

Long-Term Efficacy of a New Medical Device Containing Fernblock® and DNA Repair Enzyme Complex in the Treatment and Prevention of Cancerization Field in Patients with Actinic Keratosis

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ABSTRACT

Introduction: Actinic keratosis (AKs) are part of the cancerization field, a region adjacent to AKs containing subclinical and histologically abnormal epidermal tissue due to Ultraviolet (UV)-induced DNA damage. The photoproducts, “as consequence of DNA damage induced by UV”, are mainly cyclobutane pyrimidine dimers (CPDs). Fernblock® demonstrated in previous studies significant reduction of the number of CPDs induced by UV radiation. Photolyases are a specific group of enzymes that remove the major UV-induced DNA lesions by a mechanism called photo-reactivation.

Methods: A monocentric, prospective, controlled, and double blind interventional study was performed to evaluate the effect of a new medical device (NMD) containing a DNA-repair enzyme complex (photolyases, endonucleases and glycosilases), a combination of UV-filters, and Fernblock® in the treatment of the cancerization field in 30 AK patients after photodynamic therapy (PDT). Patients were randomized into two groups: patients receiving a standard sunscreen (SS) and patients receiving the NMD. Clinical, dermoscopic, reflectance confocal microscopy (RCM) and histological evaluations were performed.

Results: An increase of AKs was noted in all groups after three months of PDT without significant differences between them ($p=0.476$). A significant increase in the number of AKs was observed in SS group after six ($p=0.026$) and twelve months of PDT ($p=0.038$); however, this increase did not reach statistical significance in the NMD group. Regarding RCM evaluation, honeycomb pattern assessment after twelve months of PDT showed significant differences in the extension and grade of the atypia in the NMD group compared to SS group ($p=0.030$ and $p=0.026$, respectively). Concerning histopathological evaluation, keratinocyte atypia grade improved from baseline to six months after PDT in all the groups, with no statistically significant differences between the groups. Twelve months after PDT, p53 expression was significantly lower in the NMD group compared to SS group ($p=0.028$). The product was well-tolerated, with no serious adverse events reported.

Conclusion: Our results provide evidence of the utility of this NMD in the improvement of the cancerization field and in the prevention of the development of new AKs.

Keywords: Actinic keratosis; Cancerization field; Repair enzyme; Photolyases; Treatment; Prevention; Reflectance confocal microscopy

INTRODUCTION

Actinic keratosis (AK) is a common skin tumor caused by

chronic exposure to ultraviolet (UV) radiation. Recent studies have demonstrated that AKs are part of the cancerization field, a region adjacent to AKs containing subclinical and histologically

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abnormal epidermal tissue [1,2]. It has been reported that AK progression to squamous cell carcinoma (SCC) may range from 0.025% to 16% [3]. There is increasing evidence that all types of AK lesions may progress to SCC, regardless of keratinocyte atypia thickness [4]. Thus, early recognition and treatment of the cancerization field is highly recommended in order to prevent the appearance of SCC.

Cancerization field pathogenesis include UV-induced DNA damage by the formation of a high number of cyclobutane pyrimidine dimers (CPDs) and pyrimidine (6-4) photoproducts (6-4 PPs), which are the main cause of non-melanoma skin cancer [5]. Fernblock® has demonstrated in several studies efficacy in the prevention and repair of DNA damage by reducing the number of CPDs induced by UV radiation [6-9]. Photolyases are specific photorepair enzymes that are present in algae and some non-placental mammals [10]. Their function is to recognize and remove the UV-induced DNA lesions by a mechanism called photo-reactivation [11]. A growing number of clinical and experimental studies have recently demonstrated the efficacy of photolyase-based novel approaches on cancerization field; however, most of these reports include a limited number of patients, a short follow-up period, and are not double-blind studies [12-26]. Additionally, other repair enzymes such as glycosylases and endonucleases have been described and they do not need light to be activated [27,28].

A new medical device (NMD) containing a complete DNA-repair enzyme complex including photolyases, glycosylases and endonucleases, a combination of filters with full spectrum sun protection, and an extract of *Polypodium leucotomos* leaves (Fernblock®) has been recently developed (Heliocare 360°AK). To our knowledge, this is the first MD containing DNA-repair enzymes and Fernblock® that has well-known antioxidant, photoprotective, and immune-modulatory activities [29-31]. The aim of this study was to demonstrate by clinical, dermoscopic, confocal microscopy and histological evaluation the improvement in the field of cancerization, and the prevention of the appearance of new AKs, with this NMD compared to a standard sunscreen (SS) in patients with AK after photodynamic therapy (PDT) treatment.

MATERIALS AND METHODS

Design and population

This is a monocentric, prospective, controlled, and double-blind interventional study to evaluate the effect of a NMD containing a DNA-repair enzyme complex, a combination of UV-filters, and Fernblock® (Heliocare 360° AK) in the treatment of the cancerization field in 30 AK patients. Duration of the study was 12 months, with a total of six visits. An area of 25 cm² on the scalp or face was selected for evaluation in all patients. One PDT session was performed at the baseline visit (TB) on the target area, and cryotherapy was performed during follow-up visits if residual and/or new AK lesions were observed. Topical application of the sunscreen started one week after PDT. Subjects were instructed to self-apply the product three times a day on the pre-specified area.

A total of 30 patients with AK lesions on the scalp or face were

enrolled and they were randomized into two groups: 15 patients receiving a standard sunscreen (SS) and 15 patients receiving the NMD. SS was exactly the same sunscreen as the NMD (SPF 100) but without the DNA-repair enzyme complex and Fernblock®. In addition, patients were also randomized into two other groups: patients receiving treatment for a total of 6 months and a subsequent follow-up visit a 12 months (10 SS and 10 NMD), and patients receiving treatment for a total of 12 months (5 SS and 5 NMD).

Written informed consent was obtained from all patients after having read and understood the information approved by the ethics committee. The study was approved by the institutional research board and was conducted according to the Declaration of Helsinki Principles. Eligibility criteria included AK patients older than 18 years of age, the presence of at least four AKs in an area of 25 cm² on the scalp or face, and absence of previous treatments in the target area during the previous three month period. Patients were excluded from the study for pregnancy, other skin diseases requiring systemic treatment, neoplastic lesions (melanoma or non-melanoma skin cancer) in the specified area, or sensitization to one or more components of the product.

