

Prevalence and Multiplex PCR for Enterotoxin Genes of *Staphylococcus aureus* Isolates from Subclinical Mastitis and Kareish Cheese

Walid Saad Mousa^{1*}, Eman abdeen², Heba Hussein³ and Ghada Hadad⁴

¹Department of Animal Medicine and Infectious Diseases, Faculty of Veterinary Medicine, University of Sadat City, Egypt

²Department of Bacteriology, Mycology and Immunology, Faculty of Veterinary Medicine, University of Sadat City, Egypt

³Department of Food Hygiene and Control, Faculty of Veterinary Medicine, University of Sadat City, Egypt

⁴Department of Hygiene and Preventive Medicine (zoonoses), Faculty of Veterinary Medicine, University of Sadat City, Egypt

*Corresponding author: Walid Saad Mousa, Department of Animal Medicine and Infectious Diseases, Faculty of Veterinary Medicine, University of Sadat City, 32897, Egypt, Tel: 00201094647551; E-mail: walidsaadvet@yahoo.com

Received date: December 05, 2017; Accepted date: December 15, 2017; Published date: December 21, 2017

Copyright: ©2017 Saad Mousa W, et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Abstract

Aim: *Staphylococcus aureus* was categorized as a contagious pathogen incriminated in subclinical mastitis as well as in dairy products. From public health view, this organism causing food poisoning outbreaks *via* contamination of food products with its toxins. These studies highlight the prevalence of *staphylococcus aureus* among subclinical mastitic cases as well as cheese samples and the dominant enterotoxigenic genes.

Methodology: Examination of 100 samples (50 from subclinical mastitis, and 50 kareish cheese) from Sadat city, Menoufia province.

Results and interpretation: Bacteriological culturing on selective and specific medium revealed that 26.7% and 30% in subclinical mastitis and kareish cheese respectively were due to *S. aureus*. Furthermore, multiplex PCR proved to be efficient technique for detection of different enterotoxins genes. The *sea*, *seb* and *see* genes were the most prevalent genes among the tested *S. aureus* isolates. Although, no detection of *sec* and *sed* genes were observed.

Conclusion: It can be concluded that molecular characterization of *S. aureus* and its enterotoxins genes will be beneficial in designing control and preventive measures of *S. aureus* infection in human and animals.

Keywords: Enterotoxins; Food; PCR; *S. aureus*; Subclinical mastitis

Introduction

Staphylococcus aureus was responsible for wide range of illness in human and animals. In the dairy industry, it causes about 50% of intramammary infection [1,2]. The losses due to *S. aureus* subclinical mastitis includes higher losses in milk production, increase culling rate, veterinary and treatment cost [3]. This organism was frequently inhabitant in the skin of udder microbiota [4] under special circumstances, invade and colonized into udder tissues of dairy animals leading to inflammation and may produce serious form of mastitis [5]. Regarding food poisoning outbreaks, enterotoxins of *S. aureus* considered the most famous detecting in foods poisoning in human [6]. Thus assured *via* consumption of contaminated food products and milk products by *S. aureus* toxins [7]. In generally, early study of [8] reported that 14% to 40% of all human food poisoning outbreaks were associated with *staphylococcal* enterotoxins, that manifested at first by sudden onset of fever, vomiting, nausea, abdominal cramps and diarrhea [9,10]. The existence of some virulence factors as enzymes, toxins, surface antigens, and capsule have an important role in *S. aureus* pathogenicity as in intramammary infection, it depends on the induction of immunosuppression [11] through stimulating of the proinflammatory cytokines and T-cell that evoked the inflammatory features. One of these factors is enterotoxins

which proved to be more tolerance to pasteurization and higher temperature [12]. The enterotoxins constitute and reported as an important virulence agent involved in food poisoning and toxic shock syndrome [13]. The most commonly identified genes were (*SEA*, *SEB*, *SEC*, *SED*, and *SEE*) [14]. In Brazilian study the higher frequency and detection of both *see* and *seb* genes in clinical and subclinical mastitis [15]. However, other studies [16,17] have been described a new genes such as (*SEG*, *SHE*, *SEI*, *SEJ*, *SEK*, *SEL*, *SEM*, *SEN*, and *SEO*). Therefore this study aimed to estimate the prevalence and applying of multiplex PCR for identifying of enterotoxin genes from *S. aureus* isolates from subclinical mastitis and kareish cheese.

