

Natural Antivirals against Human Viruses

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Editorial

Over the past two decades there has been intense study of antivirals from plants that have antiviral activity against viruses affecting plant and animal kingdoms. This volume of Virology and Mycology has a report by Awasthi et al. [1] on the antiviral activity against plant mosaic viruses affecting commercial important bean plants. A number of viruses that can cause diseases in humans can be either neutralized or its replication can be inhibited. There has been an explosion of research in the antiviral activity in recent years against human diseases caused by pathogenic viruses, which are known to affect millions of people across the globe and cause significant morbidity and mortality. As a champion of evidence based natural products of preventive and therapeutic agents, I would like to dedicate this editorial to those who have labored on researching natural products against all odds, often experiencing a hostile and elitist opposition to the discovery of natural products. The challenges to translating the findings of natural products are many and this editorial will conclude with steps one needs to take in order to obtain regulatory approval from agencies like the FDA and the EU regulatory group. Very few of the newly found compounds are anywhere close to finding human use. The one Secomet V (the active agent of which is Fulvic acid) that has reached human observational trial stage in HIV patients reported by Van Rensburg et al. in 2010 is a precursor to more proper trials, which have been presumably underway [2]. But the promise of Secomet V, a broad spectrum natural product, first reported by Kotwal et al., [3] takes exceedingly long periods to go from bench (discovery and laboratory testing) to bedside (administered in patients). This passage of over a decade is due to the underfunding of natural products evidence based research in the western world and the low impact they make in the publication arena in top medical journals like Nature Medicine, who look down upon the struggling researchers from the natural products field. Then there are those like pomegranate, found and consumed universally and known to have health benefit and documented to have broad spectrum activity (Table 1) against enveloped viruses like HIV, Influenza, Herpes and non-enveloped food borne viruses Noroviruses, have yet to establish therapeutic levels for human use, making them less practical for realization of their valuable potential. There are a number of naturally found compounds that have been identified against medically important viruses, which together contribute to annual infections of over billion people around the globe, and have been grouped here and summarized in tables. They include the antivirals to human hepatitis viruses (Table 2), viruses causing respiratory infections (Table 3), picornaviruses which cause a number of diseases (Table 4), human herpes viruses (Table 5) and Human Immunodeficiency virus (Table 5). In addition to the medically important viruses included in the tables there are other viruses of pediatric importance viz the Rota virus which causes diarrhea leading to severe dehydration and significant infant deaths also has a natural antiviral that has been identified [4]. With such a wide range of a repertoire of identified antivirals from natural sources like plants, what are the possibilities of how they could find human use? As in the case of Fulvic acid and pomegranate juice, they could be therapeutically used to neutralize pathogenic viruses like HIV, Influenza in human tissues and thus described as the enveloped virus neutralizing compounds (EVNCs). The advantage of such compounds is that they are not susceptible to the vagaries of mutations and drug resistance due to their mode of action, which is not dependent on

the changes in amino acid sequence due to mutations [5]. EVNCs could be used in prophylaxis, as vaccine generating agents and maybe even as microbicides. Although microbicides have yet to realize their promise in providing women control over their sexual activity while ensuring that transmission is blocked. Finally, what steps will have to be taken following identification of antiviral activity? Guidelines are summarized in Table 7. The science preceding the translation has to be robust, if not the long path to human use can be costly waste of time and resources. It will be evident from the work in the field of natural antivirals that scientists from China are the leaders in the field and hopefully will continue to receive support to take on the long journey to finding their way into human use. A decade ago, in an editorial, I had envisioned that HIV could be eradicated by 2050 [49]. But amongst the medicines that could achieve that goal, I had an expectation that natural medicine would play a significant role. After reviewing the

Antiviral/Active Ingredient	Source	Antiviral Activity	Reference
Fulvic Acid	Sucrose, Carbohydrate	Enveloped viruses	[2,3,5,6]
Pomegranate Polyphenols, juice	<i>Punicagranatum</i> (Pomegranate)	Enveloped viruses, Food borne surrogate viruses	[5,7-10]
Resveratrol+	Red vine leaves, Japanese knotweed	HIV (in comb. With nucleoside analogues) EBV	[11,12]
Ethanol extract	<i>Warscewiczia coccinea</i> (Vahl) Kl. (Rubiaceae)	HIV and HBV	[13]

Table 1: Broad Spectrum Natural Antivirals against human viruses.

