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Principles and Current Topics Concerning Management of Tyrosine Kinase Inhibitor Therapy for Chronic Myelogenous Leukemia

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Abstract

Chronic myeloid leukemia (CML) is a chronic myeloproliferative disorder with leukemic cells featuring the Philadelphia (Ph1) chromosome comprising the reciprocal chromosomal translocation t(9;22)(q34;q22) and the resultant constitutive active Bcr-Abl tyrosine kinase (TK). The introduction of disease-specific molecular targeted agents, that is, TK inhibitors (TKIs) for Bcr-Abl TK, such as the first-in-class TKI imatinib mesylate (IM) or the secondgeneration TKIs (SGIs) nilotinib and dasatinib, has dramatically improved the long-term treatment outcome for CML in this century. In addition, several new SGIs, such as bosutinib or bafetinib, are under development, while ponatinib, which is active against CML refractory for all preceding TKIs, for example, CML with T315I Abl kinase domain mutation, is currently being evaluated in clinical trials. Because those TKIs have different sensitivity for Bcr-Abl mutation and profiles for adverse effects, a thorough understanding of the pharmacologic characteristics of TKIs is mandatory for their safe and effective clinical use. Recent studies have clearly shown the faster and deeper responses to SGIs, both nilotinib and dasatinib, compared with those to IM, indicating the need for a paradigm shift in approaches to TKI therapy for treatment-naïve CML. In addition, evidence accumulated during the past decade has indicated optimal methodologies for monitoring the treatment effect of TKIs, selecting the appropriate therapeutic strategies, and predicting the outcome for treatment with TKIs for individual patients with CML. In this article, we review the principles and current knowledge and topics of the various uses of TKIs for CML. We also touch upon the reason why the faster and the deeper responses to TKIs is the prerequisite for their safer use and longer-lasting positive treatment outcome for CML.

Keywords: Chronic myelogenous leukemia; Tyrosine kinase inhibitor; Imatinib; Nilotinib; Dasatinib

Introduction

Chronic myeloid leukemia (CML) is a subtype of chronic myeloproliferative disorders with leukemic cells featuring the reciprocal chromosomal translocation t(9;22)(q34;q22), also known as the Philadelphia (Ph1) chromosome, and its resultant Bcr-Abl fusion oncoprotein, which exerts constitutive tyrosine kinase (TK) activity which in turn stimulates various downstream signaling cascades for deregulated cell proliferation, cell survival, resistance to cytotoxic stimuli and genetic instability [1,2]. Until a decade ago, long-term disease control was achieved only by allogeneic hematopoietic stem cell transplantation (HSCT) with a complete cure rate of about 50-60%, while the effects on CML of pharmacologic interventions, such as the use of interferon- α (IFN- α), were rather limited [2]. However, the introduction of tyrosine kinase inhibitors (TKIs) for Bcr-Abl TK, such as the first-in-class TKI imatinib mesylate (IM), as disease-specific molecular targeted agents has dramatically improved the long-term treatment outcome for CML in this century. The IRIS (International Randomized Study of Interferon and STI571) study of chronic phase CML (CML-CP) patients which compared the effects of IM and IFN-a plus cytarabine as first-line therapy found that continuous IM treatment resulted in greater treatment success with an eight-year overall survival (OS) rate of 93% and cumulative complete cytogenetic response (CCyR) of 83%, thus demonstrating the apparent advantage of firstline IM therapy for better long-term outcomes for CML patients [3,4]. Despite these excellent results, caution is needed when interpretating the findings of this study because the analysis was based on the data for patients who successfully continued IM treatment. Indeed, subsequent reports brought out the fact that approximately 30-40% of the patients initially enrolled in the IM cohort of the IRIS study dropped out from the trial due to unsatisfactory or no response or to intolerance for IM. Moreover, even among the patients whose data were included in the analysis, 17% did not attain CCyR and 10% showed disease progression [5]. In view of these findings, comparison of much longer-term effects, *e.g.*, over at least 20~30 years, to determine long-term outcomes for IM and allogeneic HSCT remain a matter for future studies. Nevertheless, the apparently better OS rate and the notably fewer short-term life-threatening adverse events attained with IM in comparison with allogeneic HSCT, the former has come to be regarded as the first-line treatment for CML, especially in the chronic phase (CP). Moreover, the development of second-generation TKIs (SGIs), such as dasatinib and nilotinib, which can overcome IM resistance and intolerance, has been a topic of heated debate concerning the treatment for CML during the past decade. Moreover, even newer agents, such as bosutinib, bafetinib and ponatinib, are currently being developed [6,7].

In the clinical setting, an in-depth understanding of the pharmacologic characteristics of each TKI, including its kinase inhibitory ability, affinitiy for mutated Abl, as well as adverse effects and predictors for clinical outcomes, is essential for choosing the most appropriate agent for individual CML patients. This article reviews the current knowledge which is essential for the effective use of Bcr-Abl TKIs and focuses especially on the effects and limitations of IM and SGIs for CML.

Positive and Negative Aspects of the use of IM for CML

The first report regarding the dramatic preclinical effect of

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CGP57148 (later renamed STI571 and then IM), an ATP-competitive TKI which is mostly specific for the Abl protein tyrosine kinase in Bcr-Abl-positive leukemias was announced by Dr. B.J. Druker in 1996 [8]. IM inhibits the constitutive active status of autophosphorylated Abl by binding to the ATP-binding site of the inactive conformation of Bcr-Abl in which the kinase-active site is shielded by the activation loop (A-loop) [9], resulting in blocking of Bcr-Abl TK-mediated downstream signaling for leukemogenesis. In addition to Bcr-Abl TK, IM also inhibits, and even more strongly, c-KIT and PDGF-R [10,11]. As a consequence, IM inhibits cell proliferation and induces programmed cell death mainly *via* apoptosis employing Bim and Bad as the crucial pro-apoptotic BH3-only proteins [12].

Since the introduction of IM, its development as a therapeutic agent has progressed very quickly, and within a couple of years IM gained its position as the first-line therapeutic pharmacologic intervention. Besides its major benefits for CML patients, the successful development has provided new insights for a better understanding of the disease biology. Also, among the many scientific contributions made by the introduction of IM, the major progress made in research in identification of the molecular mechanisms involved in TKI resistance has provided numerous scientific insights into the development of new molecular targeted agents for CML as well as other types of cancerous diseases. However, it was found that the mechanisms for resistance against IM are different from those against conventional genotoxic agents. As early as one year after the clinical approval of IM for CML, Sawyer's group identified two major mechanisms for IM resistance, T315I genomic point mutation in Abl kinase domain (KD), which results in the conformational change of the IM binding site, and bcrabl gene amplification, which results in Bcr-Abl overexpression [13]. Soon after those epoch-making discoveries, other mechanisms for IM resistance, such as various types of Abl KD mutations [14,15], Bcr-Abl overexpression due to transcriptional upregulation or the activation of alternative kinases such as Lyn, which promotes leukemic cell proliferation and survival, were also identified [16,17]. At present, more than 100 types of Abl KD mutations have been identified as causative for modest to complete resistance against IM, and it has been demonstrated that one or more clones with mutated Abl KD may be present in approximately half of all IM-resistant CML patients. The underlying molecular mechanisms for IM resistance vary depending on the type of Abl KD mutation. For instance, T315 is essential for the hydrogen bond between IM and Abl KD, so that mutations in T315, such as T315I or T351A, result in interference with this action. Moreover, mutations in T315 lead to conformational changes in adjunctive amino acids such as K271, E286M, R362, or D381, which are also crucial for IM-binding to Abl KD. Among mutations in the phosphate-binding loop (P-loop), those at Y253, such as Y253H, also disrupt the hydrogen bond between IM and Y253, while other types of P-loop mutations, such as G250E or E255K, stabilize Bcr-Abl as an active form which is insensitive to IM. Mutations in the catalytic loop (C-loop), such as M351T or E355G, also cause conformational changes in an inactive form of Abl, thereby preventing the binding of IM. More recent studies have also identified other Bcr-Abl-unrelated mechanisms for TKI resistance, such as abnormal influx/efflux pump expression [18,19], multifactorial support by bone marrow microenvironment [20-22], deregulation of programmed cell death machineries [12,23-25], or drug insensitivity due to leukemic stem cell phenotype [26-28]. All these molecular mechanisms combine to promote IM resistance.

