluid [8].

Recently, Rottoli et al. [9,10] have conducted a 2-D analysis of BALF from SSc patients with ILD. The Authors have reported quantitative rather than qualitative differences in protein profile in the comparison between SSc patients with lung involvement and patients with other ILD forms, such as idiopathic pulmonary fibrosis or sarcoidosis [9].

\*Corresponding author: Cinzia Scambi, MD, Ph.D, Department of Medicine, Section of Rheumatology, University of Verona and AOUI, Policlinico GB Rossi, P.Ie L Scuro, 10, 37134 Verona, Italy, E-mail: cinzia.scambi@univr.it

Received November 14, 2011; Accepted January 28, 2012; Published January 31, 2012

**Citation:** Scambi C, Franceschi LD, Bosello S, Caramaschi P, Ferraccioli G, et al. (2012) Progress in Understanding the Pathogenesis of Systemic Sclerosis with Lung Involvement: The Contribution of Proteomic Studies. Rheumatology S1:004. doi:10.4172/2161-1149.S1-004

The differently expressed proteins belong to the classes of proteases, cytokines and antioxidants (Table 1). In addition, increased protein carbonyl content, as measurement of protein oxidation, was observed in SSc patients with lung involvement compared to patients with other ILD forms, suggesting a redox imbalance between oxidant generation and antioxidant mechanisms in SSc patients with lung involvement [10,11]. Higher values of complement C3b, the cleavage product of C3, were also found in BALF from SSc patients with lung damage [10]. Despite the etiopathogenic role of complement in the development of interstitial lung disease is still under investigation [12-15], a genetic study on complement system in SSc reported a strong association between SSc and the null alleles at the C4A locus in the major histocompatibility complex (MHC) [16]. In particular, the aplotype comprising C4AQ0, DR3/DR52a and DQA2 had been associated with pulmonary fibrosis in SSc patients [17].

Fietta et al. [18] compared BALF from SSc patients with and without lung disease by 2-D with MALDI-TOF and HPLC MS analysis. Among these proteins, a fragment of mtDNAtopo 1 was found only in BALF from SSc patients with ILD, while glutathione S-transferase P (GST) and cystatin SN were detectable only in SSc patients without ILD (Table 1). In addition, increased levels of calgranulin B (S100A9), a molecule promoting extracellular matrix deposition and lung fibrosis, were found in BALF from SSc patients with ILD compared to SSc patients without lung involvement, similarly to what observed in BALF from patients with idiopathic pulmonary fibrosis.

In agreement with this study, Shirahama et al. [19] identified two groups of proteins which expression was either increased as  $\alpha 2$  magroglobulin,  $\alpha 1$  antitripsin and pulmonary surfactant protein A (SPA) or decreased as heat shock proteins (HSPs) and GST. Disregulation of these proteins levels might participate in the pathogenesis of lung fibrosis in SSc patients. In fact, abnormal levels of  $\alpha 2$  magroglobulin,  $\alpha 1$  antitripsin have been related to development of alveolitis in SSc patients [20], while SPA seems to play an important role in immunological surveillance [21]. In BALF from SSc patients, the reduction in HSPs and GST levels might indicate impaired protection in SSc lungs since HSPs are molecular chaperones involved in cell protection from injury and GST is an anti-oxidant protein [22].

A proteomic analysis using Reverse Phase-High Performance Liquid Chromatography-Electrospray-Mass Spectrometry in BALF of 46 scleroderma lungs and 15 controls, allowed us to reveal the abnormal presence of Thymosin β4, β4 sulfoxide and β10 in several SSc patients (Table 1). Thymosin  $\beta$ 4 levels were significantly higher in SSc patients than in controls, but patients experiencing a worsening in the alveolar score had relatively lower BALF thymosin  $\beta$ 4 levels [23]. These data are consistent with the ability of thymosin  $\beta$ 4 to down-regulate a number of key inflammatory cytokines, like the tumor necrosis factor-a. On the other hand, Thymosin  $\beta$ 4 sulfoxide levels were higher in smokers and SSc patients with alveolitis and analyzing the progression of lung disease at one-year follow-up, we found that higher thymosin β4 levels seem to have a protective role against lung tissue damage. Another component of the thymosin family, the thymosin \$10, was also present in BALF of SSc patients, yet not different from controls, but the negative correlation between thymosin  $\beta 10$  and the diffusion lung capacity for carbon monoxide, given the anti-angiogenic properties of thymosin  $\beta 10$ , suggests a potential inhibiting role of thymosin  $\beta 10$ on the alveolar-capillary barrier function [24]. To shed further light on the angio-biologic milieu, the levels and the alveolar macrophage expression of the vascular endothelial growth factor (VEGF) have been investigated in BALF of SSc patients. Patients with high PAH have capillary loss at the capillaroscopic analysis and high circulating serum-plasma levels of VEGF, suggesting that loss of capillaries and dysregulated angiogenesis both contribute to the tissue damage [25,26].

