

The Use of Pharmaceutical Intervention as a Mechanistic Tool to Regulate Bioenergetics and Inhibit Free Radical Oxidative Stress During the Progression of Alzheimer's Disease

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Abstract

Oxidative stress is the imbalance between antioxidants and oxidants. An increase in the level of free radicals or an antioxidant deficiency can perpetuate this phenomenon. Free radical mediated oxidative stress is demonstrated in various neurodegenerative disorders such as Alzheimer's disease, Parkinson's disease, Huntington's disease, and amyotrophic lateral sclerosis. This oxidative damage can result in a mechanistic chain reaction in which free radicals can bind to alkenes and cause overall protein damage and a decrease in energy metabolism. However, this mechanism can be halted with the use of antioxidant therapy. During the progression of Alzheimer's disease, oxidative stress is continuously elevated. Current research shows several antioxidant strategies being implemented to stop oxidative stress in Alzheimer's disease such as α -lipoic acid and acetyl L-carnitine. In addition to antioxidant therapies, antiaggregants, acetylcholinesterase inhibitors, β -secretase and γ -secretase inhibitors are also being used as pharmaceutical drugs to combat the pathological hallmarks of Alzheimer's disease. This review will describe the current pharmaceutical interventions being used to regulate energy metabolism and inhibit free radical mediated oxidative stress during the progression of Alzheimer's disease.

Keywords: Alzheimer's disease; Antioxidants; Free radicals; Oxidative stress; Pharmaceutical therapeutics

Introduction

Under normal physiological conditions, there is equilibrium between antioxidants and prooxidants. When environmental factors, stressors, or disease occur, this homeostasis can become imbalanced in favor of prooxidants, resulting in a phenomenon known as oxidative stress [1]. Oxidative stress can also occur if there is an antioxidant deficiency or excess reactive oxygen/nitrogen species production [2]. The mitochondria are the key source for free radicals [3,4]. Reactive oxygen species (ROS) are molecules that contain oxygen with higher reactivity than ground state O_2 . Some examples are hydroxyl radical ($OH\cdot$), superoxide anion ($O_2^{\cdot-}$), hydrogen peroxide (H_2O_2), peroxy radical ($\cdot OOH$), hypochlorous acid ($HOCl$), and many others [1]. There are three mechanisms by which cells are protected from ROS: i) scavenging ROS and precursors; ii) binding catalytic metal ions needed for ROS formation; and iii) generating and upregulating endogenous antioxidant defenses. Oxidatively damaged proteins are often removed by the 20S proteasome. However, defects in the proteasome system can lead to elevated levels of oxidatively modified proteins. Reactive oxygen species levels increase as a function of age and are even higher in age-related neurodegenerative disorders [5]. It has been well established that oxidative stress is elevated in Alzheimer's disease [6,7], Parkinson's disease [8], amyotrophic lateral sclerosis [2], Huntington's disease [9] and other neurodegenerative disorders [6,10]. Alzheimer's disease (AD) is characterized by progressive neurodegeneration that results from a variety of possible environmental, genetic, and age-related factors [11,12]. The pathological hallmarks of the disease are extracellular senile plaques (deposits of amyloid β peptide) and intracellular neurofibrillary tangles (NFTs), but oxidative stress and metabolic abnormalities can precede these hallmarks as well the dementia that results from abnormal brain function [13,14]. While several biomarkers exist that may indicate the presence of the disease, conclusive evidence can only be established post-mortem [15]. Autopsy reveals the extent

at which neurodegeneration has occurred and is commonly described using Braak staging methods. Braak determined that Alzheimer's disease progresses from stages I to VI with the increasing presence and distribution of NFTs and neuropil threads (NTs) [16]. This post-mortem staging is used to help differentiate between the four stages of AD: preclinical AD (PCAD), mild cognitive impairment (MCI), early AD (EAD), and late stage AD (LAD). This article will discuss specifically the use of current pharmaceutical interventions as they relate to the mechanistic recovery of energy related metabolic proteins during the progression of Alzheimer's disease.

Energy Metabolism

In addition to the hallmarks of Alzheimer's disease, senile plaques composed of amyloid beta peptide and neurofibrillary tangles composed of hyperphosphorylated tau protein, this disease is also characterized by oxidative stress and metabolic dysfunction that can precede both disease hallmarks [13,14]. The intricate relationship between these two characteristics, oxidative stress and metabolism dysfunction, has historically revealed itself in a very cyclic manner. A substantial portion of oxidative stress is generated by the mitochondria during regular metabolism, and this oxidative stress also serves as a disruptor of this same metabolism as excess reactive oxygen species (ROS) and reactive nitrogen species (RNS) production begin to outweigh the

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many compensatory mechanisms aimed to keep these oxidative species regulated [17]. This is evidenced by oxidative damage to many key metabolic proteins during the preclinical stage and each of the three clinical stages of AD [18-25]. This “chicken or the egg” –type genesis of AD pathology by oxidative damage and metabolic dysfunction is by no means a perfect mechanism and many of its details have been challenged. One recent proposal brought forth by Sun describes a lack of nutrient and oxygen availability as a key factor that promotes a protective response by the neuron to down-regulate metabolism [26].

Regardless of exactly what role oxidative stress plays in the pathogenesis of AD, there is a large body of accumulating evidence that shows key proteins involved in glycolysis, TCA cycle, electron-transport chain, and antioxidant capabilities of the cell become oxidized at various stages throughout the progression of the disease [18-22,25,27,28]. A summary of these oxidized enzymes is listed in Table 1. In order to understand the progression of AD, it is important to relate changes in metabolic function with the pathological characteristics of the disease. Patients with PCAD, also referred to as asymptomatic AD (ASYAD), show no signs of cognitive decline but present some of the neuropathological symptoms of AD. Braak scores range typically between III and IV due to the presence of NFTs and NTs [29,30]. MCI can be classified as amnesic or nonamnesic to distinguish between

those with impaired memory and those without. Peterson defines MCI as being characterized by cognitive impairment and decline not related to age, no dementia, and no significant impairment in daily living [31]. Amnesic MCI patient autopsies are more common than non-amnesic patient autopsies and typically have Braak stages of III or IV, demonstrating a marked amount of neurodegeneration [32]. MCI transitions into EAD as memory continues to decline and other cognitive functions show impairment. Braak staging also increases compared to MCI, with autopsied patients having scores of IV or V. Markesbery et al. have described EAD and amnesic MCI as being virtually identical neuropathologically with the exception of neuritic plaques being elevated in EAD [33]. Dementia progresses during the transition from EAD to LAD, affecting nearly every aspect of an individual's life. Braak scores of V and VI and increased A β deposits are associated with this stage of the disease [34].