The other problem is adverse events caused by IM, which are most likely associated with its inhibitory effect on off-target kinases. Hematological adverse events, one of the most frequent complications of IM treatment, may be associated with the inhibitory effect of IM on c-KIT, while some of the non-hematologic adverse events, such as skin rash, myalgia, edema, or fluid retention, may be associated with its inhibitory effect on PDGF-R. In the IRIS study, 8% of the patients dropped out from the study during the 6-year follow-up due to intolerance to IM [5].

Development of sgis and their characteristics

To overcome the major mechanisms for IM resistance, several SGIs with more flexiblity in response to Abl KD conformational changes, higher affinity for more abundant Bcr-Abl and ability to block various leukemogenic kinases have been developed for or introduced to the treatment for CML. As of this writing, nilotinib and dasatinib have been approved for clinical use not only as second-line but also as first-line treatment, while bosutinib, bafetinib and ponatinib are undergoing clinical trials. As shown in Table 1, all SGIs possess much higher affinity for wild type Bcr-Abl TK [29-33]. While nilotinib potently inhibits the inactive form of Bcr-Abl, other SGIs are potently affect both active and inactive forms of Bcr-Abl [34]. Nilotinib is more highly selective for Bcr-Abl, and its effects on c-KIT and PDGF-R are much weaker than that on Bcr-Abl, so that it can be expected that its adverse events due to the inhibition of off-target kinases will be far less frequent than those associated with IM. While dasatinib and bosutinib exert multikinase inhibitory effects and bafetinib potently inhibits Lyn, a member of the Src kinase proteins, bosutinib does not inhibit c-KIT. Whether this insensitivity of bosutinib for c-KIT influences its anti-CML effect has not been clarified, but caution is advisable because the findings of several basic studies suggest that the concomitant blockade of Bcr-Abl and c-KIT is essential for suppressing Bcr-Abl-positive leukemias [35,36]. All SGIs possess different affinity (in vitro kinase inhibitory) profiles for mutated Abl KD. For instance, nilotinib is not effective for P-loop mutations and dasatinib not for F317 mutations [37]. Importantly, none of the TKIs except ponatinib is at all potent in inhibiting T315I, while the newest TKI ponatinib has been developed to overcome T315I mutated Bcr-Abl and has shown favorable clinical effects for CML patients who were resistant to previous TKIs regardless of the presence of T315I mutation. Thus, appropriate choice of an agent according to the type of Abl KD mutation is mandatory, especially when it is to be used as the second-line treatment for IM-resistant CML [38].

Milestone for IM treatment of treatment-naïve CM

Accumulating evidence has provided clues for information relevant for predicting long-term outcome of IM treatment as first-line treatment for CML. The European LeukemiaNet (ELN) has established guidelines for IM therapy for patients with newly diagnosed CML-CP, which make it possible to judge whether the continuation of IM treatment is optimal for individual patients at several time points during the treatment course [39]. These guidelines proposed in 2006 classify the response status into "Optimal", "Suboptimal" and "Failure" for individual patients according to the degree of response to IM, i.e., hematologic response (HR), cytogenetic response (CyR), and molecular response (MR) at 3, 6, 12 and 18 months of IM treatment (Table 2). In addition to these three classifications, the criterion "Warnings" is suggested for patients showing any additional chromosomal abnormalities in Philadelphia-positive leukemic cells at diagnosis, an increase in *bcr-abl* copy numbers at any time point, or the emergence of clones with abnormal karyotype in non-Philadelphia hematopoietic cells. More recently, the revised guidelines proposed in 2009 include an evaluation system for the effect of SGIs on IM-resistant CML-CP. In these revised guidelines, the response status has been classified into

	Imatinib	Dasatinib	Nilotinib	Bosutinib	Bafetinib	Ponatinib
Bcr-Abl conformation	Inactive only	Active & Inactive	Inactive only	Active & Inactive	Active & Inactive	Inactive only
Kinase inhibitory activity	× 1	x 100-325	x 10-30	x 50-200	x 25-55	X 200
Resistant mutations	> 100 types	V299L, T315I, T315A, F317L, F317I,	Y253H, E255K, T315I F359C, F359V	T315I, E255K, V299L	T315I	E255V?
Other target kinases	PDGFRs c-Kit	> 50 kinases	PDGFRs, ARG, c-Kit, EPHR	Lyn, Hck, Fgr, EPHR	PDGFRs, c-Kit, Lyn	PDGFR, c-Kit, Lyn, FGFR1, VEGFR
Advantage		Strongest affinity	Low cross intolerance	Not target c-Kit or PDGFs	Low cross intolerance	Effective for T315I
Disadvantage		Off target	P-loop			

 Table 1: Characteristics of tyrosine kinase inhibitors for chronic myelogenous leukemia.

Time	Failure	Suboptimal response	Failure
Diagnosis	NA	NA	High risk, del9q, ACAs in Ph cells
3M	No HR (stable disease or disease progression)	Less than CHR	NA
6M	Less than CHR, no CyR (Ph >95%)	Less than PCCyR (Ph >35%)	NA
12M	Less than PCCyR (Ph >35%)	Less than CCyR	Less than MMR
18M	Less than CCyR	Less than MMR	NA
At any time	Loss of CHR*, loss of CCyR†, mutation‡	ACA in Ph cells \S , loss of MMR \S , mutation	Any rise in transcript level; other chromosome abnormalities in Ph cells

Failure : the patient should be moved to other treatments, if available

Suboptimal response : the patient may still derive substantial benefit from continued IM treatment but because the long-term outcome is not likely to be optimal, the patient should become eligible for other treatments.

Warnings: the patient should be monitored very carefully and may become eligible for other

treatments. *To be confirmed on 2 occasions unless associated with progression to AP/BC.

To be confirmed on 2 occasions, unless associated with CHR loss or progression to AP/BC.

#High level of insensitivity to IM.