We found a statistically highly significant fall of VEGF levels in SSc patients compared to controls, a profound decrease of VEGF levels in alveolitis patients compared to the non-alveolitis ones, a strong direct correlation with the fibrotic score and an inverse correlation with neutrophil and eosinophil counts, confirming the hypothesis that the alveolo-capillary barrier can really be compromised by the lack of angiogenic factors [27].

In another study, Bargagli et al. [28] identified the macropage inhibitory factor (MIF) as multi-tasking cytokine in SSc patients with pulmonary involvement, using 2DE analysis of BALF validated by ELISA assay. The MIF expression was increased in bronchiolar epithelium and in area with active fibrosis, suggesting a possible role of MIF in progressive lung fibrosis.

Author(s)	<b>Biological material</b>	Proteomic approach	Differently expressed proteins
Rottoli P et al. [9]	BALF	2-DE	Protease, anti-protease Coagulation system Cytokines Anti-oxidant proteins Calcium binding proteins
Rottoli P et al. [10]	BALF	2-DE	Transferrin Immunoglobulin(s) Complement protein
Fietta AM et al. [18]	BALF	2DE MALDI-TOF HPLC-MS	Glutathione S-transferase P Cystatin SN
Bargagli E et al. [22]	BALF	2-DE	Calgranulin B (S100A9)
De Santis M et al. [23]	BALF	reverse phase HPLC coupled to quadrupole ion-trap MS -ESI	Thymosins β
Bargagli E et al. [28]	BALF	2-DE, ELISA	Macrophage Inhibitory Factor
Shirahama R et al. [19]	BALF	2-DE MALDI-TOF MS	α2 macroglobulin α1 antitripsin Pulmonary surfactant protein A Heat shock protein Glutathione S-transferase P
Bogatkevich GS et al. [29]	fibroblasts from BALF	2-DE/MS	Vimentin Tropomyosin Actin Disulfide isomerase Glutathione S-transferase

Table 1: Proteomic Studies on Bronchoalveolar Lavage Fluid from Patients with Ssc and Pulmonary Involvement.

Page 2 of 5

## Page 3 of 5

In a proteomic study on fibroblasts from BALF of SSc patients had been also identified 24 differently expressed protein spots, including cytoskeletal proteins (vimentin, tropomyosin and actin associated proteins) and proteins involved in redox imbalance (disulfide isomerase and GST), which may contribute to EC damage, fibroblast trafficking and tissue injury (Table 1) [29].

## Serum Interface for Proteomic Analysis of Soluble Biomarkers in SSc Patients with Lung Involvement

Serum represents an attractive biological fluid to study, since it is easily accessible and contains an enormous amount of proteins, which might be involved in the pathogenesis of SSc. In addition, serum sampling is less invasive than BALF collection.

In sera from SSc patients with lung involvement, the proteomic approach allowed the identification of new targets for anti-endothelial cell antibodies (AECA), which further support the role of EC damage as central event in the pathogenesis of the disease [30-32]. In particular, Negi et al. [32] observed a significantly higher incidence of PAH in AECA positive patients. Moreover, Tamby et al. [33] identified antigens from micro and macro-vascular endothelial cells, using 2DE and immunoblot analysis of the protein extracts from serum of patients with PAH. Recently, Dib et al. [34] identified other new target antigens for AECA, such as lamin A/C, tubulin  $\beta$  chain and vinculin, using the same proteomic approach.

The 2DE combined with immunoblot analysis of the sera from SSc patients with PAH allowed the identification of  $\alpha$ -enolase as target for anti-fibroblast antibodies, which might result from cross-reactivity after contact with either  $\alpha$ -enolase from microorganisms or tissue damage [35-37]. Another possible explanation is the reduced clearance of  $\alpha$ -enolase, which might be immunogenic but might also represent a substrate of caspase-1 that participates to activation of inflammasome (i.e.: IL-1 $\beta$ , NF-kB) [37,38].

