



Non-Linear Decline in Neuronal Density within the Cerebral Cortices and Hippocampus in very Aged Rats: Implications for Causality

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Abstract

A clear change in the slope of the declining numbers of neurons in the cerebral cortices and hippocampus could reflect the “critical” mass, where qualitative shifts occur for correlative behaviors. In the present study, numbers of neurons within the layers of different regions of the cerebral cortices and the hippocampus were counted for rats, between the ages of 30 and 940 days. A conspicuous sudden inflection in the gradual decline of neurons occurred for both regions, around 700 days of age. Partial correlation analyses indicated that the cortical decline was dependent upon loss of neurons in the hippocampus. Neurons within layers 5 and 6 of the prefrontal cortices were particularly affected. These results are consistent with human brain data and the clinical course of dementias, and suggest that a subset of less than about 25% of original neurons may drive the geriatric process. Identification of these neurons by biomolecular indicators could lead to effective interventions.

Keywords: Cerebral cortices; Hippocampus; Aging cells; Neurons; Geriatric brains; Causal connections

Introduction

Although cerebral senescence is a cell-driven process coupled to a variety of metabolic- and replication-derived processes [1,2], the numbers of cells within a volume of tissue, including the cerebrum, show emergent properties that are dependent upon cell density. The cerebral cortices [3-5], and the hippocampal formation [6-9], are two powerfully interconnected volumes of mammalian brains that are the major correlates for memory.

In human beings, these structures are also strongly associated with processes described as consciousness, self-awareness [10], and the types of episodic memory specific to autobiographical memory [8,11]. For example: retrieval of represented experiences about where and within whom memories occurred, involves a disproportional increase in activity within the right prefrontal region [5,12]. Assuming that the numbers of neurons, and their implicit connections in large part determine the amount of information that can be represented, we have been investigating the consequences of normal aging upon neuronal numbers to discern, if loss of neurons to a critical mass is required before qualitative changes in complex cognitive properties occur.

In the following analyses, we had access to brains from rats that occupied a wide ontogenetic range, from weaning to over 940 days of age. We examined the hypothesis that in very old brains, the decline in neuronal numbers would be accelerated at a specific age, and certain regions and layers would be differentially affected. The determination of the quantitative proportions of neuronal loss may lead to more clear understanding of potential subpopulations of neurons that are more vulnerable because of local sensitivity, focal or anisotropic aberrant metabolic variations, or genetic influences, in a manner similar to the DNA-determined apoptosis of specific classes of neurons within the basal ganglia, from chromosome 4 anomalies.

That a reduction in the numbers of neurons within the cerebral cortices occurs in human beings, as a function of normal geriatric progression, has been known histologically for over a hundred years [13]. Although the arguments range from minimal loss of actual cells, but instead reduced volume of previously existing magnocellular neurons [14-16], to actual apoptotic or necrotic-induced declines [17], the critical observation is that cerebral cortical neurons are fewer

in the very old cerebrum [14,18,19]. The more or less linear decline in averaged numbers of cortical neurons in human brain accelerates precipitously after 75 to 80 years of age [20]. For the hippocampal formation, as inferred by MRI measurements, the age-dependent decline in whole volume (which presumably reflects neuronal numbers and sizes), has been assumed to be more linear [21,22].

Like other mammals, the aging rat brain also shows decline in numbers or densities of cerebral cortical and hippocampal neurons [23]. Our behavioural observations of very old (>700 day old) rats indicate they display many of the characteristics of geriatric progression noted in the human being, including repetition of the same behaviour, particular deficits in memories involved with spatial relationships, and diminished species-specific interaction, important for group bonding. Young adult rats, in which seizures have been induced by lithium and pilocarpine, and who are then treated with acepromazine or other neuroleptics that promote survival, show neuronal densities in the cortices and hippocampus that indicate they are behaving in a manner similar to rats over the age of two years [24]. Here, we present clear evidence that a discrepancy in the linear slope of neuronal decline with aging occurs around 700 days of age, which is equivalent to the human age of about 75 years of age, and that layers 5 and 6 within the prefrontal cortices are particularly affected.