\$ To be confirmed on 2 occasions, unless associated with CHR or CCgR loss.

Low level of insensitivity to IM.

 Table 2: European LeukemiaNet consensus guidelines for imatinib treatment for treatment-naïve chronic myelogenous leukemia.

"Suboptimal", "Failure" and "Warnings", while "Optimal" was not included due to the lack of convincing evidence for the long-term effects of SGIs on IM-resistant patients [40]. Indeed, although promising as salvage therapy, the effects of SGIs on IM-resistant CML-CP patients have been reported to be much inferior to those on treatment-naïve CML-CP cases [41,42]. According to these guidelines, patients who do not achieve any CyR at 3 months, partial CyR (PCyR) at 6 months, or major molecular response (MMR) at 12 months are evaluated as "Failure".

The faster and deeper the response, the better the outcome may be

In brief, the ELN consensus guidelines propose that the faster and the deeper the response to IM treatment is, the more favorable the prediction is for long-term disease control of CML. In contrast, a brief look at the results of the IRIS study, which evidenced a continuous increase in the study population with cumulative incidence of CCyR as the observation period became longer, may justify expectations for the growing presence of late responders to continuous IM treatment. Indeed, in the pre-SGI era, we continued IM treatment, unless there was evidence of disease progression, even for so-called "non-optimal" responders in the hope they would turn out to be "late responders". However, several studies have clearly shown the risk for late responders to IM by demonstrating that, according to Quintas-Cardama *et al.*, late responders to IM are major candidates for future loss of response to IM [43]. These researchers revealed that, when compared with patients who achieved CCyR within 12 months of IM treatment, those who attained CCyR later were at significantly higher risks of disease progression, such as the loss of CCyR, after 60 months of treatment [43]. Similarly, de Lavallade etal. reported that failure to achieve CCyR after 12 months of IM treatment is significantly associated with later disease progression, especially after 48 months of treatment with IM. In addition to the timing of response, the depth of response is also strongly associated with long-term outcome [44]. Even for patients with CCyR within 12 to 18 months of treatment with IM, attainment of major molecular response (MMR) was significantly associated with longer maintenance of CCyR [45]. Hughes et al. stressed the importance of earlier attainment of MMR with IM treatment, showing that patients with *bcr-abl* transcripts > 10% at 6 months and > 1% at 12 months had inferior EFS and higher rate of disease progression. Their study also showed that patients who attained MMR by 18 months enjoyed markedly long-lasting responses without disease progression and with 95% EFS at 7 years [46].

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To identify the basis of the need for faster and deeper effect of TKIs for patients with CML-CP, we need to briefly review the natural history of CML. CML starts with the acquisition of clonal proliferative property by acquiring Bcr-Abl TK activity in the very early phase of the disease (early CP). This is followed during the next 5 to 10 years by various additional chromosomal and oncogenic changes unrelated to Bcr-Abl, which have a cumulative effect in a multistep manner due to genetic instability caused by Bcr-Abl TK. During this process, leukemic cells, which initially show strong addiction to Bcr-Abl TK activity for their proliferation and survival, undergo a clonal evolution and loose their dependence on Bcr-Abl TK. This evolution then results in disease progression from CP to the advanced phases, that is, the accelerated and blast crisis phases (AP and BC) [47,48]. The more and the longer residual Bcr-Abl-positive leukemic cells survive during TKI treatment, the more frequently mutations for clonal evolution are likely to occur. Once leukemic cells that are less addicted to Bcr-Abl TK emerge, they may at first create chimeras with Bcr-Abl TK-dependent clones, but later develop into major clones during TKI treatment, and eventually cause the TKI treatment to fail. In view of this scenario, it makes sense to try and reduce the chance for clonal evolution by suppressing Bcr-Abl-positive clones as much and as fast as possible with TKIs to achieve the so-called "safe haven" status. The advisability of this strategy is supported by the fact that the maintenance of CCyR for more than 3 years is associated with the longer event free survival (EFS) and a reduction in the occurrence of future adverse events from 5.4% to 0.3%[5].

SGIs as first-line and second-line treatment

On the basis of the principle that the faster and deeper the response, the better the outcome and the greater the pharmacological potency may be, how can we best take advantage of the potency of SGIs? This question may already have been answered by the findings of two large prospective trials, ENESTnd (Evaluating nilotinib efficacy and safety in clinical trials - newly diagnosed patients), which conducted a head-tohead comparison of the effects of nilitinib and IM [49] and DASISION (Dasatinib versus imatinib study in treatment-naïve CML), which conducted a head-to-head comparison of the effects of dasatinib and IM for treatment-naïve CML-CP [50]. Both studies clearly showed the faster and deeper effects of SGIs in terms of achievement of CCyR and MMR, while SGIs induced complete MR (CMR) in more patients. After 18 months of treatment, nilotinib and dasatinib had induced CMR in 21% and 13% of patients, respectively, while IM had induced CMR in only 6 to 7%. Similar results have been reported by trials conducted by GIMEMA (Gruppo Italiano Malattie Ematologiche dell'Adulto) or the M.D. Anderson Cancer Center [51]. Importantly, these studies indicated that the faster and the deeper the effects of SGIs were, the more they eventually resulted in less disease progression and longer OS periods. In the ENESTnd trial, disease progression was observed in 1/283 patients and the 12-month OS ratio was 99.3% for the nilotinib cohort, while the corresponding findings for the IM cohort were 11/283 patients and 96.9%. In the DASISION trial, disease progression at 12 months was observed in 5/269 patients in the dasatinib cohort and in 9/260 patients in the IM cohort. Further, the rates for drug cessation due to intolerance were not significantly different for IM and nilotinib or dasatinib. These data constitute sufficiently supportive evidence for the use of nilotinib and dasatinib as the first-line treatment for CML-CP. In contrast, the utility of bosutinib as the first-line treatment for CML-CP remains controversial. In the BELA (Bosutinib efficacy and safety in newly diagnosed chronic myeloid leukemia) trial, achievement of CCyR with bosutinib was not significantly superior to that with IM at 12 months (70% vs 68%), although the achievement of MMR was better for the bosutinib arm than for the IM arm (39% vs 26%). Moreover, drug cessation due to adverse events, such as severe diarrhea, was notably more frequent for the bosutinib arm than for the IM arm (19% vs 5%) [51].

In contrast to the findings for nilotinib and dasatinib as first-line treatment, their use as second-line treatment following IM treatment was not as effective. According to the results of START-C trial, after 2 months of treatment with dasatinib, CCyR and MMR had been achieved in, respectively, 45% and 37% of IM-resistant, and 78% and 78% of IM-intolerant patients with CML-CP [42]. These effects were much inferior to those obtained for treatment-naïve CML-CP, and, as was to be expected, the effects of dasatinib were rather limited for advanced phase CML [52-54]. Similarly, the effect of nilotinib as second-line treatment is also much inferior to that observed in treatment-naïve patients. The CCyR rates after 24 months of treatment with nilotinib for IM-resistant and IM-intolerant CML-CP patients were reportedly 41% and 51%, respectively, while the MMR rate obtained with nilotinib for these patients was as high as 28% [55]. These results imply that there is a need for predictors for response to second-line SGIs for the development of alternative treatment strategies.