In a study by 2DE combined with mass spectrometry and immunoblot analysis of sera from SSc patients with lung disease, Bussone et al. [39] identified new antigens involved in TGF- $\beta$ 1 pathway, which participate to fibroblast dysregulation and accumulation of extracellular matrix. Moreover, the recombinant antibody microarray permitted to identify mucin-1 and monocyte chemoatraction protein-4 (MCP-4) in serum of SSc patients, which might be the link between link inflammatory events and tissue fibrosis [40]. MCP-4 has been related to other different pathological contexts characterized by mononuclear infiltration, tissue remodeling and atherosclerosis [41,42]. Further functional studies should evaluate MCP-4 in a larger SSc patient population.

Using SELDI -TOF MS proteomic technological approach, van Bon et al. [43] identified increased levels of S100A8 (or calgranulin A) in sera from SSc patients with lung involvement, which might be a possible biomarker of chronic inflammation and progressive vascular injury.

By peptidomic approach, that combines a microamount peptideseparating method with magnetic beads and mass spectrometry analysis, Xiang et al. [44] detected in sera from SSc patients with ILD a group of short peptides with mass/charge values of 1,865, 1,778, 1,691, 1,563 and 1,450. These peptides had been identified as family members of C3f-des arginine (DRC3f) derived from C3b that might be a sign of complement system activation. Elevated levels of DRC3f and its degraded smaller fragments had been linked to vascular involvement and disease activity. In particular, the frequencies of ILD, sicca syndrome and esophageal involvement were significantly higher in patients with elevated DRC3f levels than those with normal DRC3f value. Interestingly, C3 and C4 serum levels were negatively correlated with DRC3f levels. Caccavo et al. [45] confirmed previous studies and reported an inverse correlation between high resolution CT scan (HRCT) score and serum levels of C3/C4 in SSc patients. These data had been documented in patients with autoantibodies directed against carbonic anhydrase I and/or II (CAI, CAII) and with HRCT score  $\geq$ 10. CA is a ubiquitous metalloenzyme that catalyzes the reversible hydration reaction of carbon dioxide to bicarbonate and hydrogen ions. In the lungs, CAII is mainly expressed by alveolar epithelium and is involved in respiratory gas exchange and pulmonary capillaries pH/ PCO, balance [46]. Antibodies anti-CAII were significantly increased in patients with ILD comparing to patients without ILD and healthy controls [47].

Recently, we carried out a comparative proteomic analysis of sera from SSc patients, that permitted us to identify 14 differentially expressed proteins mainly involved in EC protection and immune response [48]. We found increased concentrations of complement factor H in SSc subjects compared to healthy controls. Factor H is an important regulator of the alternative complement pathway, which normally protects self cellular surface from complement cascade. We documented a defective capacity of factor H to bind ECs and to protect them from complement mediated damage, especially in patients with pulmonary involvement. An aberrant expression of complement regulatory proteins had been already demonstrated in the endothelium of SSc patients [49] and it had been suggested that an inadequate protection from complement activation on cellular surface may be very important in the early phase of SSc disease. Complement activation leads to the formation of molecules that promote recruitment of inflammatory cells, generation of radical oxygen species and release of cytokines and chemokines, resulting in enhanced expression of EC adhesion molecules, apoptosis and tissue damage (Figure 1). The recruitment of circulating fibroblasts progenitor cells and their activation into the tissue are events that may also be facilitated by microvascular dysfunction [48,50].





Citation: Scambi C, Franceschi LD, Bosello S, Caramaschi P, Ferraccioli G, et al. (2012) Progress in Understanding the Pathogenesis of Systemic Sclerosis with Lung Involvement: The Contribution of Proteomic Studies. Rheumatology S1:004. doi:10.4172/2161-1149.S1-004

Page 4 of 5

#### Conclusions

The presented studies highlight complex and diverse pathways that intersect with each other and lead to tissue injury in the two major pulmonary syndromes associated with SSc. In general, these studies suggest an imbalance between aggressive and protective mechanisms, resulting in EC damage, amplified inflammatory response and fibroblast dysfunction in patients with lung involvement.

