

## Micropropagation and Conservation of *Rheum webbianum* Collected from Zanaskar Valley via Tissue Culture

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### Abstract

*Rheum webbianum* is one of the famous and popular medicinal plant found in Ladakh region. *Rheum webbianum* Royle is an important medicinal plant belonging to the family Polygonaceae. It is commonly known as 'Himalayan Rhubarb' in English, 'Revandchini' in Hindi, 'Xu mi da huang' in Chinese, and 'Lachhu' or 'Chu-rtsa' in Ladakh. It is native to Asia-Temperate to Asia-Tropical, from China to India, Nepal and Pakistan. In India, it is found in Himachal Pradesh, Jammu and Kashmir, Uttar Pradesh. In Jammu and Kashmir, it is grown on open slopes and shrubberies. Due to overexploitation the *Rheum webbianum* has become endangered. so, it was conserved by the tissue culture and showed best results on different phytohormones. The plant formation was observed on combination of BAP, TDZ, 2,4-D and IBA.

**Keywords:** Polygonaceae; Tissue culture; Micropropagation; Endangered; High altitudes; Medicinal plants

### Introduction

In modern medicines, the plants are either directly used as medicine or provide bioactive compounds or raw material for semi synthetic drugs or as markers for discovery of new bioactive compounds [1]. As per WHO estimation, the present demand for plant based raw material is approximately USD 14 billion per year with 15-25% annual increase. India is the second largest supplier (next to China only) of raw material of medicinal plants. In India, the medicinal plant related trade is estimated to be USD 1 billion per year [2].

In India, most of these medicinal plants (more than 90%) are collected from wild source with unplanned, unscientific and unregulated way leading to shaky, exploitative and unsustainable supply. Further, local people are unaware of morphological features, leading to mixing of different taxa or many cases leading to intentional adulteration [3,4].

India is one of the 12 mega diversity centres of the world [5]. India has diverse agroclimatic, ecological and edaphic conditions of the cold deserts of alpine Himalayas, temperate and sub-tropical parts of North-West and trans Himalayas, tropical rain forests of Western Ghats and East India, plains of Central and North India, to the dry deserts of Rajasthan and Gujarat. So, India, in general and Kashmir in particular, is rich in medicinal plant diversity, covering species diversity, genetic diversity and ecosystem diversity [6]. Out of 18, 665 Angiosperms reported from India, about 3000 are used for various medicinal purposes [4]. There is a need to study various macroscopical and microscopical characters of drugs as per WHO and pharmacopoeial guidelines [7,8].

A lot of variations in phyto-pharmaceutical products have been reported which are influenced by genetic diversity (both chromosomal and genetic), environmental factors, plant part used, developmental

stage of plant, season of collection, cultivation practices, adulteration with misidentified plant species or intentional mixing, attack of pathogens, etc. [9-16]. Further, due to multi-herbal nature of most of the herbal preparations, it is difficult to identify and quantify the active constituents.

The genus *Rheum L.* (commonly called Rhubarb), belongs to the family Polygonaceae of group Monochlamydae of Dicots, is world over represented by 60 species [17], of which only 7 species have been reported from the Indian subcontinent [18]. According to Stewart [19], all the seven species (including *R. webbianum*) reported from the Indian subcontinent, are also present in the Kashmir Himalaya. In taxonomical focus, inflorescence is pyramidal or spherical in which flowers are arranged in panicles and pedicel articulates at or below the middle. Rhubarb is derived from the Latin word "rha" (river) and "barb" (barbarian land). In ancient times, rhubarb roots were imported by the Romans from barbarian lands which were beyond the Volga or Rha River. According to Lindley's Treasury of Botany, the name is derived from the Greek rheo ('to flow'), in allusion to the purgative properties of the root. A total of 560 plant species of India are included in Red List of Threatened species by IUCN including Rheum species [20]. Rheum is a strong vigorous plant and can live long even in tough conditions. The Rheum species have been domesticated as medicinal plants for human consumption. Rheum species are perennial stout herbs. Rhubarb is a common English name of the genus Rheum. Rhubarb is a plant name for the many different species of Rheum, distributed in the temperate and sub-tropical regions of Himalaya from Kashmir to Sikkim [21] and growing wild in the mountains of the Western and North-western provinces of China and adjoining Tibetan territory. It is cultivated much in Europe and the United States. In Kashmir Himalaya, it ranges from an altitude of 1700-5500 m. It is famous for its medicinal uses. Roots are long and stout. Stem is erect, hollow, sulcate, glabrous or strigose. It is native to Asia-Temperate to Asia - Tropical, from China to India, Nepal and Pakistan. In India, it is found in Himachal Pradesh Jammu & Kashmir and Uttar Pradesh. In Jammu and Kashmir, it grows on open slopes and shrubberies in

Kashmir, Leh and Zanskar valley between 3,105-3,920 m [22]. *Rheum* also prefers a full sunny location and thrives in a moist, fertile soil. Most *Rheum* species can handle heavier clay soils better than many other herbs and makes a striking addition to the spring landscape. Rhubarb is propagated from seeds, seedlings, or pieces of rhizome.

It is used as a strong laxative and is known for its astringent effect on the mucous membranes of the mouth and nasal cavity [23]. Its astringent qualities help to improve bowel tone after it has purged the intestines, making it an excellent agent for improving the tone and health of the digestive tract [24]. Its roots have been found useful in controlling gastrointestinal hemorrhage by promoting the formation of blood platelets. This increase in the number of platelets shortens blood clotting time and is helpful in treating jaundice [25]. It, therefore, has a truly cleansing action upon the gut, removing debris and then astringing with antiseptic properties [26].

*R. webbianum* is used for treatment of indigestion, abdominal disorders, boils, wounds and flatulence [27]. It improves the memory of senile patients [28], helpful in managing cancers [29].

Tissue culture has now become a well-established technique for culturing and studying the physiological behavior of isolated plant organs, tissues, cells and protoplasts under precisely controlled physical and chemical conditions. Most of the medicinal plants either do not produce seeds or seeds are too small and do not germinate in soils. Thus, mass multiplication of disease free planting material is a general problem. In this regard, the micropropagation holds significant promise for true to type, rapid and mass multiplication, and conservation of valuable genotype under disease free conditions [30]. Cultivation of plant tissue in synthetic media offers an alternative way of producing metabolites of interest to the traditional cultivation in the fields or greenhouses [31-34]. In successful cases, cell suspension cultures can offer a repeatable method to produce secondary metabolites from elite mother plants with easily controlled conditions and with a continuous supply of material. The production of secondary metabolites can be enhanced using bioreactors. Genetic transformation may provide increased and efficient system for *in vitro* production of secondary metabolites. Tissue culture protocols have been developed for several medicinal plants, but there are many other species, which are over exploited in pharmaceutical industries and need conservation. However, genetic transformation is the most recent aspect of plant cell and tissue culture for transfer of genes with desirable trait into host plants and the recovery of transgenic plants [35]. Regeneration of disease or viral resistant plants is now achieved by employing genetic transformation technique.

## Materials and Methods

The germplasm of *R. webbianum* was collected from Ladakh and Zanaskar valley as Shown in Figure 1. The explants taken from field material, including seeds of *R. webbianum* were first washed with tap water to remove the dirt. Then transferred to the beaker, labolene and tween-20, a wetting agent, was poured and kept for 20 minutes. After 20 minutes, the beaker was covered with muslin cloth and was taken

under the running tap water so that all the labolene or tween-20 is washed out from explants. Then these were continuously shaken for proper cleaning and were rinsed in pre-autoclaved distilled water. The cleaned plant material was subjected to brief 70% alcohol rinse and soaked in different sterilant solutions of  $HgCl_2$  under laminar air flow cabinet for different time durations to get complete removal of biological contaminants. Seeds were also soaked in water and kept in refrigerator for 3 days. After 3 days, these were taken out, thoroughly washed with running tap water after cleaning them with detergent (labolene) and a few drops of tween-20 (surfactant).

The powder form of MS was used in present study. In powder form of MS medium, sucrose and agar are absent. These are useful for propagation of plant species requiring nutrients according to the recipe of standard media. In the media, DDW and sucrose was added as per instructions e.g., 30.0 g of sucrose in 1 liter of MS media. Then pH (~5.8) was balanced by using NaOH (0.1 N) and HCl (0.1 N). After balancing the pH, agar was added as per instructions i.e., 8.0 g/L in MS media.

In present study, the concentration of phytohormones was used in  $\mu M$ . The storage life of stock solutions i.e., auxins and cytokinins was upto one month under the storage temperature of 4°C.