Material and Methods

Sampling and processing of collected specimens

Milk samples were collected aseptically in a 5 ml sterile plastic tubes (50 samples). Then samples examined for subclinical mastitis by CMT that done according to [18]. Mastitic milk samples were centrifuged and sediment was streaked on specific medium. Another 50 kareish cheese, were randomly collected from supermarkets in Sadat City. Each sample about 10 gm was collected in a sterile plastic bag. All collected samples were transported in cool ice boxes and transported into the laboratory for further investigation.

Isolation and identification of *S. aureus*

The processed samples from mastitic milk and kareish cheese were streaked into Baird-Parker agar (Oxoid Ltd.) and 10% sheep-blood agar. Plates were incubated for 24-48 h at 37°C. Then, examination of the suspected colonies of *S. aureus* was done through Gram staining as according [19]. The biochemical and virulence properties as catalase, coagulase

as determined by [20], haemolysis activity on blood agar and Deoxyribonuclease (DNase) testing onto DNase agar [21]

Multiplex PCR for detection of enterotoxin genes of *S. aureus* isolates

DNA extraction: The extraction of DNA from samples was performed using the QIA amp DNA Mini kit (Qiagen Germany, GmbH) with modifications from the manufacturer's recommendations. Briefly, 200 µl of the sample suspension was incubated with 10 µl of proteinase K and 200 µl of lysis buffer at 56°C for 10 min. After incubation, 200 µl of 100% ethanol was added to the lysate. The sample was then washed and centrifuged following the

manufacturer's recommendations. Nucleic acid was eluted with 100 µl of elution buffer provided in the kit.

Oligonucleotide primer: Primers used were supplied from Metabion (Germany) are listed in table (1).

For multiplex PCR of enterotoxins, Primers were applied in a 50 µl reaction containing 25 µl of Emerald Amp Max PCR Master Mix (Takara, Japan), 1 µl of each primer of 20 pmol concentration, 16 µl of water, and 7 µl of DNA template. The reaction was performed in an Applied biosystem 2720 thermal cycler.

Analysis of the PCR products

The products of PCR were separated by electrophoresis on 1.5% agarose gel (Applichem, Germany, GmbH) in 1x TBE buffer. For gel analysis, 40 µl of the multiplex PCR products were loaded in each gel slot. Gelpilot 100 bp DNA ladder (Qiagen, Germany, GmbH) was used to determine the fragment sizes. The gel was photographed by a gel documentation system (Alpha Innotech, Biometra) and the data was analyzed through computer software.

Target gene	Primers sequences	Amplified segment (bp)	Primary Denaturation	Amplification (35 cycles)			Final extension	Reference
				Secondary denaturation	Annealing	Extension		
Sea	GGTTATCAATGTGCGGGTGG	102	94°C 5 min.	94°C 30 S	50°C 45 S	72°C 45 S	72°C 10 min.	[22]
	CGGCACTTTTTTCTCTCCGG							
Seb	GTATGGTGGTGAAGTACGAGC	164						
	CCAAATAGTGACGAGTTAGG							
Sec	AGATGAAGTAGTTGATGTGATGG	451						
	CACACTTTTAGAATCAACCG							
Sed	CCAATAATAGGAGAAATAAAG	278						
	ATTGGTATTTTTTTCGTTTC							
See	AGGTTTTTTCACAGGTCATCC	209						
	CTTTTTTTCTTCGGTCAATC							

Table 1: Primers sequences, target genes, amplicon sizes and cycling conditions.

Results

The results of enterotoxin genes in *S. aureus* isolates from subclinical mastitis milk and kareish cheese illustrated in Table 2 and Table 3, revealed that *sea* and *see* enterotoxin genes were the most prevalent among 10 *S. aureus* isolates with 60% followed by *seb* (50%), while no detection for *sec* and *sed* enterotoxin genes in any tested samples. Samples from 1-4 isolated from subclinical mastitis while

5-10 from kareish cheese. In subclinical mastitis all the detected enterotoxin genes (*sea*, *seb* and *see*) with the same percentage 20%. In contrast, the *sea* and *see* enterotoxin detected in 40% followed by *seb* enterotoxin 30% in kareish cheese.

Multiplex PCR for detection of enterotoxins genes in *S. aureus* isolates from subclinical mastitis and kareish cheese.

Negative		<i>Staphylococcus aureus</i>		Positive species	Total number of samples	Source of samples
%	Number	%	Number			
				35	100	

73.35	11	26.7%	4	15	50	Sub clinical mastitis
70%	14	30%	6	20	50	Cheese

Table 2: Prevalence of *S. aureus* isolates from subclinical mastitis and kareish cheese.

