

## Molecular Docking Studies of Wide Spectrum Targets in *Staphylococcus aureus* - An Aim towards Finding Potent Inhibitors

Balaji SR\*, Gupta KK, Anusha P and Raveena P

NTHRYS Biotech labs, Hyderabad, Andhra Pradesh, India

### Abstract

Methicillin-resistant *staphylococcus aureus* (MRSA) is a bacterium that is evolving towards adaptive changes to certain antibiotics like methicillin, oxacillin, penicillin, and amoxicillin. It is a communicable and most rapidly spreading disease worldwide. It is reported that MRSA is becoming common among children in intensive care units. This disease comes under Emerging Infectious Disease (EID). In this paper, MRSA proteome screening is done and Drug/vaccine targets are proposed based on its essentiality to the pathogen and non-homology with human proteome. Targets validation is done so that its targeting must not affect human proteome and vital pathways. Those targets which has no structure, structure prediction and validation is done and important epitopes and ligands are proposed on suitable targets.

**Keywords:** MRSA; EPA; Infection; Targets; Epitope; Drug Pathways

### Introduction

MRSA spreads infection through blood and skin. The former infection is of severe kind. But skin infections are the most common one. Skin infection appears as pustules or boils which generally form at areas of the body covered by hair like back of neck, groin, buttock, armpit, beard area of men. The degree of symptoms depends on the stage of infection. The persons who meet MRSA patients are at Risk of Acquiring MRSA Infections. In hospitals, patients has more risk of getting MRSA through catheters inserted into the skin and those patients who has undergo medical procedures like surgery. According to drug bank only three antibiotics namely Arbekacin, Meticillin and Linezolid, are approved for the treatment of MRSA. Drug targets are limited and there is an urgent need for the discovery of novel drug targets. Apart from drug target, we also need Good ligand formulation that would act as a potent inhibitors to the novel targets without affecting human proteome (Figure 1).

Any contaminated surfaces is the source of many kind of infections, MRSA is one of them. Mishandled Hospital procedures can leave immune compromised patients vulnerable to MRSA. Environmental hygienic conditions are the source of good physical and mental health. It is reported that U.S. Environmental Protection Agency (EPA) labeled Cleaners, disinfectants and sanitizers must be used in order to spread all kinds of infections.

### Material and Methodology

#### *Staphylococcus aureus* proteome screening

Prokaryotic sequence homology analysis tool (PSAT) [1] is used for finding conserved patches in all strains of *Staphylococcus aureus* and *Streptococcus*, as both are heavily involved in skin infections. *Yersinia Pestis* CO 92 is taken as reference strain. Following parameters is considered for finding conserved genes in these selected pathogens: BLAST alignment score thresholds for finding gene homologs: e-value < 10; bit score > 20; % identity > 10

#### DEG BLAST

All the conserved genes are checked for their key role to pathogens. Database of essential genes (DEG) [2] is used to screen

all the conserved genes for their vitality to the selected pathogens. This tool gives hit if query gene is showing any significant similarity with pathogen's growth, reproduction and survival.

#### NCBI Homo sapiens (human) protein BLAST

Those conserved genes which are giving hits in DEG blast were further checked in NCBI Homo sapiens (human) Protein BLAST [3]. If we are proposing target on pathogen proteome then those conserved genes must be checked if there is any similarity with human proteome because drug/vaccine targeting must not affect any of the human protein.

#### Annotation and pathways analysis

The conserved genes which are showing no significant similarity checked in NCBI Homo sapiens (human) Protein BLAST was checked for annotation and their involvement in any crucial pathway. Annotation was done by using CELLO [4] and PSORTB [5]. Pathway analysis was done in KEGG genes database [6].

#### Target proposal and structure prediction

Based on the pathway study, its essentiality and non-homolog to human proteome property of Targets, they are proposed for drug targets and epitope design. Those targets whose have no protein structure was modeled by Protein Homology/analogy Recognition Engine (PHYRE) [7].