Although it has been provided important evidence on complicated pathogenetic mechanisms involved in SSc lung damage, further studies should be carried out to evaluate the effective role of the identified molecules, especially in early stages of the disease, in order to identify new therapeutic targets.

## **Rheumatology Key Messages**

- Complex pathogenic pathways are involved in SSc with lung damage.
- Proteomic analysis provides a better understanding of pathogenesis through the identification of new molecules.

#### References

- Nikpour M, Stevens WM, Herrick AL, Proudman SM (2010) Epidemiology of systemic sclerosis. Best Pract Res Clin Rheumatol 24: 857-869.
- Nietert PJ, Silver RM (2000) Systemic sclerosis: environmental and occupational risk factors. Curr Opin Rheumatol 12: 520-526.
- Wells AU, Steen V, Valentini G (2009) Pulmonary complications: one of the most challenging complications of systemic sclerosis. Rheumatology (Oxford) 40-44.
- Trad S, Amoura Z, Beigelman C, Haroche J, Costedoat N, et al. (2006) Pulmonary arterial hypertension is a major mortality factor in diffuse systemic sclerosis, independent of interstitial lung disease. Arthritis Rheum 54: 184-191.
- De Franceschi L, Bosello S, Scambi C, Biasi D, De Santis M, et al. (2011) Proteome analysis of biological fluids from autoimmune-rheumatological disorders. Proteomics Clin Appl 5: 78-89.
- De Santis M, Bosello S, La Torre G, Capuano A, Tolusso B, et al. (2005) Functional, radiological and biological markers of alveolitis and infections of the lower respiratory tract in patients with systemic sclerosis. Respir Res 6: 96.
- Sgonc R, Gruschwitz MS, Boeck G, Sepp N, Gruber J, et al. (2000) Endothelial cell apoptosis in systemic sclerosis is induced by antibody-dependent cellmediated cytotoxicity via CD95. Arthritis Rheum 43: 2550-2562.
- Kowal-Bielecka O, Kowal K, Highland KB, Silver RM (2010) Bronchoalveolar lavage fluid in scleroderma interstitial lung disease: technical aspects and clinical correlations: review of the literature. Semin Arthritis Rheum 40: 73-88.
- Rottoli P, Magi B, Cianti R, Vagaggini C, Nikiforakis N, et al. (2005) Carbonylated proteins in bronchoalveolar lavage of patients with sarcoidosis, pulmonary fibrosis associated with systemic sclerosis and idiopathic pulmonary fibrosis. Proteomics 5: 2612-2618.
- Rottoli P, Magi B, Perari MG, Liberatori S, Nikiforakis N, et al. (2005) Cytokine profile and proteome analysis in bronchoalveolar lavage of patients with sarcoidosis, pulmonary fibrosis associated with systemic sclerosis and idiopathic pulmonary fibrosis. Proteomics 5: 1423-1430.
- Bargagli E, Olivieri C, Bennett D, Prasse A, Muller-Quernheim J, et al. (2009) Oxidative stress in the pathogenesis of diffuse lung diseases: a review. Respir Med 103: 1245-1256.
- Meliconi R, Senaldi G, Sturani C, Galavotti V, Facchini A, et al. (1990) Complement activation products in idiopathic pulmonary fibrosis: relevance of fragment Ba to disease severity. Clin Immunol Immunopathol 57: 64-73.
- Cagatay T, Bilir M, Gulbaran M, Papila C, Cagatay P (2003) The immunoglobulin and complement levels in the active pulmonary sarcoidosis. Kobe J Med Sci 49: 99-106.
- 14. Sarma VJ, Huber-Lang M, Ward PA (2006) Complement in lung disease. Autoimmunity 39: 387-394.

