

## Editorial

## Open Access

# Mycobacterial TDM: A Coat to Modulate Post Primary Pathogenesis?

**Jeffrey K. Actor\****Department of Pathology, Medical School, University of Texas-Houston Medical School, Houston, TX, USA*

**Keywords:** Post primary tuberculosis; TDM; Trehalose 6,6'-dimycolate; Cord factor

## Background

Trehalose 6,6'-dimycolate (TDM; cord factor) is the major external constituent comprising the waxy coat of *Mycobacterium tuberculosis*, and has been under active investigation for well over half a century. On one hand, this unique glycolipid imparts protective physical attributes to the organism, allowing survival outside of host cellular compartments. On the other hand, interaction within the host environment allows directed engagement of immunological processes advantageous both to host survival and to organism perpetuation. Classical modeling of TDM bioactivities has centered on innate immune parameters. However, recent scientific contributions shed light on additional facets that direct adaptive outcomes, warranting reexamination of TDM-induced activities that modulate manifestation of disease pathology in post primary clinical scenarios.

The characteristic histopathology associated with clinical primary tuberculosis manifests as a granulomatous lesion comprised of activated epithelioid macrophages cuffed by "ring-leader" lymphocytes [1,2]. The process allows functional sequestration of organisms with described (limited) focal inflammation, in essence providing a mechanism whereby hosts may carry the disease for later spread to other individuals. The abundant surface glycolipid antigens, mycolic acid and trehalose 6,6'-dimycolate, dictate much of the primary pathological response [3], with known granulomatous-inducing properties. In contrast, post primary tuberculosis (also referred to as secondary or adult tuberculosis) was recently redefined as a caseating pathology arising in immune competent individuals whereby organisms evade strong and effective systemic specific immunity [4-6]. In modern day cases, post primary tuberculosis is often associated with a lipid pneumonia, challenging long held beliefs that cavities arise from expansion of caseating granulomas [7].

## Biological Properties Associated with Cord Factor

Cord factor was identified circa 1950 to be a petroleum ether-soluble, surfactant extractable lipid constituent of virulent mycobacteria [8-12], responsible for "cording" of organisms [13]. While original studies concluded that cord factor had toxic properties [14-16], it was discovered that extracts containing trehalose 6,6'-dimycolate (TDM) were also responsible for antigenic [17] and adjuvant activities [18,19]. However, these biological properties were soon found to be dependent upon isolation, formulation and administration methodologies. While TDM has additional properties beyond its contribution to organism morphology [20], yet direct understanding of its role in clinical disease remains elusive.

There is no perfect animal model to represent the entire spectrum of human tuberculosis disease, although there are accepted ways to mimic aspects of the primary granulomatous response [21]. Implication of TDM as a major contributor to primary tuberculosis pathology has

been "validated" through use of these animal models [22-27]. There are defined molecular links for TDM to influence innate molecular responses in initiation and maintenance of granuloma related pathology [28-32], which are independent of interferon- $\gamma$  [33]. What is not particularly clear is the role for TDM in post primary tuberculosis, especially in development of disease pathology regulated via adaptive lymphocytic response.

In the 1970's antibodies that were reactive to TDM containing fractions [34] were identified with the ability to modulate pathology in model systems [35], suggesting (in hindsight) that adaptive cells were involved in higher order *in vivo* reactivity. More recent investigations revealed a hypersensitive component [36] involving multiple layers of adaptive immune function that may allow TDM effective control over local microenvironments [29,37]. Structural studies theorized that conformational restraints were required for TDM to elicit many of the higher order adaptive immune functions [32,38-40]. These theories were validated in part by studies showing that removal of the cyclopropane ring alters pathology development in model systems [41-43]. Only recently have we begun to appreciate contribution of TDM to development of lymphocytic functions [44-46] and mechanisms that allow induction of TDM-specific adaptive responses [47,48]. This now gives TDM a foothold as a mediator of adaptive functions leading to post primary tuberculosis and cavity lesion development [7].