#### Structure validation

These Target proteins were optimized from KOBAMIN [8] and Galaxy WEB server [9] and validated in Rampage [10] and Erratplot

**\*Corresponding author:** Balaji SR, Director & Research Head, NTHRYS Biotech labs, Hyderabad, Andhra Pradesh, India, Tel: 040-276 212 48; E-mail: [balajisrao@nthyrs.com](mailto:balajisrao@nthyrs.com)

**Received** January 11, 2014; **Accepted** February 13, 2014; **Published** February 22, 2014

**Citation:** Balaji SR, Gupta KK, Anusha P, Raveena P (2014) Molecular Docking Studies of Wide Spectrum Targets in *Staphylococcus aureus* - An Aim towards Finding Potent Inhibitors. Adv Tech Biol Med 2: 115. doi: [10.4172/2379-1764.1000115](https://doi.org/10.4172/2379-1764.1000115)

**Copyright:** © 2014 Balaji SR. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

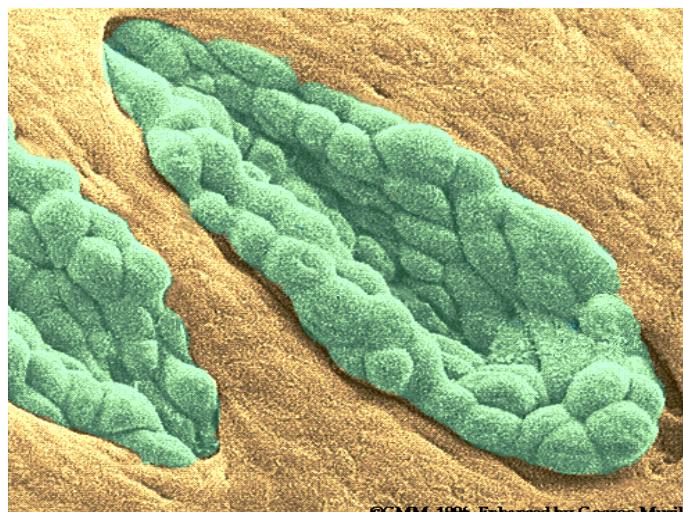


Figure 1 : MRSA.

[11], the former check the stereo chemical properties of modeled structures and latter one analyze the non-bonded interactions.

### Epitope design

Transmembrane region of proposed target was predicted with TMHMM [12]. After that, Prediction of B cell epitopes with BCpred (cutoff 0.8, 20-mer epitopes) [13]. Generally B cell epitope sequences are surface-exposed of corresponding proteins. T cell epitopes from propred [14], propred 1[15], MHCpred [16] and T epitope designer [17]. Those T cell epitopes were considered that is part of B cell epitopes and lies at Transmembrane region according to TMHMM. Predict those epitope's antigenicity from vaxijen [18]. Finally, epitopes that bound more than 13 MHC molecules in ProPred and ProPred-I with less than 100 nM IC<sub>50</sub> for DRB1\*0101 in MHCpred v2.0 and that bound >=80% of HLA molecules in T-epitope designer were selected. Proteins that were antigenic according to Vaxijen (threshold=0.4, ACC output) score above than 0.5 in VaxiJen were selected.

### Virtual screening of ligands against target proteins

All the target proteins were screened with 12 million drug-like ZINC12 + 6507 DrugBank drugs from FINDSITE-COMB [19]. Less characterized ZINC DATABASE molecules were selected for drug-likeness analysis from FAF- Drugs2 [20]. FAF results were cross checked with Osiris property explorer [21]. We modified most of the ZINC ID ligands to satify druglikeness and ADME properties. We proposed those novel ligands for each target which is following all the drug-likeness properties.

### Molecular docking studies of ADMET following ligands with target proteins

ADMET following ligands were docked with their corresponding target in particular coordinate predicted by Pocket-Finder [22] using Molegro Virtual Docker [23]. All the docking parameters and images were finalized for each ligand-protein docking.

### Results

Six targets are finalized for further study (Table 1). Protein accession number, locus tags are the unique identifier of *insilico* discovered targets. All are essential for pathogens as inferred from DEG Blast results. kdpA, opp-1B and icac structures are not available. Therefore, they must be modeled for further molecular docking studies. Pathways and Cellular localization results are shown in Table 2.