Current topics concerning the use of TKIs based on 10-year experience

Accumulating evidence has yielded important information for ways of monitoring treatment response and choice of treatment but also have given rise to new clinical questions. The next 7 sections deal with several topics regarding the use of TKIs based on a decade of experience.

1. **Optimal timing for the switch from IM to sgis:** It may be easy to decide this switch in the case of primary "Failure" for treatment with IM according to the ENL guidelines. The switch may be also not so difficult in case of intolerance IM, because cross-intolerance between IM and SGIs occurs infrequently [56,57]. The question that remains is the optimal timing for the switch from IM to SGIs in case of the loss of response in patients with initial optimal response. Kantarjian *et al.* reported that the effect of dasatinib as the second-line treatment for patients who have lost response to IM is affected by the timing of the drug switch. They classified patients into two groups according to whether they received dasatinib after the loss of major CyR (MCyR) or after the loss of McyR with concomitant loss of hematologic response (HR) when treated with IM. Although second-line dasatinib induced similar rates of complete HR (CHR)

in both groups, it induced CCyR in 72% of the first and in 42% of the second group, and 2-year progression-free survival (PFS) of 89% and 29%, respectively. These trends were not influenced by the presence of Abl KD mutations except for that of T315I [58]. These results suggest that an earlier treatment switch is recommended, at least for patients who experienced secondary "Failure" for IM.

- 2. Increase in bcr-abl copy numbers and emergence of Abl KD mutated clone despite sustained ccyr: There is a high risk of failure to achieve CCyR with TKI treatment is with the occurrence of certain events, such as an increase in bcr-abl copy numbers, emergence of Abl KD mutation and the eventual disease progression. What exactly are the effects of such events in spite of sustained CCyR? Studies so far have suggested that an elevation of bcr-abl copy numbers, including the loss of MMR, during TKI treatment is frequently associated with the emergence of a clone with mutated Abl KD even in patients with CCyR. Kantarjian et al. reported that 116 (including 40 without MMR) of 258 patients with CCyR showed definite elevation of bcrabl copy numbers, and that 48 of the 76 patients with MMR lost it during IM treatment. Importantly, disease progression was observed only when the increase in bcr-abl copy number exceeded the threshold level of MMR. In contrast, the loss of MMR was found to be associated with disease progression in 6/48 patients, even though these patients maintained their CCyR [59]. It is therefore important to assess the presence of Abl KD mutation and to consider a switch to different TKIs when bcr-abl copy numbers increase, especially in case of loss of MMR.
- 3. Biomarkers for clinical outcome of treatment-naïve CML in TKI treatment (except Abl KD mutations and *bcr-abl* copy numbers): Several biomarkers have been identified to be predictive for the clinical outcome of treatment-naïve CML patients in TKI treatment. While the Sokal score was originally established as a prediction model to predict survival of patients with conventional chemotherapies, IRIS study revealed a correlation between Sokal score and response to IM [3]. Some in vitro assays are also informative for the prediction of outcome of CML in TKI treatment. Among substrates of Bcr-Abl TK, the phosphorylation status of CrkL has been shown to be informative for the prediction of treatment outcome. White et al. has demonstrated that the relationship between the in vitro concentration of IM needed to reduce the level of phosphorylated CrkL (pCrkL) and the achievement of MMR, i.e, the lower IC50 was correlated with the higher MMR [60]. Moreover, patients who had greater than 50% in vivo inhibition of pCrkL within the first month of treatment had increased rates of MMR at 12 and 24 months [61]. IM plasma trough concentration also significantly associates with treatment outcome, namely, a minimum of trough concentration of 1002 ng/ml was associated with and optimal response [62]. The activity of drug influx pump, hOCT1, at diagnosis has been also shown to be associated with the treatment outcome of CML, when treated with IM, but not SGIs. According to two clinical trials, TIDEL (Trial of Imatinib with Dose Escalation) and TOPS (Tyrosine Kinase Inhibitor Optimization and Selectivity), it was found that MMR rates of patients with high hOCT1 activity and patients with low hOCT1 acitivty by 5 year treatment with IM were 89% and 55%, respectively [18].
- 4. Impact of load of Abl mutated clones on effects of second-line sgis: Because the *in vitro* kinase inhibitory profiles of SGIs differ, it is essential to screen Abl KD mutation to determine the optimal choice of SGIs as second-line treatment for IM-resistant patients [63,64]. An Australian group reported interesting results about this issue at the Annual Meeting of the American Society of Hematology

in 2010, showing that the load of SGI-resistant Abl KD mutated clones at baseline affects the response to SGIs. CCyR was infrequent in patients with an SGI-resistant Abl mutated clone identifiable by direct sequencing (DS) (sensitivity: 10-20%), while its frequency was 16% for patients whose clone was identifiable by high throughput chip-based mass spectrometry (Mass Spec) (sensitivity: 0.05-0.5%) after 18 months of SGI treatment. In addition, expansion of Mass Spec-detectable minor clones co-occurring with SGI-resistant Abl KD mutation was detected in 84% of patients during SGI treatment. In contrast, the CCyR rate was 42% for patients with an SGI-non-resistant Abl KD mutated clone regardless of the load of mutated clones.

The next question is whether the presence of a clone with an SGI-non-resistant Abl mutation at baseline affects the course of treatment with SGIs. In the Australian study mentioned above, it was reported that even SGI-non-resistant clones increased in 12% of patients during treatment with SGIs. Moreover, even in cases with SGI-non-resistant Abl KD mutation(s) at baseline, SGI-resistant Abl KD mutated clones emerged in 34% of patients with a single SGIsensitive Abl KD mutation and in 67% of patients with two or more different clones with SGI-sensitive Abl KD mutations. As a result, the CCyR rate for patients with a single clone was 67% and was reduced to 14% for patients with two or more different clones. At the same meeting, groups from MD Anderson Cancer Center reported further impairment of PFS and OS in patients with multiple Abl KD mutations. From the view of clonal evolution, the presence of Abl KD mutated clones indicates not only sensitivity to SGI but also to what degree leukemic cells have already acquired genetic instability at the time of investigation. Putting these findings together indicates that the identification of Abl KD mutation is indispensable, but does not satisfactorily predict the effects of SGIs. Regardless of in vitro sensitivity to SGIs, the presence of Abl KD mutations implies a genetic instability which may impair the effect of SGIs. Close monitoring of the effects of SGIs is thus required under such circumstances.