## TDM as a Mediator for Post Primary Tuberculosis

The basis for this argument has a foundation in clinical tuberculosis histopathology, from patient samples exhibiting endogenous lipid pneumonia. Reports by Hunter suggest that the rapid necrosis of tuberculous pneumonia might be due to both the activation of toxic and adaptive immunogenic properties of cord factor after contact with host lipids [4,5,7,49]. This hypothesis diverges from classical interpretation of expanding granulomas as the basis of necrotizing lesions [1,50]. Indeed, an argument may be made for a strong requirement of functional adaptive immunity in the host at the time of necrotizing conversion. The contribution of an inflammatory lipid-based process may also explain why lesions histologically appear to behave independent of one another; one lesion may progress toward cavitation while another nearby regresses. Perhaps accumulation

\*Corresponding author: Jeffrey K. Actor, Professor, Department of Pathology and Laboratory Medicine, MSB 2.214, University of Texas-Houston Medical School, 6431 Fannin, Houston, TX 77030, USA, Tel: (713) 500-5344; Fax: (713)-500-0730; E-mail: [Jeffrey.K.Actor@uth.tmc.edu](mailto:Jeffrey.K.Actor@uth.tmc.edu)

Received February 08, 2012; Accepted February 17, 2012; Published February 20, 2012

Citation: Actor JK (2012) Mycobacterial TDM: A Coat to Modulate Post Primary Pathogenesis? Mycobact Diseases 2:e106. doi:10.4172/2161-1068.1000e106

Copyright: © 2012 Actor JK. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

of lipid in alveolar macrophages becomes the tipping point in the necrotic event. Cord factor coming off organisms *in vivo* during the inflammatory process may interact with lipids to allow both toxic and antigenic events to proceed [36,51,52].

## TDM Influence on Immune Function

Host interaction with organisms must include early recognition events that mediate innate functions. Identification of TDM receptors on host monocytes strengthens these ideas. However, the direct link between interaction of TDM with these receptors on innate cells and their role in development of adaptive immunity remains unknown. Certainly this remains an open field of research towards understanding clinical transition to post primary pathology. The key to defining the role of TDM in adaptive immunity therefore must begin with understanding initial events in macrophage recognition of TDM on the surface of organisms during initial encounter. Recent discoveries link the C-type lectin Mincl [53-55] as a prime candidate receptor for TDM. This is combined, in part, by additional interactions with MARCO, TLR2 and/or CD14 which are critical for mediating activity [56] with internalized signaling events that possibly function through Card9-dependent mechanisms [57,58]. What is currently clear is that once internalized, *Mycobacterium tuberculosis* blocks maturation of phagosomes [59-61], altering molecular events critical for destruction of organisms [62] and development of intra-phagosome events critical for antigen processing [63]. TDM is directly linked to this process [64,65] and elimination of critical enzymes for TDM production [eg. attenuating fbpA] alters development of T cell immunity [66]. Indeed, over expression of these related genes products increases protective T cell response [67].

## Future Studies Warranted

The role of TDM-specific lymphocytes in development of post primary tuberculosis is unclear. Critical questions remain to identify and define lymphocyte TDM-specific responses which would allow a state of lipid pneumonia to develop into full cavitary disease. While classical T cell responses may play a role, it is important to separate glycolipid responsive NKT cell functions from this group. As mentioned above, hypersensitivity to TDM can occur under appropriate conditions [44,46]. Those reports indicated that classically defined CD4<sup>+</sup> T lymphocytes specific for TDM can greatly exacerbate pathology, at least in mice. However, one would presume that NKT would be more adept at recognizing glycolipids. Yet one report elegantly describes NKT depletion in response to TDM, rather than expected proliferation [68]. However, this investigation only takes into account primary encounter with organisms. Development of post primary pathology occurs long after initial encounter with organisms, with ample time for regeneration of reactive NKT populations. Indeed, this same report indicates upregulation of CD1d (in mice) by TDM, and it has been shown that hypersensitivity to TDM requires CD1d for pathological development [44,48,69].

TDM therefore remains an exciting molecule to study, with new questions proposed on its function in development in secondary disease. We must appreciate the polar extremes of its associated biological properties. On one hand TDM is critical for protection of the organism, both outside the host and as a mediator of intracellular phagosome maturation events. On the other hand, TDM has unique properties allowing focused innate inflammatory responses to initiate

a protective granulomatous response. We can now hope to add additional layers of complexity through [1] investigation of its function in regulating adaptive events in immune competent hosts, and, through [2] further understanding of physical interactions with host lipids to generate toxic cues related to development of cavitation and necrosis in post primary tuberculosis.