- Ward PA (1997) Recruitment of inflammatory cells into lung: roles of cytokines, adhesion molecules, and complement. J Lab Clin Med 129: 400-404.
- Briggs DC, Welsh K, Pereira RS, Black CM (1986) A strong association between null alleles at the C4A locus in the major histocompatibility complex and systemic sclerosis. Arthritis Rheum 29: 1274-1277.
- Briggs D, Stephens C, Vaughan R, Welsh K, Black C (1993) A molecular and serologic analysis of the major histocompatibility complex and complement component C4 in systemic sclerosis. Arthritis Rheum 36: 943-954.
- 18. Fietta A, Bardoni A, Salvini R, Passadore I, Morosini M, et al. (2006) Analysis of bronchoalveolar lavage fluid proteome from systemic sclerosis patients with or without functional, clinical and radiological signs of lung fibrosis. Arthritis Res Ther 8: R160.
- Shirahama R, Miyazaki Y, Okamoto T, Inase N, Yoshizawa Y (2010) Proteome analysis of bronchoalveolar lavage fluid in lung fibrosis associated with systemic sclerosis. Allergol Int 59: 409-415.
- Bouros D, Wells AU, Nicholson AG, Colby TV, Polychronopoulos V, et al. (2002) Histopathologic subsets of fibrosing alveolitis in patients with systemic sclerosis and their relationship to outcome. Am J Respir Crit Care Med 165: 1581-1586.
- Wang G, Umstead TM, Phelps DS, Al-Mondhiry H, Floros J (2002) The effect of ozone exposure on the ability of human surfactant protein a variants to stimulate cytokine production. Environ Health Perspect 110: 79-84.
- Bargagli E, Olivieri C, Prasse A, Bianchi N, Magi B, et al. (2008) Calgranulin B (S100A9) levels in bronchoalveolar lavage fluid of patients with interstitial lung diseases. Inflammation 31: 351-354.
- De Santis M, Peluso G, Inzitari R, Peluso G, Fanali C, et al. (2009) Beta thymosins in scleroderma interstitial lung disease: Biomarkers of alveolitis. Ann Rheum Dis 68: 363.
- 24. De Santis M, Inzitari R, Bosello SL, Peluso G, Fanali C, et al. (2011) Î<sup>2</sup>-Thymosins and interstitial lung disease: study of a scleroderma cohort with a one-year follow-up. Respir Res 12: 22.
- 25. Hofstee HM, Vonk Noordegraaf A, Voskuyl AE, Dijkmans BA, Postmus PE, et al. (2009) Nailfold capillary density is associated with the presence and severity of pulmonary arterial hypertension in systemic sclerosis. Ann Rheum Dis 68: 191-195.
- Distler JH, Gay S, Distler O (2006) Angiogenesis and vasculogenesis in systemic sclerosis. Rheumatology (Oxford) 26-27.
- De Santis M, Bosello S, Capoluongo E, Inzitari R, Peluso G (2011) A vascular endothelial growth factor deficiency characterizes scleroderma lung disease. Manuscript under revision
- Bargagli E, Olivieri C, Nikiforakis N, Cintorino M, Magi B, et al. (2009) Analysis of macrophage migration inhibitory factor (MIF) in patients with idiopathic pulmonary fibrosis. Respir Physiol Neurobiol 167: 261-267.
- Bogatkevich GS, Ludwicka-Bradley A, Singleton CB, Bethard JR, Silver RM (2008) Proteomic analysis of CTGF-activated lung fibroblasts: identification of IQGAP1 as a key player in lung fibroblast migration. Am J Physiol Lung Cell Mol Physiol 295: L603-L611.
- Rosenbaum J, Pottinger BE, Woo P, Black CM, Loizou S, et al. (1988) Measurement and characterisation of circulating anti-endothelial cell IgG in connective tissue diseases. Clin Exp Immunol 72: 450-456.
- Hill MB, Phipps JL, Cartwright RJ, Milford Ward A, Greaves M, et al. (1996) Antibodies to membranes of endothelial cells and fibroblasts in scleroderma. Clin Exp Immunol 106: 491-497.
- Negi VS, Tripathy NK, Misra R, Nityanand S (1998) Antiendothelial cell antibodies in scleroderma correlate with severe digital ischemia and pulmonary arterial hypertension. J Rheumatol 25: 462-466.
- 33. Tamby MC, Chanseaud Y, Humbert M, Fermanian J, Guilpain P, et al. (2005) Anti-endothelial cell antibodies in idiopathic and systemic sclerosis associated pulmonary arterial hypertension. Thorax 60: 765-772.
- 34. Dib H, Tamby MC, Bussone G, Regent A, Berezné A, et al. (2011) Targets of anti-endothelial cell antibodies in pulmonary hypertension and scleroderma. Eur Respir J.
- 35. Terrier B, Degand N, Guilpain P, Servettaz A, Guillevin L, et al. (2007) Alphaenolase: a target of antibodies in infectious and autoimmune diseases. Autoimmun Rev 6: 176-182.