All the glassware, forceps, petriplates, cotton plugs, DDW and beakers were autoclaved before using for tissue culture purposes. After boiling the media, the media was put into the glass vials (100 ml) and were also autoclaved for 20 min. at 121°C and 15 lbs/(inch)<sup>2</sup>. Sterilized explants were inoculated onto aseptic nutrient medium under laminar air flow cabinet. The whole process of inoculation was performed under highly aseptic conditions (Table 1).

After inoculation the cultures were kept in culture room. Cultures were kept for incubation under cool fluorescent tubes at day night regime of 16-hour photoperiod with light intensity of 1500-3000 lux at a constant temperature of  $25 \pm 3^\circ C$ . Relative humidity between 60-70% was maintained.

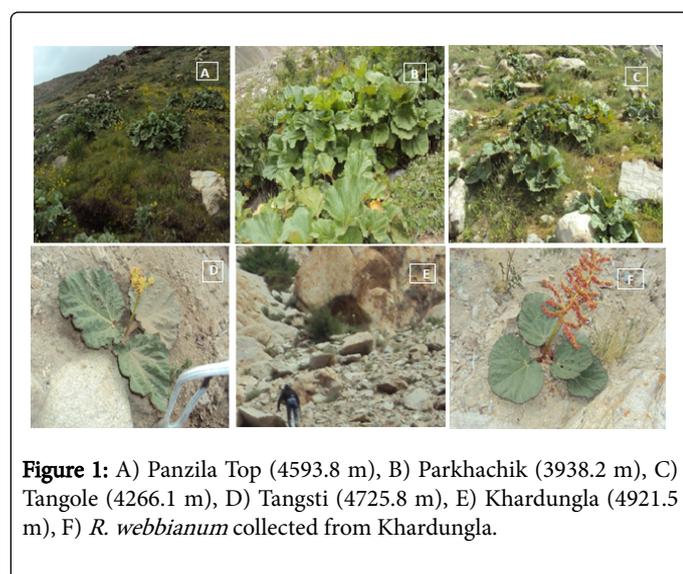
Subculturing was done after every 3-5 weeks, depending upon the organogenesis and proliferation potential of the explants of culture and was carried out in the Laminar air flow chamber under aseptic conditions. The plantlets obtained from different explants through repeated sub-cultures were finally left in culture vials with open mouth for three days in the incubation room, transferred to plastic pots containing soil-peat mixture and then taken out of incubation room of the lab. Attempts were made to acclimatize plants under the green house facility.

Ten replicates were taken for each treatment and observations were recorded at the end of every week. Data were subjected to analysis of variance using SSPP software version 17.0 (SAS Institute Inc., Cary, NC). The growth response of explants under normal conditions and also the effect of plant growth hormone response (PGPR) treatments were considered significant according to the magnitude of the F value ( $P \leq 0.005$ ).

Sr. No.	Explants (Field Material)	Chemical sterilants (Conc.) %	Duration (min)	Contamination (%)	Explants survival %
1	Seeds	$HgCl_2$ (0.02)	15	20	70

		HgCl <sub>2</sub> (0.03)	8	20	80
		HgCl <sub>2</sub> (0.05)	10	10	100
2	Leaves	HgCl <sub>2</sub> (0.02)	10	30	50
		HgCl <sub>2</sub> (0.03)	7	30	80
		HgCl <sub>2</sub> (0.05)	5	20	90
3	Petioles	HgCl <sub>2</sub> (0.01)	10	60	40
		HgCl <sub>2</sub> (0.03)	7	50	50
		HgCl <sub>2</sub> (0.05)	5	20	90
4	Leaf Midrib	HgCl <sub>2</sub> (0.2)	10	40	80
		HgCl <sub>2</sub> (0.03)	10	40	70
		HgCl <sub>2</sub> (0.05)	5	20	80
5	Shoot tips	HgCl <sub>2</sub> (0.1)	5	20	80
		HgCl <sub>2</sub> (0.1)	7	40	60
		HgCl <sub>2</sub> (0.03)	10	40	60

**Table 1:** Showing the Sterilization and contamination.



## Results

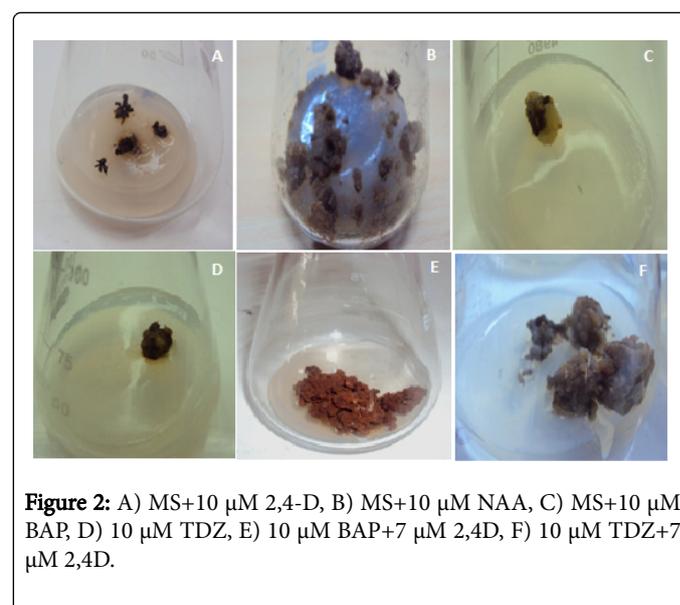
The explants (leaves, leaf midribs, petioles, rhizomes, roots and shoot tips) from field materials were washed in labolene and tween 20 and then washed under running water for 20 minutes. For surface sterilization, Mercuric chloride (HgCl<sub>2</sub>) was used. The details of the effective treatments for surface sterilization are shown in below

### Responses of different media

The different explants were cultured on different media i.e., Linsmaier and Skoog (LS), Nitsch and Nitsch (NN), Murashige and Skoog medium (MS), supplemented with different concentration and

combinations of phytohormones. But only MS medium showed good response, therefore further experiments were carried out using MS medium.

**MS medium with different concentration of auxins:** Effect of MS medium with different concentrations of auxins on callus formation from field grown plant was noted. Different explants (leaf, leaf midrib, petiole, and shoot tips) of *R. webbianum* were taken from field grown plants. The above explants were used for regeneration studies on different Auxins (2,4-D, NAA, IAA, IBA) with concentration varying from 5 μM to 10 μM.



But these field explants did not show any proper regeneration. The brownish black callus was observed on MS+10 µM 2,4-D and MS+10 µM NAA as shown in Figure 2, respectively.

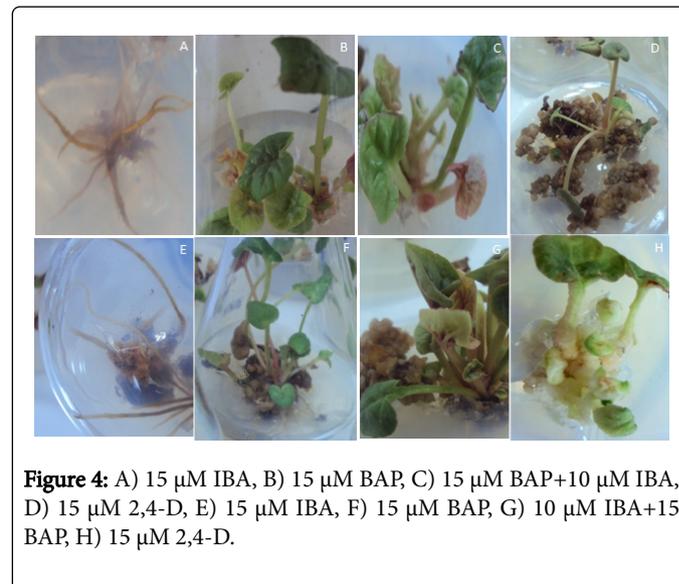
**MS medium with different concentration of cytokinins:** Effect of MS medium with different concentrations of cytokinins on callus formation from field grown plants was noted. The 50% of explants showed the callus formation on different concentrations of cytokinins i.e., MS medium with 10 µM BAP and 10 µM TDZ as shown in Figure 3, respectively.



**Figure 3:** A) 10 µM 2,4-D, B) 10 µM 2,4-D+8 µM NAA, C) 10 µM IAA+10 µM IBA, D) 10 µM BAP, E) 10 µM BAP+10 µM TDZ, F) 10 µM BAP+7 µM 2,4-D.

**MS medium with different concentration of auxins and cytokinins combination:** Effect of MS medium with different concentrations of

cytokinins and auxins on callus formation from field grown plant was noted on different combinations but the best result among all was observed on 10 µM BAP+7 µM 2,4D and 10 µM TDZ+7 µM 2,4-D as shown in Figure 4, respectively. The callus obtained was further taken for subculturing on the same combinations of phytohormones for further regeneration.