All the target proteins are suitable for epitope design as all are predicted to be localized in Membrane from CELLO and PSORTB. They can also be considered for potent drug target. Function of Target Protein is very important for Target validation. Functional annotation is done in Table 3. Epitope design on the target proteins are given in Table 4. The final epitopes following all the criteria from target proteins are given in Table 5. The structural information of refined modeled target proteins from erratplot and rampage are shown in Tables 6 and 7. The coordinates of best pockets for each target proteins are given in Table 8.

| Locus Tag | Protein ID  | Target name          | Structure/ Related (yes/no)  | DEG hits (yes/no) |
|-----------|-------------|----------------------|------------------------------|-------------------|
| SAR2165   | YP_041527.1 | kdpA                 | no                           | yes               |
| SAR2553   | YP_041904.1 | Opp-1B               | no                           | yes               |
| SAR2750   | YP_042088.1 | icac                 | no                           | yes               |
| SAR0118   | YP_039582.1 | sirA                 | Yes (Related: pdbid: 3MWF)   | yes               |
| SAR2537   | YP_041888.1 | opuCB                | Yes (Related: pdbid: 3D31 C) | yes               |
| SAS0639   | YP_042767.1 | Hypothetical protein | Yes (Related: pdbid: 3MLV L) | yes               |

Table1: Final Targets based on subtractive proteome screening.

| Target name          | CELLO          | PSORTB               | Pathway              |
|----------------------|----------------|----------------------|----------------------|
| kdpA                 | Membrane       | Cytoplasmic membrane | Two-component system |
| Opp-1B               | Membrane       | Cytoplasmic membrane | ABC transporters     |
| icac                 | Membrane       | Cytoplasmic membrane | No pathway           |
| sirA                 | Periplasmic    | Cytoplasmic membrane | ABC transporters     |
| opuCB                | Inner membrane | Cytoplasmic membrane | ABC transporters     |
| Hypothetical protein | Membrane       | Cytoplasmic membrane | No pathway           |

**Table 2:** Cellular localization and pathways of target proteins.

| Target name          | Function  | Molecular Function  |
|----------------------|---|---|
| kdpA                 | One of the components of the high-affinity ATP-driven potassium transport (or KDP) system, which catalyzes the hydrolysis of ATP coupled with the exchange of hydrogen and potassium ions | ATP binding, potassium-transporting ATPase activity                         |
| Opp-1B               | Oligopeptide transporter putative membrane permease domain  | transporter activity  |
| icac                 | Presumably involved in the export of the biofilm adhesin polysaccharide poly-beta-1,6-N-acetyl-D-glucosamine (PNAG, also referred to as PIA) across the cell membrane                     | transferase activity, transferring acyl groups other than amino-acyl groups |
| sirA                 | Iron-regulated ABC transporter siderophore-binding protein SirA   | Iron-regulated ABC transporter siderophore-binding protein SirA             |
| opuCB                | Probable glycine betaine/carnitine/choline ABC transporter opuCB  | transporter activity  |
| Hypothetical protein | Uncharacterized protein conserved in bacteria [Function unknown]  | Function unknown  |

**Table 3:** Function annotation of target proteins.

| Target name | TMHMM   | Bcpred<br>Position-Bcell epitope (confidence value)   | Propred 1<br>(Sorted in descending score)   | Propred                                | MHCpred                                    |
|-------------|---|---|---|--|--|
| kdpA        | Outside:<br>1-3<br>86-126<br>192-244<br>303-324<br>373-375<br>438-483<br>549-558<br>TMhelix:<br>4-26<br>63-85<br>127-149<br>169-191<br>245-267<br>280-302<br>325-347<br>354-372<br>376-398<br>415-437<br>484-506<br>526-548 | 462-AAANNGSGFEGLKDDTFWN (0.97)<br>335-<br>FTVITTAFTTGSVNNMHDSL<br>(0.92)<br>77-LLIVQQWLFLNPNPNHNLNQSI(0.91)<br>434<br>AFMIPGASESITNPSFHGIS (0.89) | LIVQQWLFL<br>NNGSGFEGL<br>MIPGASESI<br>AAANNGSGF<br>LLIVQQWLFL<br>IPGASESIT<br>FTVITTAFTT<br>TVITTAFTT<br>FMIPGASES<br>ITTAFTTGS<br>VITTAFTTG<br>ANNGSGFEG<br>IVQQWLFLN | FMIPGASES (10/51)                      | FMIPGASES - 72% MHC alleles are Binding.   |
| Opp-1B      | outside<br>32-106<br>162-175<br>257-275<br>TMhelix:<br>9-31<br>107-126<br>139-161<br>176-193<br>234-256<br>276-298  | 80- NFGTSYITGDPVAERIGPAF (0.99)<br>41- AQGTPNVTPELIAETNEKYG (0.91)  | GTPNNTPEL<br>TPNVTPELI<br>TSYITGDPV<br>GTSYITGDP<br>AQGTPNVTP<br>FGTSYITGD<br>NFGTSYITG<br>QGTPNVTPPE   | YITGDPV (8/51)                         | YITGDPV -100% MHC alleles are Binding.     |
| icac        | outside<br>30-43<br>102-115<br>168-186<br>234-242<br>292-305<br>Tmhelix:<br>7-29<br>44-66<br>79-101<br>116-138<br>145-167<br>187-204<br>211-233<br>243-262<br>269-291<br>306-328  | 165- YFTNNNTAFHDTVLHYYPLSE (0.7)  | NNNTAFHDTV<br>FTNNNTAFHD<br>TNNTAFHDT   | YFTNNNTAFH (14/51)<br>VLHYYPLSE (8/51) | FTNNNTAFH<br>100% MHC alleles are Binding. |