Experimental Procedures

A total of 32 cerebrums from Wistar albino rats, primarily female, were selected from our collection of brains. After weaning, the rats had been housed (2 to 3 per cage) at ambient room temperature, with a 12 hour light and dark cycle. Purina rat chow and water were available ad

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Received March 28, 2013; Accepted April 12, 2013; Published April 15, 2013

Citation: Karbowksi LM, Lafrenie RM, Persinger MA (2013) Non-Linear Decline in Neuronal Density within the Cerebral Cortices and Hippocampus in very Aged Rats: Implications for Causality. Cell Dev Biol 2: 125. doi:10.4172/2168-9296.1000125

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libitum throughout the lifespan of the rats. Selected brains had been fixed in ethanol-formalin acetic acid, as a component of our long-term aging study. Whereas rats less than 400 days had been sacrificed as controls for various studies, all of the rats older than 400 days were sacrificed when they reached the criteria for near terminal conditions, as prescribed by the University's Animal Care Committee and the policies of the Canadian Council on Animal Care.

After clearing and infiltration, the cerebrums were embedded in paraffin. They were each sectioned at 10 μm and stained with toluidine blue O, to include the same or very similar region of the occipital, parietal, temporal, and two frontal cortices, as defined by Paxinos and Watson [25]. Fp1 (M1) and Fp2 (M2) were selected because they were most representative of simple motor (Fp1) vs more "prefrontal" (Fp2) locations.

The numbers of neuronal soma in each of the six layers for each of the three caudal (temporal, parietal, occipital) locations and five layers for the frontal cortices were counted at 400X, using a 6x6 grid as reference. The summed area for the grid at 400X was equivalent to 0.09 mm^2 . Because there is no discernable layer 4 in the frontal cortices, the numbers of neurons in lower portion of layer three and upper portion of layer 5 were averaged for this layer, for the purposes of statistical analyses. The means of three separate samples within the same region of each layer for each location were computed, and employed as data. Correlation, partial correlation, analyses of variance, regression, factor analyses were completed on PC SPSS 16.

Results

Figure 1 shows the relationship between the mean numbers of neurons (per .09 mm^2), in all layers, for all the sampled regions of the cerebral cortices as a function of age. Histological examples from the prefrontal (Fp2) cortices are shown in figure 2. A similar pattern was noted (Figure 3) for the mean numbers of neurons (per 0.09 mm^2), within the CA fields of the hippocampus. Histological examples representing the age effect upon neuronal density is shown in figure 4.

The visible inflection point for accelerated diminishment of numbers of neurons after 700 days of age was assessed by regression analyses, to discern slopes. For the mean numbers cerebral cortical neurons, the slopes before and after the inflection point were -.014 and -.036, respectively (r values 0.97, 0.99, respectively). For the mean numbers of hippocampal neurons, the slopes before and after the inflection point were -.020 and -.053, respectively (r values 0.98, 0.98).