Amyloid Beta Peptide (A β) and Tau

Much progress has been made in developing transgenic mouse models of AD pathology for interpretation. Specifically, models aimed at identifying the relationship between the hallmark proteins of Alzheimer's disease, A β and tau, have provided valuable insight into the mechanisms by which these proteins interfere with normal

Enzyme	Role	Nitration ¹	HNE-Modificatio ²	Carbonylation ³	Up-Regulated ⁴	Down-Regulated ⁴	Decreased Activity ⁵
FBA	Glycolysis	MCI, EAD	LAD	EAD	PCAD, LAD		
TPI	Glycolysis	EAD, LAD	EAD				
GAPDH	Glycolysis	LAD		PCAD			
PGK	Glycolysis		MCI				
PGM	Glycolysis	EAD		EAD			
Enolase	Glycolysis	MCI, EAD, LAD	MCI, EAD, LAD	LAD	LAD		EAD
PK	Glycolysis			PCAD			MCI
LDH	LAF	LAD	MCI				MCI
Aconitase	TCA Cycle		LAD		PCAD		LAD
MDH	TCA Cycle	MCI	EAD				
COX	ETC					PCAD	
ATPase	ETC	EAD, LAD	MCI, EAD, LAD	PCAD		LAD	MCI, EAD, LAD
GDH	Metabolism/ Excitotoxicity	EAD					EAD
CK	Metabolism			LAD			LAD
Cu/Zn SOD	Antioxidant			PCAD	PCAD		EAD*
MnSOD	Antioxidant		EAD, LAD		LAD		EAD*
GST	Antioxidant	MCI	LAD		LAD		LAD
CR	Antioxidant		MCI				
Prx2	Antioxidant	EAD					
Prx6	Antioxidant	MCI	LAD				
MRP	Antioxidant	MCI	LAD				

Table 1: Oxidized proteins related to metabolism and antioxidant capacity of the cell are summarized.¹ see [19,23-25]. ² see [19,21,22,47]. ³ see [18,41,43,105,106]. ⁴ see [21,47,107]. ⁵ see [19-22,47,105]. (HNE = 4-hydroxynonenal; FBA = fructose bisphosphate aldolase; TPI = triose phosphate isomerase; GAPDH = glyceraldehyde-3-phosphate dehydrogenase; PGK = phosphoglycerate kinase; PGM = phosphoglycerate mutase; PK = pyruvate kinase; LDH = lactate dehydrogenase; MDH = malate dehydrogenase; COX = cytochrome C oxidase; ATPase = ATP synthase; GDH = glutamate dehydrogenase; CK = creatine kinase; SOD = superoxide dismutase; GST = glutathione-S-transferase; CR = carbonyl reductase; Prx = peroxiredoxin; MRP = multi-drug resistant protein; LAF = lactic acid fermentation; TCA = tricarboxylic acid; ETC = electron transport chain).

*Overall SOD activity is reduced in EAD.

mitochondrial function. Eckert highlights the synergistic roles of A β and tau on down-regulating respiratory chain enzymes while promoting oxidative stress and an increased production of hyperphosphorylated tau protein in transgenic mouse models, leading to a proposed cyclic mechanism where increased A β deposition and neurofibrillary tangle formation is facilitated by dysfunctions in the respiratory chain that are initially brought about by increased levels of A β [35]. Specifically, an increase in A β levels, but not plaque levels, is associated with a significant decrease in the mitochondrial membrane potential as well as ATP production in 3 monthold Thy-1-APP₇₅₁SL transgenic mice. This mitochondrial dysfunction becomes more prominent as the mice age, leading to elevations in ROS formation, increased susceptibility to Fe²⁺-catalyzed hydroxyl radical formation, as well as decreased respiratory chain Complex IV activity[36]. Similarly, A β fibrils induce a fivefold increase in intracellular neurofibrillary tangle formation in a tau transgenic P301L mouse model, and proteomic analyses indicate down regulation of respiratory chain Complex I, ATP synthase, triose phosphate isomerase, malate dehydrogenase, glutathione S-transferase, and glutathione peroxidase. Significant decreases in the respiratory control ratio and ATP levels were also found with 24 month old transgenic mice [37,38]. A triple transgenic mouse model (pR5/APP/PS2) combining the A β and tau pathologies to facilitate significant increases in A β and phosphorylated tau levels, leads to several mitochondrial dysfunction markers, such as decreases in the activities of respiratory chain enzymes Complex I and Complex IV at 12 months of age [39]. Transgenic mouse models such as these serve as excellent resources for determining the exact roles of A β and tau in the cascade of events leading to cell death and ultimately neurodegeneration. These studies give credit to hypotheses such as the cyclic mechanism brought forth by Eckert that place mitochondrial dysfunction at the center of early AD pathology [35].