|                      |  |  |  |                          |   |
|----------------------|--|--|--|--------------------------|---|
|                      |  |  | KNDLKDTKI<br>TTKLMGKAL<br>FQKDAKAKY<br>LGVKPVGAV<br>AAFQDAKA<br>KPVGGAESW<br>NDLKDTKIV<br>TTEIKGKPK<br>KDTTKLMGK<br>KKTESEWTS<br>LVKKTESEW<br>NDLKDTKIV<br>CSGGNSNKQS<br>KTESEWTSS<br>GCSGGNSNKQ<br>SGNSNQSS<br>DTTKLMGKA<br>MGTTEIKGKP<br>AFQKDAKAK<br>GVKPGAVE<br>DLKDTKIVG<br>VKPAVES<br>AMGTTIEKG<br>GTTEIKGKP<br>QKDAKAKYK<br>VKKTESEWT<br>GNSNKQSSD<br>TKLMGKALG |                          | LGVKPVGAV<br>68% MHC alleles are Binding.<br><br>VKPVGAVES<br>72% MHC alleles are Binding.<br><br>FQKDAKAKY<br>80% MHC alleles are Binding. |
| sirA                 | Outside:<br>Whole protein  | 20-GCSGNNSNKQSSDSKDKEETTS (0.99)<br>70-LGVKPVGAVESWTQKPKFEY (0.99)<br>149-KDTTKLMGKALGKEKEAEDL (0.97)<br>269-LVKKTESEWTSSKEWNLDA (0.97)<br>43-AMGTTIEKGKPKRVVTLYQG (0.96)<br>91-KNDLKDTKIVGQEPAPELNLEE (0.94)<br>177-AAFQKDAKAKYKDAWPLKAS (0.84) | YVGAGGLGD<br>VGAGGLGDF<br>AGGLGDFIF<br>GAGGLGDFI   | YVGAGGLGD<br>(8/51)      | YVGAGGLGD<br>72% MHC alleles are Binding.   |
| opuCB                | TMHelix<br>114 135<br>115 134<br>144 170<br>145 168<br>174 189<br>175 188<br>223 248<br>225 235<br>238 243<br>268 292<br>269 290 | 155- YVGAGGLGDFIFNGLNLNYDP (0.9)   |  |                          |   |
| Hypothetical protein | TMHelix<br>64 102<br>65 102<br>113 131<br>115 129<br>150 181<br>151 180<br>194 222<br>195 202<br>205 221                         | No B cell epitope predicted.   | No common T cell epitope   | No common T cell epitope | No epitope binding affinity prediction.   |

**Table 4:** Epitope designing on target proteins.

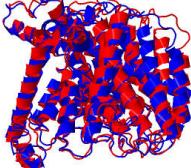
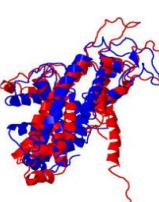
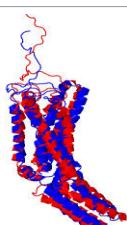
| Target name          | Final Predicted epitopes            | Vaxijen score (>0.5)       |
|----------------------|-------------------------------------|----------------------------|
| kdpA                 | FMIPGASES                           | 0.4000                     |
| Opp-1B               | YITGDPV                             | 1.2802                     |
| icac                 | FTNNNTAFH                           | 0.3954                     |
| sirA                 | LGVKPVGAV<br>VKPVGAVES<br>FQKDAKAKY | 0.0443<br>0.0286<br>1.9244 |
| opuCB                | YVGAGGLGD                           | 1.5458                     |
| Hypothetical protein | No predicted epitope                | No predicted epitope       |

