

Editorial

Open Access

On the Potential Use of Genetically Modified Mesenchymal Stem Cells to Treat Articular Cartilage Defects

Magali Cucchiariini*

Saarland University Medical Center, Homburg/Saar, Germany

Damage to articular cartilage, the tissue that allows for a smooth gliding of the articulating surface of a joint (and to withstand high loads), is an unsolved problem, especially following joint trauma [1]. As adult, hyaline cartilage is avascular, and without a lymphatic drainage, it does not have access to reparative cells that may be brought locally in response to injury. Defects (partial or full-thickness) of the cartilage thus do not heal on themselves and instead, tend to become persistent and progress inevitably, often towards osteoarthritis [2].

Current options for cartilage repair are diverse (marrow-stimulating techniques, transplantation of tissue or cells, replacement surgery), yet none can reliably regenerate the natural functions of the cartilage, including its integrity (type-II and not type-I collagen), the adherence of the repair tissue to the surrounding cartilage, and the ability of withstanding mechanical stress over time.

Considerable efforts have been made to improve the repair processes and in particular, the promise of treating cartilage defects by cell- and gene-based approaches has attracted much attention [3-7]. Indeed, administration of candidate genes, rather than recombinant factors with short half-lives, might be a means to promote sustained repair like for disorders showing a gradual progression. The development of strategies using the implantation of genetically modified mesenchymal stem cells (MSCs) as therapeutic platforms might prove beneficial to enhance cartilage repair during surgical procedures. As a matter of fact, MSCs might be better suited for this purpose than committed cells such as chondrocytes that require more invasive methods of extraction and tend to lose their phenotype upon culture. Also, MSCs have the ability to recapitulate lineage transitions originally involved in mesenchymal tissue formation (cartilage, bone, fat tissue), among which a potential for chondrogenesis [8-10]. MSCs further display critical tropic activities and demonstrate, interestingly, a broad panel of immunomodulatory properties. MSCs can be isolated from the bone marrow, bone, adipose tissue, muscle, synovium, periosteum, and perichondrium. It is important to note that genetic modification of such cells might also allow to overcome some of the limitations still associated with their use, like the large amount of cells needed for application *in vivo*, the relatively low percentage of which enter proper chondrogenic pathways to produce a functional, reparative tissue, and the decline of lifespan and potency seen in pathological conditions (in OA patients, for instance) [11,12]. For cartilage repair, a challenge will be also to maintain the cells in a prehypertrophic state that avoids premature terminal differentiation, hypertrophy, and ossification [10,13-15].

Genetic manipulation of MSCs to enhance their potency for chondrogenesis and cartilage repair has been attempted using different gene delivery vectors, including nonviral systems as well as adenoviral, retro-/lentiviral, and recombinant adeno-associated viral (rAAV) vectors, without altering their multilineage potential [16-25]. Beneficial effects upon MSC chondrogenic differentiation *in vitro* have been reported by transfer of gene sequences coding for growth (TGF- β , BMPs, IGF-I, FGF-2) and transcription factors (SOX5, SOX6, SOX9, ZNF145), or signaling molecules (hedgehogs, PTHrP) [6,26,27]. Most remarkably, several experimental studies have evidenced the

feasibility of applying genetically modified MSCs (commonly as cell suspensions, three-dimensional cultures, coagulates, in conjunction with biomaterials, or even as tissue grafts) to successfully improve the repair of cartilage defects *in vivo* using many of the gene candidates cited above [19,21,28-36]. It remains to be seen now whether this approach will meet decisive success in patients, as no known clinical trial is ongoing for the treatment of articular cartilage defects by administration of gene-modified MSCs.

