

Editorial

Open Access

Potential Applications of Epigallocatechin Gallate-Fatty Acid Derivatives as Antiviral Agents

Kunihiro Kaihatsu*

The Institute of Scientific and Industrial Research, Osaka University, 8-1 Mihogaoka, Ibaraki, Osaka 567-0047, Japan

Keywords: Epigallocatechin gallate; EGCG-fatty acid monoester; DNA virus; RNA virus; Membrane permeability; Chemical stability; Antiviral activity

Abbreviations

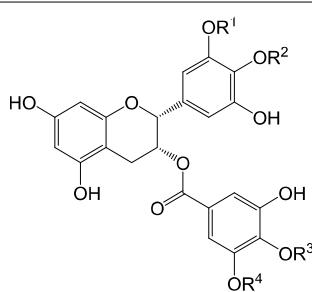
EGCG: (-)-Epigallocatechin-3-O-gallate; MDCK: Madin-Darby Canine Kidney; HSV: Herpes Simplex Virus; HPV: Human Papilloma Virus; HBV: Hepatitis B Virus; HCV: Hepatitis C Virus; HIV-1: Human Immunodeficiency Virus-1; EGCG-C₁₆: EGCG-Monopalmitate; EC₅₀: 50% Effective Concentration

Editorial

Epigallocatechin gallate (EGCG) (Figure 1a) is the major catechin component of green tea (*Camellia sinensis*). It has been shown to have antiviral properties against DNA viruses such as herpes simplex virus (HSV; *Herpesviridae*) [1], adenovirus (*Adenoviridae*) [2], human papilloma virus (HPV; *Papovaviridae*) [3], and hepatitis B virus (HBV; *Hepadnaviridae*) [4], and RNA viruses such as influenza virus (*Orthomyxoviridae*) [5], ebola virus (*Filoviridae*) [6], hepatitis C virus (HCV; *Flaviviridae*) [7] and human immunodeficiency virus-1 (HIV-1; *Retroviridae*) [8].

The antiviral mechanisms of the actions of EGCG vary depending on its target virus. According to previous reports, EGCG directly interacts with the lipid membrane and the viral membrane proteins of influenza virus [5] and HCV [7] and interferes with viral entry or membrane fusion steps. EGCG has also been shown to inhibit late infection stages including reverse-transcription [8], transcription [9], integration [8,10], cell signal transduction [11] and protein chaperoning [6]. Although these broad antiviral activities of EGCG are attractive, its poor chemical stability under physiological conditions [12] and low bioavailability (<0.3%) *in vivo* [13] are drawbacks preventing its use in its native form.

There have been several reports in which the chemical, biological



- a) EGCG: R¹=R²=R³=R⁴=H
b) EGCG-C₁₆: One of R¹, R², R³, or R⁴ = CO(CH₂)₁₄CH₃, Others=H

Figure 1: Chemical structure of epigallocatechin gallate (EGCG) and its fatty acid derivatives. (a) A major green tea catechin, epigallocatechin gallate. (b) EGCG-C₁₆ that possesses a palmitoyl group at R¹, R², R³, or R⁴.

and antiviral properties of EGCG were improved by chemical modification. Kouno et al. introduced an *n*-octadecyl group at the 4'-position of EGCG using isocyanate chemistry [14]. They found that EGCG modified with a C₁₈ alkyl group possesses increased hydrophobicity and lipid membrane affinity relative to natural EGCG. Mori et al. developed a method to introduce fatty acids into the B-ring or D-ring of EGCG using lipase-catalyzed transesterification [15]. They found that the effectiveness of EGCG-fatty acid monoesters against influenza A/Puerto Rico/8/33 (H1N1) virus was increased in an alkyl chain length-dependent manner in Madin-Darby canine kidney (MDCK) cells [15]. Among EGCG-saturated fatty acid monoesters, EGCG-monopalmitate (EGCG-C₁₆) (Figure 1b) showed the highest anti-influenza virus activity [15]. Although EGCG-C₁₆ exhibited a slightly (3.2-fold) higher cytotoxic effect to MDCK cells, it showed significantly (23.5-fold) higher anti-influenza virus activity against the A/Puerto Rico/8/34(H1N1) strain [16]. EGCG-C₁₆ inhibited human-, swine- and avian-pathogenic influenza A viruses, and the EC₅₀ was between 10 nM and 61 nM, a 7.1-fold to 44-fold lower concentration than unmodified EGCG [16]. A lethal dose of avian-influenza A/H5N2 virus pretreated with EGCG-C₁₆ completely prevented the death of embryonated chicken eggs inoculated with the virus by an allantoic cavity route [16].