Sample	Results				
	Sea	Seb	Sec	Sed	See
1	-	+	-	-	-
2	+	-	-	-	+
3	+	+	-	-	+
4	-	-	-	-	-
5	+	-	-	-	+
6	+	-	-	-	+
7	+	+	-	-	+
8	-	+	-	-	-
9	-	+	-	-	-
10	+	-	-	-	+

Table 3: Distribution and prevalence of enterotoxin genes among *S. aureus* isolates from subclinical mastitis milk and kareish cheese.

Discussion

Mastitis constitutes the major economic and productive disease in dairy cows all over the world. Mastitic milk can possess a serious hazard to human consumers due to higher bacterial count or toxins [23,24]. The incidence of subclinical mastitis form has become more prevalent 15-40 times than clinical form [23] so, this form of mastitis may act as continues source of new infection to mammary glands [25] that often ended by serious form of clinical mastitis [26]. One of the adverse effects on the milk quality due to subclinical mastitis is the decreasing of shelf life period of raw milk [27]. Many studies proved that *Staphylococcus aureus* is a major bacteria existed in bovine mastitis [28]. This study revealed that the prevalence rate of *S. aureus* from subclinical mastitis was 26.7%. Nearly similar findings in Egypt reported by [29,30] they estimated *S. aureus* prevalence were 28.2% and 29.16% respectively. Additionally, [31] recorded (29%) in Bangladesh. On the other hand, higher prevalence (55.58%) was reported in the study of [32]. However, [33] mentioned that approximately 30%-40% of all mastitis cases caused by *S. aureus*. In contrast, the prevalence rate in kareish cheese was 30% in our study.

These nearly in contact with [34] who detected *S. aureus* isolates in 27% from 200 milk and cheese samples in Iran. Although, a little lower prevalence rate 25% had been recovered from 100 Iranian white and feta cheese samples by [35]. The higher prevalence 75% and 46% has been obtained by [36,37] respectively. The lower prevalence (10%, 9.5%, and 7.7%) were observed in different countries in the studies [38-40] in Iran, turkey and European countries respectively. The observed variation in prevalence of *S. aureus* especially in milk products may be reflect the level of sanitary measures in milk manufacture or due to the differences in technological methods in

cheese manufacture[35]. Regarding *Staphylococcus* food poisoning that considered a major etiology of gastroenteritis in human [41]. This depends on the ability of *Staphylococcus aureus* to produce toxins or virulence factors that facilitated the disease occurrence [42]. The *S. auerus* enterotoxins (SE), particular SEA-SEE were the most classical discovered genes in cattle, sheep and goats milk [1,43] Showed that the host environment played a role in the adaptation of *S. aureus* in target host through production of SE. Modern molecular techniques as multiplex PCR has successfully adapted for detection of enterotoxins in *S. aureus* isolates from subclinical mastitis milk and Karich-cheese was done in this work as showed in figure 1 and table (3). It's clear that *sea* and *see* enterotoxin genes were the most prevalent among the tested *S. aureus* isolates followed by *seb*, while no detection for *sec* and *sed* enterotoxin genes.

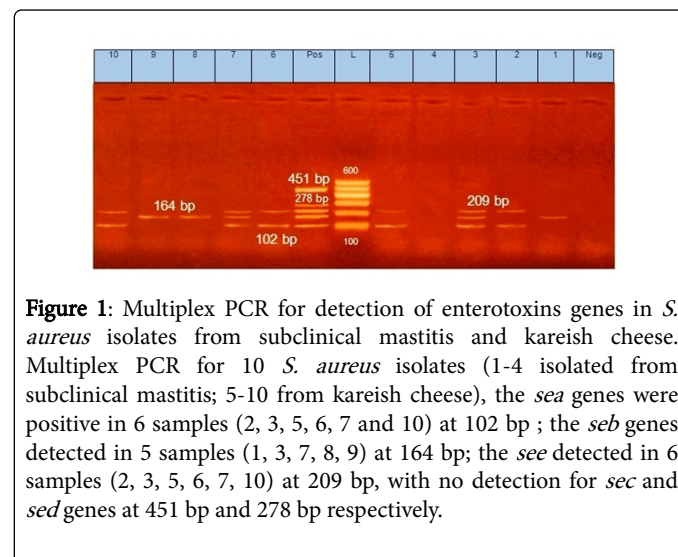


Figure 1: Multiplex PCR for detection of enterotoxins genes in *S. aureus* isolates from subclinical mastitis and kareish cheese. Multiplex PCR for 10 *S. aureus* isolates (1-4 isolated from subclinical mastitis; 5-10 from kareish cheese), the *sea* genes were positive in 6 samples (2, 3, 5, 6, 7 and 10) at 102 bp ; the *seb* genes detected in 5 samples (1, 3, 7, 8, 9) at 164 bp; the *see* detected in 6 samples (2, 3, 5, 6, 7, 10) at 209 bp, with no detection for *sec* and *sed* genes at 451 bp and 278 bp respectively.