Antiviral/Active Ingredient	Source	Antiviral Activity	Reference
Sesquiterpenoids and alkaloids	Roots of <i>Alangium chinense</i>	HCV	[14]
Oleanolic acid and ursolic acid	<i>Fructus Ligustri Lucidi</i>	HCV	[15]
Longumosides and amide alkaloids	<i>Piper Longum</i>	HBV	[16]
epigallocatechin Lucidone	Green Tea	HBV	[17]
Lectins	<i>Lindera erythrocarpa</i>	HCV	[18]
3-hydroxy caruilignan	Red&Blue green algae <i>Swietenia macrophylla</i> stems	HCV HCV	[19,20]
Ladanein (BJ486K), a flavonoid	<i>Marrubium peregrinum</i>	All HCV genotypes	[21]
Hypericin	John's Wort	HCV	[22]

Table 2: Natural Antivirals against human hepatitis viruses.

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Antiviral/Active Ingredient	Source	Antiviral Activity	Reference
Cimifugin Carnosic acid	<i>Cimicifuga foetida</i> , <i>Rosmarinus officinalis</i>	RSV RSV	[23,24]
Polybromocatechol	Rhodophyta, <i>Neorhododendron laevigatum</i>	Rhinovirus	[25]
Sesquiterpenecoumarins	<i>Ferula assafoetida</i>	Influenza	[26]
Herbal marine compound (HESA-A)	Herbal marine	Influenza	[27]
Chalcones	Glycyrrhiza leaflets	Influenza	[28]
Liquid extract	Elderberry	Influenza	[29]
diarylheptanoids	<i>Alpinia katsumadai</i>	Influenza	[30]

Table 3: Natural Antivirals against human respiratory viruses

Antiviral/Active Ingredient	Source	Antiviral Activity	Reference
Diterpenoids, sesquiterpenoids	Roots of <i>Illicium majus</i>	Coxsackie B3	[31]
Sesquiterpenoids and alkaloids	Roots of <i>Alangium chinense</i>	Coxsackie B3	[14]
Polysaccharides	<i>Azadirachta indica</i>	Poliovirus	[32]
Gallic acid	<i>Woodfordia fruticosa</i> flowers	Enterovirus 71	[33]
Raoulic acid	<i>Raoulia australis</i>	Picornaviruses	[34]
Punicagalin		Enterovirus 71	[35]

Table 4: Natural Antivirals against Picornaviruses.

Antiviral/Active Ingredient	Source	Antiviral Activity	Reference
Sulfated galactans	<i>Cryptopleuramarosa</i> (red sea weed)	HSV-1 and HSV-2 replication in Vero	[36]
Rhamnogalacturonan	Cell wall Pectin polysaccharides	HCMV cytotoxicity	[37]
Chebulagic acid and punicalagin (Hydrolyzable Tannins)	<i>Terminalia chebula</i> Retz (dried fruits)	HSV-1 and HSV-2 entry and spread	[38]
Verbascoside	Lepechinia species	HSV-1 and HSV-2	[39]
Bark extract	<i>Azadirachta indica</i> (Neem)	HSV-1	[39]
Extract	Blackberry	HSV-1	[40]

Table 5: Natural Antivirals against human Herpes viruses.

Antiviral/Active Ingredient	Source	Antiviral Activity	Reference
Griffithsin	<i>Griffithsia</i> (Red algae)	HIV clade C in comb. With Tenofovir, maraviroc and enfuvirtide	[41]
Quinoline alkaloids, Theaflavins	<i>Euodiaroxburghiana</i>	HIV entry	[42]
Diterpenes, Extract	<i>Coleus forskohlii</i> , Dandelion, <i>Coleus parvifolius</i>	HIV replication, RT, HIV integrase	[43,44]
Niruriside	Phyllanthum	HIV REV/RRE inhib.	[45]
Coumarin & xanthone	Calophyllum	HIV RT	[46]
Recombinant MAP 30	Bitter melon	HIV	[47,48]

Table 6: Natural Antivirals against HIV.