## References

1. Dannenberg A (1999) Tuberculosis and nontuberculosis mycobacterial infections. In *Pathophysiology: basic aspects*. D. Schlossberg, ed. W. B. Saunders, Philadelphia. 26-27.
2. Paige C, Bishai W R (2010) Penitentiary or penthouse condo: the tuberculous granuloma from the microbe's point of view. *Cell Microbiol* 12: 301-309.
3. Goren M, Brennan P (1979) Mycobacterial lipids: chemistry and biologic activities. In *Tuberculosis*. G. Youmans, ed. W.B. Saunders, Philadelphia. 63-193.
4. Hunter RL (2011) On the pathogenesis of post primary tuberculosis: The role of bronchial obstruction in the pathogenesis of cavities. *Tuberculosis* 1: S6-S10.
5. Hunter RL (2011) Pathology of post primary tuberculosis of the lung: an illustrated critical review. *Tuberculosis* 91: 497-509.
6. Gideon HP, Flynn JL (2011) Latent tuberculosis: what the host "sees"? *Immunol Res* 50: 202-212.
7. Hunter RL, Jagannath C, Actor JK (2007) Pathology of postprimary tuberculosis in humans and mice: contradiction of long-held beliefs. *Tuberculosis* 87: 267-278.
8. Middlebrook G, Dubos RJ, Pierce C (1947) Virulence and Morphological Characteristics of Mammalian Tuberle Bacilli. *J Exp Med* 86: 175-184.
9. Bloch H (1950) Studies on the virulence of tubercle bacilli; the relationship of the physiological state of the organisms to their pathogenicity. *J Exp Med* 92: 507-526.
10. Bloch H (1950) Studies on the virulence of tubercle bacilli; isolation and biological properties of a constituent of virulent organisms. *J Exp Med* 91: 197-218.
11. Bloch H, Noll H (1953) Studies on the virulence of tubercle bacilli; variations in virulence effected by tween 80 and thiosemicarbazone. *J Exp Med* 97: 1-16.
12. Bloch H, Noll H (1955) Studies on the virulence of Tuberle bacilli; the effect of cord factor on murine tuberculosis. *Br J Exp Pathol* 36: 8-17.
13. Sorkin E, Erlenmeyer H, Bloch H (1952) Purification of a lipid material ('cord factor') obtained from young cultures of tubercle bacilli. *Nature* 170: 124.
14. Bloch H, Sorkin E, Erlenmeyer H (1953) A toxic lipid component of the tubercle bacillus (cord factor). I. Isolation from petroleum ether extracts of young bacterial cultures. *Am Rev Tuberc* 67: 629-643.
15. Yarkoni E, Rapp HJ (1979) Influence of oil and Tween concentrations on enhanced endotoxin lethality in mice pretreated with emulsified trehalose-6,6'-dimycolate (cord factor). *Infect Immun* 24: 571-572.
16. Sakurai T, Saiki I, Ishida H, Takeda K, Azuma I (1989) Lethal toxicity and adjuvant activities of synthetic TDM and its related compounds in mice. *Vaccine* 7: 269-274.
17. Brennan PJ, Goren MB (1977) Mycobacterial glycolipids as bacterial antigens. *Biochem Soc Trans* 5: 1687-1693.
18. Bekierkunst A, Levij IS, Yarkoni E, Vilkas E, Adam A, et al. (1969) Granuloma formation induced in mice by chemically defined mycobacterial fractions. *J Bacteriol* 100: 95-102.
19. Saito R, Tanaka A, Sugiyama K, Azuma I, Yamamura Y (1976) Adjuvant effect of cord factor, a mycobacterial lipid. *Infect Immun* 13: 776-781.
20. Hunter RL, Venkataprasad N, Olsen MR (2006) The role of trehalose dimycolate (cord factor) on morphology of virulent *M. tuberculosis* in vitro. *Tuberculosis (Edinb)* 86: 349-356.
21. Padilla-Carlin D J, McMurray D N, Hickey A J (2008) The guinea pig as a model of infectious diseases. *Comp Med* 58: 324-340.