- 5. Factors other than Abl KD mutation which impact the effects of sgis as second-line treatment: The in vitro kinase inhibitory profile cannot fully determine the clinical outcome of second-line SGI treatment. No effects of SGIs can of course be expected in the presence of T315I mutation, while the CyR rates attainable in the clinical setting with second-line SGIs are not significantly different for patients with SGI-moderately sensitive and patients with SGIsensitive Abl KD mutation, indicating that factors other than Abl KD mutations also play a role in determining clinical outcomes [37,65,66]. Accumulating evidence has made it clear that poor risk factors for second-line SGIs include non-MCyR with IM at 12 months, more than 4% basophils in peripheral leukocytes at the switch to SGIs, hemoglobin less than 12g/dL, the presence of additional chromosomal abnormality, poor Sokal score, or neutropenia during IM treatment [67,68]. All these poor risk factors show a positive association with clonal evolution and disease progression in CML, and the evidence presented here suggests the importance of disease control with TKIs whenever CML clones prove to be addictive to Bcr-Abl TK.
- 6. **Monitoring of effect of second-line sgis during treatment:** The long-term effect of second-line SGI is also predictable by monitoring the effect of ongoing SGI treatment. Because the effects of SGIs as second-line treatment are limited, it is essential to determine whether the ongoing SGI treatment is appropriate. Several findings have demonstrated the importance of treatment response after 3 months of SGI therapy. Hughes et al. demonstrated that the monitoring of

bcr-abl copy numbers can help predict possible attainment of CCyR and MMR by using second-line SGIs. In this study, the rates of CCyR and MMR for patients who showed reduction of *bcr-abl* copy numbers to $\leq 1\%$, 1~10% and >10% after 3 months of SGI treatment were 91% and 86%, 56% and 55%, and 11% and 4%, respectively, after 24 months of SGI treatment [69]. Another study showed that the attainment of at least minor CyR after 3 months of SGI treatment was associated with later attainment of MCyR and better OS and EFS [70]. These findings may prove useful for deciding whether to switch treatment to another SGI or even to allogeneic HSCT.

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- 7. The fight against T315I: At the timing of writing, no FDA-approved Abl-specific TKIs had been proven effective for leukemic cells with T315I Abl KD mutation, so that allogeneic HSCT remains one of the most effective strategies against T315I [71,72]. However, several new agents designed to fight T315I are under development. One of the most promising of these agents is ponatinib (formerly known as AP24534). Ponatinib is similar to IM and nilotinib in terms of its limited binding ability to the inactive conformation of Bcr-Abl but was developed as a scaffold that did not need to create a hydrogen bond with T315, and would therefore be effective for inhibiting Abl KD with T315I mutation. Another difference from IM and nilotinib is that ponatinib is active against P-loop mutations, such as E255K, Y253F, or G250, and, while it is effective against Src, Lyn, c-KIT, VEGFR2, PDGF-R and FGFR1, it is most effective against wild type Abl among a series of target kinases [33]. Cortes et al. reported the results of a finalized Phase I study of the efficacy of ponatinib for patients with advanced Philadelphia-positive leukemias, including 64 CML patients refractory to one to three TKIs and 10 with Philadelphia-positive acute lymphoblastic leukemia (ALL) [7]. Of the 38 CML-CP patients, 66% attained MCyR and 53% CCyR, of the 12 CML-CP patients with T315I, all attained CHR and MCyR and 89% CCyR, and MMR was attained by 42% of CML-CP patients, including 7 with T315I. Finally, adverse events were tolerable in most cases. A phase II study of ponatinib (Ponatinib Ph+ acute lymphoblastic leukemia (ALL) and CML Evaluation or PACE) is currently under way. Moreover, several other agents, such as DCC-2036, a switch pocket inhibitor [73], or danusertib (PHA-739358), XL-228 and AT-9283, aurora kinase inhibitors, are being developed as potent therapeutics for IM/SGI-refractory CML, including patients with T315I Abl KD mutation [7,74]. Danusertib is a pan Aurora and Abl inhibitor and has been evaluated in a Phase I study for 12 advanced phase CML and 11 Ph1-positive ALL patients, including 15 with T315I mutation. Danusertib induced hematologic response in five and cytogenetic (any) response in three patients in those series. XL-228, a multi-kinase inhibitor for Aurora A, Abl, IFG-1R, Src and Lyn, has been also evaluated in Phase I study for 27 patients with CML and Ph1-positive ALL, 10 of which had T315I mutation. In this trial, objective responses have been reported in some patients, including patients with T315I. The effect of the pan-Aurora, Abl, FLT3 and JAK2 inhibitor AT9283 has been also reported against CML patients who were refractory to IM and dasatinib [74]. The final results of those trials are awaited.
- 8. **Complete cure with tkis?** A recent study by a French group of patients who maintained complete MR (CMR) for more than two years by IM treatment has shown the possibility of cure by IM treatment for some patients. Even in case of relapse following the cessation of IM treatment, the reintroduction of IM was effective for those patients. A higher probability of sustained CMR following the cessation of IM treatment was observed for patients with a low Sokal risk score and longer IM treatment (≧ 50 months) [75]. These results

suggest two important hypotheses. One is that low-risk, perhaps early phase, CML-CP, which is highly addictive to Bcr-Abl TK, may be curable with intense and long-term TKI treatment. The other is that even for non-cured patients, the long-term and deep suppression of leukemic clones may reduce the risk of clonal evolution which may cause the loss of ddiction to Bcr-Abl TK activity. The faster and the deeper curative effects of SGIs on newly diagnosed CML-CP clearly point to the urgent need to investigate the potential potency of SGIs for a complete cure of CML.

Future perspective of pharmacologic interventions for cure of CML

Finally, we address the possible new treatment strategies by the combination of TKI and other anti-leukemic agents for cure of CML. Rationally, a strategy which blocks Bcr-Abl signaling as the main pathway using TKIs and the simultaneous blockade of collateral signaling pathway by partner agents may provide the best chance for disease elimination. Especially, combination strategies which completely eliminate leukemic stem cells (LSCs) may be desirable, because CML LCSs are insensitive to IM and persist as the cause of relapse. Considering this aspect, the combination of agents those potentially target molecules essential for LSC survival and maintenance may be of interest. Homoharringtonine (HHT), a Chinese evergreenderived alkaloid cephalotaxine, simultaneously inhibits Mcl-1 and β-catenin, those are crucial for the survival of LSCs, and it indeed effectively kills LSCs in preclinical model [77,78]. According to the result of the trial for the combination of IM, G-CSF and HHT against advanced phase IM-resistant CML, this combinatory strategy was most likely feasible [76]. It will be also interesting to look at the combinatory effects of IM and HHT for CML-CP. Also, the combination of IM and decitabine, a DNA methyltransferase inhibitor, has been shown to be well-tolerable and active against advanced phase CML [79]. The effects of DNA methyl transferase inhibitors on the gene expression modification in LSCs are expected to enhance the clinical effect of IM in CML-CP. Farnesyl transferase inhibitor (FTI) which potentially inactivates Ras-related proteins and MEK inhibitor have been shown to be active against CML LCSs [80], and the combination of lonafartinib, a member of FTIs, and IM has been shown to be well-tolerated against CML patients who failed IM monotherapy in Phase I trial [81]. It would be of interest to investigate the effect of this combination against CML-CP as the first-line setting.

Conclusion

In this article, we reviewed the current knowledge regarding TKI treatment for CML. The success of TKI treatment for CML constitutes the paradigm of molecular targeted therapy for malignant diseases and has shown the way towards the development of new treatment for cancers as well as how problems involving new therapeutics can be tackled. Making further progress on the basis of the new and updated information is becoming increasingly important for both clinicians and researchers.