Methods and evaluation

Clinical, dermoscopy and RCM evaluations were performed at TB before PDT and during all follow-up visits (one week post-PDT-T0, one month post-PDT-T1, three months post-PDT-T3, six months post-PDT-T6, one year post-PDT-T12). Additionally, Investigator Global Assessment (IGA), Patient Global Assessment (PGA) and Actinic Keratosis Quality of Life (AKQoL) were also evaluated. A biopsy was taken for histopathological assessment at TB and at the end of the treatment period (T6 and T12).

Clinical and dermoscopic evaluation

An area of 25 cm² on the scalp or face was selected for evaluation at basal visit, and subsequently revised in each follow-up using a plastic wrap. At baseline this area was subdivided in four quadrants and AKs of each of the quadrants were annotated. Number, size (mm) and grade (I/II/III) of the AK lesions were collected at all visits. Clinical and dermoscopic pictures were taken at each follow-up visit.

Reflectance confocal microscopy evaluation

Reflectance confocal microscopy (RCM) images were obtained using the VivaScope 1500® device (MAVIG GmbH, Munich, Germany). All RCM images were obtained and evaluated by the same investigator. A specific spot in the pre-specified area was marked in the plastic wrap in order to perform RCM assessments at the same site in all visits. RCM uses a low-power 830 nm laser beam that generates horizontal sections of the skin of 1.0 µm lateral resolution up to approximately 200 µm in depth. Instruments and acquisition procedures have been previously described [32].

At each spot, 50 images of 500 × 500 µm were acquired starting above the stratum corneum until papillar dermis (termed “vivastack”). Moreover, a minimum of 4 mosaics (termed as “vivablock”), with a maximum area of 8 × 8 mm, were obtained

per lesion, one in the stratum corneum level, one in the epidermis (stratum granulosum/spinosum), one at the dermoepidermal junction (DEJ), and one in papillary dermis. Videos at dermal level were acquired to evaluate enlarged vessels.

The same investigator without regard to patient's data retrospectively evaluated all recorded confocal images. All RCM evaluated parameters are specified in Table 1.

Histopathology evaluation

Two 3 mm-punch biopsies were performed in all patients at TB and T6 visits in two sites of cancerization field (without visible AK lesions).

Additionally, in the subgroup of patients with treatment until T12 (5 SS and 5 NMD) a third biopsy was done in another pre-specified site of the precancerous field. All specimens were processed using the standard histopathologic method of paraffin embedding sectioning and hematoxylin-eosin (HE) staining.

Evaluated histopathological characteristics are specified in Table 2. Additionally, histochemical and immunohistochemical (IHQ) studies for p53, Ki67 and orcein were conducted on all biopsies. Criteria used for evaluation of immunohistochemistry evaluation are specified in Table 3.

Statistical analysis

Quantitative variables were summarized by their mean and standard deviation (SD), and categorical variables by relative and absolute frequencies. The relationship between the categorical variables was evaluated using χ^2 test, and for quantitative variables

Kruskal-Wallis test was used. P values <0.05 were considered statistically significant. All computations were performed using the SPSSv24 statistical package (Chicago, IL).

RESULTS

Demographic features of the patients

A total of 30 patients were included in the study, and all but one concluded the complete study period. The baseline characteristics of the patients in both treatment groups did not differ significantly (Table 4). Twenty-two (73.4%) were males and 8 (26.6%) females. The age ranged from 59 to 91, with a mean age of 75 years (\pm 7.3). With regard to skin phototype, 70% (21/30) were phototype II, 26.7% (8/30) phototype III, and one patient (3.3%) phototype IV.

Clinical evaluation

At baseline evaluation (TB), the total number of AKs ranged from 4 to 13, without significant differences between groups (Figures 1 and 2). One week after PDT (T0) clinical clearance of 95% of all AKs was noted. From a dermoscopic point of view, an improvement in scaling, erythema, pigmentation and the presence of follicular plugs was observed in all the patients. A slight increase in AKs was noted in both groups at T3 without significant differences between them ($p=0.476$). Regarding T6, a significant increase in the number of AKs compared to T0 was observed in the SS group ($p=0.026$), but there was no statistically significant increase in the NMD group group. In addition, at T6 the number of AKs was lower compared to T3

Table 1: Parameters for RCM evaluation.

Location	Features	Scoring
Epidermis	Parakeratosis	0= absent; 1 \leq 10%; 2=10-25%; 3 \geq 25%
	Atypical honeycomb pattern (extension)	0= absent; 1 \leq 10%; 2=10-25%; 3 \geq 25%
	Atypical honeycomb pattern (grade)	0=absent; 1=slightly atypical; 2=moderate atypia; 3=severe atypia
	Mottled pigmentation	0= absent; 1 \leq 10%; 2=10-25%; 3 \geq 25%
	Inflammatory Infiltrate	0= absent; 1 \leq 10%; 2=10-25%; 3 \geq 25%
DEJ	Polycyclic papillary contours	0= absent; 1 \leq 10%; 2=10-25%; 3 \geq 25%
	Inflammatory Infiltrate	0= absent; 1 \leq 10%; 2=10-25%; 3 \geq 25%
Dermis	Round vessels	0= absent; 1 \leq 10%; 2=10-25%; 3 \geq 25%
	Solar elastosis	0=absent; 1=slight solar elastosis; 2=moderate-severe solar elastosis

Table 2: Evaluated histopathological characteristics.

Location	Features	Scoring
Epidermis	Epidermis thickness	Evaluated as a quantitative variable (mm)
	KIN	0= absent; 1=I (atypical basal and suprabasal cells); 2=II (two-thirds thickness); 3=III (all layers)
Dermis	Dermis thickness	Evaluated as a quantitative variable (mm)
	Inflammatory Infiltrate	0=absent; 1=focal; 2=diffuse
	Solar elastosis	0=absent; 1=slight solar elastosis; 2=moderate-severe solar elastosis

Table 3: Evaluated immunohistochemical studies.

IHQ	Scoring
Orcein	(0=absent; 1=focal; 2=diffuse)
Ki67	Evaluated in epidermis as a quantitative variable
p53	Evaluated in epidermis as a quantitative variable

Table 4: Baseline characteristics of the patients.