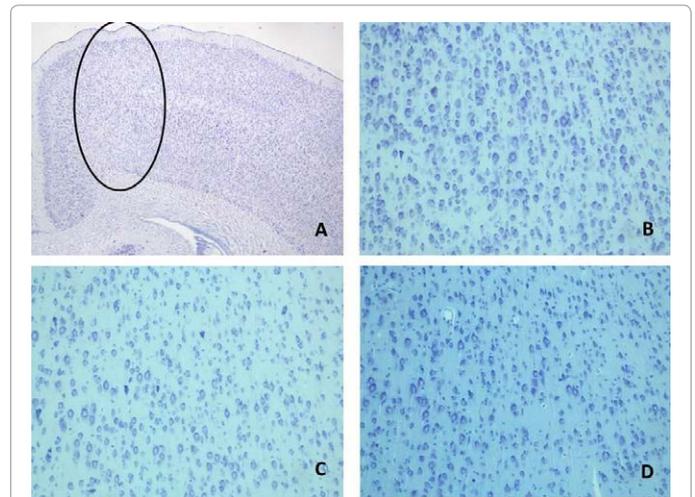
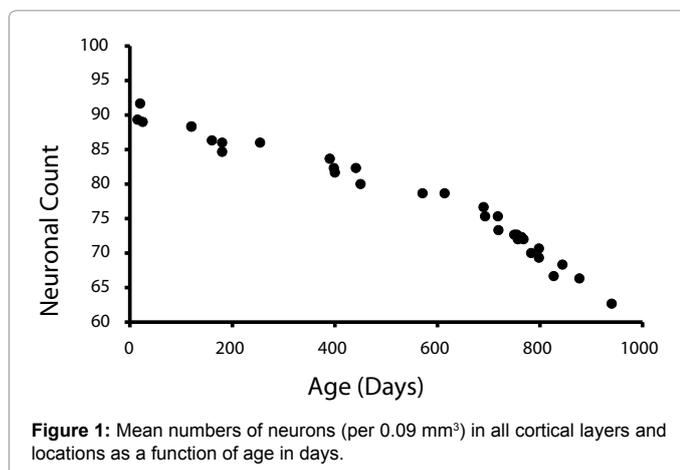


Figure 2: A) 40X magnification (toluidine Blue O) view of the Fp2 area used for comparing neuronal counts in aging rats B) 200X magnification view of Fp2 young aged rats C) 200X magnification view of Fp2 middle aged rats D) 200X magnification view of Fp2 old aged rats.

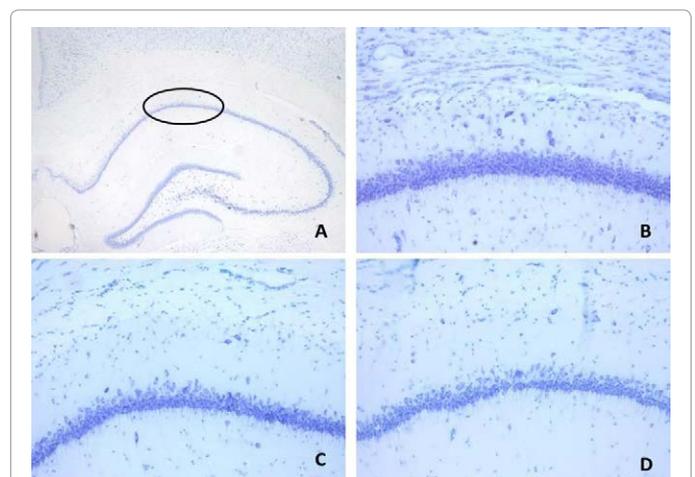
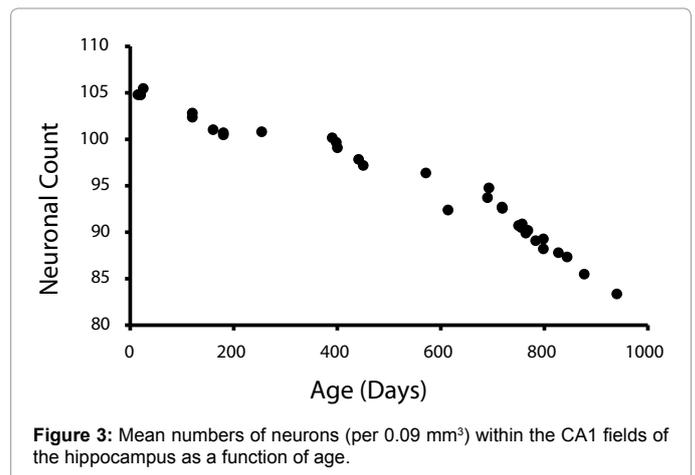


Figure 4: A) 40X magnification (toluidine Blue O) view of the hippocampus with the CA1 area indicated for comparing neuronal counts in aging rats B) 200X magnification view of CA1 hippocampus in young aged rats C) 200X magnification view of CA1 hippocampus in middle aged rats D) 200X magnification view of CA1 hippocampus in old aged rats.