Glycolysis

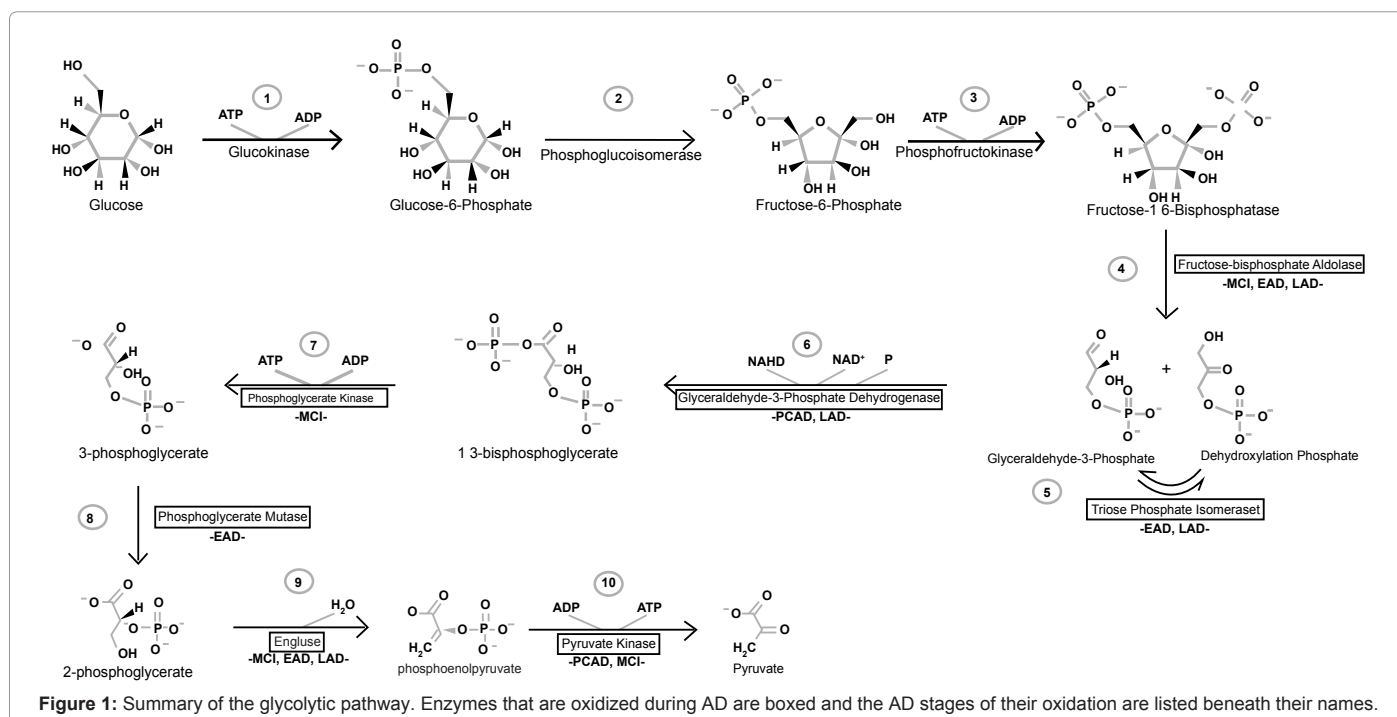
Glycolytic enzymes are the most heavily targeted metabolic

proteins throughout the progression of AD. Although glycolysis does not produce the majority of the energy for the cell, it generates pyruvate, which can serve as a starting material for other metabolic pathways. Proteomics research has found 70% of the enzymes involved in glycolysis to be oxidized by ROS and RNS species [18,19,21,22,25,27,40-43]. These enzymes are fructose-bisphosphate aldolase (FBA), triose phosphate isomerase (TPI), glyceraldehyde-3-phosphate dehydrogenase (GAPDH), phosphoglycerate kinase (PGK), phosphoglycerate mutase (PGM), enolase, and pyruvate kinase (PK). Figure 1 illustrates which enzymes are oxidized in glycolysis during the progression of AD.

TCA Cycle

The TCA cycle accounts for approximately 90% of the energy produced for the cell. Thus, preserving the integrity of the enzymes involved in this metabolic pathway is integral to meeting the cell's energy demands. Neurons have very specific homeostatic requirements, such as maintaining phospholipid asymmetry, calcium regulation, and synaptic functioning. Overall energy demand for these processes in the brain is very high, as evidenced by the fact that the brain accounts for approximately 2% of body weight but uses 20% of body's glucose and over 30% of the inspired oxygen. Proteomics research has found 25% of the enzymes directly involved in the TCA cycle to become significantly oxidized during the progression of AD [19,21,23]. These enzymes are aconitase (ACO) and malate dehydrogenase (MDH).

In addition to its function in the TCA cycle, MDH is also involved in the malate-aspartate shuttle, which is partially responsible for efficient transfer of NADH equivalent electrons across the inner mitochondrial membrane. It is also worth noting that oxaloacetate can also be formed from pyruvate via pyruvate carboxylase, but pyruvate formation is potentially hindered in the early stages of AD by the oxidation of PK in PCAD and MCI and reduced PK activity in MCI [18,22,44]. Glutamate dehydrogenase (GDH) can form α -ketoglutarate (α -KG), another key substrate in the TCA cycle and malate-aspartate



shuttle, via deamination of glutamate to yield one NADH molecule. GDH is oxidized in EAD, resulting in a reduction of enzyme activity [20]. Because GDH is responsible for keeping glutamate levels down, loss of GDH function can increase glutamate levels and lead to excitotoxicity [45]. Thus, the oxidation of MDH, PK, and GDH early in the progression of Alzheimer's disease could limit the availabilities of acetylCoA, oxaloacetate, and α -KG, limit the efficient transfer of NADH equivalent electrons across the inner mitochondrial membrane, and lead to excitotoxicity. This implicates MDH, PK, and GDH oxidation in the early metabolic dysfunction and neuronal loss that characterize AD.

Other Metabolic Processes

Protein oxidation is also involved in lactic acid fermentation, creatine phosphorylation, and ATP synthesis at various stages of AD. Lactic acid fermentation, an anaerobic method of pyruvate reduction by lactate dehydrogenase (LDH), generates lactic acid for use in gluconeogenesis as well as recycling NAD^+ back into glycolysis. The free NAD^+ generated by LDH can be very important when the cell requires immediate energy at the expense of glucose via glycolysis, thus potentially implicating the oxidation of LDH in metabolic dysfunction in early stages of AD progression. Creatine kinase catalyzes the reversible phosphorylation of creatine to phosphocreatine by ATP in the intermembrane space of the mitochondria to transport high-energy phosphate groups into the cytosol. This plays a large role in cell energy homeostasis by giving the cell access to the ATP generated by mitochondria. ATP synthase (ATPase), also known as Complex V of the electron transport chain, is a multi-subunit protein involved in the substrate-level phosphorylation of ADP in the mitochondrial matrix. Figure 2 illustrates the function of ATPase. The generation of ATP by ATPase is central to proper metabolism. Unfortunately, oxidation of the components of ATPase is highly involved in each stage during the progression of AD, leading to decreased enzyme activity during MCI, EAD, and LAD [18-22,46]. Due to its role in metabolism and

oxidized during each stage of AD, ATPase is likely a key player in the progression of this disease.

