

Personalized Medicine through Circulating Tumor Cells: The New Dimension in Cancer Research

Shailender Singh Kanwar^{1,2}, Lianette Rivera^{1,2} and Sunitha Nagrath^{1,2,3*}

¹Department of Chemical Engineering, College of Engineering, University of Michigan, Ann Arbor, Michigan-48109, USA

²Bio-interfaces Institute, North Campus Research Center, University of Michigan, Ann Arbor, Michigan-48109, USA

³Translational Oncology Program, University of Michigan, Ann Arbor, Michigan-48109, USA

Outcomes in some of the current clinical trials could be misleading as they fail to represent the high genetic variability seen among cancer patients, leading to averaged responses taken from broader patient groups. In fact, clinicians and drug makers both agree that no group of trial volunteers could ever match the extraordinary biological diversity of the drug's eventual consumers i.e., patients [1]. With such challenges the American Cancer Society anticipates that 580,350 Americans will die of cancer this year, resulting in nearly 1,600 people per day [2]. Since metastasis is a major cause of such high mortality rate, elucidating this enigmatic process can potentially help to change the current paradigm of cancer diagnosis, treatment and monitor efficacy of therapies. Emerging evidence indicates the important roles of Circulating Tumor Cells (CTCs) in the spread of cancers and metastasis [3,4]. CTCs are the cells shed by primary tumor into circulation and known to be key players in metastasis [5,6]. Recent reports also show that these cells could be a good surrogate biomarker for not only prognosis, but also for cancer detection and development of personalized treatment [7-9]. This editorial briefly examines the current perspectives in using CTCs as a liquid biopsy to develop personalized targeted therapies for advancing the field of cancer research.

The first critical step in using CTCs as biomarker is the optimal isolation of these cells. This step represents a major technological challenge, since CTCs make up a minute fraction of the total number of cells circulating in blood, 1-10 CTCs per mL of whole blood compared to a few million white blood cells and a billion red blood cells. In consequence, a plethora of novel technologies have emerged in the past years for detecting and isolating CTCs [5,10,11]. These techniques are classified based on properties that distinguish CTCs from surrounding hematopoietic cells, such as physical properties and biological properties. Physical properties include size, density, electric charges, and deformability; while biological properties mainly focus on cell surface protein expression and viability [12-15]. For either of these strategies, it is imperative that developing an optimal CTC isolation method needs to meet following criteria: (1) high recovery, (2) high purity of CTCs by removal of all other blood components, and (3) high system throughput to assure practical application of large sample volume [16].

Currently, Cell Search[®] Method is the only FDA approved system and the "gold standard" for emerging CTC isolation methods, however, many questions remained unanswered regarding the general clinical acceptance, validation and optimal characterization assay for CTCs [17,18]. Cell Search[®] Method is a surface expression based method, using antibodies against epithelial cell adhesion marker (EpCAM) [19]. As in any other biological based method, its major limitation is potential loss of any subpopulation of CTCs that do not express the EpCAM-antigens used in the capture protocol. Recognizing this limitation, ongoing efforts are directed towards methods that are independent of EpCAM expression exploiting CTC's biological and physical properties as previously mentioned [20-22].

Once optimal enrichment of CTCs is achieved, the second critical step involves the practical use of CTCs for advancing research from the bench-to-bedside. CTCs isolated from newly diagnosed cancer patients as well as those undergoing treatments, can be used to unravel the mysteries behind disease progression by elucidating key-cellular interactions of metastasis through functional assays [8,12]. Understanding the science of CTCs therefore leads the way to personalized medicine. Despite of technological advancements made in this field, a standardized clinical approach to identify CTCs and its characterization is yet to be found. The challenge arises because of the extremely low number CTCs present in blood (limiting their availability for downstream assays) and therein the erogeneity [9,23].

Researchers have identified new methods to characterize CTCs based on quantifying the levels of protein expression, mRNA expression and chromosomal abnormalities. For examples, it has shown that protein expression of HER2 correlate well with gene amplification when assessed in parallel by Fluorescence In Situ Hybridization (FISH) on cytospun CTCs obtained from cell lines and patient samples [24,25]. Additionally, other clinical studies have utilized mRNA expression profile of CTCs using Reverse Transcription Polymerase Chain Reaction (RT-PCR) mainly using commercially available Adna Test[®] (AdnaGen AG, langenhagen) [26,27]. However, as mentioned earlier, most of these assays are yet to be validated.

Other characterization assays include performing FISH at RNA and DNA level to analyze tumor-specific markers and chromosomal aberrations to identify gene amplifications and translocations [23]. Considering the genomic characterization, single cell isolation of CTCs employing microfluidic approaches have demonstrated to have major clinical importance [5]. Ongoing research is currently focused on gaining extensive data-characteristics of few CTCs through combining immunofluorescence-based assays using different fluorophores, image cytometry and FISH as well as PCR-based multiplex assays [8,28-30].