Variable	Total (n=30)
Epidemiological features	N# (%)
Age [Mean (\pm SD)]	75.3 (\pm 6.7)
Sex [N (%)]	
• Male	22 (73.4)
• Female	8 (26.6)
Family history of Cancer [(N%)]	
• Yes	5 (16.7)
• No	25 (83.3)
Personal history of Cancer [(N%)]	
• Yes	3 (10.0)
• No	27 (90.0)
Clinical features	N# (%)
Cutaneous phototype [(N%)]	
• I	0(0)
• II	21 (70)
• III	8 (26.7)
• IV	1 (3.3)
Actinic Keratosis Number [Mean (\pm SD)]	6.5 (\pm 1.7)
Actinic keratosis grade [Mean (\pm SD)]	1.6 (\pm 0.6)
Mean Actinic keratosis diameter [Mean (\pm SD)]	7.2 (\pm 3.4)



Figure 1: (A) Clinical evaluation at TB (B) Clinical evaluation at T3 (C) Clinical evaluation at T6 and (D) Clinical evaluation at T12 in a patient in the SS group.

in the NMD group but was higher in the SS group (Figures 1 and 2). A significant increase in AK number was identified in SS group at T12 compared to T0 ($p=0.038$). Conversely, in the NMD group the increase was not-significant ($p=0.083$). None of the evaluated patients presented any malignant neoplasm during the follow-up of the study. The NMD was well-tolerated, with just ocular itching reported in 3.3% (1/30) of the patients. No serious adverse events were reported.

Regarding the subjective clinical evaluation, PGA score (intense

improvement) was higher in the NMD group as compared to the SS one at T6 (80% in NMD group vs 64% in SS) and T12 (60% vs 33%). Concerning IGA, there were no significant differences at T3 and T6, but a higher score in the NMD group was observed at T12 (100% vs 83%). No differences were identified regarding AKQoL.

Confocal microscopy evaluation

At baseline (TB), no statistically significant differences were identified in any of the evaluated parameters between study

groups (Table 5). Atypia in the honeycomb pattern was mild in 40% (12/30), moderate in 53.3% (16/30), and severe in 6.7% (2/30) of patients (Figure 3). An improvement in

atypical honeycomb pattern was observed one month after PDT (T1) in both groups (p=0.003), with absence of atypia in 20% (6/30), mild atypia in 66.6% (20/30), moderate atypia

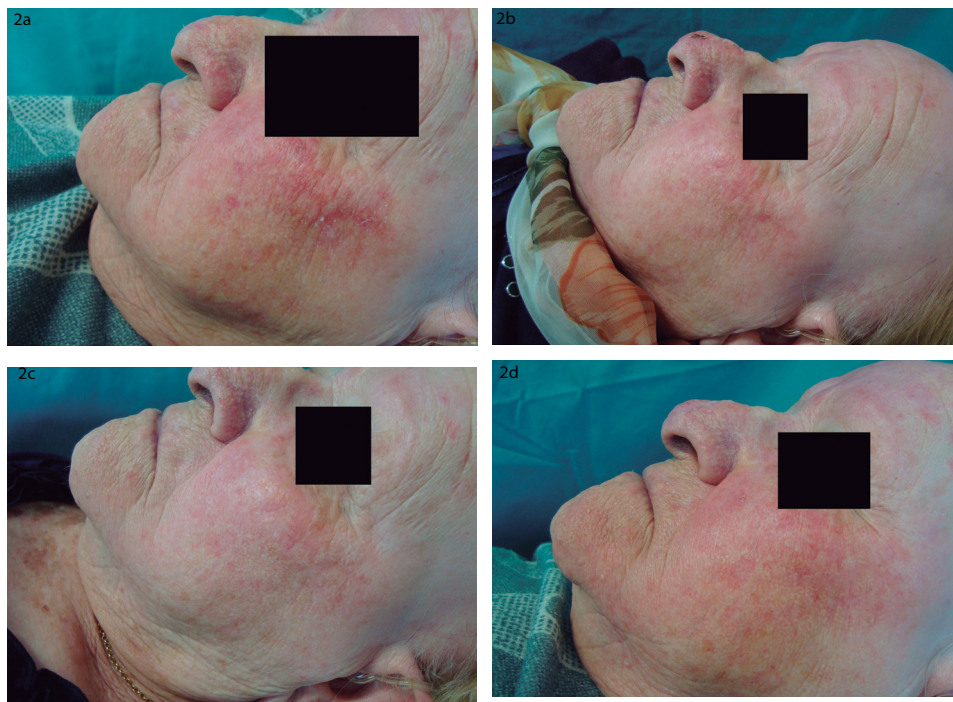


Figure 2: (A) Clinical evaluation at TB (B) Clinical evaluation at T3 (C) Clinical evaluation at T6 and (D) Clinical evaluation at T12 in a patient in the NMD group.

Table 5: Confocal microscopy evaluation.