Antioxidant Enzymes

Since metabolic function is the primary source of ROS production, and because these ROS play such a significant role in metabolic protein function, antioxidant enzymes are thus implicated in the progression of AD [17]. Several antioxidant enzymes exist to maintain a reductive environment in the cytosol and mitochondria. A number of these proteins are oxidized during AD. Specifically, glutathione-S-transferase (GST), carbonyl reductase (CR), multidrug resistant protein 3, various superoxide dismutase (SOD) isoenzymes, and peroxiredoxins (Prx) are all oxidized at various stages of the disease [19-23,47]. GST detoxifies toxic species such as HNE so it is crucial for maintaining the integrity of proteins susceptible to HNE oxidation. Like GST, CR can also reduce HNE as well as carbonyl containing compounds. Superoxide is generated as a result of electron passage through the electron transport chain, so keeping mitochondrial SOD levels is very important for protecting metabolic proteins from oxidation.

Pharmaceutical Interventions

While there are several treatment options for Alzheimer's disease, current therapeutic interventions are limited in their functionality, measurements, and effectiveness. There are currently no options available that stop or reverse the disease. Treating the two neuropathological hallmarks of AD, extracellular senile plaques and intracellular neurofibrillary tangles, has become a major area of focus for drug development [48-50]. Other treatment options aim to counteract the symptoms of AD by increasing neurotransmitter levels (specifically acetylcholine) or acting on neurotransmitter receptors to limit excitotoxicity [51-54]. Because metabolic abnormalities can precede the cognitive impairments associated with AD, and because cytosolic and mitochondrial oxidative stress are well-documented in the disease, early pharmaceutical and diet intervention in these areas is of utmost importance [13,14,55,56]. A general overview of treatment options is listed in Table 2.

Treatment of Senile Plaques

Treatment of senile plaques involves decreasing the formation and aggregation of the $\text{A}\beta$ peptide, the main component of these plaques. In relation to oxidative damage, especially mitochondrial oxidative damage, early treatments for reducing $\text{A}\beta$ formation may play a critical role in slowing the progression of AD. Amyloid precursor protein (APP) is a transmembrane protein that plays a role in long term potentiation, neuronal plasticity, and memory loss [57]. Cleavage of APP is accomplished by three different enzymes (Figure 3). Specifically, β -secretase and γ -secretase are involved in cleaving APP at the two sites required to produce the insoluble $\text{A}\beta$ peptide, so they are targets for inhibition to decrease $\text{A}\beta$ formation [58,59]. APP cleavage by γ -secretase is nonspecific and forms $\text{A}\beta$ peptides of varying lengths. The peptides with lengths of 40 amino acids ($\text{A}\beta_{40}$) and 42 amino acids ($\text{A}\beta_{42}$) play a central role in plaque formation because their ratio has been implicated in how $\text{A}\beta$ aggregation occurs [60]. $\text{A}\beta_{42}$ acts as an insoluble seed that promotes the aggregation of $\text{A}\beta$ peptides, whereas $\text{A}\beta_{40}$ has been shown to inhibit $\text{A}\beta$ deposit formation [61-63]. Thus, methods to decrease the $\text{A}\beta_{42}/\text{A}\beta_{40}$ ratio via modulating γ -secretase activity may become very useful strategies in treating AD as they evolve

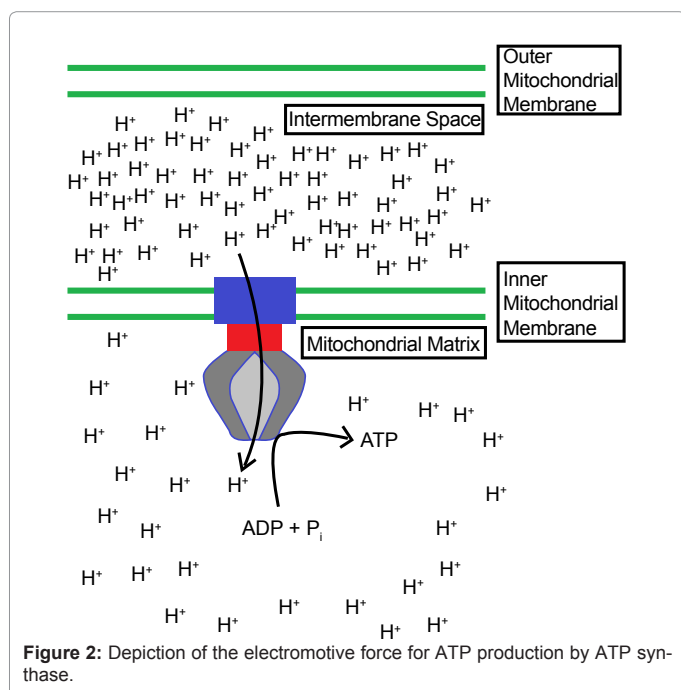
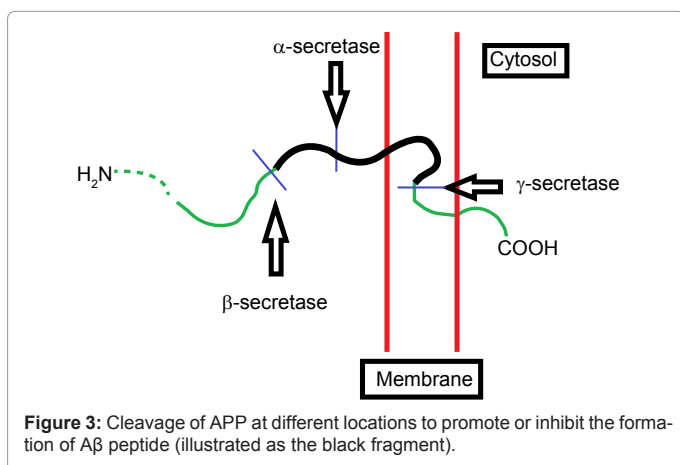


Figure 2: Depiction of the electromotive force for ATP production by ATP synthase.