**Figure 4:** A) 15 µM IBA, B) 15 µM BAP, C) 15 µM BAP+10 µM IBA, D) 15 µM 2,4-D, E) 15 µM IBA, F) 15 µM BAP, G) 10 µM IBA+15 µM BAP, H) 15 µM 2,4-D.

#### Seed germination of *R. webbianum*

Seed germination studies were carried out using MS medium with different concentrations of phytohormones. Various combinations were used for germination, but the best results observed are given in Table 2 and Figures 3A-3C.

Sr. No.	Phytohormones (Conc. in µM)	% seed germination	%Plant formation
1.	10.0 µM 2,4-D	40	20
2.	10.0 µM 2,4-D+8.0 µM NAA	40	20
3.	10.0 µM IAA+10.0 µM IBA	40	30
4.	10.0 µM BAP	50	40
5.	10.0 µM BAP+10.0 µM KN	60	50
6.	10.0 µM BAP+10.0 µM TDZ	60	50
8.	10.0 µM BAP+10 µM TDZ+10 µM IBA	60	40
9.	15.0 µM BAP+10.0 µM TDZ+10.0 µM 2,4-D	70	50

**Table 2:** Response of seed germination on different media combinations.

These seedlings of *R. webbianum* were further used for regeneration studies as source of various explants i.e., leaves, leaf midrbs, petioles and shoot tips.

#### Regeneration studies on *R. webbianum* using different explants from tissue culture raised seedlings

Callus formation was observed from all the four explants (leaf, leaf mid rib, petiole and shoot tip explants) at different concentrations of

auxins and cytokinins individually as well as in combinations after 4-6 weeks of initial inoculation. The weight of callus was in the range of 3.0 g to 6.0 g.

#### Leaf culture, shoot tip culture, petiole culture, leaf midrib culture

Indirect organogenesis was observed after further subculturing of callus on same media combinations i.e., regeneration of shoots and

roots were observed after 4-6 weeks of sub culturing as shown in Tables 3-5.

Only rooting was observed on IAA and IBA, when used individually, and the rooting in large number was found on 15 µM IBA with 70% response (Figure 3D). However, only shoot regeneration was observed at low concentrations (5-10 µM) of BAP and Kn. The different explants from seedling were used for regeneration studies. After 2 to 3 weeks callus formation was observed. Regeneration of

shoots was observed when the above callus was sub cultured on the same medium after four weeks. The shoot regeneration was observed at higher concentration of auxins i.e., 15 µM 2,4-D and 15 µM NAA using different explants as shown in the Figure 5 refer to Table 3. The regenerated shoot number was maximum approximately 5 per explants (Figure 3E) in case of leaf midrib and shoot tip explants at 15 µM 2,4-D (Figure 3F). However, no shooting was observed in case of higher concentration of IBA and IAA. Only rooting was observed from callus.

Sr. No.	Explants	2,4-D (µM)	NAA (µM)	IAA (µM)	IBA (µM)	Shoot No.	Shoot length (cm)	Leaf shoot No./	Root No.	Root length (cm)	Response	% response
1.	Leaf											
		12	-	-	-	a3.7 ± 0.5	a1.7 ± 0.4	b4.9 ± 0.5	ab2.1 ± 0.5	a1.5 ± 0.4	Indirect shooting/ rooting	60
		15	-	-	-	b4.5 ± 0.5	ab2.1 ± 0.5	bc5.1 ± 0.5	b2.5 ± 0.5	a1.7 ± 0.5	do	60
		-	12	-	-	a3.2 ± 0.4	a1.5 ± 0.5	a3.9 ± 0.5	a1.9 ± 0.4	a1.4 ± 0.3	do	60
		-	15	-	-	a3.9 ± 0.5	a1.9 ± 0.3	b4.7 ± 0.5	c3.2 ± 0.5	b2.3 ± 0.1	do	60
		-	-	12	-	nil	nil	nil	d4.2 ± 0.3	b2.5 ± 0.3	indirect rooting only	50
		-	-	15	-	nil	nil	nil	d4.8 ± 0.5	b 2.9 ± 0.5	do	50
		-	-	-	12	nil	nil	nil	e5.1 ± 0.3	c 3.2 ± 0.5	do	50
		-	-	-	15	nil	nil	nil	e5.4 ± 0.5	c 3.5 ± 0.5	do	50
2.	Leaf mid rib											
		12	-	-	-	a3.8 ± 0.4	a1.8 ± 0.4	cb5.1 ± 0.5	b2.2 ± 0.5	a1.7 ± 0.5	indirect minimum shooting/rooting	60
		15	-	-	-	b4.9 ± 0.5	b2.5 ± 0.5	c5.6 ± 0.5	b2.7 ± 0.5	a1.8 ± 0.5	do	60
		-	12	-	-	a3.4 ± 0.5	a1.7 ± 0.5	b4.2 ± 0.5	ba2.1 ± 0.5	a1.7 ± 0.5	do	do
		-	15	-	-	b4.3 ± 0.5	b2.2 ± 0.1	b4.9 ± 0.5	c3.4 ± 0.5	b2.5 ± 0.5	do	do
		-	-	12	-	nil	nil	nil	d4.4 ± 0.5	b 2.7 ± 0.5	indirect rooting only	50
		-	-	15	-	nil	nil	nil	d4.8 ± 0.5	c 3.2 ± 0.5	do	50
		-	-	-	12	nil	nil	nil	e5.4 ± 0.5	c 3.4 ± 0.5	do	50
		-	-	-	15	nil	nil	nil	e5.7 ± 0.5	c 3.7 ± 0.6	do	50
3.	Petiole											
		12	-	-	-	a3.4 ± 0.3	a1.4 ± 0.3	b4.5 ± 0.	a1.8 ± 0.3	a1.3 ± 0.2	minimum shooting/ rooting	60

		15	-	-	-	b4.2 ± 0.2	a1.8 ± 0.3	b4.9 ± 0.5	b2.2 ± 0.1	a1.5 ± 0.2	do	60
		-	12	-	-	a3.1 ± 0.2	a1.3 ± 0.2	a3.7 ± 0.5	a1.7 ± 0.4	a1.2 ± 0.3	do	60
		-	15	-	-	a3.7 ± 0.5	a1.7 ± 0.3	b4.5 ± 0.5	cb3.1 ± 0.5	ab2.1 ± 0.1	do	60
		-	-	12	-	nil	nil	nil	3.8 ± 0.2	b 2.3 ± 0.1	indirect rooting only	50
		-	-	15	-	nil	nil	nil	d4.5 ± 0.5	b 2.6 ± 0.5	do	50
		-	-	-	12	nil	nil	nil	d4.8 ± 0.3	c3.1 ± 0.1	do	50
		-	-	-	15	nil	nil	nil	e5.1 ± 0.4	c3.3 ± 0.2	do	50
4.	Shoot tip											
		12	-	-	-	a3.9 ± 0.5	a1.9 ± 0.4	c5.2 ± 0.3	b2.4 ± 0.3	a1.7 ± 0.4	minimum shooting/ rooting	60
		15	-	-	-	b4.8 ± 0.5	b2.4 ± 0.5	c5.8 ± 0.5	b2.7 ± 0.5	a1.9 ± 0.5	do	60
		-	12	-	-	a3.5 ± 0.4	a1.7 ± 0.5	b4.5 ± 0.4	b2.5 ± 0.3	a1.7 ± 0.5	minimum shooting/ rooting	60
		-	15	-	-	b4.3 ± 0.5	b2.3 ± 0.2	b4.9 ± 0.5	c3.6 ± 0.5	b2.5 ± 0.3	do	60
		-	-	12	-	nil	nil	nil	d4.7 ± 0.5	b 2.9 ± 0.5	indirect rooting only	50
		-	-	15	-	nil	nil	nil	e5.5 ± 0.5	c 3.5 ± 0.5	do	50
		-	-	-	12	nil	nil	nil	e5.7 ± 0.3	c3.8 ± 0.5	do	50
		-	-	-	15	nil	nil	nil	e5.9 ± 0.7	d4.2 ± 0.5	do	50

**Table 3:** Indirect organogenesis on MS medium with different concentrations of auxins on multiple shoot regeneration/rooting from callus obtained from different explants of *R. webbianum*. Values are represented as mean ± SD (n=10), Data was analyzed by ANOVA using Duncan's multiple range test (SPSS17.0); the values with different superscript along the columns are statistically significant at P<0.005, Data scored after 12 weeks of culture period.