Features	TB		p value	T1		p value	T3		p value	T6		p value	T12		p value
	SS	NMD		SS	NMD		SS	NMD		SS	NMD		SS	NMD	
Epidermis															
Parakeratosis															
None	5 (55.6)	4 (44.4)	0.174	7 (58.3)	5 (41.7)	0.355	3 (60)	2 (40)	0.674	4 (66.7)	2 (33.3)	0.403	1 (20)	2 (40)	0.49
<10%	4 (44.4)	5 (55.6)		0 (0)	0 (0)		0 (0)	0 (0)		1 (100)	0 (0)		0 (0)		
10-25%	0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)		
>25%	0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)		
Atypical HP (extension)															
None	0 (0)	0 (0)	0.097	4 (66.7)	2 (33.3)	0.512	4 (50)	4 (50)	0.589	3 (50)	3 (50)	0.991	0 (0)	4 (80)	0.03
<10%	5 (33.3)	10 (66.7)		8 (42.1)	11 (57.9)		6 (42.9)	8 (57.1)		9 (47.4)	10 (52.6)		3 (60)	1 (20)	
10-25%	9 (64.3)	5 (35.7)		3 (60)	2 (20)		4 (66.7)	2 (33.3)		2 (50)	2 (50)		2 (40)	0 (0)	
>25%	0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)	
Atypical HP (grade)															
None	0 (0)	0 (0)	0.311	4 (66.7)	2 (33.3)	0.091	4 (50)	4 (50)	0.809	3 (50)	3 (50)	0.472	0 (0)	4 (80)	0.026
Mild atypia	4 (33.3)	8 (66.7)		7 (35.0)	13 (65.0)		8 (50)	8 (50)		9 (56.3)	7 (43.8)		2 (40)	1 (20)	
Moderate atypia	10 (62.5)	6 (37.5)		3 (100)	0 (0)		2 (50)	2 (50)		2 (28.6)	5 (71.4)		3 (60)	0 (0)	
Severe atypia	1 (50)	1 (50)		1 (100)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)	
Mottled pigmentation															
None	0 (0)	2 (100)	0.339	1 (25)	3 (75)	0.041	0 (0)	2 (100)	0.014	0 (0)	1 (100)	0.034	0 (0)	0 (0)	0.117
<10%	6 (46.2)	7 (53.8)		7 (38.9)	11 (61.1)		4 (25)	9 (75)		2 (18.2)	9 (81.8)		2 (40)	5 (100)	
10-25%	8 (57.1)	6 (42.9)		7 (87.5)	1 (12.5)		11 (78.6)	3 (21.4)		11 (73.3)	4 (26.7)		2 (40)	0 (0)	
>25%	1 (100)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		1 (50)	1 (50)		1 (20)	0 (0)	

Inflammatory infiltrate															
None	9 (50)	9 (50)		9 (47.4)	10 (52.6)		14 (56)	11 (44)		12 (50)	12 (50)		5 (100)	5 (100)	
<10%	3 (33.3)	6 (67.6)	0.135	2 (50)	2 (50)	0.917	0 (0)	2 (100)	0.228	2 (40)	3 (60)	0.535	0 (0)	0 (0)	NS
10-25%	3 (100)	0 (0)		4 (57.1)	3 (42.9)		0 (0)	1 (100)		0 (0)	0 (0)		0 (0)	0 (0)	
>25%	0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)	
DE junction															
Polycyclic papillary contours															
None	7 (43.8)	9 (56.3)		8 (42.1)	11 (57.9)		6 (40)	9 (60)		7 (46.7)	8 (53.3)		1 (20)	5 (100)	
<10%	7 (58.3)	5 (41.7)	0.747	7 (70.0)	3 (30.0)	0.215	7 (70)	3 (30)	0.187	3 (50)	3 (50)	0.759	4 (80)	0 (0)	0.024
10-25%	1 (50)	1 (50)		0 (0)	1 (100)		1 (100)	0 (0)		4 (57.1)	3 (42.9)		0 (0)	0 (0)	
>25%	0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	2 (100)		0 (0)	0 (100)		0 (0)	0 (0)	
Dermis															
Inflammatory Infiltrate															
None	6 (42.9)	8 (57.1)		7 (53.8)	6 (46.2)		7 (43.8)	9 (56.3)		12 (50)	12 (50)		2 (40)	5 (100)	
<10%	8 (61.5)	5 (38.5)	0.519	6 (42.9)	8 (57.1)	0.706	6 (54.5)	5 (45.5)	0.511	1 (33.3)	2 (66.7)	0.291	3 (60)	0 (0)	0.083
10-25%	1 (33.3)	2 (67.7)		2 (66.7)	1 (33.3)		1 (100)	0 (0)		1 (50)	1 (50)		0 (0)	0 (0)	
>25%	0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)	
Round vessels															
None	12 (52.2)	10 (47.8)		12 (52.2)	11 (47.8)		12 (50)	12 (50)		12 (80)	12 (80)		5 (100)	4 (80)	
<10%	2 (50)	2 (50)	0.385	0 (0)	3 (100)	0.224	1 (33.3)	2 (66.7)	0.512	1 (6.7)	2 (13.3)	0.861	0 (0)	1 (20)	0.292
10-25%	0 (0)	2 (100)		2 (66.7)	1 (33.3)		1 (100)	0 (0)		1 (6.7)	1 (6.7)		0 (0)	0 (0)	
>25%	1 (6.7)	0 (0)		1 (100)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)		0 (0)	0 (0)	

in 10% (3/30), and severe atypia in 3.3% (1/30) of patients. Although no statistically significant differences between the groups were detected at T1, we observed a trend ($p=0.076$) with a predominance of mild or absent atypia in the NMD group compared to the SS group, in which more cases of moderate atypia and one case of severe atypia were identified. No statistically significant differences in the honeycomb pattern were identified at T3 and T6 visits. Conversely, honeycomb pattern assessment at T12 showed significant differences in the extension and grade of the atypia in the NMD group compared to the SS group ($p=0.030$ and $p=0.026$, respectively) (Figure 4). Honeycomb pattern score was also significantly higher in the SS group compared to the NMD one at T12 ($p=0.01$). Furthermore, a significant reduction in the honeycomb pattern score at T12 compared to T0 was seen in the NMD group ($p=0.034$), but not in the SS group ($p=0.276$).

In addition, at T12 we evaluated the atypical honeycomb pattern in SS and NMD patients who received treatment for 12 months as compared to those who received it for 6 months only. Both extension and grade of atypia were significantly higher in those in the NMD group that received just 6 months of treatment compared to 12 months treatment ($p=0.004$ and $p=0.004$, respectively), but these differences were not observed in the SS group ($p=0.651$ and $p=0.592$, respectively).

With respect to other parameters, we observed an increase in the epidermal and dermal inflammatory infiltrate at T0 due to the PDT treatment which declined progressively at T3 and T6 to even lower levels at TB. Moreover, at T12 there was a trend regarding dermal inflammatory infiltrate that was absent in the

NMD group compared to the SS group ($p=0.083$). Additionally, a higher prevalence of polycyclic papillary contours was detected at T12 in the SS group compared to the NMD group ($p=0.024$). No statistically significant differences were identified in the assessment of the stratum corneum and the presence of round blood vessels.