Even though the sample size was limited to 32 brains, an exploratory factor analyses was completed for numbers of neurons in all cortical areas and layers, in order to find general relationships. A total of 6 factors emerged with the first factor (Eigen value=15.55; 52% variance explained), along with the second factor (Eigen value=2.39; 8% of variance explained), and third (Eigen value=1.78; 6% of variance explained), accommodating two-thirds of all of the variance. The loading coefficients (correlations) of each variable for the three factors are shown in table 1. Given the sample size, we accepted only $r_s > 0.7$ as significant, statistically. In general, the common theme involved shared variance between neurons in layers 3 through 6 for most layers. Factor analyses for the three hippocampal CA fields (not shown), resulted in one factor.

The relationship between the factor scores for the first factor, for the numbers of neurons in the cerebral cortices, in comparison with the factor score for hippocampal neurons, are compared in figure 5. Although the initial raw scores for numbers of neurons differed between the hippocampus and the cortices, the best-fit central tendency of the z-scores for the decline of the two populations with age were effectively identical.

The intercorrelation coefficients were: age-cortices: -0.98, age-hippocampus: -0.97, and hippocampus-cortices: 0.99. To address this "identity" problem, partial correlation analyses (all dfs=29) were completed. After removal of the shared variance with the third variable, the first order partial correlations were: age-cortices: -0.38 ($p < .05$), hippocampus-cortices: -0.27 (n.s.), and age-hippocampus: 0.77 ($p < .001$).

Area	Factor 1	Factor 2	Factor 3
F1 L1	0.187	0.558	0.055
F1 L2	0.051	0.430	0.703
F1 L3	0.941	0.208	0.087
F1 L4	0.861	0.294	0.154
F1 L5	0.966	0.160	0.067
F1 L6	0.929	0.226	0.106
F2 L1	0.217	0.725	0.145
F2 L2	0.389	0.633	0.088
F2 L3	0.838	0.428	0.107
F2 L4	0.791	0.327	0.173
F2 L5	0.909	0.346	0.045
F2 L6	0.940	0.246	0.132
T L1	0.141	0.093	0.030
T L2	0.518	-0.130	0.535
T L3	0.771	0.009	0.325
T L4	0.643	0.243	-0.070
T L5	0.923	0.226	-0.011
T L6	0.920	0.228	0.129
P L1	0.581	0.412	-0.251
P L2	0.212	0.062	0.832
P L3	-0.034	-0.149	-0.119
P L4	0.687	-0.094	0.312
P L5	0.764	0.473	0.138
P L6	0.606	0.269	0.372
O L1	0.190	0.045	0.093
O L2	0.519	0.594	0.273
O L3	0.650	0.256	0.205
O L4	0.789	-0.203	0.034
O L5	-0.037	0.137	-0.039
O L6	0.775	-0.042	0.171

Table 1: Loading (correlation coefficients) for the various layers and areas for the first three major factors for numbers of neurons.