The effect of cytokinins were studied using different explants as shown in Figure 6 refer to the Table 4. The maximum number of shoots regeneration from callus was observed on 15 µM of cytokinins i.e.,

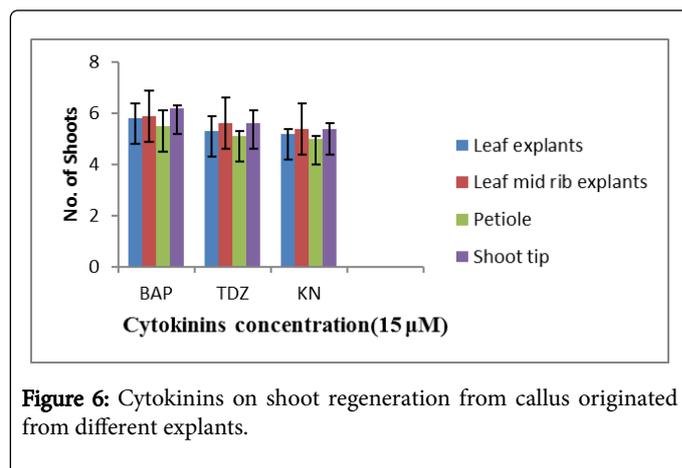
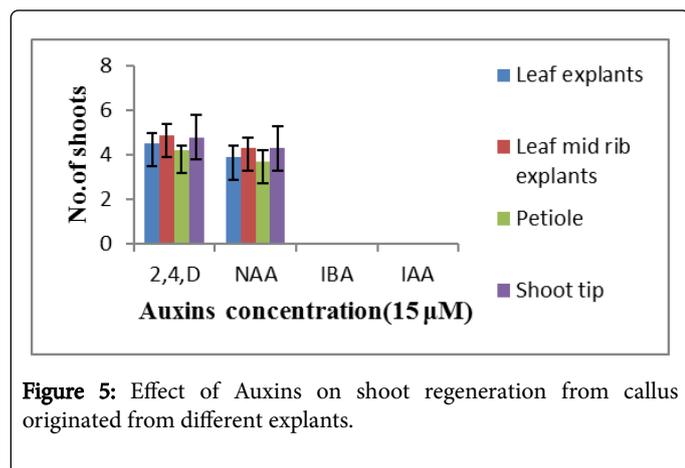
BAP, TDZ and Kn from all the explants. However, in case of shoot tip explants the number of regenerated shoots was maximum i.e., 6 shoots per explants at 15 µM BAP.

Sr. No.	Explants	BAP (µM)	TDZ (µM)	KN (µM)	Z (µM)	Shoot No.	Shoot length (cm)	Leaf shoot No./	Root No.	Root length (cm)	Response	% response
1.	Leaf											
		12	-	-	-	c5.2 ± 0.5	ab2.1 ± 0.4	dc6.1 ± 0.5	Nil	Nil	indirect shooting only	70
		15	-	-	-	c5.8 ± 0.6	b2.2 ± 0.5	d6.9 ± 0.5	Nil	nil	indirect shooting maximum	70

		-	12	-	-	b4.8 0.2	± a1.8 ± 0.3	c5.8 ± 0.5	a1.7 0.3	± a1.2 0.4	± infirect shooting/rooting	70
		-	15	-	-	c5.3 0.6	± ab2.1 ± 0.2	dc6.1 ± 0.3	a1.9 0.5	± a1.5 0.1	± indirect maximum shooting/rooting	70
		-	-	12	-	b4.9 0.3	± 1.3 ± 0.2	5.3 ± 0.4	1.6 0.3	± a1.2 0.5	± indirect shooting/rooting	70
		-	-	15	-	c5.2 0.2	± a1.5 ± 0.3	c5.7 ± 0.2	a1.8 0.3	± a1.4 0.2	± indirect maximum shooting/rooting	70
2.	Leaf mid rib											
		12	-	-	-	c5.5 0.5	± b2.3 ± 0.4	d6.4 ± 0.5	Nil	Nil	indirect shooting only	70
		15	-	-	-	c5.9 0.5	± b2.4 ± 0.5	e7.2 ± 0.1	a1.5 0.3	± a1.7 0.2	± do	70
		-	12	-	-	b4.9 0.3	± a1.9 ± 0.5	c5.9 ± 0.5	a1.9 0.4	± a1.4 0.5	± do	70
		-	15	-	-	c5.6 0.6	± b2.4 ± 0.5	d6.4 ± 0.3	b2.5 0.5	± a1.7 0.3	± do	70
		-	-	12	-	c5.1 0.3	± a1.6 ± 0.2	c5.7 ± 0.4	a1.8 0.5	± a1.4 0.5	± do	70
		-	-	15	-	c5.4 0.2	± a1.7 ± 0.3	dc6.1 ± 0.2	a1.9 0.4	± a1.6 0.4	± do	70
3.	Petiole											
		12	-	-	-	bc5.0 0.1	± a1.9 ± 0.4	c5.8 ± 0.5	Nil	Nil	indirect shooting only	60
		15	-	-	-	c5.5 0.6	± a1.2 ± 0.5	d6.9 ± 0.5	Nil	nil	do	60
		-	12	-	-	b4.6 0.2	± a1.6 ± 0.3	c5.7 ± 0.5	a1.5 0.3	± a1.1 0.2	± indirect shooting/rooting	60
		-	15	-	-	bc5.1 0.2	± a1.9 ± 0.3	c5.9 ± 0.3	a1.7 0.5	± a1.3 0.1	± do	60
		-	-	12	-	b4.7 0.3	± a1.2 ± 0.1	5.1 ± 0.3	a1.4 0.2	± a1.1 0.2	± do	60
		-	-	15	-	bc5.0 0.1	± a1.3 ± 0.2	c5.5 ± 0.2	a1.6 0.3	± a1.2 0.1	± do	60
4.	Shoot tip											
		12	-	-	-	c5.4 0.3	± b2.5 ± 0.4	d6.4 ± 0.5	Nil	Nil	indirect shooting only	70
		15	-	-	-	d6.2 0.1	± b2.5 ± 0.3	e7.3 ± 0.5	a1.9 ± 0 ± 3	± a1.5 0.2	± maximum indirect shooting/rooting	70
		-	12	-	-	c5.2 0.2	± b2.3 ± 0.2	d6.2 ± 0.5	a1.9 0.5	± a1.4 0.2	± do	70
		-	15	-	-	c5.6 0.5	± b2.3 ± 0.2	d6.4 ± 0.4	b2.3 0.5	± a1.7 0.2	± do	70
		-	-	12	-	c5.2 0.1	± 1.6 ± 0.2	5.7 ± 0.3	1.9 0.5	± 1.5 ± 0.5	do	70

		-	-	15	-	c5.4 ± 0.2	a1.7 ± 0.3	c5.9 ± 0.2	ba2.1 ± 0.1	a1.6 ± 0.4	do	70
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**Table 4:** Indirect organogenesis on MS medium with different concentrations of cytokinins on multiple shoot regeneration/rooting from callus obtained from different explants of *R. webbianum*. Values are represented as mean ± SD (n=10), Data was analyzed by ANOVA using Duncan's multiple range test (SPSS17.0); the values with different superscript along the columns are statistically significant at P<0.005, Data scored after 12 weeks of culture period.