References

1. O'Driscoll SW (1998) The healing and regeneration of articular cartilage. *J Bone Joint Surg Am* 80: 1795-1812.
2. Buckwalter JA, Mankin HJ (1998) Articular cartilage repair and transplantation. *Arthritis Rheum* 41: 1331-1342.
3. Ansboro S, Greiser U, Barry F, Murphy M (2012) Strategies for improved targeting of therapeutic cells: implications for tissue repair. *Eur Cell Mater* 23: 310-318.
4. Chen FH, Rousche KT, Tuan RS (2006) Technology Insight: adult stem cells in cartilage regeneration and tissue engineering. *Nat Clin Pract Rheumatol* 2: 373-382.
5. Cucchiariini M, Madry H (2005) Gene therapy for cartilage defects. *J Gene Med* 7: 1495-1509.
6. Cucchiariini M, Venkatesan JK, Ekici M, Schmitt G, Madry H (2012) Human mesenchymal stem cells overexpressing therapeutic genes: from basic science to clinical applications for articular cartilage repair. *Biomed Mater Eng* 22: 197-208.
7. Madry H, Cucchiariini M (2011) Clinical potential and challenges of using genetically modified cells for articular cartilage repair. *Croat Med J* 52: 245-261.
8. Baksh D, Song L, Tuan RS (2004) Adult mesenchymal stem cells: characterization, differentiation, and application in cell and gene therapy. *J Cell Mol Med* 8: 301-316.
9. Caplan AI (2007) Adult mesenchymal stem cells for tissue engineering versus regenerative medicine. *J Cell Physiol* 213: 341-347.
10. Prockop DJ (2009) Repair of tissues by adult stem/progenitor cells (MSCs): controversies, myths, and changing paradigms. *Mol Ther* 17: 939-946.
11. Murphy JM, Dixon K, Beck S, Fabian D, Feldman A, et al. (2002) Reduced chondrogenic and adipogenic activity of mesenchymal stem cells from patients with advanced osteoarthritis. *Arthritis Rheum* 46: 704-713.
12. Muschler GF, Nitto H, Boehm CA, Easley KA (2001) Age- and gender-related changes in the cellularity of human bone marrow and the prevalence of osteoblastic progenitors. *J Orthop Res* 19: 117-125.

*Corresponding author: Magali Cucchiariini, Ph.D., Associate Professor in Molecular Biology, Center of Experimental Orthopaedics, Saarland University Medical Center, Kirbergerstr. Homburg/Saar, Germany, Tel: ++49-6841-1624987; Fax: ++49-6841-1624988; E-mail: mmcucchiariini@hotmail.com

Received September 17, 2012; Accepted September 26, 2012; Published October 01, 2012

Citation: Cucchiariini M (2012) On the Potential Use of Genetically Modified Mesenchymal Stem Cells to Treat Articular Cartilage Defects. *Orthop Muscul Syst* 1:e106. doi:[10.4172/2161-0533.1000e106](https://doi.org/10.4172/2161-0533.1000e106)

Copyright: © 2012 Cucchiariini M. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

13. Augello A, De Bari C (2010) The regulation of differentiation in mesenchymal stem cells. *Hum Gene Ther* 21: 1226-1238.
14. Hollander AP, Dickinson SC, Kafienah W (2010) Stem cells and cartilage development: complexities of a simple tissue. *Stem Cells* 28: 1992-1996.
15. Tuan RS (2006) Stemming cartilage degeneration: adult mesenchymal stem cells as a cell source for articular cartilage tissue engineering. *Arthritis Rheum* 54: 3075-3078.
16. Allay JA, Dennis JE, Haynesworth SE, Majumdar MK, Clapp DW, et al. (1997) LacZ and interleukin-3 expression in vivo after retroviral transduction of marrow-derived human osteogenic mesenchymal progenitors. *Hum Gene Ther* 8: 1417-1427.
17. Cucchiarini M, Madry H (2010) Genetic modification of mesenchymal stem cells for cartilage repair. *Biomed Mater Eng* 20: 135-143.
18. Elsler S, Schetting S, Schmitt G, Kohn D, Madry H, et al. (2012) Effective, safe nonviral gene transfer to preserve the chondrogenic differentiation potential of human mesenchymal stem cells. *J Gene Med* 14: 501-511.
19. Grande DA, Mason J, Light E, Dines D (2003) Stem cells as platforms for delivery of genes to enhance cartilage repair. *J Bone Joint Surg Am* 85-A: 111-116.
20. Ito H, Goater JJ, Tiyapatanaputi F, Rubery PT, O'Keffe RJ, et al. (2004) Light-activated gene transduction of recombinant adeno-associated virus in human mesenchymal stem cells. *Gene Ther* 11: 34-41.
21. Mason JM, Breitbart AS, Barcia M, Porti D, Pergolizzi RG, et al. (2000) Cartilage and bone regeneration using gene-enhanced tissue engineering. *Clin Orthop Relat Res*: S171-S178.
22. Mosca JD, Hendricks JK, Buyaner D, Davis-Sproul J, Chuang LC, et al. (2000) Mesenchymal stem cells as vehicles for gene delivery. *Clin Orthop Relat Res*: S71-S90.
23. Palmer GD, Steinert A, Pascher A, Gouze E, Gouze JN, et al. (2005) Gene-induced chondrogenesis of primary mesenchymal stem cells *in vitro*. *Mol Ther* 12: 219-228.
24. Stender S, Murphy M, O'Brien T, Stengaard C, Ulrich-Vinther M, et al. (2007) Adeno-associated viral vector transduction of human mesenchymal stem cells. *Eur Cell Mater* 13: 93-99.
25. Van Damme A, Thorez L, Ma L, Vandeburgh H, Eyckmans J, et al. (2006) Efficient lentiviral transduction and improved engraftment of human bone marrow mesenchymal cells. *Stem Cells* 24: 896-907.
26. Cucchiarini M, Ekici M, Schetting S, Kohn D, Madry H (2011) Metabolic activities and chondrogenic differentiation of human mesenchymal stem cells following recombinant adeno-associated virus-mediated gene transfer and over expression of fibroblast growth factor 2. *Tissue Eng Part A* 17: 1921-1933.
27. Venkatesan JK, Ekici M, Madry H, Schmitt G, Kohn D, et al. (2012) SOX9 gene transfer via safe, stable, replication-defective recombinant adeno-associated virus vectors as a novel, powerful tool to enhance the chondrogenic potential of human mesenchymal stem cells. *Stem Cell Res Ther* 3: 22.
28. Cao L, Yang F, Liu G, Yu D, Li H, et al. (2011) The promotion of cartilage defect repair using adenovirus mediated Sox9 gene transfer of rabbit bone marrow mesenchymal stem cells. *Biomaterials* 32: 3910-3920.
29. Goomer RS, Deftos LJ, Terkeltaub R, Maris T, Lee MC, et al. (2001) High-efficiency non-viral transfection of primary chondrocytes and perichondrial cells for ex-vivo gene therapy to repair articular cartilage defects. *Osteoarthritis Cartilage* 9: 248-256.
30. Guo X, Zheng Q, Yang S, Shao Z, Yuan Q, et al. (2006) Repair of full-thickness articular cartilage defects by cultured mesenchymal stem cells transfected with the transforming growth factor beta1 gene. *Biomed Mater* 1: 206-215.
31. Ivkovic A, Pascher A, Hudetz D, Maticic D, Jelic M, et al. (2010) Articular cartilage repair by genetically modified bone marrow aspirate in sheep. *Gene Ther* 17: 779-789.
32. Katayama R, Wakitani S, Tsumaki N, Morita Y, Matsushita I, et al. (2004) Repair of articular cartilage defects in rabbits using CDMP1 gene-transfected autologous mesenchymal cells derived from bone marrow. *Rheumatology (Oxford)* 43: 980-985.
33. Kubo S, Cooper GM, Matsumoto T, Phillipi JA, Corsi KA, et al. (2009) Blocking vascular endothelial growth factor with soluble Flt-1 improves the chondrogenic potential of mouse skeletal muscle-derived stem cells. *Arthritis Rheum* 60: 155-165.
34. Kuroda R, Usas A, Kubo S, Corsi K, Peng H, et al. (2006) Cartilage repair using bone morphogenetic protein 4 and muscle-derived stem cells. *Arthritis Rheum* 54: 433-442.
35. Liu TM, Guo XM, Tan HS, Hui JH, Lim B, et al. (2011) Zinc-finger protein 145, acting as an upstream regulator of SOX9, improves the differentiation potential of human mesenchymal stem cells for cartilage regeneration and repair. *Arthritis Rheum* 63: 2711-2720.
36. Pagnotto MR, Wang Z, Karpie JC, Ferretti M, Xiao X, et al. (2007) Adeno-associated viral gene transfer of transforming growth factor-beta1 to human mesenchymal stem cells improves cartilage repair. *Gene Ther* 14: 804-813.