22. Baba T, Natsuhara Y, Kaneda K, Yano I (1997) Granuloma formation activity and mycolic acid composition of mycobacterial cord factor. *Cell Mol Life Sci* 53: 227-232.
23. Perez RL, Roman J, Roser S, Little C, Olsen M, et al. (2000) Cytokine message and protein expression during lung granuloma formation and resolution induced by the mycobacterial cord factor trehalose-6,6'-dimycolate. *J Interferon Cytokine Res* 20: 795-804.
24. Perez RL, Roman J, Staton GW Jr, Hunter RL (1994) Extravascular coagulation and fibrinolysis in murine lung inflammation induced by the mycobacterial cord factor trehalose-6,6'-dimycolate. *Am J Respir Crit Care Med* 149: 510-518.
25. Behling CA, Perez RL, Kidd MR, Staton GW Jr, Hunter RL (1993) Induction of pulmonary granulomas, macrophage procoagulant activity, and tumor necrosis factor-alpha by trehalose glycolipids. *Ann Clin Lab Sci* 23: 256-266.
26. Hamasaki N, Isowa K, Kamada K, Terano Y, Matsumoto T, et al. (2000) In vivo administration of mycobacterial cord factor (Trehalose 6, 6'-dimycolate) can induce lung and liver granulomas and thymic atrophy in rabbits. *Infect Immun* 68: 3704-3709.
27. Yarkoni E, Rapp HJ (1977) Granuloma formation in lungs of mice after intravenous administration of emulsified trehalose-6,6'-dimycolate (cord factor): reaction intensity depends on size distribution of the oil droplets. *Infect Immun* 18: 552-554.
28. Sakai Y, Uchida K, Nakayama H (2011) Histopathological features and expression profiles of cytokines, chemokines and SOCS family proteins in trehalose 6,6'-dimycolate-induced granulomatous lesions. *Inflamm Res* 60: 371-378.
29. Abbott AN, Guidry TV, Welsh KJ, Thomas AM, Kling MA, et al. (2009) 11beta-hydroxysteroid dehydrogenases are regulated during the pulmonary granulomatous response to the mycobacterial glycolipid trehalose-6,6'-dimycolate. *Neuroimmunomodulation* 16: 147-154.
30. Welsh KJ, Abbott AN, Hwang SA, Indigo J, Armitage LY, et al. (2008) A role for tumour necrosis factor-alpha, complement C5 and interleukin-6 in the initiation and development of the mycobacterial cord factor trehalose 6,6'-dimycolate induced granulomatous response. *Microbiology* 154: 1813-1824.
31. Borders CW, Courtney A, Ronen K, Pilar Laborde-Lahoz M, Guidry TV, et al. (2005) Requisite role for complement C5 and the C5a receptor in granulomatous response to mycobacterial glycolipid trehalose 6,6'-dimycolate. *Scand J Immunol* 62: 123-130.
32. Ryll R, Kumazawa Y, Yano I (2001) Immunological properties of trehalose dimycolate (cord factor) and other mycolic acid-containing glycolipids--a review. *Microbiol Immunol* 45: 801-811.
33. Takimoto H, Maruyama H, Shimada KI, Yakabe R, Yano I, et al. (2006) Interferon-gamma independent formation of pulmonary granuloma in mice by injections with trehalose dimycolate (cord factor), lipoarabinomannan and phosphatidylinositol mannosides isolated from *Mycobacterium tuberculosis*. *Clin Exp Immunol* 144: 134-141.
34. Kato M (1973) Immunochemical properties of anti-cord factor antibody. *Infect Immun* 7: 9-13.
35. Kato M (1973) Effect of anti-cord factor antibody on experimental tuberculosis in mice. *Infect Immun* 7: 14-21.
36. Bekerkunst A, Yarkoni E (1973) Granulomatous hypersensitivity to trehalose-6,6'-dimycolate (cord factor) in mice infected with BCG. *Infect Immun* 7: 631-638.
37. Abbott AN, Welsh KJ, Hwang SA, Płoszaj P, Choudhury T, et al. (2011) IL-6 mediates 11betaHSD type 2 to effect progression of the mycobacterial cord factor trehalose 6,6'-dimycolate-induced granulomatous response. *Neuroimmunomodulation* 18: 212-225.
38. Behling CA, Bennett B, Takayama K, Hunter RL (1993) Development of a trehalose 6,6'-dimycolate model which explains cord formation by *Mycobacterium tuberculosis*. *Infect Immun* 61: 2296-2303.
39. Fujita Y, Okamoto Y, Uenishi Y, Sunagawa M, Uchiyama T, et al. (2007) Molecular and supra-molecular structure related differences in toxicity and granulomatogenic activity of mycobacterial cord factor in mice. *Microb Pathog* 43: 10-21.