Histopathology evaluation

At baseline, hyperkeratosis with parakeratosis, pleomorphism of keratinocyte nuclei, mild to moderate inflammatory infiltrate and solar elastosis were observed in all patients. Of all the cohort, 77.5% (31/40) showed KIN I, 22.5% (9/40) KIN II, and none showed KIN III. KIN grade did not show any statistically significant differences between the groups at TB ($p=0.413$) or T6 ($p=0.831$). Although not significant, keratinocyte atypia grade improved from the TB visit to T6 in both groups (Figure 5). Regarding T12, no significant differences between the KIN grade of SS and the NMD group ($p=0.738$) were noted. Epidermal and dermal thickness evaluation did not show any statistically significant differences.

Concerning histochemistry and immunohistochemistry, no statistically significant differences were identified in orcein and Ki67 proliferation index at TB, T6 and T12, however an improvement was noted in Ki67 proliferation index in both groups. Although there was a tendency towards decreased expression of P53 with respect to TB, no significant differences were observed at T6 ($p=0.611$). However, p53 expression was significantly lower in the NMD group as compared to SS ($p=0.028$) at T12 (Figure 5).

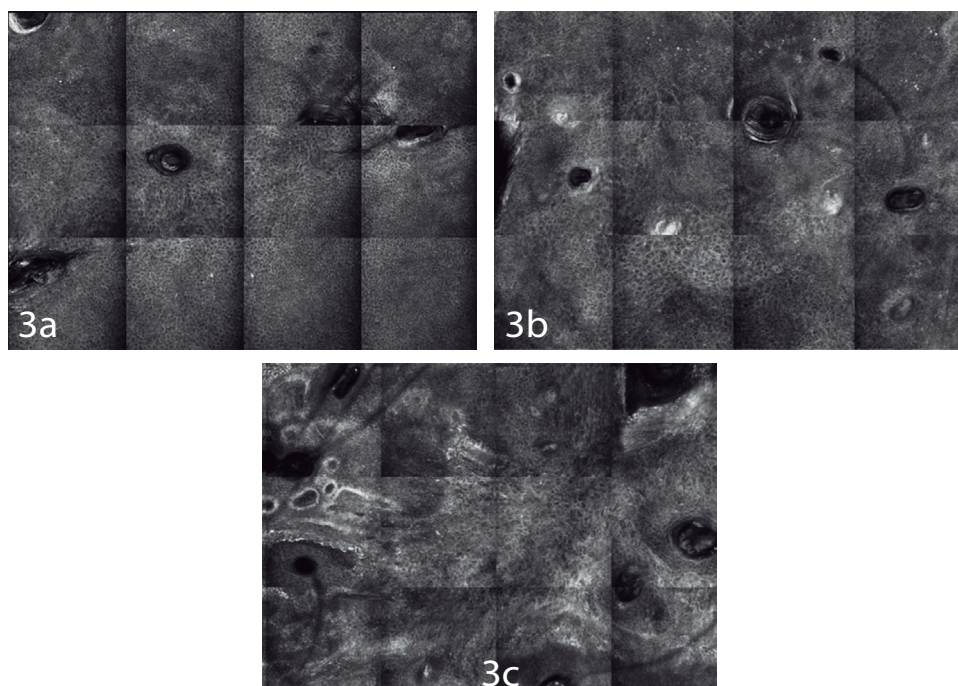


Figure 3: (A) Mild atypia in the honeycomb pattern at TB, (B) Moderate atypia in the honeycomb pattern at TB, (C) Severe atypia in the honeycomb pattern at TB.

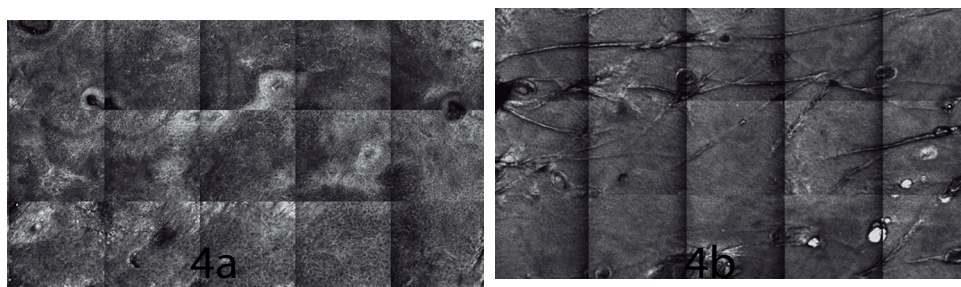


Figure 4: (A) Moderate atypia in the honeycomb pattern at T12 in a patient in the SS group. (B) Absence of atypia in the honeycomb pattern at T12 in a patient in the NMD group.

DISCUSSION

AK lesions are premalignant skin conditions that represent one of the most frequent dermatological consultations. Several field-directed treatments are available with high clearance rates [33], although relapse in patients with chronic exposure to UV radiation are very frequent.

In recent years, several studies have demonstrated the utility of photolyase-based sunscreens in the treatment of the cancerization field [12-26]. Consistent with these studies, we have demonstrated the efficacy of a NMD with a complete DNA-repair enzyme complex and Fernblock® in the prevention of the appearance of new AK lesions. Results from our cohort demonstrate a lower number of AK lesions in patients treated in the NMD group as compared to SS after six and twelve months. In this regard, Eibenschutz et al. also performed a randomized and parallel-group study with a photolyase-based device and demonstrated a significant reduction of new AK lesions, however their trial was not double-blind [34]. In addition, Moscarella et al. recently conducted a randomized, double-blind and parallel-group study, and described a significantly lower percentage of patients who presented new lesions, but just taking into account in the analysis the subgroup of patients with <10 AK lesions

[26]. Of note, during the twelve-month treatment period, none of the patients in our cohort developed any skin neoplasms in the study area.

In addition to clinical and dermoscopic evaluation, we performed RCM assessment as a novel noninvasive technique that provides *in vivo* evaluation of the epidermis and superficial dermis at a cellular resolution in real time. RCM enables repeated imaging of the same lesion over time, which can be used for monitoring efficacy of non-invasive treatments. It has demonstrated to be helpful not only in identifying and grading AK lesions, but also in monitoring AK treatment outcomes [35-38]. In this regard, several studies have been performed with RCM to evaluate the efficacy of field-directed treatments such as 5-fluorouracil [39], ingenol mebutate [40], PDT [41], diclofenac [42], imiquimod [43], and surgery [44]. However, just two studies have used RCM to evaluate improvement in cancerization field by other MD with DNA-repair enzymes [19,26]. Puig et al. [18] demonstrated a decreased in stratum corneum alterations, and an improvement in atypical honeycomb pattern four weeks after topical application of a photolyase-based sunscreen, however they studied a small number of patients (n=13), and they did not compare with a SS group.