## Citation: Scambi C, Franceschi LD, Bosello S, Caramaschi P, Ferraccioli G, et al. (2012) Progress in Understanding the Pathogenesis of Systemic Sclerosis with Lung Involvement: The Contribution of Proteomic Studies. Rheumatology S1:004. doi:10.4172/2161-1149.S1-004

Page 5 of 5

- Terrier B, Tamby MC, Camoin L, Guilpain P, Broussard C, et al. (2008) Identification of target antigens of antifibroblast antibodies in pulmonary arterial hypertension. Am J Respir Crit Care Med 177: 1128-1134.
- 37. Pontillo A, Di Toro N, Edomi P, Shadlow A, Ammadeo A, et al. (2011) Anti-αenolase Antibodies in Serum from Pediatric Patients Affected by Inflammatory Diseases: Diagnostic and Pathogenetic Insights. Int J Rheumatol 2011: 870214.
- Shao W, Yeretssian G, Doiron K, Hussain SN, Saleh M (2007) The caspase-1 digestome identifies the glycolysis pathway as a target during infection and septic shock. J Biol Chem 282: 36321-36329.
- Bussone G, Mouthon L (2011) Interstitial lung disease in systemic sclerosis. Autoimmun Rev 10: 248-255.
- 40. Carlsson A, Wuttge DM, Ingvarsson J, Bengtsson AA, Sturfelt G, et al. (2011) Serum protein profiling of systemic lupus erythematosus and systemic sclerosis using recombinant antibody microarrays. Mol Cell Proteomics 10: M110.
- Barinka C, Prahl A, Lubkowski J (2008) Structure of human monocyte chemoattractant protein 4 (MCP-4/CCL13). Acta Crystallogr D Biol Crystallogr 64: 273-278.
- Breland UM, Michelsen AE, Skjelland M, Folkersen L, Krohg-Sørensen K, et al. (2010) Raised MCP-4 levels in symptomatic carotid atherosclerosis: an inflammatory link between platelet and monocyte activation. Cardiovasc Res 86: 265-273.
- 43. Van Bon L LA, Wittkowski H, van Heerde W, Vonk M, van den Berg W, et al. (2010) Poteomic analysis of systemic sclerosis serum identifies the toll-like receptor agonist S100A8/A9 as a novel possible pathogenic marker. J Transl Med 8: P67.

- 44. Xiang Y, Matsui T, Matsuo K, Shimada K, Tohma S, et al. (2007) Comprehensive investigation of disease-specific short peptides in sera from patients with systemic sclerosis: complement C3f-des-arginine, detected predominantly in systemic sclerosis sera, enhances proliferation of vascular endothelial cells. Arthritis Rheum 56: 2018-2030.
- 45. Caccavo D, Afeltra A, Rigon A, Vadacca M, Zobel BB, et al. (2008) Antibodies to carbonic anhydrase in patients with connective tissue diseases: relationship with lung involvement. Int J Immunopathol Pharmacol 21: 659-667.
- 46. Esbaugh AJ, Tufts BL (2006) The structure and function of carbonic anhydrase isozymes in the respiratory system of vertebrates. Respir Physiol Neurobiol 154: 185-198.
- Alessandri C, Bombardieri M, Scrivo R, Viganego F, Conti F, et al. (2003) Anticarbonic anhydrase II antibodies in systemic sclerosis: association with lung involvement. Autoimmunity 36: 85-89.
- 48. Scambi C, La Verde V, De Franceschi L, Barausse G, Poli F, et al. (2010) Comparative proteomic analysis of serum from patients with systemic sclerosis and sclerodermatous GVHD. Evidence of defective function of factor H. PLoS One 5: e12162.
- 49. Venneker GT, van den Hoogen FH, Boerbooms AM, Bos JD, Asghar SS (1994) Aberrant expression of membrane cofactor protein and decay-accelerating factor in the endothelium of patients with systemic sclerosis. A possible mechanism of vascular damage. Lab Invest 70: 830-835.
- Abraham DJ, Krieg T, Distler J, Distler O (2009) Overview of pathogenesis of systemic sclerosis. Rheumatology (Oxford) 3-7.

This article was originally published in a special issue, Lung Involvement in Scleroderma handled by Editor(s). Dr. Galina Bogatkevich, Medical University of South Carolina, USA