Signs and Symptoms	Treatment	Effect
Senile Plaque Formation	β -secretase inhibitor	Decreases the formation of A β by inhibiting cleavage of APP by β -secretase [51]
	γ -secretase inhibitor	Decreases the formation of A β by inhibiting cleavage of APP by γ -secretase [52]
	PKC activator	Decreases the formation of A β by increasing cleavage of APP within the A β domain by α -secretase via up-regulation by PKC activation [53]
	Antiaggregant	Inhibits aggregation of A β into oligomers [54,55]
	Vaccination	Causes an immune response by which the body targets A β for degradation [56-58]
	ApoE inhibitor	Allows for removal of A β by blocking attachment of ApoE to A β [59]
Neurofibrillary Tangle	CDK5 inhibitor	Inhibits phosphorylation of tau protein by CDK5 [60]
	GSK-3 inhibitor	Inhibits phosphorylation of tau protein by GSK-3 [61]
	ApoE inhibitor	May inhibit phosphorylation of tau protein by GSK-3 via signal blocking [62,63]
	Antiaggregant	Inhibits aggregation of tau protein [64]
Low Acetylcholine Levels	AChE inhibitor	Increases levels of acetylcholine by inhibiting its degradation by AChE [45-48]
Oxidative Stress	Antioxidant	Breaks or inhibits free radical chain reactions by scavenging unpaired electrons [65,66]

Table 2: A summary of current treatment options for AD and their effects. (A β = amyloid beta peptide; AChE = acetylcholinesterase; ApoE = apolipoprotein E; APP = amyloid precursor protein; CDK5 = cyclin-dependent kinase 5; GSK-3 = glycogen synthase kinase 3; PKC = protein kinase C).

[60]. Activation of α -secretase also inhibits A β formation by cleaving APP within the A β domain [64]. Once formed, A β aggregates form oligomers of various sizes as well as the plaques that are traditionally associated with AD. Interference with this aggregation takes place at each stage of the process, with several compounds targeting early aggregation and others targeting larger assemblies of oligomers [65,66]. Apolipoprotein E (ApoE) is a lipoprotein used to in triglyceride catabolism. Although several alleles exist for this lipoprotein, the E4 allele is a risk factor for AD. Inhibiting apolipoprotein E has also been explored, as this protein hinders the removal of A β [67]. A relatively new area of research involves vaccination against A β , which has shown promising results in mouse models, and to some extent, in human trials [68-70]. Petrushina demonstrated that this type of immunotherapy can selectively decrease the formation of insoluble A β plaques without affecting the levels of soluble A β or causing cerebral microhemorrhages that have previously been encountered [70,71]. However, human clinical trials highlighted other dangers associated with immunotherapy when several patients undergoing this treatment strategy developed meningoencephalitis, causing the trials to be halted. While levels of A β plaques were found to be lower in treated patients at autopsy, cerebral inflammation was also found and cognitive decline was not prevented as a result of plaque clearance [69].



Treatment of Neurofibrillary Tangles

Neurofibrillary tangles are composed of hyperphosphorylated tau protein. Similar to treatment options for senile plaques, reduction of NFTs has been investigated using enzyme inhibitors as well as antiaggregant compounds that block the aggregation of tau protein [72]. Hyperphosphorylation of tau has been attributed to re-entry into the cell cycle by neurons, which has been observed in AD, because tau hyperphosphorylation is common during embryonic cell development but not in differentiated cells [73]. Hyperphosphorylation of tau is known to occur via cyclin-dependent kinase 5 (CDK5) and glycogen synthase kinase 3 (GSK-3), both of which have been targeted for inhibition [74,75]. ApoE may also become a target for inhibition specifically for its role in signaling GSK-3 [76,77]. Recent research has shown a direct relationship between cell cycle dysregulation (likely as a result of oxidative stress), hyperphosphorylation of tau, and AD progression [78]. Thus, treating only NFT formation, instead of upstream regulation defects, may only slow the progression of AD and not prevent or reverse the disease.

Neurotransmitter Availability

When neurons die as a result of neurodegeneration, levels of neurotransmitters inevitably fall and consequently promote cognitive decline. In particular, acetylcholine has been the major neurotransmitter targeted for increase by means of acetylcholinesterase (AChE) inhibition. In 1985, Davies highlights the central role of acetylcholine in learning and cognitive function [79]. For this reason, 80% of FDA-approved treatments (i.e. Exelon[®], Razadyne[®], Cognex[®], and Aricept[®]) for cognitive impairment in Alzheimer's disease target the degradation of acetylcholine by the enzyme acetylcholinesterase, with the fifth treatment, Namenda[®], antagonizing N-methyl-D-aspartate (NMDA) receptors to decrease excitotoxicity [51-54]. While increasing the availability of neurotransmitters may temporarily increase cognition, this type of treatment addresses AD-related symptoms and not the pathology of the disease.

Antioxidant Therapies

Oxidative stress manifests itself through a variety of mechanisms during the progression of AD. More importantly, oxidative stress

precedes both traditional pathological hallmarks of AD, senile plaque formation and neurofibrillary tangles, leading in part to the formation of the "two-hit hypothesis" postulated by Zhu in 2004 [14,80,81]. This hypothesis places oxidative stress and cell cycle dysregulation at the forefront of the pathological onset of AD. Changes in metabolism have also been observed to occur before the appearance of the cognitive deficits of AD, and these kinds of changes can be caused by mtDNA mutation after exposure to reactive oxygen species (ROS) via changes in mitochondrial fission and fusion processes [13,82-84]. Control of mitochondrial fission and fusion is known to directly affect energy production and ROS formation [82,85]. Thus there is a potential cyclic mechanism by which early oxidative stress causes changes in mitochondrial fission and fusion control via mtDNA mutation, which in turn creates more ROS and exacerbates the shift toward an oxidative environment that the cell can't overcome. A similar mechanism introduced in 2000 incorporates energy insufficiency and Ca^{2+} dyshomeostasis into the mechanism by which ROS and mitochondrial damage perpetuate one another [86]. These mechanisms offer insight into how important and possibly manageable the progression of AD can be if biomarkers for these disruptions in cell homeostasis are discovered early enough.

Successful attempts at reducing oxidative stress, reducing changes in mitochondrial dynamics, and restoring cognitive function have used a supplemental combination of the antioxidants acetyl L-carnitine (ALCAR) and R- α -lipoic acid (LA) [87-91]. These endogenous compounds are involved in maintaining efficient metabolism of glucose and fatty acids. Structures for ALCAR and LA are shown in Figure 4. L-carnitine is involved in transporting long-chain fatty acids into the mitochondria for β -oxidation and transporting shorter fatty acid chains out of the mitochondria. By transporting short-chain fatty acids out of the mitochondria, L-carnitine is also involved in freeing coenzyme A (CoASH) for use in the TCA cycle via carnitine acyltransferase (CAT). LA is required as a coenzyme for pyruvate dehydrogenase and α -ketoglutarate dehydrogenase. ALCAR and LA have been shown to work synergistically to improve CAT binding and activity, which are reduced in aged rat brain [89]. ALCAR alone has also been shown to increase antioxidant enzyme activity as well as total antioxidant capacity in the plasma of healthy people, making it a viable candidate for early, safe intervention long before biomarkers for AD appear [92].