1. Test for antiviral activity in a large number of natural sources/extracts in an appropriate cell culture assay.
2. Determine the highest concentration at which there is no cell toxicity.
3. Test for reduction of infectious virus counts
4. Identify the mechanism of action.
5. Identify the compound/s that has the activity and determine the structure.
6. Chemically synthesize the compound, determine the structure and test for activity.
7. Perform safety, efficacy and toxicity assays on the active compound in an experimental animal model
8. Go through the national regulatory bodies to perform observational and phase 1-4 human trials

Table 7: Steps in potential therapeutic use of natural antivirals against disease causing human viruses.

progress in just the past 3 year, I am optimistic but the determination to take on the challenges may have to be very strong and the support from their respective countries to the scientists on this frontier has to be absolute and unconditional.

References

- Awasthi LP, Singh SP, Verma HN, Kluge S (2013) Further studies on the Antiviral Agent isolated from Host plants, pre-treated with Boerhaaviadiffusa Glycoprotein. VirolMycol 3:124.
- Van Rensburg CEJ, Gandy JJ and Snyman (2010) Letter: An observational trial: patient profile of users of Secomet V. SA FamPract 52: 165.
- Kotwal GJ, Kaczmarek JN, Leivers S, Ghebremariam YT, Kulkarni AP, et al. (2005) Anti-HIV, anti-poxvirus, and anti-SARS activity of a nontoxic, acidic plant extract from the Trifolium species Secomet-V/anti-vac suggests that it contains a novel broad-spectrum antiviral. Ann N Y AcadSci 1056: 293-302.
- Tam KI, Roner MR (2011) Characterization of in vivo anti-rotavirus activities of saponin extracts from Quillajasaponaria Molina. Antiviral Res 90: 231-241.
- Kotwal GJ (2008) Genetic diversity-independent neutralization of pandemic viruses (e.g. HIV), potentially pandemic (e.g. H5N1 strain of influenza) and carcinogenic (e.g. HBV and HCV) viruses and possible agents of bioterrorism (variola) by enveloped virus neutralizing compounds (EVNCs). Vaccine 26: 3055-3058.
- Ballardin M, Scarpato R, Kotwal GJ, Barale R (2005) In vitro mutagenicity studies of the antiretrovirals AZT, Didanosine, and 3TC and a plant antiviral extract Secomet-V derived from the Trifolium species. Ann N Y AcadSci 1056: 303-310.
- Neurath AR, Strick N, Li YY, Debnath AK (2004) Punicagranatum (Pomegranate) juice provides an HIV-1 entry inhibitor and candidate topical microbicide. BMC Infect Dis 4: 41.
- Neurath AR, Strick N, Li YY, Debnath AK (2005) Punicagranatum (pomegranate) juice provides an HIV-1 entry inhibitor and candidate topical microbicide. Ann N Y AcadSci 1056: 311-327.
- Su X, Sangster MY, D'Souza DH (2010) In vitro effects of pomegranate juice and pomegranate polyphenols on foodborne viral surrogates. Foodborne Pathog Dis 7: 1473-1479.
- Sundararajan A, Ganapathy R, Huan L, Dunlap JR, Webby RJ, et al. (2010) Influenza virus variation in susceptibility to inactivation by pomegranate polyphenols is determined by envelope glycoproteins. Antiviral Res 88: 1-9.
- Heredia A, Davis C, Redfield R (2000) Synergistic inhibition of HIV-1 in activated and resting peripheral blood mononuclear cells, monocyte-derived macrophages, and selected drug-resistant isolates with nucleoside analogues combined with a natural product, resveratrol. J Acquir Immune DeficSyndr 25:246-255.
- De Leo A, Arena G, Lacanna E, Oliviero G, Colavita F, et al. (2012) Resveratrol inhibits Epstein Barr Virus lytic cycle in Burkitt's lymphoma cells by affecting multiple molecular targets. Antiviral Res 96: 196-202.
- Quintero A, Fabbro R, Maillo M, Barrios M, Milano MB, et al. (2011) Inhibition of hepatitis B virus and human immunodeficiency virus (HIV-1) replication by Warscewiczia coccinea (Vahl) Kl. (Rubiaceae) ethanol extract. Nat Prod Res 25: 1565-1569.
- Zhang Y, Liu YB, Li Y, Ma SG, Li L, et al. (2013) Sesquiterpenes and alkaloids from the roots of Alangium chinense. J Nat Prod 76: 1058-1063.
- Kong L, Li S, Liao Q, Zhang Y, Sun R, et al. (2013) Oleanolic acid and ursolic acid: novel hepatitis C virus antivirals that inhibit NS5B activity. Antiviral Res 98: 44-53.
- Jiang ZY, Liu WF, Zhang XM, Luo J, Ma YB, et al. (2013) Anti-HBV active constituents from Piper longum. Bioorg Med ChemLett 23: 2123-2127.