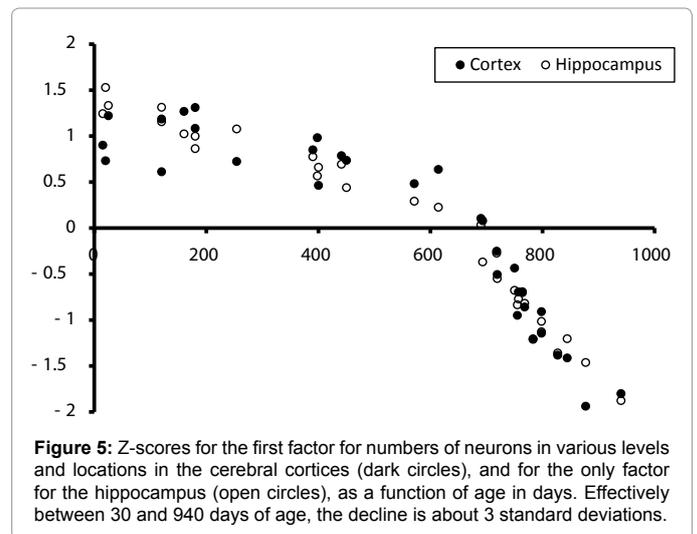


Figure 6 shows a comparison of the mean numbers of neurons within the five locations, as a function of aging. The Fp2 and temporal cortices displayed the most congruent decrease (in slope), while the occipital cortices displayed the least decrease, although the negative drift was still apparent.

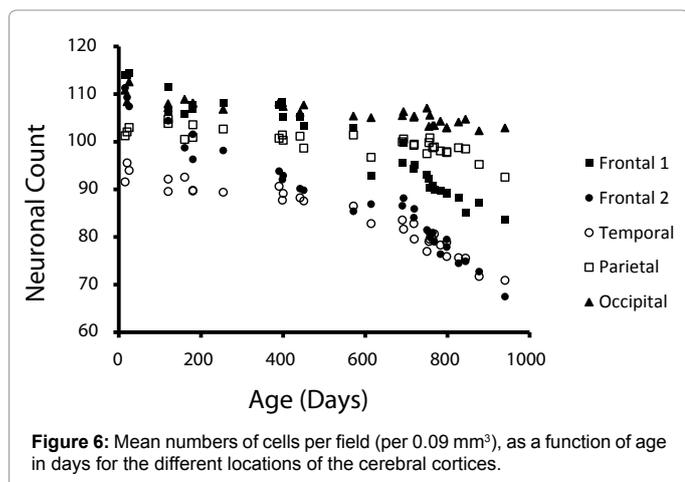
In order to discern recondite interactions between numbers of neurons within cerebral layers and locations as a function of age, the brains were designated to 4 groups: adolescent, young (3) adult (6), middle aged adult (6), and very old adult (17). The mean numbers of cells per layer (for all regions) and mean numbers of cells per region (layers combined), were analyzed by analyses of variance. Because group differences (all dfs=3,28) were highly significant statistically (all $F_s > 8.00$), omega-squared estimates (effect sizes) were obtained for comparison. The results are shown in table 2.

A three way analyses of variance, with two levels repeated (layers and locations), and one between subject level (age), showed a significant ($F(3,28)=61.83$, $p < .001$; eta-squared=0.87) main effect for age differences. All of the two way and the three way interactions were statistically significant; for example: the interaction between age, location and layer was quite strong ($F(60,560)=22.17$, $p < .001$).

Discussion

The aging cell, including the neuron, undergoes a variety of biochemical modifications from internal (molecular pathways) and external (vascular availability) sources that result in diminished metabolic capacity [26,27]. The consequences are the classic "die back" of more distal dendrites, and general reduction of soma volume. The removal of dendritic connections, the presumed major correlate of information storage within the cerebral volume, has been considered to be the major cause of the loss of behavioral capacity and cognitive abilities [28,29]. These changes in the human being become very conspicuous after 75 years of age, and increase at an accelerated rate compared to previous decades.

The results of our quantitative analyses for geriatric rat brain are remarkably consistent with the gross clinical observations noted in human populations [30]. The mean numbers of neurons as a function of age, within both the cerebral cortices and the hippocampal formation, displayed a clear inflection downwards at around 700 days of age. The slopes were similar for both populations of neurons. Quantitatively after the age of 700 days, the loss increased by a factor



Area	Omega ² %	Pups		Young		Middle-Age		Old	
		M	SD	M	SD	M	SD	M	SD
Layer 1	62	18	1	14	2	12	1	12	2
Layer 2	47	75	1	70	2	69	1	67	4
Layer 3	79	151	2	156	1	142	1	135	4
Layer 4	67	112	2	109	1	107	1	102	3
Layer 5	87	135	1	133	2	126	3	113	5
Layer 6	85	145	1	143	1	133	4	114	7
Frontal 1	89	114	1	108	2	105	2	91	4
Frontal 2	85	109	2	101	4	91	3	80	6
Temporal	83	94	2	91	1	88	1	78	4
Parietal	56	102	1	103	2	101	1	98	2
Occipital	74	111	2	108	1	107	1	104	1