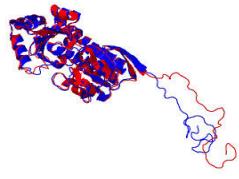
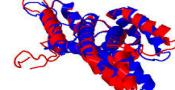
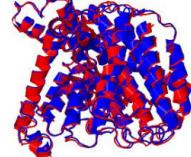
Note: red color indicates non promising peptides whereas green color indicates promising one.

**Table 5:** Final Epitopes on target proteins.

| Target Name/Protein ID            | Ramachandran parameters | Errat plot quality factor |
|-----------------------------------|-------------------------|---------------------------|
| kdpA/ YP_041527.1                 | <b>RMSD</b> 2.290       | 78.46                     |
|                                   | Clash score 20.9        |                           |
|                                   | Poor rotamers 3.5       |                           |
|                                   | Rama favored 90.6       |                           |
| Opp-1B/ YP_041904.1               | <b>RMSD</b> 8.941       | 82.7                      |
|                                   | Clash score 15.7        |                           |
|                                   | Poor rotamers 0.4       |                           |
|                                   | Rama favored 94.2       |                           |
| Icac/ YP_042088.1                 | <b>RMSD</b> 5.597       | 93.59                     |
|                                   | Clash score 17.4        |                           |
|                                   | Poor rotamers 2.2       |                           |
|                                   | Rama favored 95.7       |                           |
| sirA/ YP_039582.1                 | <b>RMSD</b> 4.377       | 86.00                     |
|                                   | Clash score 8.6         |                           |
|                                   | Poor rotamers 1.0       |                           |
|                                   | Rama favored 98.2       |                           |
| opuCB/ YP_041888.1                | <b>RMSD</b> 5.944       | 94.44                     |
|                                   | Clash score 10.9        |                           |
|                                   | Poor rotamers 1.2       |                           |
|                                   | Rama favored 96.2       |                           |
| Hypothetical protein/ YP_042767.1 | <b>RMSD</b> 0.749       | 77.03                     |
|                                   | Clash score 17.0        |                           |
|                                   | Poor rotamers 1.0       |                           |
|                                   | Rama favored 91.2       |                           |

**Table 6:** Structural information of target proteins.

| Target name         | Refined modeled target structures   |
|---------------------|---|
| kdpA/ YP_041527.1   |  <p>Number of residues in favoured region (~98.0% expected): 504 ( 90.6%)<br/>     Number of residues in allowed region (~2.0% expected) : 36 ( 6.5%)<br/>     Number of residues in outlier region : 16 (2.9%)</p> |
| Opp-1B/ YP_041904.1 |  <p>Number of residues in favoured region (~98.0% expected): 284 ( 93.7%)<br/>     Number of residues in allowed region (~2.0% expected) : 14 ( 4.6%)<br/>     Number of residues in outlier region : 5 (1.7%)</p>   |
| Icac/ YP_042088.1   |  <p>Number of residues in favoured region (~98.0% expected) : 334 ( 96.0%)<br/>     Number of residues in allowed region (~2.0% expected) : 12 ( 3.4%)<br/>     Number of residues in outlier region : 2 (0.6%)</p> |

|                                   |  |   |
|-----------------------------------|--|---|
| sirA/ YP_039582.1                 |  | Number of residues in favoured region (~98.0% expected) : 322 ( 98.2%)<br>Number of residues in allowed region (~2.0% expected) : 6 ( 1.8%)<br>Number of residues in outlier region : 0 ( 0.0%)   |
| opuCB/ YP_041888.1                |   | Number of residues in favoured region (~98.0% expected) : 201 ( 96.2%)<br>Number of residues in allowed region (~2.0% expected) : 7 ( 3.3%)<br>Number of residues in outlier region : 1 ( 0.5%)   |
| Hypothetical protein/ YP_042767.1 |  | Number of residues in favoured region (~98.0% expected) : 506 ( 91.0%)<br>Number of residues in allowed region (~2.0% expected) : 37 ( 6.7%)<br>Number of residues in outlier region : 13 ( 2.3%) |