40. Yarkoni E, Rapp HJ (1978) Toxicity of emulsified trehalose-6,6'-dimycolate (cord factor) in mice depends on size distribution of mineral oil droplets. *Infect Immun* 20: 856-860.
41. Glickman MS, Cox JS, Jacobs WR Jr (2000) A novel mycolic acid cyclopropane synthetase is required for cording, persistence, and virulence of *Mycobacterium tuberculosis*. *Mol Cell* 5: 717-727.
42. Rao V, Gao F, Chen B, Jacobs WR Jr, Glickman MS (2006) Trans-cyclopropanation of mycolic acids on trehalose dimycolate suppresses *Mycobacterium tuberculosis*-induced inflammation and virulence. *J Clin Invest* 116: 1660-1667.
43. Rao V, Fujiwara N, Porcelli SA, Glickman MS (2005) *Mycobacterium tuberculosis* controls host innate immune activation through cyclopropane modification of a glycolipid effector molecule. *J Exp Med* 201: 535-543.
44. Guidry TV, Hunter RL Jr, Actor JK (2006) CD3+ cells transfer the hypersensitive granulomatous response to mycobacterial glycolipid trehalose 6,6'-dimycolate in mice. *Microbiology* 152: 3765-3775.
45. Yamagami H, Matsumoto T, Fujiwara N, Arakawa T, Kaneda K, et al. (2001) Trehalose 6,6'-dimycolate (cord factor) of *Mycobacterium tuberculosis* induces foreign-body- and hypersensitivity-type granulomas in mice. *Infect Immun* 69: 810-815.
46. Guidry TV, Hunter RL Jr, Actor JK (2007) Mycobacterial glycolipid trehalose 6,6'-dimycolate-induced hypersensitive granulomas: contribution of CD4+ lymphocytes. *Microbiology* 153: 3360-3369.
47. Oiso R, Fujiwara N, Yamagami H, Maeda S, Matsumoto S, et al. (2005) Mycobacterial trehalose 6,6'-dimycolate preferentially induces type 1 helper T cell responses through signal transducer and activator of transcription 4 protein. *Microb Pathog* 39: 35-43.
48. Guidry TV, Olsen M, Kil KS, Hunter RL Jr, Geng YJ, et al. (2004) Failure of CD1D-/- mice to elicit hypersensitive granulomas to mycobacterial cord factor trehalose 6,6'-dimycolate. *J Interferon Cytokine Res* 24: 362-371.
49. Syed SS, Hunter RL Jr (1997) Studies on the toxic effects of quartz and a mycobacterial glycolipid, trehalose 6,6'-dimycolate. *Ann Clin Lab Sci* 27: 375-383.
50. Rich A (1951) The pathogenesis of tuberculosis. Charles C. Thomas, Springfield.
51. Geisel RE, Sakamoto K, Russell DG, Rhoades ER (2005) In vivo activity of released cell wall lipids of *Mycobacterium bovis* bacillus Calmette-Guerin is due principally to trehalose mycolates. *J Immunol* 174: 5007-5015.
52. Rhoades ER, Geisel RE, Butcher BA, McDonough S, Russell DG (2005) Cell wall lipids from *Mycobacterium bovis* BCG are inflammatory when inoculated within a gel matrix: characterization of a new model of the granulomatous response to mycobacterial components. *Tuberculosis (Edinb)* 85: 159-176.
53. Schoenen H, Bodendorfer B, Hitchens K, Manzanero S, Werninghaus K, et al. (2010) Cutting edge: Mincle is essential for recognition and adjuvanticity of the mycobacterial cord factor and its synthetic analog trehalose-dibehenate. *J Immunol* 184: 2756-2760.
54. Ishikawa E, Ishikawa T, Morita YS, Toyonaga K, Yamada H, et al. (2009) Direct recognition of the mycobacterial glycolipid, trehalose dimycolate, by C-type lectin Mincle. *J Exp Med* 206: 2879-2888.
55. Matsunaga I, Moody DB (2009) Mincle is a long sought receptor for mycobacterial cord factor. *J Exp Med* 206: 2865-2868.
56. Bowdish DM, Sakamoto K, Kim MJ, Kroos M, Mukhopadhyay S, et al. (2009) MARCO, TLR2, and CD14 are required for macrophage cytokine responses to mycobacterial trehalose dimycolate and *Mycobacterium tuberculosis*. *PLoS Pathog* 5: e1000474.
57. Werninghaus K, Babiak A, Gross O, Hölscher C, Dietrich H, et al. (2009) Adjuvanticity of a synthetic cord factor analogue for subunit *Mycobacterium tuberculosis* vaccination requires FcRgamma-Syk-Card9-dependent innate immune activation. *J Exp Med* 206: 89-97.
58. LeibundGut-Landmann S, Gross O, Robinson MJ, Osorio F, Slack EC, et al.