Future Directions

The role of inflammation in the pathogenesis of AD has become a central target for therapeutic intervention, involving such pharmaceuticals as non-steroidal anti-inflammatory drugs (NSAIDs), immunotherapy options, and other means of destroying A β plaques

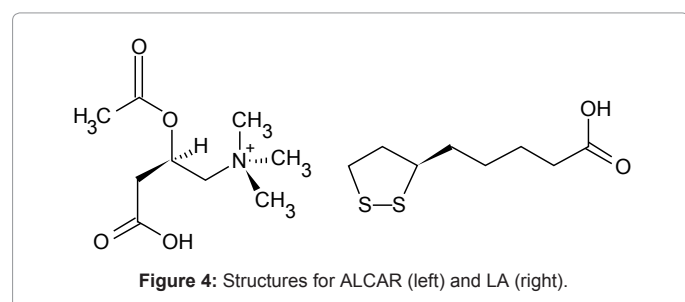


Figure 4: Structures for ALCAR (left) and LA (right).

[66,68,93]. The specific roles of various conformations of A β peptide as neurotoxins as well as mediators of beneficial cellular processes significantly impact the direction of anti-A β treatments [94]. Current literature suggests that progress has been made in developing A β antibodies that are specific to soluble and insoluble conformations [95]. A multi-treatment approach may prove to be invaluable in the coming years, as indicated by Gotz, as therapeutic strategies targeting tau protein also become more sophisticated [38]. This is evidenced by several transgenic mouse models of individual and combined A β - and tau-induced AD pathologies in relation to mitochondrial dysfunction [35-37,39,96-99]. Given the natural abilities of certain antioxidants to protect mitochondria from ROS-induced oxidative damage, future therapeutic strategies would likely benefit from treatments targeting not only the reduction of AD pathological hallmarks, but also the strengthening of protective mechanisms that are inherent to the neuron [91].

Conclusions

Oxidative stress occurs when there is an imbalance between oxidants and antioxidants in a system. Free radicals from ROS and RNS are highly elevated in during the progression of Alzheimer's disease, while antioxidant enzymes have shown loss of protein function, reduced activity, and sometimes compensatory up-regulation to meet cellular demands. Therefore, increased oxidative stress can result from these free radicals causing cognitive decline in PCAD, MCI, EAD, and AD patients supporting the hypothesis of free radicals as an underlying contributor to Alzheimer's disease [100-103]. Among the metabolic and antioxidant enzymes reviewed, the glycolytic enzymes are the primary targets of oxidative damage. Because glycolysis is an upstream process relative to the TCA cycle and electron transport chain, glycolytic enzyme oxidation and accompanying loss of activity may serve as a trigger for downstream metabolic defects that occur early in the progression of AD. Although there are several hurdles in developing successful treatments for Alzheimer's disease [104], namely understanding the pathogenesis and successfully approaching the factor(s) involved in progression, early intervention and treatment efficacy are the most important factors that influence changes in the progression of AD. Current treatments have been met with little effectiveness, although vaccination against A β and early supplementation of ALCAR and LA may provide more promising results in the future. Combining therapies such as these at an early stage may prove to be a very effective therapeutic strategy once more progress has been made in clarifying AD pathogenesis.

References

- Halliwel B (2006) Oxidative stress and neurodegeneration: where are we now? *J Neurochem* 97: 1634-1658.
- Ferrante RJ, Browne SE, Shinobu LA, Bowling AC, Baik MJ, et al. (1997) Evidence of increased oxidative damage in both sporadic and familial amyotrophic lateral sclerosis. *J Neurochem* 69: 2064-2074.
- Miquel J, Economos AC, Fleming J, Johnson JE (1980) Mitochondrial role in cell aging. *Exp Gerontol* 15: 575-591.
- Sastre J, Pallardo FV, Garcia de la Asuncion J, Vina J (2000) Mitochondria, oxidative stress and aging. *Free Radic Res* 32: 189-198.
- Aruoma OI, Kaur H, Halliwel B (1991) Oxygen free radicals and human diseases. *J R Soc Health* 111: 172-177.
- Butterfield DA, Kanski J (2001) Brain protein oxidation in age-related neurodegenerative disorders that are associated with aggregated proteins. *Mech Ageing Deve* 122: 945-962.