**Table 7:** Modeled proteins.

| Target name         | Best pocket with coordinates  |
|---------------------|---|
| Icac/ YP_042088.1   | 1st site coordinates predicted by Pocket-Finder<br>Min Coords: (-16, -47, 24)<br>Max Coords: (-4, -33, 37)  |
| Opp-1B/ YP_041904.1 | Best pocket residues predicted by FINDSITE comb<br>TEMPLATE 1L9HA<br>RES 288 1.00 -0.21 HIS<br>RES 293 1.00 -0.93 ASP<br>RES 318 1.00 -0.51 LEU<br>RES 319 1.00 -0.37 GLY<br>RES 322 1.00 -0.43 ILE<br>DNAE 1.994 |
|                     | 1st site coordinates predicted by Pocket-Finder<br>Min Coords: (-39, 12, -6)<br>Max Coords: (-21, 29, 15)   |

|                                   |   |
|-----------------------------------|---|
| opuCB/ YP_041888.1                | 1st site coordinates predicted by Pocket-Finder<br>Min Coords: (-35, -74, 64)<br>Max Coords: (-17, -62, 79)   |
|                                   | Best pocket residues predicted by FINDSITE comb<br>TEMPLATE 3PUVG03<br>TEMPLATE 3PUXG03<br>TEMPLATE 3PUWG01<br>TEMPLATE 3QBIB02<br>TEMPLATE 1QBZA00<br>TEMPLATE 1IW6A00<br>TEMPLATE 1CWQB08<br>RES 32 0.43 0.00 ILE<br>RES 35 0.43 0.00 VAL<br>RES 36 0.57 0.00 PRO<br>RES 128 0.57 0.00 LEU<br>RES 129 0.71 0.00 PRO<br>RES 132 0.71 0.00 LEU<br>DNAE 2.093  |
| kdpA/ YP_041527.1                 | 1st site coordinates predicted by Pocket-Finder<br>Min Coords: (-7, -16, -15)<br>Max Coords: (9, -3, 5)   |
|                                   | Best pocket residues predicted by FINDSITE comb<br>TEMPLATE 1M75B00<br>TEMPLATE 1F0YA00<br>TEMPLATE 1F12A00<br>TEMPLATE 3HDHA00<br>RES 8 1.00 0.00 THR<br>RES 9 1.00 0.00 MET<br>RES 12 1.00 0.00 MET<br>RES 16 1.00 0.00 VAL<br>RES 20 0.75 0.00 TYR<br>RES 46 1.00 0.00 ILE<br>DNAE 1.749   |
| sirA/ YP_039582.1                 | 1st site coordinates predicted by Pocket-Finder<br>Min Coords: (-5, -4, -16)<br>Max Coords: (17, 11, -1)  |
|                                   | Best pocket residues predicted by FINDSITE comb<br>TEMPLATE 3MWFA00<br>TEMPLATE 3NU1B00<br>TEMPLATE 3NU1A00<br>TEMPLATE 2J6IA00<br>RES 81 1.00 0.00 TRP<br>RES 82 0.50 0.00 THR<br>RES 84 0.50 0.00 LYS<br>RES 104 0.50 0.00 PRO<br>RES 124 0.50 0.00 VAL<br>RES 125 0.50 0.00 ARG<br>RES 145 0.50 0.00 VAL<br>RES 201 1.00 0.00 ARG<br>RES 206 0.75 0.00 ARG<br>RES 208 0.75 0.00 TYR<br>RES 238 0.50 0.00 ILE<br>RES 258 0.50 0.00 VAL<br>RES 260 0.50 0.00 SER<br>RES 262 1.00 0.00 PRO<br>RES 263 1.00 0.00 ASN<br>RES 299 0.50 0.00 ASP<br>RES 300 0.50 0.00 GLU<br>RES 301 0.50 0.00 ILE<br>RES 304 0.50 0.00 ASN<br>RES 305 0.50 0.00 LEU<br>DNAE -0.912 |
| Hypothetical protein/ YP_042767.1 | 1st site coordinates predicted by Pocket-Finder<br>Min Coords: (-12, 12, -19)<br>Max Coords: (4, 31, 2)   |
|                                   | Best pocket residues predicted by FINDSITE comb<br>TEMPLATE 2C2FA<br>RES 6 1.00 -0.20 TRP<br>RES 18 1.00 -0.20 VAL<br>RES 19 1.00 -0.20 GLY<br>RES 21 1.00 -0.20 ILE<br>RES 22 1.00 -0.29 LYS<br>DNAE 0.634   |