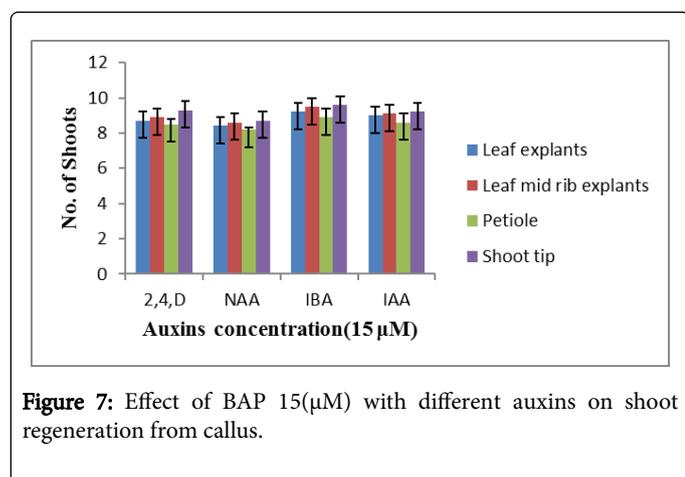


Sr. No.	Cytokinin	Auxins				Shoot No.	Shoot length (cm)	Leaf no.	Root No	Root length (cm)	Response	% Response	
		BAP (μM)	2,4-D (μM)	NAA (μM)	IBA (μM)								IAA (μM)
1.	Leaf												
		15	15				d8.7 ± 0.5	c3.5 ± 0.2	g9.9 ± 0.5	c3.5 ± 0.2	b2.7 ± 0.5	Multiplication elongation/ rooting	70
		15		15			d8.4 ± 0.5	bc3.1 ± 0.2	g9.1 ± 0.3	c3.7 ± 0.3	b2.9 ± 0.5	do	70
		15			15		e9.2 ± 0.5	c3.5 ± 0.3	h10.5 ± 0.5	e5.7 ± 0.5	cd4.1 ± 0.3	do	70
		15				15	e9.0 ± 0.5	c3.0 ± 0.3	h10.1 ± 0.5	e5.4 ± 0.5	c3.9 ± 0.2	do	70
2.	Mid rib												
		15	15				d8.9 ± 0.5	c3.6 ± 0.2	hg10.1 ± 0.5	c3.6 ± 0.2	b2.9 ± 0.5	do	70
		15		15			d8.6 ± 0.5	c3.3 ± 0.2	g9.3 ± 0.4	c3.9 ± 0.3	c3.3 ± 0.5	do	70
		15			15		e9.5 ± 0.5	c3.7 ± 0.3	h10.7 ± 0.5	e5.9 ± 0.5	d4.3 ± 0.3	do	70
		15				15	e9.1 ± 0.5	c3.2 ± 0.3	h10.2 ± 0.5	e5.3 ± 0.5	d4.1 ± 0.3	do	
3.	Petiole												
		15	15				d8.5 ± 0.3	c3.2 ± 0.2	g9.7 ± 0.5	c3.3 ± 0.2	c2.5 ± 0.3	do	70

	15		15			d8.2 ± 0.1	b2.9 ± 0.2	f8.8 ± 0.4	c3.5 ± 0.2	c2.7 ± 0.3	do	70
	15		15			d8.9 ± 0.5	c3.6 ± 0.3	g9.9 ± 0.5	e5.6 ± 0.3	c3.9 ± 0.3	do	70
	15		15			d8.6 ± 0.5	c3.3 ± 0.3	g9.7 ± 0.5	e5.3 ± 0.3	c3.5 ± 0.3	do	70
4.	Shoot tip											
	15	15				e9.3 ± 0.5	c3.7 ± 0.2	h10.5 ± 0.5	c3.7 ± 0.5	b2.9 ± 0.5	do	70
	15		15			d8.7 ± 0.5	c3.4 ± 0.2	h10.3 ± 0.3	c3.9 ± 0.5	c3.2 ± 0.1	do	70
	15		15			e9.6 ± 0.5	c3.8 ± 0.3	h10.8 ± 0.5	e5.9 ± 0.5	d4.4 ± 0.3	do	70
	15		15			e9.2 ± 0.5	c3.4 ± 0.3	h10.4 ± 0.5	e5.6 ± 0.5	d4.1 ± 0.3	do	70

**Table 5:** Indirect organogenesis on MS medium with 15 µM BAP and different concentrations of auxins on multiple shoot regeneration/rooting from callus obtained from different explants of *R. webbianum*. Values are represented as mean ± SD (n=10), Data was analyzed by ANOVA using Duncan's multiple range test (SPSS17.0); the values with different superscript along the columns are statistically significant at P<0.005, Data scored after 12 weeks of culture period.

The effect of combination of phytohormones i.e., cytokinins with auxins on fixed concentration of 15 µM was studied as shown in the Figure 7 refer to the Table 5. The shoot tip showed maximum shoot regeneration from callus on 15 µM BAP with 15 µM IBA i.e., 10 shoots per explants (Figure 8).



**Figure 7:** Effect of BAP 15(µM) with different auxins on shoot regeneration from callus.

The different concentrations and combinations of phytohormones were used for regeneration studies from different explants of *R. webbianum*, out of which the 15 µM BAP with 15 µM IBA combination showed good results. The regenerated shoot number on 15 µM concentration of BAP with different auxins i.e., 2,4-D, NAA, IBA and IAA was found to be more than 8 per explants in all the explants (Figure 9).



**Figure 8:** 10 µM BAP+10 µM TDZ+10 µM IBA.



**Figure 9:** 15 µM BAP+10 µM TDZ+10 µM 2,4-D.

### Direct organogenesis from different *in-vitro* explants of *R. webbianum*

The plants developed from callus of explants (originated from seedlings) were used as a source of *in-vitro* explants which were further used for regeneration studies i.e., direct organogenesis with minimum callus formation on different combinations of phytohormones. The different explants used for direct regeneration were leaf, leaf midrib, petiole and shoot tip.

The *in-vitro* explants were sub-cultured on different concentrations of phytohormones starting from 5  $\mu\text{M}$  to 15  $\mu\text{M}$  but the best results were found on high concentrations of phytohormones. The regeneration data collected from above experiments was analyzed by ANOVA using Duncan's multiple test as shown in Tables 6-8.

The plant formation on auxins from all the *in-vitro* explants was found on MS+15  $\mu\text{M}$  2,4-D and from cytokinin's was found on MS+15  $\mu\text{M}$  BAP with 70% response (Figure 10) and the combination of phytohormones i.e., 15  $\mu\text{M}$  BAP+10  $\mu\text{M}$  IBA, complete plant formation was also observed with 80% response (Figure 4A). Only rooting was observed on IAA and IBA when used individually and the rooting was maximum on 15  $\mu\text{M}$  IBA with 70% response (Figure 4B). It was observed that at low concentrations of BAP and KN only shooting was observed.



Figure 10: 15  $\mu\text{M}$  2,4-D.

The effect of 15  $\mu\text{M}$  of auxins on different *in-vitro* explants was studied for shoot regeneration as shown in Figure 11 refer to Table 6. The different concentrations (5 to 15  $\mu\text{M}$ ) were used, but the maximum shoot regeneration was observed at 15  $\mu\text{M}$ . The maximum shoot regeneration was found in case of leaf midrib and shoot tip i.e., 5 shoots per explants without callus formation (Figures 4C and 4D). No shoot regeneration was found in case of IAA and IBA, only rooting was observed even at higher concentrations.

Sr. No.	Explants	2,4-D ( $\mu\text{M}$ )	NAA ( $\mu\text{M}$ )	IAA ( $\mu\text{M}$ )	IBA ( $\mu\text{M}$ )	Shoot No.	Shoot length (cm)	Leaf shoot No./	Root No.	Root length (cm)	Response	% response
1.	Leaf											
		12	-	-	-	a3.8 ± 0.5	a1.8 ± 0.4	c5.2 ± 0.3	b2.3 ± 0.2	a1.7 ± 0.4	minimum shooting/ rooting	60
		15	-	-	-	b4.6 ± 0.4	b2.3 ± 0.5	c5.4 ± 0.5	b2.7 ± 0.5	a1.9 ± 0.3	do	60
		-	12	-	-	a3.4 ± 0.4	a1.7 ± 0.5	ab4.2 ± 0.3	b2.3 ± 0.4	a1.7 ± 0.3	do	60
		-	15	-	-	b4.3 ± 0.2	b2.3 ± 0.2	b4.9 ± 0.5	c3.4 ± 0.5	b2.5 ± 0.3	do	60
		-	-	12	-	nil	nil	nil	4.9 ± 0.5	c3.2 ± 0.4	direct rooting only	50
		-	-	15	-	nil	nil	nil	5.6 ± 0.5	c3.5 ± 0.5	do	50
		-	-	-	12	nil	nil	nil	5.2 ± 0.5	d4.2 ± 0.8	do	50
		-	-	-	15	nil	nil	nil	6.3 ± 0.5	d4.5 ± 0.5	do	50
2.	Leaf mid rib											
		12	-	-	-	a3.9 ± 0.4	b2.3 ± 0.4	c5.3 ± 0.5	b2.4 ± 0.5	a1.9 ± 0.5	minmum shooting/ rooting	60
		15	-	-	-	c5.2 ± 0.3	b2.7 ± 0.5	c5.8 ± 0.5	b2.9 ± 0.5	ab2.2 ± 0.5	maximum shooting/ rooting	60