Table 2: Descriptive values for the average number of neurons (per 0.09 mm³) per layer and per region (all layers combined).

of about 2.5 (250%), in both the cerebral cortices, in general, and the hippocampal formation. Comparisons of the factor scores that removed the individual differences in values for neurons in these two locations indicated the intrinsic age-related processes were effectively identical (Figure 3).

The decline in factor scores, representing the shared variance in numbers of neurons within the CA hippocampal fields and the cerebral cortices, was equivalent to 3 standard deviations over the age span of the youngest brain (30 days of age) to the oldest brain (940 days). This translated into the quantitative equivalent of a loss of about 25% of total mean numbers of neurons in the cerebral cortices and the hippocampus. In other words, the majority of neuronal numbers compared to very young brains were still present, despite the loss of capacity in these very old rats. This quantitative proportion would be consistent with the contention, there is a “critical” mass or essential number of neurons required for the emergent properties that define the qualitative aspects of a species.

Although the marked congruence of the non-linear decline in factor scores for both the cerebral cortices and hippocampus were similar, the results of the partial correlation analyses were revealing. When three or more variables are strongly intercorrelated, there is often one intercorrelation whose presence is epiphenomenal, because of its recondite relation to other variables. For example: the bivariately obvious association between cholesterol and certain pathologies of the lumen of arteries was found to be actually more related to triglycerides. The cholesterol connection was because these levels were

primarily determined by the triglycerides, which were (at the time) less discernable.

In the present study, the residuals from covarying for the other locus of neurons, before either the hippocampal formation or cerebral cortices, was correlated with age, showed that association between age and neurons in the latter was no longer significant. However, when the shared variance with neurons within the cortices was first removed, the hippocampal-age connection remained strongly correlated and statistically significant.

Classical interpretation of such a relationship suggests that the decline of neurons in the hippocampal formation is primarily responsible for the similarly proportional decline of neurons within the cerebral cortices. Such an interpretation would be consistent with the general models, that the cerebral cortices evolved as an additional volume that allows “storage” of information [3,15], first represented and then accessed from the hippocampal formation [31], following the transient electrical lability of about 20 min, during which time, spine emergence occurs in those regions affected by the experience [23]. It would also be consistent with the fact that the cortical thickness has not changed appreciably in the evolution of mammals, and that the increased volume of the hippocampal formation and cerebral volume are strongly associated [1,8,12].

The dependence of neuronal density or a subpopulation of neurons, within the cerebral cortices upon hippocampal neurons, could accommodate the general patterns of degenerative activity that occurs during multiple forms of senile dementia. Denuding of the hippocampal formation from other brain regions is the first major stage of senility, particularly presenile dementias [32]. Indeed, the loss of neurons within the basal nucleus of Meynert, and the diminished cholinergic input to the cerebral cortices, particularly the prefrontal regions, are strong parallel occurrences. Our results suggest that loss of neurons within the hippocampal formation would be the initiating process that affected the subcortical telencephalic-cerebral cortical network. We predict that the neurochemical identification of the neurons most lost in the aging brain would be those related to cholinergic- dominated neurons.

The other major change we observed in very old rats is that the prefrontal cortices and the temporal cortices were disproportionately affected by the latter stages of aging. The slope of the decline in neurons was visually obvious compared to other areas, such as the occipital cortices, after 700 days of age. If analogous changes occur in the human brain, then one would expect the greater loss of accuracy and potency of autobiographical memory, because of its strong reliance upon the left prefrontal region for encoding, and the right prefrontal region for organization of the reconstruction of memory [33].