In subclinical mastitis enterotoxin genes (*sea*, *seb* and *see*) were detected in 20%. In contrast, the *sea* and *see* enterotoxin detected in 40% followed by *seb* enterotoxin 30% in Karich-cheese. These were parallel to results of [44] who revealed the *sea* gene was the most prevalent gene, followed by *seb*, *sec*, *sed* and *see*. Furthermore, [45] found that *sea* gene was the prevalent SEs (36.7%) followed by *seb* (17.4%), *see* (16.5%), *sec-1* (11.01%), and *sed* (6.4%). Although, disconnected with [46] reported lower prevalence (1.6%) of *sea* gene. The absence of *sec* gene in our study can be explained that the *sec* gene has three subtypes (*sec1*, *sec2*, and *sec3*) that classified according to antigenic properties or the diversity in the sequencing of enterotoxin C [47]. There was no detection of *sed* gene in our findings. This almost observed in the result of [48] and [44] found that *sed* was the lowest dominant enterotoxins genes in very low percentage 0.5%*

Conclusion

Our results concluded that *Staphylococcus aureus* was identified as one of the contagious cause of intramammary infection in the dairy

farm as well as food poisoning outbreak worldwide. The pathogenicity of this organism relies on some virulence agents as enterotoxins that exert a toxic effect on host cell or in food products. Several enterotoxins genes, particularly, (*sea, seb, sec, sed and see*) genes were the most identified from *S. aureus* isolates of milk origin.. These genes appear to have a critical role in the pathogenicity of *S. aureus* in subclinical mastitis and food poisoning cases. Further studies about the using of these genes in genotyping of *S. aureus* isolates of milk origin are needed in future.