		-	12	-	-	a3.6 ± 0.5	a1.9 ± 0.5	b4.4 ± 0.3	b2.3 ± 0.5	a1.8 ± 0.3	minimum shooting/ rooting	60
		-	15	-	-	b4.5 ± 0.3	b2.4 ± 0.1	c5.3 ± 0.3	c3.6 ± 0.5	b2.7 ± 0.5	do	60
		-	-	12	-	nil	nil	nil	e5.1 ± 0.5	c3.5 ± 0.4	direct rooting only	50
		-	-	15	-	nil	nil	nil	e5.8 ± 0.5	c3.8 ± 0.5	do	50
		-	-	-	12	nil	nil	nil	e5.5 ± 0.5	d4.5 ± 0.5	do	50
		-	-	-	15	nil	nil	nil	f6.5 ± 0.5	d4.7 ± 0.5	do	50
3.	Petiole											
		12	-	-	-	a3.6 ± 0.5	a1.6 ± 0.3	a4.7 ± 0.5	a1.9 ± 0.3	a1.5 ± 0.2	minimum shooting/ rooting	60
		15	-	-	-	b4.4 ± 0.2	a1.9 ± 0.3	c5.3 ± 0.3	b2.4 ± 0.1	a1.7 ± 0.2	do	60
		-	12	-	-	a3.4 ± 0.2	a1.4 ± 0.2	a3.9 ± 0.5	a1.9 ± 0.4	a1.4 ± 0.3	do	60
		-	15	-	-	a3.8 ± 0.5	a1.9 ± 0.3	b4.7 ± 0.5	c3.3 ± 0.5	b2.4 ± 0.3	do	60
		-	-	12	-	nil	nil	nil	d4.1 ± 0.2	b2.7 ± 0.3	direct rooting only	50
		-	-	15	-	nil	nil	nil	e 5.0 ± 0.2	c3.2 ± 0.5	do	50
		-	-	-	12	nil	nil	nil	e5.2 ± 0.3	c3.3 ± 0.5	do	50
		-	-	-	15	nil	nil	nil	e 5.5 ± 0.6	c3.6 ± 0.5	do	50
4.	Shoot tip											
		12	-	-	-	b4.3 ± 0.5	b2.3 ± 0.4	c5.4 ± 0.3	b2.6 ± 0.3	a1.9 ± 0.4	direct shooting/rooting	60
		15	-	-	-	c5.2 ± 0.1	b2.6 ± 0.5	c5.9 ± 0.3	b2.9 ± 0.5	b2.3 ± 0.5	do	60
		-	12	-	-	a3.7 ± 0.4	a1.9 ± 0.5	a4.7 ± 0.4	b2.7 ± 0.3	a1.9 ± 0.5	do	60
		-	15	-	-	b4.5 ± 0.5	b2.5 ± 0.2	c5.3 ± 0.2	c3.8 ± 0.5	b2.7 ± 0.3	do	60
		-	-	12	-	nil	nil	nil	e5.3 ± 0.2	b 2.7 ± 0.5	only rooting	50
		-	-	15	-	nil	nil	nil	e 5.1 ± 0.1	c3.1 ± 0.2	do	50
		-	-	-	12	nil	nil	nil	e 5.2 ± 0.3	c3.4 ± 0.5	do	50

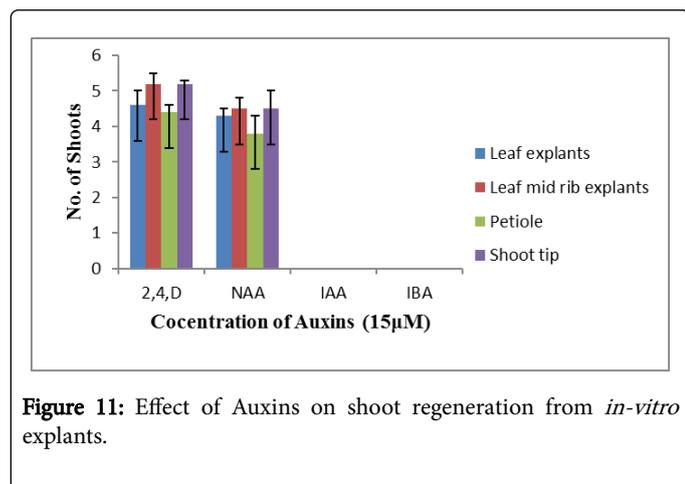
		-	-	-	15	nil	nil	nil	e 5.6 ± 0.5	c3.7 ± 0.5	do	50
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**Table 6:** Direct organogenesis on MS medium with different concentrations of auxins on multiple shoot regeneration/rooting from *in-vitro* explants of *R. webbianum*. Values are represented as mean ± SD (n=10), Data was analyzed by ANOVA using Duncan's multiple range test (SPSS17.0); the values with different superscript along the columns are statistically significant at P<0.005, Data scored after 12 weeks of culture period.

Sr. No.	Explants	BAP (µM)	TDZ (µM)	KN (µM)	Z (µM)	Shoot No.	Shoot length (cm)	Leaf shoot No./	Root No.	Root length (cm)	Response	% response
1.	Leaf											
		12	-	-	-	c5.4 ± 0.3	b2.4 ± 0.2	d6.3 ± 0.5	nil	Nil	direct shooting only	70
		15	-	-	-	c5.9 ± 0.4	b2.4 ± 0.3	e7.2 ± 0.1	nil	nil	do	70
		-	12	-	-	b4.9 ± 0.2	a1.9 ± 0.3	c5.9 ± 0.5	a1.9 ± 0.3	a1.4 ± 0.3	direct shooting/ rooting	70
		-	15	-	-	c5.4 ± 0.3	b2.3 ± 0.2	d6.3 ± 0.3	b2.3 ± 0.4	a1.7 ± 0.3	do	70
		-	-	12	-	c5.0 ± 0.5	a1.3 ± 0.2	c5.4 ± 0.3	a1.7 ± 0.5	a1.2 ± 0.5	do	70
		-	-	15	-	c5.4 ± 0.3	a1.7 ± 0.3	c5.9 ± 0.5	b2.4 ± 0.5	a1.6 ± 0.4	do	70
2.	Leaf mid rib											
		12	-	-	-	c5.7 ± 0.5	b2.5 ± 0.4	d6.6 ± .5	nil	Nil	direct maximum shooting only	70
		15	-	-	-	d6.2 ± 0.3	b2.6 ± 0.3	e7.4 ± 0.5	a1.7 ± 0.3	a1.9 ± 0.2	direct shooting/ rooting	70
		-	12	-	-	c5.4 ± 0.2	b2.3 ± 0.5	d6.3 ± 0.5	b2.4 ± 0.3	a1.6 ± 0.2	direct shooting / rooting	70
		-	15	-	-	c5.8 ± 0.3	b2.6 ± 0.5	d6.6 ± 0.3	b2.7 ± 0.5	a1.9 ± 0.3	do	70
		-	-	12	-	c5.3 ± 0.1	a1.7 ± 0.2	c5.9 ± 0.4	b 2.1 ± 0.3	a1.6 ± 0.2	do	70
		-	-	15	-	c5.6 ± 0.2	a1.9 ± 0.3	d6.3 ± 0.5	b2.3 ± 0.4	a1.8 ± 0.4	do	70
3.	Petiole											
		12	-	-	-	c5.3 ± 0.4	b2.5 ± 0.3	d6.2 ± 0.1	nil	Nil	direct shooting only	70
		15	-	-	-	c5.7 ± 0.5	a1.4 ± 0.3	e7.3 ± 0.5	nil	nil	direct shooting only	70
		-	12	-	-	b4.8 ± 0.3	a1.8 ± 0.3	c5.9 ± 0.3	a1.7 ± 0.3	a1.3 ± 0.2	direct shooting/ rooting	70
		-	15	-	-	c5.3 ± 0.2	b2.3 ± 0.5	d6.4 ± 0.5	a1.9 ± 0.5	a1.6 ± 0.3	do	70
		-	-	12	-	b4.8 ± 0.3	a1.3 ± 0.2	c5.5 ± 0.4	a1.6 ± 0.2	a1.2 ± 0.1	do	70
		-	-	15	-	c5.3 ± 0.4	a1.5 ± 0.2	c5.7 ± 0.2	a1.8 ± 0.3	a1.4 ± 0.1	do	70
4.	Shoot tip											
		12	-	-	-	c5.6 ± 0.3	b2.7 ± 0.4	d6.6 ± 0.5	nil	Nil	direct shooting only	70
		15	-	-	-	d6.4 ± 0.4	b2.7 ± 0.3	e7.6 ± 0.5	b2.5 ± 0.3	a1.7 ± 0.2	direct shooting/ rooting	70
		-	12	-	-	c5.4 ± 0.2	b2.5 ± 0.2	d6.5 ± 0.5	b2.3 ± 0.5	a1.6 ± 0.2	do	70
		-	15	-	-	c5.8 ± 0.4	b2.5 ± 0.2	d6.6 ± 0.4	b2.4 ± 0.5	a1.8 ± 0.2	do	70