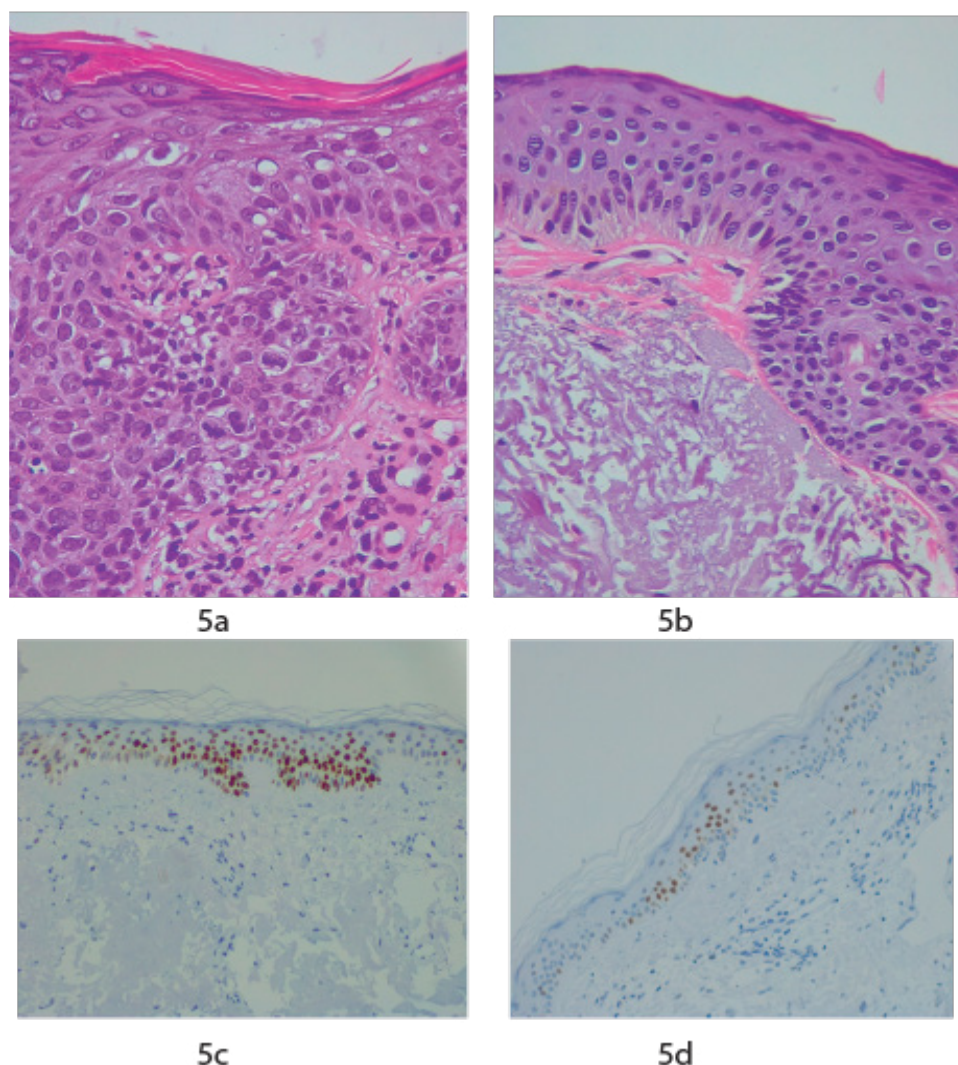


Figure 5: (A) Severe keratinocyte atypia occupying the lower two-thirds of the epidermis (KIN II) at baseline visit, (B) Absence of keratinocyte atypia at T6, (C) Intense p53 expression at TB in a patient in the NMD group, (D) Decrease in p53 expression at T12 in the same patient.

Consistent with our study, Moscarella et al. described similar results regarding RCM parameters in both groups at one, three and six months of follow-up [26]. Similarly, in our cohort we did not identify significant differences at T1, T3, and T6, however honeycomb pattern assessment at T12 showed significant differences in the extension and grade of atypia, as well as in honeycomb pattern score in the NMD group with respect to the SS group. Thus, our results provide evidence for the benefit to honeycomb pattern after long-term use of our NMD. Strikingly, we also identified by RCM a decrease in the inflammatory infiltrate after treatment, with absence of dermal inflammatory infiltrate in the NMD group, and persistence in the SS group. To our knowledge, none of these previous studies have detected an improvement in this parameter that also represents an improvement in the cancerization field.

Concerning histopathological evaluation, we demonstrated an improvement in keratinocyte atypia grade after treatment, although no differences were detected between groups. In addition, our results provide further support for the utility of DNA-repair enzymes in p53 expression improvement. There is strong evidence for the role of p53 in UV-induced skin carcinogenesis. P53 is a tumor suppressor gene that regulates DNA replication in response to DNA damage from UV

exposure. Several reports have demonstrated that there is a significant increase in p53 expression levels in AKs and SCCs [45]. Furthermore, reduction in p53 expression levels has been reported after photolyase-based approaches [46]. In our study, a tendency to reduced p53 expression was noted at T6 and T12 as compared to TB visit. In addition, p53 expression was significantly lower in the NMD group as compared to SS at T12. In this regard, Puig et al. [18] did not find any significant changes in p53 expression after one-month treatment, but they suggested the necessity of long-term follow-up studies. As previously reported, a high level of expression of Ki67 was identified in our cohort at baseline, however, our study failed to demonstrate significant improvement in this proliferation index after treatment with the NMD.

CONCLUSION

In conclusion, our results show the utility of the NMD in the improvement of the cancerization field and in the prevention of the development of new AKs. Early treatment of the cancerization field with the present NMD (Heliocare 360° AK), could be useful to prevent the development of AK lesions and SCC. Long-term studies with a larger number of patients are needed to confirm the efficacy of these findings.