Current beliefs in oncology are shifting to personalized medicine. Therefore, an approach of personalized medicine for developing drugs in cancer therapy seems imperative than ever

***Corresponding author:** Sunitha Nagrath, Assistant Professor, Department of Chemical Engineering, University of Michigan, 2300 Hayward Street, Ann Arbor, Michigan-48109, USA, Tel: 734-647-7985; E-mail: snagrath@umich.edu

Received August 27, 2013; **Accepted** August 29, 2013; **Published** September 04, 2013

Citation: Kanwar SS, Rivera L, Nagrath S (2013) Personalized Medicine through Circulating Tumor Cells: The New Dimension in Cancer Research. J Develop Drugs 2: e128. doi:10.4172/2329-6631.1000e128

Copyright: © 2013 Kanwar SS, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

before. CTCs analysis, often described as a “liquid biopsy”, could serve as a companion diagnostic for the pharmaceutical industry by incorporating CTCs based biomarkers as endpoints in future clinical trial design [31]. Accordingly, CTCs are currently implemented into more than 400 clinical trials at which the integration of CTCs into immunotherapeutic approaches seems to be very promising [32].

In the pharmaceutical industries, which are associated with a high failure rate, significant savings are possible if companies can assess efficacy at an early stage in clinical trials [33] through the easily accessible CTCs, which can represent better the dynamic changes of tumor progression. The dynamic changes in CTC counts before initiation of therapy and during (neo) adjuvant therapy provide strong prognostic value in evaluating real-time information on the efficacy of therapy. CTCs as surrogate endpoints could provide new insights into the complex mechanism of drug resistance, allowing clinicians to take individual therapy decision when change in the treatment course is needed. New ongoing trials investigating the use of CTCs as predictive markers are expected to provide great insights about its measurable benefits for the individual patients [34,35]. Conjoining the approaches of single CTC isolation with genomic profiling, it has become possible to accurately identify the disruptive molecular pathway(s) at any given time, thereby, leading to individualized clinical decisions. Hence, a genuine personalized treatment approach in cancer patients utilizing CTCs seems to be feasible in near future.

Although, the presence of CTCs is known for a long time they became the subject of active research only in last two decades [36]. Significantly, the research in past decade has been instrumental in bringing forth CTCs to clinic. However, CTCs are far from being a mainstream clinical assay. We acknowledge a strong need of benchmarks for CTCs isolation and characterization in order to realize their full potential in clinical setting and for tailoring the individual therapies. Nonetheless, the study of CTCs poses a direct impact upon society by presenting novel ways to address some of the major hurdles in cancer research such as early detection, real-time monitoring for drug efficacy, and patient screening for enabling better clinical trials. CTCs will potentially foster the further understanding of the pharmacodynamics, thereby, facilitating testing of novel therapeutic targets for the advancement of personalized medicine.