Oliveira et al. reported that EGCG-C₁₆ inhibited the adsorption step of herpes simplex virus infection in Vero cells [17]. Zhao et al. reported that EGCG-C₁₆ exhibits significantly higher antiviral activity against porcine reproductive and respiratory syndrome virus than natural EGCG and ribavirin as both pre-treatment and post-treatment [18]. These reports suggest that the pronounced antiviral activities of EGCG-fatty acid derivatives were due to increased permeability through both the viral membrane and the cell membrane.

Although EGCG interferes with different types of viral infections, it is easily oxidized or hydrolyzed under physiological conditions because of the reactive hydroxyl groups in the gallyl moiety in the B-ring and the galloyl moiety in the D-ring [19,20]. However, addition of a palmitoyl group to the B-ring or D-ring prolonged its half-life in a cell culture medium approximately seven-fold [21].

These improved chemical, biological, and antiviral activities of

***Corresponding author:** Kunihiro Kaihatsu, The Institute of Scientific and Industrial Research, Osaka University, 8-1 Mihogaoka, Ibaraki, Osaka 567-0047, Japan, Tel: +81-6-6879-8472; Fax: +81-6-6879-8474; Email: kunihiro@sanken.osaka-u.ac.jp

Received July 08, 2015; **Accepted** July 09, 2015; **Published** July 16, 2015

Citation: Kaihatsu K (2015) Potential Applications of Epigallocatechin Gallate-Fatty Acid Derivatives as Antiviral Agents. J Antivir Antiretrovir 7: Iv-Ivi. doi:10.4172/jaa.1000e127

Copyright: © 2015 Kaihatsu K. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

this EGCG-fatty acid monoester should expand its application as an antiviral agent and may allow us to combat emerging drug-resistant viruses more effectively.