References

- Rumpold H, Webersinke G (2011) Molecular pathogenesis of Philadelphiapositive chronic myeloid leukemia - is it all BCR-ABL? . Curr Cancer Drug Targets. 11: 3-19.
- Goldman JM (2010) Chronic myeloid leukemia: a historical perspective. Semin Hematol. 47: 302-311.
- Druker BJ, Guilhot F, O'Brien SG, Gathmann I, Kantarjian H et al. (2006) Fiveyear follow-up of patients receiving imatinib for chronic myeloid leukemia. N Engl J Med. 355: 2408-2417.

- O'Brien SG, Guilhot F, Larson RA, Gathmann I, Baccarani M et al. (2003) Imatinib compared with interferon and low-dose cytarabine for newly diagnosed chronic-phase chronic myeloid leukemia. N Engl J Med 348: 994-1004.
- Hochhaus A, O'Brien SG, Guilhot F, Druker BJ, Branford S et al. (2009) Sixyear follow-up of patients receiving imatinib for the first-line treatment of chronic myeloid leukemia. Leukemia. 23: 1054-1061.
- O'Hare T, Deininger MW, Eide CA, Clackson T, Druker BJ (2011) Targeting the BCR-ABL signaling pathway in therapy-resistant Philadelphia chromosomepositive leukemia. Clin Cancer Res 17:212-221.
- Santos FP, Quintás-Cardama A (2011) New drugs for chronic myelogenous leukemia. Curr Hematol Malig Rep 6:96-103.
- Druker BJ, Tamura S, Buchdunger E, Ohno S, Segal GM et al. (1996) Effects of a selective inhibitor of the Abl tyrosine kinase on the growth of Bcr-Abl positive cells. Nat Med 2: 561-566.
- Schindler T, Bornmann W, Pellicena P, Miller WT, Clarkson B et al. (2000) Structural mechanism for STI-571 inhibition of abelson tyrosine kinase. Science 289: 1938-1942.
- Heinrich MC, Griffith DJ, Druker BJ, Wait CL, Ott KA et al (2000) Inhibition of c-kit receptor tyrosine kinase activity by STI 571, a selective tyrosine kinase inhibitor. Blood 96: 925-932.
- Buchdunger E, Cioffi CL, Law N, Stover D, Ohno-Jones S Abl(2000) proteintyrosine kinase inhibitor STI571 inhibits in vitro signal transduction mediated by c-kit and platelet-derived growth factor receptors. J Pharmacol Exp Ther 295: 139-145.
- Kuroda J, Puthalakath H, Cragg MS, Kelly PN, Bouillet P et al (2006) Bim and Bad mediate imatinib-induced killing of Bcr/Abl+ leukemic cells, and resistance due to their loss is overcome by a BH3 mimetic. Proc Natl Acad Sci U S A 103: 14907-14912.
- Gorre ME, Mohammed M, Ellwood K, Hsu N, Paquette R et al. (2001) Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. Science 293: 876-880.
- Hochhaus A, La Rosée P, Müller MC, Ernst T, Cross NC (2011) Impact of BCR-ABL mutations on patients with chronic myeloid leukemia. Cell Cycle 10: 250-260.
- Nardi V, Azam M, Daley GQ (2004) Mechanisms and implications of imatinib resistance mutations in BCR-ABL. Curr Opin Hematol 11: 35-43.
- Nimmanapalli R, O'Bryan E, Huang M, Bali P, Burnette PK et al. Molecular characterization and sensitivity of STI-571 (imatinib mesylate, Gleevec)resistant, Bcr-Abl-positive, human acute leukemia cells to SRC kinase inhibitor PD180970 and 17-allylamino-17-demethoxygeldanamycin. (2002) Cancer Res 62: 5761-5769.
- Dai Y, Rahmani M, Corey SJ, Dent P, Grant S A (2004) Bcr/Abl-independent, Lyn-dependent form of imatinib mesylate (STI-571) resistance is associated with altered expression of Bcl-2. J Biol Chem 279: 34227-34239.
- White DL, Dang P, Engler J, Frede A, Zrim S et al. (2010) Functional activity of the OCT-1 protein is predictive of long-term outcome in patients with chronicphase chronic myeloid leukemia treated with imatinib. J Clin Oncol 28: 2761-2767.
- Kuroda J, Kimura S, Segawa H, Kobayashi Y, Yoshikawa T et al. (2003) The third-generation bisphosphonate zoledronate synergistically augments the anti-Ph+ leukemia activity of imatinib mesylate. Blood 102: 2229-2235.
- Jin L, Tabe Y, Konoplev S, Xu Y, Leysath CE et al. (2008) CXCR4 up-regulation by imatinib induces chronic myelogenous leukemia (CML) cell migration to bone marrow stroma and promotes survival of quiescent CML cells. Mol Cancer Ther 7: 48-58.
- Filippi I, Naldini A, Carraro F (2011) Role of the hypoxic microenvironment in the antitumor activity of tyrosine kinase inhibitors. Curr Med Chem 18:2885-2892.
- Nair RR, Tolentino J, Hazlehurst LA (2010) The bone marrow microenvironment as a sanctuary for minimal residual disease in CML. Biochem Pharmacol 80: 602-612.
- San José-Eneriz E, Agirre X, Jiménez-Velasco A, Cordeu L (2009) Epigenetic down-regulation of BIM expression is associated with reduced optimal responses to imatinib treatment in chronic myeloid leukaemia. Eur J Cancer 45:1877-1889.
- 24. Kamitsuji Y, Kuroda J, Kimura S, Toyokuni S, Watanabe K (2008) The Bcr-Abl

Page 6 of 8

kinase inhibitor INNO-406 induces autophagy and different modes of cell death execution in Bcr-Abl-positive leukemias. Cell Death Differ 15: 1712-1722.