REFERENCES

1. Slaughter DP, Southwick HW, Smejkal W. Field cancerization in oral stratified squamous epithelium. *Cancer*. 1953;6:963-968.
2. Stern RS, Bolshakov S, Nataraj AJ, Ananthaswamy HN. P53 mutation in nonmelanoma skin cancers occurring in psoralen ultraviolet A-treated patients: evidence for heterogeneity and field cancerization. *J Invest Dermatol*. 2002;119:522-526.
3. Glogau RG. The risk of progression to invasive disease. *J Am Acad Dermatol*. 2000;42:23-24.
4. Fernández-Figueras MT, Carrato C, Sáenz X, Puig L, Musulen E, Ferrándiz C, et al. Actinic keratosis with atypical basal cells (AK I) is the most common lesion associated with invasive squamous cell carcinoma of the skin. *J Eur Acad Dermatol Venereol*. 2015;29:991-997.
5. Jans J, Schul W, Sert YG, Rijkse Y, Rebel H, Eker AP, et al. Powerful skin cancer protection by a CPD-photolyase transgene. *Curr Biol*. 2005;15: 105-115.
6. Philips N, Dulaj L, Upadhyay T. Cancer cell growth and extracellular matrix remodeling mechanism of ascorbate; beneficial modulation by P. leucotomos. *Anticancer Res*. 2009;29:3233-3238.
7. Zattra E, Coleman C, Arad S, Helms E, Levine D, Bord E, et al. Polypodium leucotomos extract decreases UV-induced Cox-2 expression and inflammation, enhances DNA repair, and decreases mutagenesis in hairless mice. *Am J Pathol*. 2009;175:1952-1961.
8. Auriemma M, Di Nicola M, Gonzalez S, Piaserico S, Capo A, Amerio P. Polypodium leucotomos supplementation in the treatment of scalp actinic keratosis: could it improve the efficacy of photodynamic therapy? *Dermatol Surg*. 2015;41: 898-902.
9. Kohli I, Shafi R, Isedeh P, Griffith JL, Al-Jamal MS, Silpa-Archa N, et al. The impact of oral Polypodium leucotomos extract on ultraviolet B response: A human clinical study. *J Am Acad Dermatol*. 2017;77: 33-41.
10. Rastogi RP, Richa, Kumar A, Tyagi MB, Sinha RP. Molecular mechanisms of ultraviolet radiation-induced DNA damage and repair. *J Nucleic Acids*. 2010;2010:592980.
11. Garinis GA, Jans J, van der Horst GT. Photolyases: capturing the light to battle skin cancer. *Future Oncol*. 2006;2:191-199.
12. DeBoyes T, Kouba D, Ozog D, Fincher E, Moy L, Iwata K. Reduced number of actinic keratoses with topical application of DNA repair enzyme creams. *J Drugs Dermatol*. 2010;1519-1521.
13. Berardesca E, Bertona M, Altabas K, Altabas V, Emanuele E. Reduced ultraviolet-induced DNA damage and apoptosis in human skin with topical application of a photolyase-containing DNA repair enzyme cream: clues to skin cancer prevention. *Mol Med Rep*. 2012;5:570-574.
14. Emanuele E, Altabas V, Altabas K, Berardesca E. Topical application of preparations containing DNA repair enzymes prevents ultraviolet-induced telomere shortening and c-FOS proto-oncogene hyperexpression in human skin: an experimental pilot study. *J Drugs Dermatol*. 2013; 12:1017-1021.
15. Piaserico S, Milani M. Efficacia clinica della fotoliasi topica dopo terapia fotodinamica in soggetti con cheratosi attiniche: studio prospettico randomizzato intrapaziente. *G Ital Dermatol Venereol*. 2012;147:109.
16. Puig-Butillé JA, Malveyh J, Potrony M, Trullas C, Garcia-García F, Dopazo J, et al. Role of CPL-17 in restoring skin homeostasis in cutaneous field of cancerization: effects of topical application of a film-forming medical device containing photolyase and UV filters. *Exp Dermatol*. 2013;22:494-496.
17. Puviani M, Barcella A, Milani M. Efficacy of a photolyase-based device in the treatment of cancerization field in patients with actinic keratosis and non-melanoma skin cancer. *G Ital Dermatol Venereol*. 2013;148:693-698.
18. Puig S, Puig-Butillé JA, Díaz MA, Trullas C, Malveyh J. Field Cancerization Improvement with Topical Application of a Film-Forming Medical Device Containing Photolyase and UV Filters in Patients with Actinic Keratosis, a Pilot Study. *J Clin Exp Dermatol Res*. 2014;5:220.
19. Giustini S, Miraglia E, Berardesca E, Milani M, Calvieri S. Preventive long-term effects of a topical film-forming medical device with ultra-high UV protection filters and DNA repair enzyme in xeroderma pigmentosum: a retrospective study of eight cases. *Case Rep Dermatol*. 2014;6:222-226.
20. Rstom SA, Martinez B, Abdalla Z, Rezza GG, Paschoal FM. Avaliação da ação de creme contendo fotoliasi em lipossomas e filtro solar FPS100 na queratose actínica da face: estudo clínico, dermatoscópico e por microscopia confocal. *Surg Cosmet Dermatol*. 2014;6:226-231.
21. Laino L, Elia F, Desiderio F, Scarabello A, Sperduti I, et al. The efficacy of a photolyase-based device on the cancerization field: a clinical and thermographic study. *J Exp Clin Cancer Res*. 2015;34:84.
22. Carducci M, Pavone PS, De Marco G, Lovati S, Altabas V, Altabas K, et al. Comparative Effects of Sunscreens Alone vs Sunscreens Plus DNA Repair Enzymes in patients with Actinic Keratosis: Clinical and Molecular Findings from a 6-Month, Randomized, Clinical Study. *J Drugs Dermatol*. 2015;14:986-990.
23. Eibenschutz L, Silipo V, De Simone P, Buccini PL, Ferrari A, Carbone A, et al. A 9-month, randomized, assessor-blinded, parallel-group study to evaluate clinical effects of film-forming medical devices containing photolyase and sun filters in the treatment of field cancerization compared with sunscreen in patients after successful photodynamic therapy for actinic keratosis. *Br J Dermatol*. 2016;175:1391-1393.
24. Navarrete-Dechent C, Molgó M. The use of a sunscreen containing DNA-photolyase in the treatment of patients with field cancerization and multiple actinic keratosis: a case series. *Dermatol Online J*. 2017;23:1-3.
25. Moscarella E, Argenziano G, Longo C, Aladren S. Management of cancerization field with a medical device containing photolyase: a randomized, double-blind, parallel-group pilot study. *J Eur Acad Dermatol Venereol*. 2017;31:386-427.
26. Ba X, Aguilera-Aguirre L, Rashid QT, Bacsí A, Radak Z, Sur S, et al. (2014). The role of 8-oxoguanine DNA glycosylase-1 in inflammation. *Int J Mol Sci* 15:16975-16997.
27. Yarosh D, Klein J, O'Connor A, Hawk J, Rafal E, Wolf P, et al. (2001). Effect of topically applied T4 endonuclease V in liposomes on skin cancer in xeroderma pigmentosum: a randomised study. *Xeroderma Pigmentosum Study Group. Lancet*. 2001;357:926-929.
28. Middelkamp-Hup MA, Pathak MA, Parrado C, Goukassian D, Rius-Díaz F, Mihm MC, et al. Oral Polypodium leucotomos extract decreases ultraviolet-induced damage of human skin. *J Am Acad Dermatol*. 2004;51:910-918.
29. Middelkamp-Hup MA, Pathak MA, Parrado C, Garcia-Caballero T, Rius-Díaz F, Fitzpatrick TB, et al. Orally administered Polypodium leucotomos extract decreases psoralen-UVA-induced phototoxicity, pigmentation, and damage of human skin. *J Am Acad Dermatol*. 2004;50:41-49.
30. Torricelli P, Fini M, Fanti PA, Dika E, Milani M, et al. Protective effects of Polypodium leucotomos extract against UVB-induced damage in a model of reconstructed human epidermis. *Photodermatol Photoimmunol Photomed*. 2017; 33:156-163.