References

1. Isaacs CE, Wen GY, Xu W, Jia JH, Rohan L, et al. (2008) Epigallocatechin gallate inactivates clinical isolates of herpes simplex virus. *Antimicrob Agents Chemother* 52: 962-970.
2. Weber JM, Ruzindana-Umunyana A, Imbeault L, Sircar S (2003) Inhibition of adenovirus infection and adenain by green tea catechins. *Antiviral Res* 58: 167-173.
3. He L, Zhang E, Shi J, Li X, Zhou K, et al. (2013) (-)-Epigallocatechin-3-gallate inhibits human papillomavirus (HPV)-16 oncoprotein-induced angiogenesis in non-small cell lung cancer cells by targeting HIF-1α. *Cancer Chemother Pharmacol* 71: 713-725.
4. He W, Li LX, Liao QJ, Liu CL, Chen XL (2011) Epigallocatechin gallate inhibits HBV DNA synthesis in a viral replication - inducible cell line. *World J Gastroenterol* 17: 1507-1514.
5. Nakayama M, Suzuki K, Toda M, Okubo S, Hara Y, et al. (1993) Inhibition of the infectivity of influenza virus by tea polyphenols. *Antiviral Res* 21: 289-299.
6. Reid SP, Shurtleff AC, Costantino JA, Tritsch SR, Retterer C, et al. (2014) HSPA5 is an essential host factor for Ebola virus infection. *Antiviral Res* 109: 171-174.
7. Ciesek S, von Hahn T, Colpitts CC, Schang LM, Friesland M, et al. (2011) The green tea polyphenol, epigallocatechin-3-gallate, inhibits hepatitis C virus entry. *Hepatology* 54: 1947-1955.
8. Yamaguchi K, Honda M, Ikigai H, Hara Y, Shimamura T (2002) Inhibitory effects of (-)-epigallocatechin gallate on the life cycle of human immunodeficiency virus type 1 (HIV-1). *Antiviral Res* 53: 19-34.
9. Chang LK, Wei TT, Chiu YF, Tung CP, Chuang JY, et al. (2003) Inhibition of Epstein-Barr virus lytic cycle by (-)-epigallocatechin gallate. *Biochem Biophys Res Commun* 301: 1062-1068.
10. Jiang F, Chen W, Yi K, Wu Z, Si Y, et al. (2010) The evaluation of catechins that contain a galloyl moiety as potential HIV-1 integrase inhibitors. *Clin Immunol* 137: 347-356.
11. Yan Z, Yong-Guang T, Fei-Jun L, Fa-Qing T, Min T, et al. (2004) Interference effect of epigallocatechin-3-gallate on targets of nuclear factor kappaB signal transduction pathways activated by EB virus encoded latent membrane protein 1. *Int J Biochem Cell Biol* 36: 1473-1481.
12. Lam WH, Kazi A, Kuhn DJ, Chow LM, Chan AS, et al. (2004) A potential prodrug for a green tea polyphenol proteasome inhibitor: evaluation of the peracetate ester of (-)-epigallocatechin gallate [(-)-EGCG]. *Bioorg Med Chem* 12: 5587-5593.
13. Kohri T, Matsumoto N, Yamakawa M, Suzuki M, Nanjo N, et al. (2001) Metabolic fate of (-)-[4-3H] epigallocatechin gallate in rats after oral administration. *J Agric Food Chem* 49: 4102-4112.
14. Tanaka T, Kusano R, Kouno I (1998) Synthesis and antioxidant activity of novel amphipathic derivatives of tea polyphenol. *Bioorg Med Chem Lett* 8: 1801-1806.
15. Mori S, Miyake S, Kobe T, Nakaya T, Fuller SD, et al. (2008) Enhanced anti-influenza A virus activity of (-)-epigallocatechin-3-O-gallate fatty acid monoester derivatives: effect of alkyl chain length. *Bioorg Med Chem Lett* 18: 4249-4252.
16. Kaihatsu K, Mori S, Matsumura H, Daidoji T, Kawakami C, et al. (2009) Broad and potent anti-influenza virus spectrum of epigallocatechin-3-O-gallate-monopalmitate. *J Mol Genet Med* 3: 195-197.
17. de Oliveira A, Adams SD, Lee LH, Murray SR, Hsu SD, et al. (2013) Inhibition of herpes simplex virus type 1 with the modified green tea polyphenol palmitoyl-epigallocatechin gallate. *Food Chem Toxicol* 52: 207-215.
18. Zhao C, Liu S, Li C, Yang L, Zu Y (2014) In vitro evaluation of the antiviral activity of the synthetic epigallocatechin gallate analog-epigallocatechin gallate (EGCG) palmitate against porcine reproductive and respiratory syndrome Virus. *Viruses* 6: 938-950.
19. Mizooku Y, Yoshikawa M, Tsuneyoshi T, Arakawa R (2003) Analysis of oxidized epigallocatechin gallate by liquid chromatography/mass spectrometry. *Rapid Commun Mass Spectrom* 17: 1915-1918.
20. Severino JF, Goodman BA, Kay CWM, Stolze K, Tunega D, et al. (2009) Free radicals generated during oxidation of green tea polyphenols: Electron paramagnetic resonance spectroscopy combined with density functional theory calculations. *Free Radic Biol Med* 46: 1076-1088.
21. Jung JH, Yun M, Choo EJ, Kim SH, Jeong MS, et al. (2015) A derivative of epigallocatechin-3-gallate induces apoptosis via SHP-1-mediated suppression of BCR-ABL and STAT3 signalling in chronic myelogenous leukaemia. *Br J Pharmacol* 172: 3565-3578.