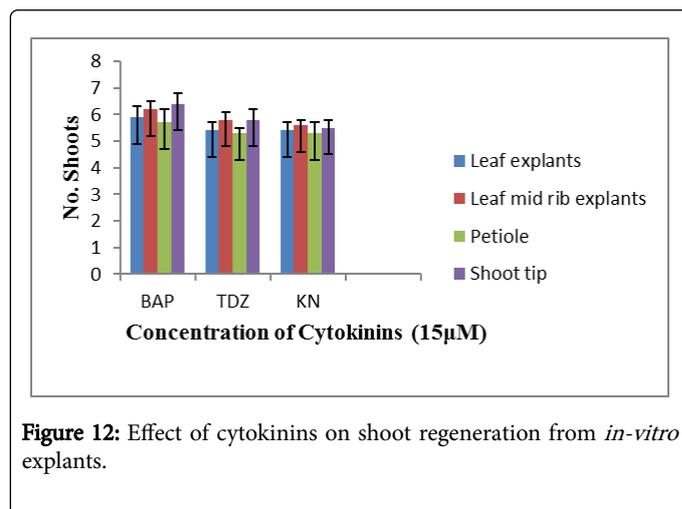
		-	-	12	-	c5.4 ± 0.2	a1.8 ± 0.2	d6.1 ± 0.3	b2.3 ± 0.1	a1.7 ± 0.3	do	70
		-	-	15	-	c5.5 ± 0.3	a1.9 ± 0.3	d6.2 ± 0.5	b2.4 ± 0.1	a1.8 ± 0.4	do	70

**Table 7:** Direct organogenesis on MS medium with different concentrations of cytokinins on multiple shoot regeneration/rooting from *in-vitro* explants of *R. webbianum*. Values are represented as mean ± SD (n=10), Data was analyzed by ANOVA using Duncan’s multiple range test (SPSS17.0); the values with different superscript along the columns are statistically significant at P<0.005, Data scored after 12 weeks of culture period.



**Figure 11:** Effect of Auxins on shoot regeneration from *in-vitro* explants.

The Figure 12 shows data taken from Table 7, in which maximum shoot regeneration was observed on 15 µM cytokinins i.e., BAP, TDZ and KN. The different concentrations of cytokinins were used for regeneration studies, out of which 15 µM BAP showed maximum shoot regeneration on different explants. The maximum shoot number was observed on 15 µM BAP using shoot tip explants i.e., 6 shoots per explants (Figure 4E).



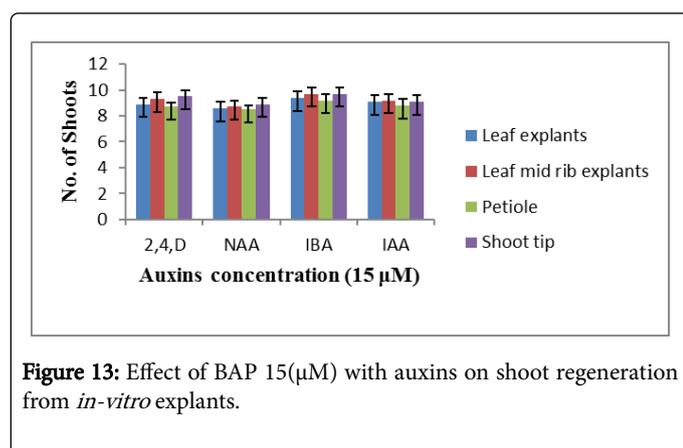
**Figure 12:** Effect of cytokinins on shoot regeneration from *in-vitro* explants.

Based on the data available in the Table 8, the following Figure 13 shows the effect of combination of phytohormones i.e., cytokinins (15 µM BAP) with 15 µM auxins, (2,4-D, NAA, IAA and IBA) on regeneration from different explants. However, different concentrations were used, but maximum shoot regeneration was observed on 15 µM. This was also observed that the combination of 15 µM BAP with 15 µM IBA showed best results i.e., 10 shoots per explants, in case of leaf midrib and shoot tip explants along with rooting (Figure 4F).

Sr. No.	Cytokinin	Auxins				Shoot No.	Shoot length (cm)	Leaf no.	Root No	Root length (cm)	Response	% Response
		BAP (µM)	2,4-D (µM)	NAA (µM)	IBA (µM)							
1.	Leaf											
	15	15	-			d8.9 ± 0.5	c3.7 ± 0.2	g10.3 ± 0.5	c3.7 ± 0.4	b2.9 ± 0.5	multiplication/ elongation/ rooting	70
	15		15			d8.6 ± 0.5	c3.3 ± 0.2	f9.4 ± 0.3	c3.9 ± 0.3	c3.2 ± 0.5	do	70
	15		-	15		e9.4 ± 0.5	c3.7 ± 0.3	g10.7 ± 0.5	e5.9 ± 0.5	d4.4 ± 0.3	do	80
	15		-		15	e9.1 ± 0.5	c3.3 ± 0.3	g10.2 ± 0.5	e5.5 ± 0.5	d4.1 ± 0.3	do	80
2.	Mid rib											
	15	15				f9.3 ± 0.5	c3.7 ± 0.2	g10.4 ± 0.5	c3.8 ± 0.4	c3.3 ± 0.5	do	70

	15		15		e8.7 ± 0.5	c3.5 ± 0.3	f9.5 ± 0.4	d4.3 ± 0.2	c3.5 ± 0.4	do	70
	15		15		f9.7 ± 0.5	c3.9 ± 0.3	g10.9 ± 0.5	f6.3 ± 0.5	d4.5 ± 0.3	do	80
	15		15		f9.2 ± 0.5	c3.5 ± 0.3	g10.6 ± 0.5	f6.1 ± 0.5	d4.2 ± 0.3	do	80
3.	Petiole										
	15	15			e8.7 ± 0.3	c3.4 ± 0.2	f9.9 ± 0.5	c3.5 ± 0.2	b2.7 ± 0.3	do	70
	15		15		e8.5 ± 0.3	c3.4 ± 0.2	f9.3 ± 0.5	c3.7 ± 0.2	b2.9 ± 0.3	do	70
	15		-	15	f9.2 ± 0.5	c3.8 ± 0.3	g10.3 ± 0.5	e5.8 ± 0.3	d4.3 ± 0.2	do	80
	15		-	15	e8.8 ± 0.5	c3.5 ± 0.3	f9.9 ± 0.5	e5.4 ± 0.3	d4.1 ± 0.2	do	80
4.	Shoot tip										
	15	15			f9.5 ± 0.5	c3.9 ± 0.2	g10.7 ± 0.5	c3.9 ± 0.5	c3.3 ± 0.2	do	70
	15		15		e8.9 ± 0.5	c3.6 ± 0.2	g10.5 ± 0.3	d4.2 ± 0.5	c3.4 ± 0.2	do	70
	15		-	15	f9.7 ± 0.5	c3.9 ± 0.3	g10.8 ± 0.5	f6.2 ± 0.5	d4.6 ± 0.3	do	80
	15		-	15	f9.1 ± 0.5	c3.4 ± 0.3	g10.3 ± 0.5	e5.8 ± 0.5	d4.3 ± 0.3	do	80

**Table 8:** Direct organogenesis on MS medium with 15 µM BAP and different concentrations of auxins on multiple shoot regeneration/rooting from *in-vitro* explants of *R. webbianum*. Values are represented as mean ± SD (n=10), Data was analyzed by ANOVA using Duncan's multiple range test (SPSS17.0); the values with different superscript along the columns are statistically significant at P<0.005, Data scored after 12 weeks of culture period.



**Figure 13:** Effect of BAP 15(µM) with auxins on shoot regeneration from *in-vitro* explants.

## Discussion

In present studies, the main aim was the micro-propagation of the *R. webbianum* as it is vulnerable and now endangered species. These have vanished from lower altitudes and now found only on high altitudes of Kashmir Himalaya, so the micropropagation is best way towards conservation of these plants. *R. webbianum* showed good growth response i.e., upto 70% as it was collected from very cold region

i.e., Ladakh, which is also known as cold desert and remains, covered with snow for 8 months in a year.