31. Rajadhyaksha M, Grossman M, Esterowitz D, Webb RH, Anderson RR. In vivo confocal scanning laser microscopy of human skin: melanin provides strong contrast. *J Invest Dermatol.* 1995;104:946-952.
32. Gupta AK, Paquet M. Network meta-analysis of the outcome 'participant complete clearance' in nonimmunosuppressed participants of eight interventions for actinic keratosis: a follow-up on a Cochrane review. *Br J Dermatol.* 2013;169:250-259.
33. Ahlgrim-Siess V, Laimer M, Rabinovitz HS, Oliviero M, Wellenhof RH, et al. Confocal Microscopy in Skin Cancer. *Curr Dermatol Rep.* 2018;7:105-118.
34. Zalaudek I, Piana S, Moscarella E, Longo C, Zendri E, Castagnetti F, et al. Morphologic grading and treatment of facial actinic keratosis. *Clin Dermatol.* 2014;32:80-87.
35. Fernandez Figueras MT. From actinic keratosis to squamous cell carcinoma: pathophysiology revisited. *J Eur Acad Dermatol Venereol.* 2017;31:5-7.
36. Rishpon A, Kim N, Scope A, Porges L, Oliviero MC, Braun RP, et al. Reflectance confocal microscopy criteria for squamous cell carcinomas and actinic keratoses. *Arch Dermatol.* 2009;145:766-772.
37. Pellacani G, Longo C. Reflectance confocal microscopy: a crucial role for actinic keratosis treatment monitoring. *J Eur Acad Dermatol Venereol.* 2018;32:1055.
38. Ulrich M, Reinhold U, Falqués M, Rodriguez Azeredo R, Stockfleth E. Use of reflectance confocal microscopy to evaluate 5-fluorouracil 0.5%/salicylic acid 10% in the field-directed treatment of subclinical lesions of actinic keratosis: subanalysis of a Phase III, randomized, double-blind, vehicle-controlled trial. *J Eur Acad Dermatol Venereol.* 2018;32:390-396.
39. López Estebanz JL, Pampin Franco A, Gamo Villegas R, Floristán U. Monitoring Ingenol Mebutate Gel Treatment of Actinic Keratoses by Reflectance Confocal Microscopy. *Acta Derm Venereol.* 2017;97:646-648.
40. Seyed Jafari SM, Timchik T, Hunger RE. In vivo confocal microscopy efficacy assessment of daylight photodynamic therapy in actinic keratosis patients. *Br J Dermatol.* 2016;175:375-381.
41. Malvey J, Roldán-Marín R, Iglesias-García P, et al. Monitoring treatment of field cancerisation with 3% diclofenac sodium 2.5% hyaluronic acid by reflectance confocal microscopy: a histologic correlation. *Acta Derm Venereol.* 2015;95:45-50.
42. Ulrich M, Krueger-Corcoran D, Roewert-Huber J, et al. (2010). Reflectance confocal microscopy for noninvasive monitoring of therapy and detection of subclinical actinic keratoses. *Dermatology* 220:15-24. [PMID: 19907131]
43. Richtig E, Ahlgrim-Siess V, Koller S, Gerger A, Horn M, Smolle J. Follow-up of actinic keratoses after shave biopsy by in-vivo reflectance confocal microscopy-a pilot study. *J Eur Acad Dermatol Venereol.* 2010; 24:293-298.
44. Einspahr JG, Xu MJ, Warneke J, Saboda K, Ranger-Moore J, Bozzo P, et al. Reproducibility and expression of skin biomarkers in sun-damaged skin and actinic keratoses. *Cancer Epidemiol Biomarkers Prev.* 2006;15:1841-1848.
45. Vidal-Asensi S, Gutiérrez-Ortega C, Domingo Deagustín-Vázquez D, Ughelli-Yampey G. Photolyase sunscreen decreases expression of p53 and Ki67 in comparison to standard 50 SPF. *J Am Acad Dermatol.* 2012; 66:AB156.
46. Da Silva TA, Coelho G, Lorenzetti Bocca A, Figueiredo Cavalcante Neto F. Expression of apoptotic, cell proliferation regulatory, and structural proteins in actinic keratosis and their association with dermal elastosis. *J Cutan Pathol.* 2007;34:315-323.