17. Xu J, Wang J, Deng F, Hu Z, Wang H (2008) Green tea extract and its major component epigallocatechingallate inhibits hepatitis B virus in vitro. *Antiviral Res* 78: 242-249.
18. Chen WC, Wang SY, Chiu CC, Tseng CK, Lin CK, et al. (2013) Lucidone suppresses hepatitis C virus replication by Nrf2-mediated heme oxygenase-1 induction. *Antimicrob Agents Chemother* 57: 1180-1191.
19. Takebe Y, Saucedo CJ, Lund G, Uenishi R, Hase S, et al. (2013) Antiviral lectins from red and blue-green algae show potent in vitro and in vivo activity against hepatitis C virus. *PLoS One* 8: e64449.
20. Wu SF, Lin CK, Chuang YS, Chang FR, Tseng CK, et al. (2012) Anti-hepatitis C virus activity of 3-hydroxy caruillignan C from *Swieteniamacrophylla* stems. *J Viral Hepat* 19: 364-370.
21. Haid S, Novodomská A, Gentzsch J, Grethe C, Geuenich S, et al. (2012) A plant-derived flavonoid inhibits entry of all HCV genotypes into human hepatocytes. *Gastroenterology* 143: 213-22.e5.
22. Jacobson JM, Feinman L, Liebes L, Ostrow N, Koslowski V, et al. (2001) Pharmacokinetics, safety, and antiviral effects of hypericin, a derivative of St. John's wort plant, in patients with chronic hepatitis C virus infection. *AntimicrobAgents Chemother* 45:517-524.
23. Wang KC, Chang JS, Lin LT, Chiang LC, Lin CC (2012) Antiviral effect of cimicifugin from *Cimicifugafoetida* against human respiratory syncytial virus. *Am J Chin Med* 40: 1033-1045.
24. Shin HB, Choi MS, Ryu B, Lee NR, Kim HI, et al. (2013) Antiviral activity of carnosic acid against respiratory syncytial virus. *Virol J* 10: 303.
25. Park SH, Song JH, Kim T, Shin WS, Park GM, et al. (2012) Anti-human rhinoviral activity of polybromocatechol compounds isolated from the rhodophyta, *Neorhodomelaaculeata*. *Mar Drugs* 10: 2222-2233.
26. Lee CL, Chiang LC, Cheng LH, Liaw CC, Abd El-Razek MH, et al. (2009) Influenza A (H1N1) Antiviral and Cytotoxic Agents from *Ferula assa-foetida*. *J Nat Prod* 72: 1568-1572.
27. He W, Han H, Wang W, Gao B (2011) Anti-influenza virus effect of aqueous extracts from dandelion. *Virol J* 8: 538.
28. Dao TT, Nguyen PH, Lee HS, Kim E, Park J, et al. (2011) Chalcones as novel influenza A (H1N1) neuraminidase inhibitors from *Glycyrrhizainflata*. *Bioorg Med ChemLett* 21: 294-298.
29. Krawitz C, Mraheil MA, Stein M, Imirzalioglu C, Domann E, et al. (2011) Inhibitory activity of a standardized elderberry liquid extract against clinically-relevant human respiratory bacterial pathogens and influenza A and B viruses. *BMC Complement Altern Med* 11:16.
30. Grienke U, Schmidtknecht M, Kirchmair J, Pfarr K, Wutzler P, et al. (2010) Antiviral potential and molecular insight into neuraminidase inhibiting diarylheptanoids from *Alpinia katsumadai*. *J Med Chem* 53: 778-786.
31. Wang YD, Zhang GJ, Qu J, Li YH, Jiang JD, et al. (2013) Diterpenoids and sesquiterpenoids from the roots of *Illiciummajus*. *J Nat Prod* 76: 1976-1983.
32. Faccin-Galhardi LC, Yamamoto KA, Ray S, Ray B, CarvalhoLinhares RE, et al. (2012) The in vitro antiviral property of *Azadirachta indica* polysaccharides for poliovirus. *J Ethnopharmacol* 142: 86-90.