7. Zhu X, Lee HG, Casadesus G, Avila J, Drew K, et al. (2005) Oxidative imbalance in Alzheimer's disease. *Mol Neurobiol* 31: 205-217.
8. Bowling AC, Beal MF (1995) Bioenergetic and oxidative stress in neurodegenerative diseases. *Life Sci* 56: 1151-1171.
9. Beal MF (1995) Aging, energy, and oxidative stress in neurodegenerative diseases. *Ann Neurol* 38: 357-366.
10. Butterfield DA, Reed T, Perluigi M, De Marco C, Coccia R, et al. (2006) Elevated protein-bound levels of the lipid peroxidation product, 4-hydroxy-2-nonenal, in brain from persons with mild cognitive impairment. *Neurosci Lett* 397: 170-173.
11. Launer LJ, Andersen K, Dewey ME, Letenneur L, Ott A, et al. (1999) Rates and risk factors for dementia and Alzheimer's disease: results from EURODEM pooled analyses. EURODEM Incidence Research Group and Work Groups. European Studies of Dementia. *Neurology* 52: 78-84.
12. Grant WB, Campbell A, Itzhaki RF, Savory J (2002) The significance of environmental factors in the etiology of Alzheimer's disease. *J Alzheimers Dis* 4: 179-189.
13. Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, et al. (1996) Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med* 334: 752-758.
14. Nunomura A, Perry G, Aliev G, Hirai K, Takeda A, et al. (2001) Oxidative damage is the earliest event in Alzheimer disease. *J Neuropathol Exp Neurol* 60: 759-767.
15. (1997) Consensus recommendations for the postmortem diagnosis of Alzheimer's disease. The National Institute on Aging, and Reagan Institute Working Group on Diagnostic Criteria for the Neuropathological Assessment of Alzheimer's Disease. *Neurobiol Aging* 18: S1-2.
16. Braak H, Braak E (1991) Neuropathological staging of Alzheimer-related changes. *Acta neuropathol* 82: 239-259.
17. Beckman KB, Ames BN (1998) Mitochondrial aging: open questions. *Ann N Y Acad Sci* 854: 118-127.
18. Aluise CD, Robinson RA, Cai J, Pierce WM, Markesbery WR, et al. (2011) Redox proteomics analysis of brains from subjects with amnesic mild cognitive impairment compared to brains from subjects with preclinical Alzheimer's disease: insights into memory loss in MCI. *J Alzheimers Dis* 23: 257-269.
19. Reed TT, Pierce WM, Markesbery WR, Butterfield DA (2009) Proteomic identification of HNE-bound proteins in early Alzheimer disease: Insights into the role of lipid peroxidation in the progression of AD. *Brain Res* 1274: 66-76.
20. Reed TT, Pierce WM, Turner DM, Markesbery WR, Butterfield DA (2009) Proteomic identification of nitrated brain proteins in early Alzheimer's disease inferior parietal lobule. *J Cell Mol Med* 13: 2019-2029.
21. Perluigi M, Sultana R, Cenini G, Di Domenico F, Memo M, et al. (2009) Redox proteomics identification of 4-hydroxynonenal-modified brain proteins in Alzheimer's disease: Role of lipid peroxidation in Alzheimer's disease pathogenesis. *Proteomics Clin Appl* 3: 682-693.
22. Reed T, Perluigi M, Sultana R, Pierce WM, Klein JB, et al. (2008) Redox proteomic identification of 4-hydroxy-2-nonenal-modified brain proteins in amnesic mild cognitive impairment: insight into the role of lipid peroxidation in the progression and pathogenesis of Alzheimer's disease. *Neurobiol Dis* 30: 107-120.
23. Sultana R, Reed T, Perluigi M, Coccia R, Pierce WM, et al. (2007) Proteomic identification of nitrated brain proteins in amnesic mild cognitive impairment: a regional study. *J Cell Mol Med* 11: 839-851.
24. Sultana R, Boyd-Kimball D, Poon HF, Cai J, Pierce WM, et al. (2006) Redox proteomics identification of oxidized proteins in Alzheimer's disease hippocampus and cerebellum: An approach to understand pathological and biochemical alterations in AD. *Neurobiol Aging* 27: 1564-1576.
25. Castegna A, Thongboonkerd V, Klein JB, Lynn B, Markesbery WR, et al. (2003) Proteomic identification of nitrated proteins in Alzheimer's disease brain. *Journal of neurochemistry* 85: 1394-1401.
26. Sun J, Feng X, Liang D, Duan Y, Lei H (2011) Down-Regulation of Energy Metabolism in Alzheimer's Disease is a Protective Response of Neurons to the Microenvironment. *J Alzheimers Dis* 28: 389-402.
27. Sultana R, Boyd-Kimball D, Cai J, Pierce WM, Klein JB, et al. (2007) Proteomics analysis of the Alzheimer's disease hippocampal proteome. *J Alzheimers Dis* 11: 153-164.
28. Sultana R, Perluigi M, Butterfield DA (2006) Protein oxidation and lipid peroxidation in brain of subjects with Alzheimer's disease: insights into mechanism of neurodegeneration from redox proteomics. *Antioxid Redox signal* 8: 2021-2037.
29. West MJ, Kawas CH, Stewart WF, Rudow GL, Troncoso JC (2004) Hippocampal neurons in pre-clinical Alzheimer's disease. *Neurobiol Aging* 25: 1205-1212.
30. Jicha GA, Abner EL, Schmitt FA, Kryscio RJ, Riley KP, et al. (2011) Preclinical AD Workgroup staging: pathological correlates and potential challenges. *Neurobiol Aging* 33: 622.
31. Petersen RC (2004) Mild cognitive impairment as a diagnostic entity. *J intern med* 256: 183-194.
32. Markesbery WR (2010) Neuropathologic alterations in mild cognitive impairment: a review. *J Alzheimers Dis* 19: 221-228.
33. Markesbery WR, Schmitt FA, Kryscio RJ, Davis DG, Smith CD, et al. (2006) Neuropathologic substrate of mild cognitive impairment. *Arch Neurology* 63: 38-46.
34. Braak H, Braak E (1997) Frequency of stages of Alzheimer-related lesions in different age categories. *Neurobiol Aging* 18: 351-357.
35. Eckert A, Schulz KL, Rhein V, Gotz J (2010) Convergence of amyloid-beta and tau pathologies on mitochondria in vivo. *Mol Neurobiol* 41: 107-114.
36. Hauptmann S, Scherping I, Drose S, Brandt U, Schulz KL, et al. (2009) Mitochondrial dysfunction: an early event in Alzheimer pathology accumulates with age in AD transgenic mice. *Neurobiol Aging* 30: 1574-1586.
37. Gotz J, Chen F, van Dorpe J, Nitsch RM (2001) Formation of neurofibrillary tangles in P3011 tau transgenic mice induced by Abeta 42 fibrils. *Science* 293: 1491-1495.
38. Gotz J, Ittner A, Ittner LM (2012) Tau-targeted treatment strategies in Alzheimer's disease. *Br J Pharmacol* 165: 1246-1259.
39. Rhein V, Song X, Wiesner A, Ittner LM, Baysang G, et al. (2009) Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proc Natl Acad Sci USA* 106: 20057-20062.
40. Butterfield DA, Perluigi M, Sultana R (2006) Oxidative stress in Alzheimer's disease brain: new insights from redox proteomics. *Eur J Pharmacol* 545: 39-50.
41. Castegna A, Aksenov M, Aksenova M, Thongboonkerd V, Klein JB, et al. (2002) Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part I: creatine kinase BB, glutamine synthase, and ubiquitin carboxy-terminal hydrolase L-1. *Free Radic Biol Med* 33: 562-571.
42. Sultana R, Poon HF, Cai J, Pierce WM, Merchant M, et al. (2006) Identification of nitrated proteins in Alzheimer's disease brain using a redox proteomics approach. *Neurobiol Dis* 22: 76-87.
43. Sultana R, Perluigi M, Newman SF, Pierce WM, Cini C, et al. (2010) Redox proteomic analysis of carbonylated brain proteins in mild cognitive impairment and early Alzheimer's disease. *Antioxid Redox signal* 12: 327-336.
44. Butterfield DA, Poon HF, St Clair D, Keller JN, Pierce WM, et al. (2006) Redox proteomics identification of oxidatively modified hippocampal proteins in mild cognitive impairment: insights into the development of Alzheimer's disease. *Neurobiol Dis* 22: 223-232.
45. Facheris M, Beretta S, Ferrarese C (2004) Peripheral markers of oxidative stress and excitotoxicity in neurodegenerative disorders: tools for diagnosis and therapy? *J Alzheimers Dis* 6: 177-184.
46. Sultana R, Perluigi M, Butterfield DA (2006) Redox proteomics identification of oxidatively modified proteins in Alzheimer's disease brain and in vivo and in vitro models of AD centered around Abeta(1-42). *J Chromatogr B Analyt Technol Biomed Life Sci* 833: 3-11.
47. Sultana R, Butterfield DA (2004) Oxidatively modified GST and MRP1 in