References

- Zschöck M, Kloppert B, Wolter W, Hamann HP, Lämmle CH (2005) Pattern of enterotoxin genes *seg, she, sei* and *sej* positive *Staphylococcus aureus* isolated from bovine mastitis. Vet Microbiol 108: 243-249.
- Taverna F, Negri A, Piccinini R, Zecconi A, Nonnis S, et al. (2007) Characterization of cell wall associated proteins of a *S aureus* isolated from bovine mastitis. Vet Microbiol 119: 240-247.
- Middleton JR, Fox LK, Gay JM, Tyler JW, Besser TE (2002) Use of pulsed-field gel electrophoresis for detecting differences in *Staphylococcus aureus* strain populations between dairy herds with different cattle importation practices. Epidemiol Infect 129: 387-395.
- Bobos S, Vidic B (2005) Mammary gland of ruminant's pathomorphology (in Serbian) Monography Poljoprivredni fakultet Novi Sad Naučni Institut za veterinarstvo "Novi Sad" Serbia Novi Sad.
- Rajic Savic N, Katic V, Velebit B (2014) Characteristics of coagulase positive *Staphylococci* isolated from milk in cases of subclinical mastitis. Acta Vet-Belgrade 64: 115-123.
- Gilmour A, Harvey J (1990) *Staphylococci* in milk and milk products. J Appl Bacteriol 69: 147-166.
- Chiang YC, Liao WW, Fan CM, Pai WY, Chiou CS, et al. (2008) PCR detection of *Staphylococcal* enterotoxins (SEs) N O P Q R U and survey of SE types in *Staphylococcus aureus* isolates from food-poisoning cases in Taiwan. Int J Food Microbiol 121: 66-73.
- Holmberg SD, Blake PA (1984) *Staphylococcal* food poisoning in the United States New facts and old misconceptions. JAMA 251: 487-489.
- Omoe K, Hu DL, Takahashi Omoe H, Nakane A, Shinagawa K (2003) Identification and characterization of a new *Staphylococcal* enterotoxin-related putative toxin encoded by two kinds of plasmids. Infect Immun 71: 6088-6094.
- Le Loir Y, Baron F, Gautier M (2003) *Staphylococcus aureus* and food poisoning. Genet Mol Res 2: 63.
- Akineden Ö, Annemüller C, Hassan AA, Lämmle C, Wolter W, et al. (2001) Toxin genes and other characteristics of *Staphylococcus aureus* isolates from milk of cows with mastitis. Clin Diagn Lab Immunol 8: 959-964.
- Asao T, Kumeda Y, Kawai T (2003) An extensive outbreak of *Staphylococcal* food poisoning due to low-fat milk in Japan: estimation of enterotoxin A in the incriminated milk and powdered skim milk. Epidemiol Infect 130: 33-40.
- Orwin PM, Fitzgerald JR, Leung DY, Gutierrez JA, Bohach GA, et al. (2003) Characterization of *Staphylococcus aureus* enterotoxin L. Infect Immun 71: 2916-2919.
- Letertre C, Perelle S, Dilasser F, Fach P (2003) Identification of a new putative enterotoxin SEU encoded by the *egc* cluster of *Staphylococcus aureus*. J Appl Microbiol 95: 38-43.
- Nader AF, Ferreira LM, Amaral LA (2007) Production of enterotoxins and toxic shock syndrome toxin by *Staphylococcus aureus* strains isolated from bovine mastitis. Arq Bras Med Vet Zootec 59: 1316-1318.
- Orwin PM, Leung DY, Donahue HL, Novick RP, Schlievert PM (2001) Biochemical and biological properties of *Staphylococcal* enterotoxin K. Infect Immun 69: 360-366.
- Omoe K, Hu DL, Omoe HT, Nakane A, Shinagawa K (2005) Comprehensive analysis of classical and newly described *Staphylococcal* superantigenic toxin genes in *Staphylococcus aureus* isolates. FEMS Microb Lett 246: 191-198.
- Schalm O, Noorlander O (1957) Experiments and observations leading to development of California Mastitis Test. J Am Vet Med Assoc 130: 199-204.
- Quinn PJ, Carter ME, Makrkey BK, Carter GR (1994) Clinical veterinary microbiology. Europ Limited Lynton House London 109-126.
- APHA (1992) Standard Methods for the Examination of dairy products. AGRIS.
- Murray PR, Baron EJ, Jorgensen JH (2003) Manual of Clinical Microbiology 8th ed. Clin Infec Dis 38: 1199-1200.
- Mehrotra M, Wang G, Johnson M (2000) Multiplex PCR for detection of genes for *Staphylococcus aureus* enterotoxins exfoliative toxins toxic shock syndrome toxin 1 and methicillin resistance. J Clin Microbiol 38: 1032-1035.
- Seegers H, Fourichon C, Beaudou F (2003) Production effects related to mastitis and mastitis economics in dairy cattle herds. Vet Res 34: 475-491.
- González RN, Wilson DJ (2003) Mycoplasmal mastitis in dairy herds. Vet Clin North Am Food Anim Pract 19: 199-221.
- Oliver SP, Gillespie BE, Headrick SJ, Moorehead H, Lunn P, et al. (2004) Efficacy of extended ceftiofur intramammary therapy for treatment of subclinical mastitis in lactating dairy cows. J Dairy Sci 87: 2393-2400.
- Reksen O, Solverød L, Branscum AJ, Osteras O (2006) Relationships between milk culture results and treatment for clinical mastitis or culling in Norwegian dairy cattle. J Dairy Sci 89: 2928-2937.
- Busato A, Trachsel P, Schällibaum M, Blum JW (2000) Udder health and risk factors for subclinical mastitis in organic dairy farms in Switzerland. Prev Vet Med 44: 205-220.
- Haran K, Godden SM, Boxrud D, Jawahir S, Bender JB, et al. (2011) Prevalence and Characterization of *Staphylococcus aureus* Including Methicillin-Resistant *Staphylococcus aureus* Isolated from Bulk Tank Milk from Minnesota Dairy Farms. J Clin Microbiol 50: 688-695.
- Enany ME, Younes S, AL gammal AM, Salem M, aEl Dieb HA (2013) Prevalence of coagulase (*coa*) gene and *mec A* gene of *S aureus* isolated from bovine clinical mastitis Seuz Canal. Vet Med 147-157.
- Eman Abdeen E, Mousa W, Heba H, Saher R (2015) PCR for detection of virulence and antibiotics resistance genes of *Staphylococcus aureus* from clinical mastitis in Egypt. Int J Bas App Sci 4: 315-319.
- Islam MA, Kabir SM, Rahman MT (2016) Molecular detection and characterization of *Staphylococcus aureus* isolated from raw milk sold in different markets of Bangladesh. Bangl J Vet Med 14: 277-282.
- Marija P, Stanko B, Branko V, Zoran R, Vera K, et al. (2016) Prevalence and molecular characterization of enterotoxin-producing strains of *Staphylococcus aureus* isolated from serbian dairy cows. Acta Veterinaria Beograd 66: 466-477.
- Bedane A, Kasim G, Yohannis T, Habtamu T, Asseged B, et al. (2012) Study on Prevalence and Risk Factors of Bovine Mastitis in Borana Pastoral and Agro-Pastoral Settings of Yabello District Borana Zone Southern Ethiopia American-Eurasian. J Agric and Environ Sci 12: 1274-1281.
- Saadat YR, Imani Fooladi AA, Shapouri R, Hosseini MM, Deilami Khiabani Z (2014) Prevalence of enterotoxigenic *Staphylococcus aureus* in organic milk and cheese in Tabriz, Iran. Iran J Microbiol 5: 345-359.
- Arefi F, Mohsenzadeh M, Razmyar J (2013) Isolation antimicrobial susceptibility and *mecA* gene analysis of methicillin-resistant *Staphylococcus aureus* in Iranian white cheeses. Iran J Vet Res Shiraz Uni 127-131.
- Jørgensen HJ, Mork T, Caugant DA, Kearns A, Rorvik LM (2005) Genetic Variation among *Staphylococcus aureus* Strains from Norwegian Bulk Milk. Appl Environ Microbiol 71: 8352-8361.
- Marhamatizadeh MH, Karim G, Nikafrooz R, Peikar J (2006) Survey on the white traditional cheese by *Staphylococcus aureus* in Kazeroun In: 16th National Congress of Iran. Food Industry Gorgan Iran 1-10.