References

- Leaf C (2013) Do clinical trials work? *The New York Times-Sunday Review*.
- Cancer Facts & Figures 2013.
- Onstenk W, Gratama JW, Foekens JA, Sleijfer S, et al. (2013) Towards a personalized breast cancer treatment approach guided by circulating tumor cell (CTC) characteristics. *Cancer Treat Rev* 39: 691-700.
- Magbanua MJ, Sosa EV, Roy R, Eisenbud LE, Scott JH, et al. (2013) Genomic profiling of isolated circulating tumor cells from metastatic breast cancer patients. *Cancer Res* 73: 30-40.
- Kanwar SSG, Meggie MG, Sunitha N (2013) Bio-MEMS (Microfluidics) for CTC detection in cancer patients in microfluidics, nanotechnology and disease biomarkers for personalized medicine applications. E.J.H.W. Muhammad J. A. Shiddiky, Sakandar Rauf and Matt Trau, Editor 2013, Nova Science Publishers, USA.
- Koenigsberg R, Obermayr E, Bises G, Pfeiler G, Gneist M, et al. (2011) Detection of EpCAM positive and negative circulating tumor cells in metastatic breast cancer patients. *Acta Oncol* 50: 700-710.
- Nieva JJ, Kuhn P (2012) Fluid biopsy for solid tumors: a patient's companion for lifelong characterization of their disease. *Future Oncol* 8: 989-998.
- Sieuwerts AM, Jeffrey SS (2012) Multiplex molecular analysis of CTCs. *Recent Results Cancer Res* 195: 125-140.
- Maheswaran S, Sequist LV, Nagrath S, Ullkus L, Brannigan B, et al. (2008) Detection of mutations in EGFR in circulating lung-cancer cells. *N Engl J Med* 359: 366-377.
- Zhang Z, Nagrath S (2013) Microfluidics and cancer: are we there yet? *Biomed Microdevices* 15: 595-609.
- Hughes AD, King MR (2012) Nanobiotechnology for the capture and manipulation of circulating tumor cells. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 4: 291-309.
- Mueller V, Stahmann N, Riethdorf S, Rau T, Zabel T, et al. (2005) Circulating tumor cells in breast cancer: correlation to bone marrow micrometastases, heterogeneous response to systemic therapy and low proliferative activity. *Clin Cancer Res* 11: 3678-3685.
- Mohamed H, Murray M, Turner JN, Caggana M (2009) Isolation of tumor cells using size and deformation. *J Chromatogr A* 1216: 8289-8295.
- Tan SJ, Yobas L, Lee GY, Ong CN, Lim CT (2009) Microdevice for the isolation and enumeration of cancer cells from blood. *Biomed Microdevices* 11: 883-892.
- Nagrath S, Sequist LV, Maheswaran S, Bell DW, Irimia D, et al. (2007) Isolation of rare circulating tumour cells in cancer patients by microchip technology. *Nature* 450: 1235-1239.
- Li P, Stratton ZS, Dao M, Ritz J, Huang TJ (2013) Probing circulating tumor cells in microfluidics. *Lab Chip* 13: 602-609.
- Strati A, Kasimir-Bauer S, Markou A, Parisi C, Lianidou ES (2013) Comparison of three molecular assays for the detection and molecular characterization of circulating tumor cells in breast cancer. *Breast Cancer Res* 15: R20.
- Parkinson DR, Dracopoli N, Petty BG, Compton C, Cristofanilli M, et al. (2012) Considerations in the development of circulating tumor cell technology for clinical use. *J Transl Med* 10: 138.
- Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, et al. (2004) Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *N Engl J Med* 351: 781-791.
- Lin HK, Zheng S, Williams AJ, Balic M, Groshen S, et al. (2010) Portable filter-based microdevice for detection and characterization of circulating tumor cells. *Clin Cancer Res* 16: 5011-5018.
- Krebs MG, Hou JM, Sloane R, Lancashire L, Priest L, et al. (2012) Analysis of circulating tumor cells in patients with non-small cell lung cancer using epithelial marker-dependent and -independent approaches. *J Thorac Oncol* 7: 306-315.
- Hou HW, Warkiani ME, Khoo BL, Li ZR, Soo RA, et al. (2013) Isolation and retrieval of circulating tumor cells using centrifugal forces. *Sci Rep* 3: 1259.
- Yu M, Bardia A, Wittner BS, Stott SL, Smas ME, et al. (2013) Circulating breast tumor cells exhibit dynamic changes in epithelial and mesenchymal composition. *Science* 339: 580-584.
- Ignatiadis M, Rothé F, Chaboteaux C, Durbecq V, Rouas G, et al. (2011) HER2-positive circulating tumor cells in breast cancer. *PLoS One* 6: e15624.
- Punnoose EA, Atwal SK, Spoerke JM, Savage H, Pandita A, et al. (2010) Molecular biomarker analyses using circulating tumor cells. *PLoS One* 5: e12517.
- Sieuwerts AM, Mostert B, Bolt-de Vries J, Peeters D, de Jongh FE, et al. (2011) mRNA and microRNA expression profiles in circulating tumor cells and primary tumors of metastatic breast cancer patients. *Clin Cancer Res* 17: 3600-3618.
- Lianidou ES, Markou A (2011) Circulating tumor cells in breast cancer: detection systems, molecular characterization, and future challenges. *Clin Chem* 57: 1242-1255.
- Navin N, Kendall J, Troge J, Andrews P, Rodgers L, et al. (2011) Tumour evolution inferred by single-cell sequencing. *Nature* 472: 90-94.
- Strati A, Markou A, Parisi C, Politaki E, Mavroudis D, et al. (2011) Gene expression profile of circulating tumor cells in breast cancer by RT-qPCR. *BMC Cancer* 11: 422.
- Powell AA, Talasz AH, Zhang H, Coram MA, Reddy A, et al. (2012) Single cell profiling of circulating tumor cells: transcriptional heterogeneity and diversity from breast cancer cell lines. *PLoS One* 7: e33788.
- Balic M, Williams A, Lin H, Datar R, Cote RJ (2013) Circulating tumor cells: from bench to bedside. *Annu Annu Rev Med* 64: 31-44.
- Gorges TM, Pantel K (2013) Circulating tumor cells as therapy-related biomarkers in cancer patients. *Cancer Immunol Immunother* 62: 931-939.

-
33. Paul SM, Mytelka DS, Dunwiddie CT, Persinger CC, Munos BH, et al. (2010) How to improve R&D productivity: the pharmaceutical industry's grand challenge. *Nat Rev Drug Discov* 9: 203-214.
34. Riethdorf S, Mueller V, Zhang L, Rau T, Loibl S, et al. (2010) Detection and HER2 expression of circulating tumor cells: prospective monitoring in breast cancer patients treated in the neoadjuvant GeparQuattro trial. *Clin Cancer Res* 16: 2634-2645.
35. Witzel I, Loibl S, von Minckwitz G, Eidtmann H, Fehm T, et al. (2012) Predictive value of HER2 serum levels in patients treated with lapatinib or trastuzumab --a translational project in the neoadjuvant GeparQuinto trial. *Br J Cancer* 107: 956-960.
36. Ashworth TR (1869) A case of cancer in which cells similar to those in the tumors were seen in the blood after death. *Australian Medical Journal* 14: 146-149.