In order to prevent and to avoid the contamination to carry out tissue culture experiments, the explants surface sterilization was done using different sterilants. The seeds and field grown plants were sterilized with different concentrations of HgCl<sub>2</sub> and NaOCl and good results were achieved on HgCl<sub>2</sub>. In contrast to our results, Farzami and Ghorbanli [36] performed sterilization of seeds of *R. ribes* L. after immersing them in 0.3% of benomil solution for 1 h, and then transferred to ethanol for 1 minute and then to NaOCl (50%) for 10 minutes. In present study, it was observed that the seeds of *Rheum webbianum* showed 100% germination on MS medium when treated with different phytohormones, but according to Farzami and Ghorbanli [37] the seedling formation on MS basal medium in *R. ribes* was achieved after chilling for three days with GA<sub>3</sub>. No GA<sub>3</sub> was used in the present study. Combinations of different phytohormones were used on different explants. Field grown material showed only callus formation, especially in leaf. The seedlings were further used as explants. Different parts of seedlings were used i.e., leaf and petiole. The callus formed was soft, friable, regenerative and green. The callus was sub-cultured further to get regeneration. After sub culturing the callus on different phytohormones, it showed organogenesis. Both rooting and shooting was also observed. In order to get direct rooting,

these *in-vitro* shoots were also sub-cultured on different concentrations of phytohormones.

*R. webbianum* showed best shoot regeneration on concentration of 15  $\mu\text{M}$  BAP and 15  $\mu\text{M}$  2,4-D and also on combination of 15  $\mu\text{M}$  BAP with 15  $\mu\text{M}$  IBA on MS medium. These results are in line with that of Lal and Ahuja [38] who reported the *in vitro* propagation of Indian rhubarb (*R. emodi*) on MS medium supplemented with BAP and IBA. Our results were also supported by various workers like Roggemans and Claes [39], Roggemans and Boxus [40] (*R. rhaponticum*), Farzami and Ghorbanli [36] (*R. ribes*), Thomas and Sharon [41] (*R. rhubarbarum*, *R. rhaponticum*, *R. undulatum*), Malik *et al.* [42] (*R. emodi*) who also reported formation of multiple shoots on MS supplemented with BAP and IBA. However, Verma *et al.* [43] reported formation of multiple shoots in *R. emodi* on MS with BAP and IAA. On the other hand, Walkey [44] reported plantlet formation of *R. rhaponticum* using LS medium instead of MS medium.

Shoot multiplication of *R. webbianum* on MS medium from leaf was also observed when the medium was supplemented with different concentrations of BAP, Kinetin or TDZ. Similar results were obtained by Walkey and Mathews [45], Roggemans and Boxus [40] (*R. rhaponticum*) on MS medium when supplemented with Kinetin; but Lal and Ahuja [38] reported that Kinetin did not show any response when shoot tips of *R. emodi* were cultured on it. It seems that it is basically the endogenous level of hormones that play a vital role in exhibiting the regeneration potential of a species. In *R. webbianum*, it was noticed that BAP shows better results than Kinetin, TDZ and Zeatin, and it has also been reported by Murashige [46] and Street [47] which is attributed to group localized at N6 position of cytokinin [48]. Lal and Ahuja [38] in *R. emodi* has reported decrease in multiple shoot formation with increase in concentration of BAP. Chin [49] also reported that low concentration of TDZ are effective for micropropagation, which is in contrast to our results.

Number of shoots, size of shoots and leaf showed increase with increase in concentration of phytohormones. The maximum shoot length in *Rheum* species was observed on BAP 15  $\mu\text{M}$ +IAA 15  $\mu\text{M}$ ; BAP 15  $\mu\text{M}$ +IBA 15  $\mu\text{M}$ ; Kinetin, TDZ, 2,4-D and NAA 15  $\mu\text{M}$  combined with IAA and IBA showed maximum shoot formation and elongation. These results are similar to the reports of Hu and Wang, and with Wareing and Phillips. Shoot formation on BAP and Kinetin is again supported by Lal and Ahuja [38,50], Malik *et al.* [51] reported shoot regeneration from leaves of *R. emodi* using BAP in MS medium. Rooting on MS medium was achieved with different phytohormones which is strikingly different from those who report rooting on basal medium without using any phytohormones. Walkey and Mathews [45] in *R. rhaponticum*, Malik *et al.* [42] in *R. emodi*. In present studies, the maximum rooting was observed on IBA and IAA; and 2,4-D and NAA. These results are similar to those of Roggemans and Claes [39] and Roggemans and Boxus [40] who initiated roots in *R. rhaponticum* on MS+IBA, Lal and Ahuja [38,52] in *R. emodi* on MS+IBA; Ishimaru *et al.* [53] in *R. palmatum* on MS+NAA; Thomas and Sharon on MS+IAA in *R. rhubarbarum*, *R. rhaponticum*, *R. undulatum*, but the reports of Farzami and Ghorbanli [36] showed the formation of maximum roots on half strength MS+IBA in *R. ribes*. When BAP was used in high concentration i.e., 15  $\mu\text{M}$ , shooting as well as low rooting was recorded in *R. webbianum*. Similarly, when 2,4-D and NAA was used in high concentration low shooting and high rooting was observed. In present studies, IAA and IBA showed good response for rooting and BAP showed good response for shoot formation in *R. webbianum*, and this result is more or less similar to the report given by Parveen *et al.* [54].

The *in-vitro* raised plantlets were transferred first to plastic pots and later on to nursery where survival 50% for *R. webbianum* which is quite contrary to the results of Lal and Ahuja [38,52] who reported hardening of *in-vitro* raised plantlets of *R. emodi* with 89%-90% survival rate, in *R. emodi* [40].

## Conclusion

As we already know plants are very essential and important part of our daily lives. All living creatures are directly and indirectly dependent on plants. The plants are micro-biosynthetic factories for a variety of compounds which are their secondary metabolites. The medicinal properties are attributed to specific combination of bioactive phytochemicals. The medicinal plants are being used by human being since time immemorial and we get reference to these in ancient literature. As per WHO estimates, about 80% world population still relies on traditional system of medicine. Plant based drugs are preferred over synthetic ones because these are cheap, with less side effects and biocompatibilities with the human bodies. Due to recent resurgence in interest in plant-based drugs, their demand has increased manifold. Further, most of these plants are collected from the wild source in unplanned and unscientific way with destructive harvesting without following for sustainable supply. India is one of the largest suppliers of raw materials of medicinal plants with a foreign trade of over USD 1 billion per annum and has a great potential for cultivation of these medicinal plants.

*Rheum* with 60 species in the world, is represented by 7 species in high altitudinal area of North India. The different species are demarcated on the basis of various morphological characters of stem leaf and inflorescence. Due to high medicinal value, it is being over exploited and has been included in red list of threatened species of IUCN, thus necessitating the need for conservation and cultivation. The medicinal value of rhubarb is attributed to anthraquinones or their derivatives such as rhein, emodin, aloe-emodin, chrysaphanol, physcion and rutin.

*R. webbianum* are endangered species and they have vanished from lower altitudes. At present, they have shown their presence only on high altitudes of Kashmir Himalaya, so the various tissue culture techniques including micropropagation are best way towards conservation of this plant. Besides, efficient use these technologies can provide the germplasms required for ex situ conservation and cultivation of these species. In the present study, different media were used i.e., MS medium, LS medium and Nitsch & Nitsch medium. But the MS medium showed best results. Seed germination with complete seedling was observed on MS medium with different concentrations of phytohormones in which shooting and rooting both was observed. The best results for plant regeneration were obtained in BAP and IBA. The different concentrations of cytokinins and auxins were used. The best results were observed on BAP, 2,4-D, TDZ, Kinetin, IBA and IAA. Further, sub-culturing was done using small portions of leaf and petiole in different concentrations of phytohormones with MS medium. The best results were observed on BAP, 2,4-D. The green compact, friable, regenerative and soft callus was formed after sub culturing. The callus obtained from different explants was further sub cultured on different concentrations of phytohormones with MS medium. The plant formation was observed on different concentrations of BAP, 2,4-D, KN and NAA and TDZ. But the best result were found on high concentrations of phytohormones i.e., 15  $\mu\text{M}$ , especially BAP.

*R. webbianum* was treated with combination of phytohormones i.e., 15  $\mu$ M BAP with 15  $\mu$ M IBA, the maximum shoot regeneration was observed from leaf callus and petiole callus. For direct organogenesis, these explants were sub cultured again. The maximum direct shoot regeneration was found on 15  $\mu$ M 2,4-D from leaf and petiole. While using cytokinins, 15  $\mu$ M BAP showed maximum shoot regeneration on leaf. The best result was found on combination of phytohormones i.e., 15  $\mu$ M BAP with 15  $\mu$ M IBA. The response of this species was observed upto 70%. Hardening of *in-vitro* raised plants was done by gradual shifting from plastic pots containing autoclaved soil, sand and peat mixed with MS medium to the soil pots with covering of transparent polythene paper placed in the culture room for 15 days and then these pots transferred to net house and kept for 10 days. After 10 days all the plants were taken out from pots and were sown directly in soil of net house. The plants showed good growth response for conservation.

### Author's Contributions

All authors equally participated in designing experiments analysis and interpretation of data. All authors read and approved the final manuscript.

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