The layers of the cortices that were most affected were 5 and 6. They contain the neurons that are intimately associated with the feedback loops from the thalamic nuclei, and to the caudate-putamen [34]. These loops are important for stimulus-response association for various sensory modalities [34]. From some perspectives, they might also be considered the base information from which the upper cortical layers and their greater interhemispheric interactivity access information that contributes to higher cognitive and behavioural function.

Enhanced loss of neurons within the lower cortical layers could also be associated with general diminishment of activation to sensory stimuli [34]. This effect would be congruent with the fMRI studies that showed older individuals show less activation to stimuli [32]. With less activation to “novel” stimuli, more complex and sensitive pathways

could be more prone to a type of “transneuronal” degeneration from diminished input. Decline in prefrontal neurons in the lower layers would affect the intrinsic circuitry of the basal ganglia.

Quantitative studies of neurons within the layers of the cerebral cortices from different regions are labour-intensive and time consuming. Although chemical indicators are attractive, and molecular pathways within aging cells populations are critical to understanding the biochemistry of senescence, the absolute numbers of neurons and how they reflect normal aging, might contain sufficient macroscopic information and patterns that reflect the details within the bimolecular pathways, for which some interventions could be developed.

Acknowledgements

We thank the Laurentian University Neuroscience Research Group, whose dedication for maintaining the health and longevity of the rats, despite pressures from external bodies, cost many of them their careers and opportunities.

Author Contributions

Mr. Lukasz M Karbowksi completed the histology and cytometry. Dr. Michael A Persinger and Robert M Lafrenie were involved with the design of the experiment, the data analyses, and the composition of the manuscript.