- 25. Bellodi C, Lidonnici MR, Hamilton A, Helgason GV, Soliera AR et al. (2009) Targeting autophagy potentiates tyrosine kinase inhibitor-induced cell death in Philadelphia chromosome-positive cells, including primary CML stem cells. J Clin Invest 119: 1109-1123.
- 26. Holtz MS, Forman SJ, Bhatia R (2005) Nonproliferating CML CD34+ progenitors are resistant to apoptosis induced by a wide range of proapoptotic stimuli. Leukemia 19: 1034-1041.
- 27. Chen Y, Peng C, Sullivan C, Li D, Li S (2010) Critical molecular pathways in cancer stem cells of chronic myeloid leukemia. Leukemia. 24: 1545-1554.
- Nicholson E, Holyoake T. The chronic myeloid leukemia stem cell. (2009) Clin Lymphoma Myeloma 9: S376-381.
- 29. Shah NP, Tran C, Lee FY, Chen P, Norris D et al. CL (2004) Overriding imatinib resistance with a novel ABL kinase inhibitor. Science 305: 399-401.
- Weisberg E, Manley PW, Breitenstein W, Brüggen J, Cowan-Jacob SW (2005) Characterization of AMN107, a selective inhibitor of native and mutant Bcr-Abl. Cancer Cell. 7: 129-141.
- 31. Golas JM, Arndt K, Etienne C, Lucas J, Nardin D (2003) SKI-606, a 4-anilino-3-quinolinecarbonitrile dual inhibitor of Src and Abl kinases, is a potent antiproliferative agent against chronic myelogenous leukemia cells in culture and causes regression of K562 xenografts in nude mice. Cancer Res 63: 375-381.
- Kimura S, Naito H, Segawa H, Kuroda J, Yuasa T et al (2005) . NS-187, a potent and selective dual Bcr-Abl/Lyn tyrosine kinase inhibitor, is a novel agent for imatinib-resistant leukemia. Blood 106: 3948-3954.
- 33. O'Hare T, Shakespeare WC, Zhu X, Eide CA, Rivera VM et al. (2009) AP24534, a pan-BCR-ABL inhibitor for chronic myeloid leukemia, potently inhibits the T315I mutant and overcomes mutation-based resistance. Cancer Cell 16: 401-412.
- 34. Tokarski JS, Newitt JA, Chang CY, Cheng JD, Wittekind M et al. (2006) The structure of Dasatinib (BMS-354825) bound to activated ABL kinase domain elucidates its inhibitory activity against imatinib-resistant ABL mutants. Cancer Res 66: 5790-5797.
- Belloc F, Airiau K, Jeanneteau M, Garcia M, Guérin E et al. (2009) The stem cell factor-c-KIT pathway must be inhibited to enable apoptosis induced by BCR-ABL inhibitors in chronic myelogenous leukemia cells. Leukemia 23: 679-685.
- Wong S, McLaughlin J, Cheng D, Zhang C, Shokat KM et al. (2004) Sole BCR-ABL inhibition is insufficient to eliminate all myeloproliferative disorder cell populations. Proc Natl Acad Sci U S A 101(50):17456-17461.
- 37. Jabbour E, Kantarjian HM, Jones D, Reddy N, O'Brien Set al. (2008) Characteristics and outcome of chronic myeloid leukemia patients with F317L BCR-ABL kinase domain mutation after therapy with tyrosine kinase inhibitors. Blood 112: 4839-4842.
- Jabbour E, Soverini S (2009) Understanding the role of mutations in therapeutic decision making for chronic myeloid leukemia. Semin Hematol 46: S22-26.
- 39. Baccarani M, Saglio G, Goldman J, Hochhaus A, Simonsson B et al.(2006) Evolving concepts in the management of chronic myeloid leukemia: recommendations from an expert panel on behalf of the European LeukemiaNet. Blood 108: 1809-1820.
- Baccarani M, Cortes J, Pane F, Niederwieser D, Saglio G et al. (2009) Chronic myeloid leukemia: an update of concepts and management recommendations of European LeukemiaNet. J Clin Oncol 2009 27: 6041-6051.
- 41. Kantarjian HM, Giles F, Gattermann N, Bhalla K, Alimena G et al.(2007) Nilotinib (formerly AMN107), a highly selective BCR-ABL tyrosine kinase inhibitor, is effective in patients with Philadelphia chromosome-positive chronic myelogenous leukemia in chronic phase following imatinib resistance and intolerance. Blood 110: 3540-3546.
- 42. Hochhaus A, Kantarjian HM, Baccarani M, Lipton JH, Apperley JF et al. (2007) Dasatinib induces notable hematologic and cytogenetic responses in chronicphase chronic myeloid leukemia after failure of imatinib therapy. Blood 109: 2303-2309.
- 43. Quintás-Cardama A, Kantarjian H, Jones D, Shan J, Borthakur G et al.(2009) Delayed achievement of cytogenetic and molecular response is associated with increased risk of progression among patients with chronic myeloid leukemia in early chronic phase receiving high-dose or standard-dose imatinib therapy. Blood 113: 6315-6321.

44. de Lavallade H, Apperley JF, Khorashad JS, Milojkovic D, Reid AG et al.(2008) Imatinib for newly diagnosed patients with chronic myeloid leukemia: incidence of sustained responses in an intention-to-treat analysis. J Clin Oncol 26: 3358-3363.

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- 45. Marin D, Milojkovic D, Olavarria E, Khorashad JS, de Lavallade H et al.(2008) European LeukemiaNet criteria for failure or suboptimal response reliably identify patients with CML in early chronic phase treated with imatinib whose eventual outcome is poor. Blood. 112: 4437-4444.
- 46. Hughes TP, Hochhaus A, Branford S, Müller MC, Kaeda JS et al.(2010) Long-term prognostic significance of early molecular response to imatinib in newly diagnosed chronic myeloid leukemia: an analysis from the International Randomized Study of Interferon and STI571 (IRIS). Blood 116: 3758-3765.
- Cortes J, O'Dwyer ME Clonal evolution in chronic myelogenous leukemia. Hematol Oncol Clin North Am 18: 671-684.
- Radich JP, Dai H, Mao M, Oehler V, Schelter J et al. (2006) Gene expression changes associated with progression and response in chronic myeloid leukemia. Proc Natl Acad Sci U S A 103: 2794-2799.
- Saglio G, Kim DW, Issaragrisil S, le Coutre P, Etienne G et al. (2010) Nilotinib versus imatinib for newly diagnosed chronic myeloid leukemia.N Engl J Med 362: 2251-2259.
- Kantarjian H, Shah NP, Hochhaus A, Cortes J, Shah S et al. (2010) Dasatinib versus imatinib in newly diagnosed chronic-phase chronic myeloid leukemia. N Engl J Med 362: 2260-2270.
- Kantarjian HM, Baccarani M, Jabbour E, Saglio G, Cortes JE (2011) Second-Generation Tyrosine Kinase Inhibitors: The Future of Frontline CML Therapy. Clin Cancer Res 17: 1674-1683.
- 52. Kantarjian H, Pasquini R, Lévy V, Jootar S, Holowiecki J et al.(2009) Dasatinib or high-dose imatinib for chronic-phase chronic myeloid leukemia resistant to imatinib at a dose of 400 to 600 milligrams daily: two-year follow-up of a randomized phase 2 study (START-R). Cancer 115: 4136-4147.
- Apperley JF, Cortes JE, Kim DW, Roy L, Roboz GJ et al. (2009) Dasatinib in the treatment of chronic myeloid leukemia in accelerated phase after imatinib failure: the START a trial. J Clin Oncol 27: 3472-3479.
- Quintás-Cardama A, Cortes JE, O'Brien S, Ravandi F, Borthakur G et al. (2009) Dasatinib early intervention after cytogenetic or hematologic resistance to imatinib in patients with chronic myeloid leukemia. Cancer. 115: 2912-2921.
- Kantarjian H, Giles F, Wunderle L, Bhalla K, O'Brien S et al. (2006) Nilotinib in imatinib-resistant CML and Philadelphia chromosome-positive ALL. N Engl J Med 354: 2542-2551.
- Cortes JE, Hochhaus A, le Coutre PD, Rosti G, Pinilla-Ibarz J et al. (2011) Minimal cross-intolerance with nilotinib in patients with chronic myeloid leukemia in chronic or accelerated phase who are intolerant to imatinib. Blood 117: 5600-5606.
- 57. Kobayashi Y, Sakamaki H, Fujisawa S, Ando K, Yamamoto K et al. Lack of nonhematological cross intolerance of dasatinib to imatinib in imatinib-intolerant patients with Philadelphia chromosome positive chronic myeloid leukemia or acute lymphatic leukemia: a retrospective safety analysis. Int J Hematol 93: 745-749.
- Kantarjian H, O'Brien S, Talpaz M, Borthakur G, Ravandi F et al. (2007) Outcome of patients with Philadelphia chromosome-positive chronic myelogenous leukemia post-imatinib mesylate failure. Cancer 109: 1556-1560.
- 59. Kantarjian HM, Shan J, Jones D, O'Brien S, Rios MB et al. Significance of increasing levels of minimal residual disease in patients with Philadelphia chromosome-positive chronic myelogenous leukemia in complete cytogenetic response. J Clin Oncol 27: 3659-3663.
- White D, Saunders V, Lyons AB, Branford S, Grigg Aet al. (2005) In vitro sensitivity to imatinib-induced inhibition of ABL kinase activity is predictive of molecular response in patients with de novo CML. Blood 106: 2520-2526.
- White D, Saunders V, Grigg A, Arthur C, Filshie R et al. (2007) Measurement of in vivo BCR-ABL kinase inhibition to monitor imatinib-induced target blockade and predict response in chronic myeloid leukemia. J Clin Oncol 25: 4445-4451.
- Picard S, Titier K, Etienne G, Teilhet E, Duci et al.(2007) Trough imatinib plasma levels are associated with both cytogenetic and molecular responses to standard-dose imatinib in chronic myeloid leukemia.Blood 109: 3496-3499.
- Hughes T, Saglio G, Branford S, Soverini S, Kim DW et al.(2009) Impact of baseline BCR-ABL mutations on response to nilotinib in patients with chronic myeloid leukemia in chronic phase. J Clin Oncol 27: 4204-4210.