33. Choi HJ, Song JH, Park KS, Baek SH (2010) In vitro anti-enterovirus 71 activity of gallic acid from *Woodfordiafruticosa* flowers. *LettApplMicrobiol* 50: 438-440.
34. Choi HJ, Lim CH, Song JH, Baek SH, Kwon DH (2009) Antiviral activity of raoulic acid from *Raoulia australis* against Picornaviruses. *Phytomedicine* 16: 35-39.
35. Yang J, Li L, Tan S, Jin H, Qiu J, et al. (2012) A natural theaflavins preparation inhibits HIV-1 infection by targeting the entry step: potential applications for preventing HIV-1 infection. *Fitoterapia* 83: 348-355.
36. Carlucci MJ, Scolaro LA, Errea MI, Matulewicz MC, Damonte EB (1997) Antiviral activity of natural sulphatedgalactans on herpes virus multiplication in cell culture. *Planta Med* 63: 429-432.
37. Steinmassl M, Anderer FA (1996) Enhancement of human NK and LAK cytotoxicity against HCMV-infected cells by rhamnogalacturonan: specificity of reaction. *Viral Immunol* 9: 27-34.
38. Lin LT, Chen TY, Chung CY, Noyce RS, Grindley TB, et al. (2011) Hydrolyzable tannins (chebulagic acid and punicalagin) target viral glycoprotein-glycosaminoglycan interactions to inhibit herpes simplex virus 1 entry and cell-to-cell spread. *J Virol* 85:4386-4398.
39. Martins FO, Esteves PF, Mendes GS, Barbi NS, Menezes FS, et al. (2009) Verbascoside isolated from *Lepechiniaspeciosa* has inhibitory activity against HSV-1 and HSV-2 in vitro. *Nat Prod Commun* 4: 1693-1696.
40. Tiwari V, Darmani NA, Yue BY, Shukla D (2010) In vitro antiviral activity of neem (*Azadirachta indica* L.) bark extract against herpes simplex virus type-1 infection. *Phytother Res* 24: 1132-1140.
41. Danaher RJ, Wang C, Dai J, Mumper RJ, Miller CS (2011) Antiviral effects of blackberry extract against herpes simplex virus type 1. *Oral Surg Oral Med Oral Pathol Oral RadiolEndod* 112: e31-35.
42. Féir G, Palmer KE, Schols D (2011) Synergistic activity profile of griffithsin in combination with tenofovir, maraviroc and enfuvirtide against HIV-1 clade C. *Virology* 417: 253-258.
43. McCormick JL, McKee TC, Cardellina JH 2nd, Boyd MR (1996) HIV inhibitory natural products. 26. Quinoline alkaloids from *Euodiaroxburghiana*. *J Nat Prod* 59: 469-471.
44. Han H, He W, Wang W, Gao B (2011) Inhibitory effect of aqueous Dandelion extract on HIV-1 replication and reverse transcriptase activity. *BMC Complement Altern Med* 11: 112.
45. Tewtrakul S, Miyashiro H, Nakamura N, Hattori M, Kawahata T, et al. (2003) HIV-1 integrase inhibitory substances from *Coleus parvifolius*. *Phytother Res* 17: 232-239.
46. Qian-Cutrone J, Huang S, Trimble J, Li H, Lin PF, et al. (1996) Niruriside, a new HIV REV/RRE binding inhibitor from *Phyllanthusniruri*. *J Nat Prod* 59: 196-199.
47. Dharmaratne HR, Tan GT, Marasinghe GP, Pezzuto JM (2002) Inhibition of HIV-1 reverse transcriptase and HIV-1 replication by *Calophyllumcoumarins* and xanthenes. *Planta Med* 68: 86-87.
48. Lee-Huang S, Huang PL, Chen HC, Huang PL, Bourinbaier A, et al. (1995) Anti-HIV and anti-tumor activities of recombinant MAP30 from bitter melon. *Gene* 161: 151-156.
49. Kotwal GJ (2004) HIV treatment and eradication in South Africa. *J R Soc Med* 97: 1-2.