38. Imani Fooladi AA, Tavakoli HR, Naderi A (2010) Detection of enterotoxigenic *Staphylococcus aureus* isolates in domestic dairy products. Iran J Microbiol 2: 137-142.
39. Can YH, Celik HT (2012) Detection of enterotoxigenic and antimicrobial resistant *Staphylococcus aureus* in Turkish cheese. Food Control 24: 100-103.
40. Akineden O, Hassan A, Schneider E, Usleber E (2008) Enterotoxigenic properties of *Staphylococcus aureus* isolated from goats milk cheese. Int J Food Microbiol 124: 211-216.
41. Scherrer D, Corti S, Muehlherr JE, Zweifel C, Stephan R (2004) Phenotypic and genotypic characteristics of *Staphylococcus aureus* isolates from raw bulk-tank milk samples of goats and sheep. Vet Microbiol 101: 101-107.
42. Haveri M, Roslof A, Rantala L, Pyorala S (2007) Virulence genes of bovine *Staphylococcus aureus* from persistent and non-persistent intramammary infections with different clinical characteristics. J Appl Microbiol 103: 993-1000.
43. Banks MC, Kamel NS, Zabriskie JB, Larone DH, Ursea D, et al. (2003) *Staphylococcus aureus* express unique superantigens depending on the tissue source. J Infect Dis 187: 77-86.
44. Priscila LM, Danilo FMR, Luiza P, Lisiane AM, Maria AVPB, et al. (2016) Detection of Enterotoxigenic Potential and Determination of Clonal Profile in *Staphylococcus aureus* and Coagulase-Negative *Staphylococci* Isolated from Bovine Subclinical Mastitis in Different Brazilian States. Toxins 8: 104.
45. Seyoum ET, Mekonene TK, Woldetsadik DA, Zewudie BM, Wondwossen A, et al. (2016) Enterotoxin gene profile of *Staphylococcus aureus* isolates recovered from bovine milk produced in central Ethiopia. J Infect Dev Ctries 10: 138-142.
46. Ertas N, Gonulalan Z, YildirimY, Kum E (2010) Detection of *Staphylococcus aureus* enterotoxins in sheep cheese and dairy desserts by multiplex PCR technique. Int J Food Microbiol 15: 74-77.
47. Balaban N, Rasooly A (2000) *Staphylococcal* enterotoxins. Int J Food Microbiol 61: 1-10.
48. Calsolari RAO, Pereira VCP, Júnior JPA, Cunha MLRS (2011) Determination of toxigenic capacity by RT-PCR in coagulase-negative *Staphylococci* and *Staphylococcus aureus* isolated from newborns in Brazil. Microbiol Immunol 55: 394-407.