References

- Hof PR, Morrison JH (2004) The aging brain: morphomolecular senescence of cortical circuits. *Trends Neurosci* 27: 607-613.
- Kremen WS, O'Brien RC, Panizzon MS, Prom-Wormley E, Eaves LJ, et al. (2010) Salivary cortisol and prefrontal cortical thickness in middle-aged men: A twin study. *Neuroimage* 53: 1093-1102.
- Bear MF (1996) A synaptic basis for memory storage in the cerebral cortex. *Proc Natl Acad Sci U S A* 93: 13453-13459.
- Petrides M (1985) Deficits on conditional associative-learning tasks after frontal- and temporal-lobe lesions in man. *Neuropsychologia* 23: 601-614.
- Smith ML, Milner B (1981) The role of the right hippocampus in the recall of spatial location. *Neuropsychologia* 19: 781-793.
- Auer RN, Jensen ML, Whishaw IQ (1989) Neurobehavioural deficit due to ischemic brain damage limited to half of the CA1 sector of the hippocampus. *J Neurosci* 9: 1641-1647.
- Buffalo EA, Reber PJ, Squire LR (1998) The human perirhinal cortex and recognition memory. *Hippocampus* 8: 330-339.
- Eichenbaum H, Schoenbaum G, Young B, Bunsey M (1996) Functional organization of the hippocampal memory system. *Proc Natl Acad Sci U S A* 93: 13500-13507.
- Squire LR, Ojemann JG, Miezin FM, Petersen SE, Videen TO, et al. (1992) Activation of the hippocampus in normal humans: a functional anatomical study of memory. *Proc Natl Acad Sci U S A* 89: 1837-1841.
- Cave CB, Squire LR (1991) Equivalent impairment of spatial and nonspatial memory following damage to the human hippocampus. *Hippocampus* 1: 329-340.
- Klatsky RL, Loomis JM, Beall AC, Chance SS, Golledge RG (1998) Spatial updating of self-position and orientation during real, imagined and virtual locomotion. *Psychol Sci* 9: 1648-1659.
- Henson RNA, Shallice T, Dolan RJ (1999). Right prefrontal cortex and episodic memory retrieval: a functional MRI test of the monitoring hypothesis. *Brain* 122: 1367-1381.
- Haug H, Knebel G, Mecke E, Orün C, Sass NL (1981) The aging of cortical cytoarchitectonics in the light of stereological investigations. *Prog Clin Biol Res* 59B: 193-197.
- Henderson G, Tomlinson BE, Gibson PH (1980) Cell counts in human cerebral cortex in normal adults throughout life using an image analysing computer. *J Neurol Sci* 46: 113-136.
- Dekaban AS (1978) Changes in brain weights during the span of human life: relation of brain weights to body heights and body weights. *Ann Neurol* 4: 345-356.
- Matsumae M, Kikinis R, Mórocz IA, Lorenzo AV, Sándor T, et al. (1996) Age-related changes in intracranial compartment volumes in normal adults assessed by magnetic resonance imaging. *J Neurosurg* 84: 982-991.
- Raz N, Briggs SD, Marks W, Acker JD (1999) Age-related deficits in generation and manipulation of mental images: II. The role of dorsolateral prefrontal cortex. *Psychol Aging* 14: 436-444.
- Brody H (1955) Organization of the cerebral cortex. III. A study of aging in the human cerebral cortex. *J Comp Neurol* 102: 511-516.
- Terry RD, DeTeresa R, Hansen LA (1987) Neocortical cell counts in normal human adult aging. *Ann Neurol* 21: 530-539.
- Meier-Ruge W, Hunziker O, Iwango P, Reichlmleiter K, Sandoz P (1978) Alteration of morphological and neurochemical parameter of the brain due to normal aging. In: *Senile dementia: A Biochemical approach*, Nandy K (Ed.), Elsevier, New York, USA.
- Düzel E, Habib R, Rotte M, Guderian S, Tulving E, et al. (2003) Human hippocampal and parahippocampal activity during visual associative recognition memory for spatial and nonspatial stimulus configurations. *J Neurosci* 23: 9439-9444.
- West MJ (1993) Regionally specific loss of neurons in the aging human hippocampus. *Neurobiol Aging* 14: 287-293.
- Diamond MC, Johnson RE, Ingham CA (1975) Morphological changes in the young, adult and aging rat cerebral cortex, hippocampus, and diencephalon. *Behav Biol* 14: 163-174.
- Peredery O, Persinger MA, Parker G, Mastrosov L (2000) Temporal changes in neuronal dropout following inductions of lithium/pilocarpine seizures in the rat. *Brain Res* 881: 9-17.
- Paxinos G, Watson C (1986) *The rat brain in stereotaxic coordinates*. (2nd edn), Academic Press, California, USA.
- Chung HY, Cesari M, Anton S, Marzetti E, Giovannini S, et al. (2009) Molecular inflammation: underpinnings of aging and age-related diseases. *Ageing Res Rev* 8: 18-30.
- Tranah GJ (2011) Mitochondrial-nuclear epistasis: implications for human aging and longevity. *Ageing Res Rev* 10: 238-252.
- Feldman ML, Dowd C (1975) Loss of dendritic spines in aging cerebral cortex. *Anat Embryol (Berl)* 148: 279-301.
- Timiras PS (1994) *Aging of the nervous system: functional changes*. In: *Physiological basis of aging and geriatrics*, Timiras PS (Ed.), CRC Press Inc., Florida, USA.
- Wong TP (2002) Aging of the cerebral cortex. *MJM* 6: 104-113.
- Burgess N, Maguire EA, O'Keefe J (2002) The human hippocampus and spatial and episodic memory. *Neuron* 35: 625-641.
- Wolf H, Grunwald M, Kruggel F, Riedel-Heller SG, Angerhöfer S, et al. (2001) Hippocampal volume discriminates between normal cognition; questionable and mild dementia in the elderly. *Neurobiol Aging* 22: 177-186.
- Buckner RL, Petersen SE (1996) What does neuroimaging tell us about the role of the prefrontal cortex in memory retrieval? *Seminars in Neuroscience* 8: 47-55.
- Finch DM (1996) Neurophysiology of converging synaptic inputs from the rat prefrontal cortex, amygdala, midline thalamus, and hippocampal formation onto single neurons of the caudate/putamen and nucleus accumbens. *Hippocampus* 6: 495-512.