DNAE stands for DNA binding pockets (-ve value is most favorable)

**Table 8:** Best pockets and its coordinates.

## References

1. Fong C, Rohmer L, Radey M, Wasnick M, Brittnacher MJ (2008) PSAT: a web tool to compare genomic neighborhoods of multiple prokaryotic genomes. *BMC Bioinformatics* 9: 170.
2. Zhang R, Lin Y (2009) DEG 5.0, a database of essential genes in both prokaryotes and eukaryotes. *Nucleic Acids Res* 37: D455-D458.
3. NCBI Homo sapiens (human) Protein BLAST.
4. Yu CS, Lin CJ, Hwang JK (2004) Predicting subcellular localization of proteins for Gram-negative bacteria by support vector machines based on n-peptide compositions. *Protein Sci* 13: 1402-1406.
5. Yu NY, Wagner JR, Laird MR, Melli G, Rey S, et al. (2010) PSORTb 3.0: Improved protein subcellular localization prediction with refined localization subcategories and predictive capabilities for all prokaryotes. *Bioinformatics* 26: 1608-1615.
6. Kanehisa M, Goto S, Sato Y, Furumichi M, Tanabe M (2012) KEGG for integration and interpretation of large-scale molecular data sets. *Nucleic Acids Res* 40: D109-D114.
7. Kelley LA, Sternberg MJ (2009) Protein structure prediction on the Web: a case study using the Phyre server. *Nat Protoc* 4: 363-371.
8. Rodrigues JP, Levitt M, Chopra G (2012) KoBaMIN: a knowledge-based minimization web server for protein structure refinement. *Nucleic Acids Res* 40: W323-W328.
9. Ko J, Park H, Heo L, Seok C (2012) GalaxyWEB server for protein structure prediction and refinement. *Nucleic Acids Res* 40: W294-W297.
10. Rampage.
11. Colovos C, Yeates TO (1993) Verification of protein structures: patterns of nonbonded atomic interactions. *Protein Sci* 2: 1511-1519.
12. TMHMM v 2.0.
13. Chen J, Liu H, Yang J, Chou KC (2007) Prediction of linear B-cell epitopes using amino acid pair antigenicity scale. *Amino Acids* 33: 423-428.
14. Singh H, Raghava GP (2003) ProPred1: prediction of promiscuous MHC Class-I binding sites. *Bioinformatics* 19: 1009-1014.
15. Singh H, Raghava GP (2001) ProPred: prediction of HLA-DR binding sites. *Bioinformatics* 17: 1236-1237.
16. Guan P, Doytchinova IA, Zygouri C, Flower DR (2003) MHCPred: bringing a quantitative dimension to the online prediction of MHC binding. *Appl Bioinformatics* 2: 63-66.
17. Kangueane P, Sakharkar MK (2005) T-Epitope Designer: A HLA-peptide binding prediction server. *Bioinformation* 1: 21-24.
18. Doytchinova IA, Flower DR (2007) VaxiJen: a server for prediction of protective antigens, tumour antigens and subunit vaccines. *BMC Bioinformatics* 8: 4.
19. Zhou H, Skolnick J (2013) FINDSITE(comb): a threading/structure-based, proteomic-scale virtual ligand screening approach. *J Chem Inf Model* 53: 230-240.
20. Lagorce D, Sperandio O, Galons H, Miteva MA, Villoutreix BO (2008) FAF-Drugs2: free ADME/tox filtering tool to assist drug discovery and chemical biology projects. *BMC Bioinformatics* 9: 396.
21. Osiris property explorer.
22. Hendlich M, Rippmann F, Barnickel G (1997) LIGSITE: automatic and efficient detection of potential small molecule-binding sites in proteins. *J Mol Graph Model* 15: 359-363, 389.
23. Thomsen R, Christensen MH (2006) MolDock: a new technique for high-accuracy molecular docking. *J Med Chem* 49: 3315-3321.