- al. (2007) Syk- and CARD9-dependent coupling of innate immunity to the induction of T helper cells that produce interleukin 17. *Nat Immunol* 8: 630-638.
59. Deretic V, Fratti RA (1999) Mycobacterium tuberculosis phagosome. *Mol Microbiol* 31: 1603-1609.
60. Fratti RA, Vergne I, Chua J, Skidmore J, Deretic V (2000) Regulators of membrane trafficking and Mycobacterium tuberculosis phagosome maturation block. *Electrophoresis* 21: 3378-3385.
61. Russell DG (2001) Mycobacterium tuberculosis: here today, and here tomorrow. *Nat Rev Mol Cell Biol* 2: 569-577.
62. Sturgill-Koszycki S, Schlesinger PH, Chakraborty P, Haddix PL, Collins HL, et al. (1994) Lack of acidification in Mycobacterium phagosomes produced by exclusion of the vesicular proton-ATPase. *Science* 263: 678-681.
63. Katti MK, Dai G, Armitige LY, Rivera Marrero C, Daniel S, et al. (2008) The Delta fbpa mutant derived from Mycobacterium tuberculosis H37Rv has an enhanced susceptibility to intracellular antimicrobial oxidative mechanisms, undergoes limited phagosome maturation and activates macrophages and dendritic cells. *Cell Microbiol* 10: 1286-1303.
64. Indrigo J, Hunter RL Jr, Actor JK (2002) Influence of trehalose 6,6'-dimycolate (TDM) during mycobacterial infection of bone marrow macrophages. *Microbiology* 148: 1991-1998.
65. Indrigo J, Hunter RL Jr, Actor JK (2003) Cord factor trehalose 6,6'-dimycolate (TDM) mediates trafficking events during mycobacterial infection of murine macrophages. *Microbiology* 149: 2049-2059.
66. Roche CM, Smith A, Lindsey DR, Meher A, Schluns K, et al. (2011) The DeltafbpA attenuated candidate vaccine from Mycobacterium tuberculosis, H37Rv primes for a stronger T-bet dependent Th1 immunity in mice. *Tuberculosis (Edinb)* 1: S96-S104.
67. Lindsey DR, Dhandayuthapani S, Jagannath C (2009) Anti-tuberculosis immunity induced in mice by vaccination with Mycobacterium smegmatis over-expressing Antigen 85B is due to the increased influx of IFNgamma-positive CD4 T cells into the lungs. *Tuberculosis (Edinb)* 1: S46-S48.
68. Ryll R, Watanabe K, Fujiwara N, Takimoto H, Hasunuma R, et al. (2001) Mycobacterial cord factor, but not sulfolipid, causes depletion of NKT cells and upregulation of CD1d1 on murine macrophages. *Microbes Infect* 3: 611-619.
69. Actor JK, Olsen M, Hunter RL Jr, Geng YJ (2001) Dysregulated response to mycobacterial cord factor trehalose-6,6'-dimycolate in CD1D-/ mice. *J Interferon Cytokine Res* 21: 1089-1096.