Page 8 of 8

- 64. Jabbour E, Jones D, Kantarjian HM, O'Brien S, Tam C, Koller C et al. (2009) Long-term outcome of patients with chronic myeloid leukemia treated with second-generation tyrosine kinase inhibitors after imatinib failure is predicted by the in vitro sensitivity of BCR-ABL kinase domain mutations. Blood 114: 2037-2043.
- Branford S, Melo JV, Hughes TP (2009) Selecting optimal second-line tyrosine kinase inhibitor therapy for chronic myeloid leukemia patients after imatinib failure: does the BCR-ABL mutation status really matter? Blood 114: 5426-5435.
- 66. Ahn JS, Kim YK, Lee SR, Yu L, Yang DH et al. (2010) Coexisting with clonal evolution and BCR-ABL mutant in CML patients treated with second-generation tyrosine kinase inhibitors predict the discrepancy of in vitro drug sensitivity. Cancer Res Treat 42: 37-41.
- 67. Milojkovic D, Nicholson E, Apperley JF, Holyoake TL, Shepherd P et al.(2010) Early prediction of success or failure of treatment with secondgeneration tyrosine kinase inhibitors in patients with chronic myeloid leukemia. Haematologica 95: 224-231.
- 68. Kim TD, Türkmen S, Schwarz M, Koca G, Nogai H et al.(2010) Impact of additional chromosomal aberrations and BCR-ABL kinase domain mutations on the response to nilotinib in Philadelphia chromosome-positive chronic myeloid leukemia. Haematologica 95: 582-588
- Hughes TP, Branford S (2009) Monitoring disease response to tyrosine kinase inhibitor therapy in CML. Hematology Am Soc Hematol Educ Program: 477-487.
- 70. Tam CS, Kantarjian H, Garcia-Manero G, Borthakur G, O'Brien S et al. (2008) Failure to achieve a major cytogenetic response by 12 months defines inadequate response in patients receiving nilotinib or dasatinib as second or subsequent line therapy for chronic myeloid leukemia. Blood 112: 516-518.
- 71. Jabbour E, Cortes J, Santos FP, Jones D, O'Brien S et al. (2011) Results of allogeneic hematopoietic stem cell transplantation for chronic myelogenous leukemia patients who failed tyrosine kinase inhibitors after developing BCR-ABL1 kinase domain mutations. Blood 117: 3641-3647.
- 72. Yamamoto M, Kuroda J, Uchiyama H, Kobayashi T, Tsutsumi Y et al.(2010) Allogenic bone marrow transplantation with fludarabine/busulfan16 conditioning regimen and dasatinib maintenance therapy for elderly Philadelphia-positive acute/advanced leukemia patients. Leuk Res 34: e111-112.

- Chan WW, Wise SC, Kaufman MD, Ahn YM, Ensinger CI et al.(2011) Conformational Control Inhibition of the BCR-ABL1 Tyrosine Kinase, Including the Gatekeeper T315I Mutant, by the Switch-Control Inhibitor DCC-2036. Cancer Cell 19: 556-568.
- Moore AS, Blagg J, Linardopoulos S, Pearson AD (2010) Aurora kinase inhibitors: novel small molecules with promising activity in acute myeloid and Philadelphia-positive leukemias. Leukemia 24: 671-678.
- 75. Mahon FX, Réa D, Guilhot J, Guilhot F, Huguet F et al.(2010) Discontinuation of imatinib in patients with chronic myeloid leukaemia who have maintained complete molecular remission for at least 2 years: the prospective, multicentre Stop Imatinib (STIM) trial. Lancet Oncol 11: 1029-1035.
- 76. Fang B, Li N, Song Y, Han Q, Zhao RC (2010) Standard-dose imatinib plus low-dose homoharringtonine and granulocyte colony-stimulating factor is an effective induction therapy for patients with chronic myeloid leukemia in myeloid blast crisis who have failed prior single-agent therapy with imatinib. Ann Hematol 89: 1099-1105.
- Kuroda J, Kamitsuji Y, Kimura S, Ashihara E, Kawata E et al.(2008) Antimyeloma effect of homoharringtonine with concomitant targeting of the myeloma-promoting molecules, Mcl-1, XIAP, and beta-catenin. Int J Hematol 87: 507-515.
- Chen Y, Hu Y, Michaels S, Segal D, Brown D et al. (2009) Inhibitory effects of omacetaxine on leukemic stem cells and BCR-ABL-induced chronic myeloid leukemia and acute lymphoblastic leukemia in mice. Leukemia 23: 1446-1454.
- 79. Oki Y, Kantarjian HM, Gharibyan V, Jones D, O'brien S et al. (2007) Phase II study of low-dose decitabine in combination with imatinib mesylate in patients with accelerated or myeloid blastic phase of chronic myelogenous leukemia. Cancer 109: 899-906.
- Pellicano F, Simara P, Sinclair A, Helgason GV, Copland M et al.(2011) The MEK inhibitor PD184352 enhances BMS-214662-induced apoptosis in CD34+ CML stem/progenitor cells. Leukemia 25: 1159-1167.
- Cortes J, Jabbour E, Daley GQ, O'Brien S, Verstovsek S (2007) Phase 1 study of lonafarnib (SCH 66336) and imatinib mesylate in patients with chronic myeloid leukemia who have failed prior single-agent therapy with imatinib. Cancer 110: 1295-1302.

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