- Alzheimer's disease brain: implications for accumulation of reactive lipid peroxidation products. *Neurochem Res* 29: 2215-2220.
48. Aguzzi A, O'Connor T (2010) Protein aggregation diseases: pathogenicity and therapeutic perspectives. *Nat rev Drug discov* 9: 237-248.
49. Grill JD, Cummings JL (2010) Current therapeutic targets for the treatment of Alzheimer's disease. *Expert rev Neurother* 10: 711-728.
50. Selkoe DJ, Schenk D (2003) Alzheimer's disease: molecular understanding predicts amyloid-based therapeutics. *Annu Rev Pharmacol Toxicol* 43: 545-584.
51. Geldmacher DS (2004) Donepezil (Aricept) for treatment of Alzheimer's disease and other dementing conditions. *Expert Rev Neurother* 4: 5-16.
52. Litvinenko IV, Sakharovskaia AA (2009) [Results of the open multicenter prospective study of safety and tolerability of rivastigmine (exelon) in different titration regimes in mild and moderate Alzheimer's disease]. *Zh Nevrol Psikhiatr Im S S Korsakova* 109: 29-35.
53. Summers WK (2000) Tacrine (THA, Cognex(R)). *J Alzheimers Dis* 2: 85-93.
54. Woodruff-Pak DS, Tobia MJ, Jiao X, Beck KD, Servatius RJ (2007) Preclinical investigation of the functional effects of memantine and memantine combined with galantamine or donepezil. *Neuropsychopharmacology* 32: 1284-1294.
55. Pappolla MA, Omar RA, Kim KS, Robakis NK (1992) Immunohistochemical evidence of oxidative [corrected] stress in Alzheimer's disease. *Am J Pathol* 140: 621-628.
56. Yakes FM, Van Houten B (1997) Mitochondrial DNA damage is more extensive and persists longer than nuclear DNA damage in human cells following oxidative stress. *Proc Natl Acad Sci U S A* 94: 514-519.
57. Turner PR, O'Connor K, Tate WP, Abraham WC (2003) Roles of amyloid precursor protein and its fragments in regulating neural activity, plasticity and memory. *Prog Neurobiol* 70: 1-32.
58. Lanz TA, Karmilowicz MJ, Wood KM, Pozdnyakov N, Du P, et al. (2006) Concentration-dependent modulation of amyloid-beta in vivo and in vitro using the gamma-secretase inhibitor, LY-450139. *J Pharmacol Exp Ther* 319: 924-933.
59. Vassar R, Bennett BD, Babu-Khan S, Kahn S, Mendiaz EA, et al. (1999) Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science* 286: 735-741.
60. Yin YI, Bassit B, Zhu L, Yang X, Wang C, et al. (2007) {gamma}-Secretase Substrate Concentration Modulates the Abeta42/Abeta40 Ratio: IMPLICATIONS FOR ALZHEIMER DISEASE. *J Biol Chem* 282: 23639-23644.
61. Kim J, Onstead L, Randle S, Price R, Smithson L, et al. (2007) Abeta40 inhibits amyloid deposition in vivo. *J Neurosci* 27: 627-633.
62. Deng Y, Tarassishin L, Kallhoff V, Peethumongsin E, Wu L, et al. (2006) Deletion of presenilin 1 hydrophilic loop sequence leads to impaired gamma-secretase activity and exacerbated amyloid pathology. *J Neurosci* 26: 3845-3854.
63. Jarrett JT, Berger EP, Lansbury PT (1993) The carboxy terminus of the beta amyloid protein is critical for the seeding of amyloid formation: implications for the pathogenesis of Alzheimer's disease. *Biochemistry* 32: 4693-4697.
64. Hooper NM, Turner AJ (2002) The search for alpha-secretase and its potential as a therapeutic approach to Alzheimer's disease. *Curr Med Chem* 9: 1107-1119.
65. Maezawa I, Hong HS, Liu R, Wu CY, Cheng RH, et al. (2008) Congo red and thioflavin-T analogs detect Abeta oligomers. *J Neurochem* 104: 457-468.
66. McLaurin J, Kierstead ME, Brown ME, Hawkes CA, Lambermon MH, et al. (2006) Cyclohexanehexol inhibitors of Abeta aggregation prevent and reverse Alzheimer phenotype in a mouse model. *Nat Med* 12: 801-808.
67. Liu Q, Wu WH, Fang CL, Li RW, Liu P, et al. (2011) Mapping ApoE/Abeta binding regions to guide inhibitor discovery. *Mol Biosyst* 7: 1693-1700.
68. Asuni AA, Boutajangout A, Scholtzova H, Knudsen E, Li YS, et al. (2006) Vaccination of Alzheimer's model mice with Abeta derivative in alum adjuvant reduces Abeta burden without microhemorrhages. *Euro J Neurosci* 24: 2530-2542.
69. Kuzuhara S (2010) Treatment strategy of Alzheimer's disease: pause in clinical trials of Abeta vaccine and next steps. *Brain Nerve* 62: 659-666.
70. