

**Open Access** 

## Novel Vaccinology Approaches against Ticks

## Yasser E. Shahein\*

Molecular Biology Department, National Research Centre, Egypt

Over the past few years, the science of developing vaccines have had, and continue to prevent diseases. Veterinary vaccines have a major role in reducing animal suffering, protection of animal health and enhance efficient animal productivity. Ticks are considered among the most important disease transmitting vectors second to mosquitoes. Tick borne diseases affect both human and livestock animals in many tropical and subtropical areas [1,2]. Ticks transmit several viral infections, bacterial and protozoan diseases causing Encephalitis, Crimean-Congo Hemorrhagic fever, Kyasanur forest disease, Lyme disease, Rocky Mountain spotted fever, monocytic and granulocytic ehrlichiosis, and bebesiosis.

Control of ticks and tick borne diseases has passed several steps started by using the biological control and the use of acaricides which showed partial success and many drawbacks on the environment and animal productivity. Other strategies had evolved using immunization protocols of native purified proteins followed by the cloning of the gut cell surface molecule BM86; the most successful vaccine against the cattle tick *Rhipicephalus microplus* with some limitations against non *Rhipicephalus* species.

Recently, other approaches have been developed based on the genomics of the tick species. The genome sequence of the target tick species will explore potential protein coding sequences that can be used as candidate vaccines [3]. Vaccinology in the genome era is considered to be the second revolution in the 20<sup>th</sup> century [4]. This approach is known as the reverse vaccinology which utilizes the information of the entire genome of the parasite to predict and localize potential functional proteins through the open reading frames in the genome. These potential candidates are then cloned and expressed and finally their immunogenicity is validated in animal models.

In ticks, the genome size varies between  $\sim 10^9$  bp (*Amblyomma americanum*) and  $\sim 7 \times 10^9$  bp (*Rhipicephalus microplus*). The efforts to characterize the genomes of other ticks like *Ixodes scapularis* and the current studies implying the use of cDNA libraries of *R. annulatus* [5,6], *R. microplus* [7], *A. variegatum* [8], *I scapularis* [9] in the identification of novel molecules, will impact on the discovery of new tick-protective antigens. The use of the information deduced from the open reading frames in conjugation with functional analysis using technologies such as RNAi, bioinformatics, mutagenesis, transcriptomics, proteomics will allow for rapid discovery of novel tick vaccines.

High-throughput studies for some tick species have been accomplished such as the deer tick; *I. scapularis*, blacklegged tick; *Ixodes pacificus*, the Rocky mountain wood tick; *Dermacentor andersoni*, the tropical bont tick; *Amblyomma variegatum*, the castor bean tick; *Ixodes ricinus* [10], *Rhipicephalus haemaphysaloides* [11], the Gulf coast tick; *Amblyomma maculatum* [12] and the coarse bontlegged tick; *Hyalomma marignatum rufipes* [13]. The authors have investigated the sialotranscriptome and proteome in tick salivary glands of previously mentioned tick species. Several protein families have been identified as shown in table 1. These annotated databases can be used in the development of new strategies to control tick infestations and the discovery of new candidates for animal vaccination.

Two projects studying the sialotranscriptome of the cattle and camel ticks; *Rhipicephalus annulatus and Hyalomma dromedarii*, respectively, are currently in progress.

Tick species	Proteins/Genes
	Secreted genes Housekeeping genes
(Salivary glands CDNA library from adult fed females)	Unknown group or genes Transposable elements Metalloproteases Proteins with antimicrobial functions Basic tail proteins Madanin thrombin inhibitor proteins Lipocalin proteins Glycin rich proteins Mucins proteins Immunity related proteins
Ixodes ricinus	Housekeeping genes Reverse transcriptase-like proteins
(Salivary glands cDNA libraries from: unfed, 24hours fed, partially 4 days fed, and fully engorged-7 days fed female ticks)	Collagen-like secreted proteins Basic tail secreted proteins Kunitz domain containing proteins 18.9 kDa group (related to feeding) RGD-domain containing proteins WC-containing proteins (unique proteins) Histamine-binding proteins Metalloproteases a-2 macroglobulin Calreticulin Defensins Neuropeptide-like proteins Carboxypeptidase inhibitors Ixoderin B Phospholipases
Rhipicephalus haemaphysaloides	Signal transduction proteins
(Proteins extracted from salivary glands of fed and unfed adult females)	Cell apoptosis factor Molecular chaperones Synthesizing immunosuppressant proteins

Table 1: Examples of annotated protein families or genes identified in some tick species.

## References

- Jongejan F, Uilenberg G (1994) Ticks and control methods. Rev Sci Tech 13: 1201-1226.
- Estrada-Peña A, Jongejan F (1999) Ticks feeding on humans: a review of records on human-biting Ixodoidea with special reference to pathogen transmission. Exp Appl Acarol 23: 685-715.
- Kanampalliwar AM, Rajkumar S, Girdhar A, Archana T (2013) Reverse Vaccinology: Basics and Applications. J Vaccines Vaccin 4: 194.
- Lobanova LM (2010) New Strategies for Vaccine Development. Program in Vaccinology and Immunotherapeutics. SPCV 2: e4.
- Shahein YE (2008) Molecular cloning and expression of a larval immunogenic protein from the cattle tick Boophilus annulatus. Vet Immunol Immunopathol 121: 281-289.

\*Corresponding author: Yasser E. Shahein, Molecular Biology Department, National Research Centre, Egypt, Tel: +20 23337121; E-mail: yassershahein\_nrc@yahoo.com

Received December 15, 2013; Accepted December 18, 2013; Published December 20, 2013

Citation: Shahein YE (2013) Novel Vaccinology Approaches against Ticks. Clon Transgen 3: e107. doi:10.4172/2168-9849.1000e107

**Copyright:** © 2013 Shahein YE. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Page 2 of 2

- Shahein YE, El-Rahim MT, Hussein NA, Hamed RR, El-Hakim AE, et al. (2010) Molecular cloning of a small heat shock protein (sHSPII) from the cattle tick Rhipicephalus (Boophilus) annulatus salivary gland. Int J Biol Macromol 47: 614-622.
- Guerrero FD, Miller RJ, Rousseau ME, Sunkara S, Quackenbush J, et al. (2005) BmiGI: a database of cDNAs expressed in Boophilus microplus, the tropical/southern cattle tick. Insect Biochem Mol Biol 35: 585-595.
- Nene V, Lee D, Quackenbush J, Skilton R, Mwaura S, et al. (2002) AvGI, an index of genes transcribed in the salivary glands of the ixodid tick Amblyomma variegatum. Int J Parasitol 32: 1447-1456.
- Ribeiro JM, Alarcon-Chaidez F, Francischetti IM, Mans BJ, Mather TN, et al. (2006) An annotated catalog of salivary gland transcripts from Ixodes scapularis ticks. Insect Biochem Mol Biol 36: 111-129.

- Chmelar J, Anderson JM, Mu J, Jochim RC, Valenzuela JG, et al. (2008) Insight into the sialome of the castor bean tick, Ixodes ricinus. BMC Genomics 9: 233.
- 11. Xiang F, Zhang JW, Zhou YZ, Li Z, Gong HY, et al. (2009) Proteomic analysis of proteins in the salivary glands of the fed and unfed female tick Rhipicephalus haemaphysaloides. Agr Sciences China 8: 121-127.
- 12. Karim S, Singh P, Ribeiro JM (2011) A deep insight into the sialotranscriptome of the gulf coast tick, Amblyomma maculatum. PLoS One 6: e28525.
- Francischetti IM, Anderson JM, Manoukis N, Pham VM, Ribeiro JM (2011) An insight into the sialotranscriptome and proteome of the coarse bontlegged tick, Hyalomma marginatum rufipes. J Proteomics 74: 2892-2908.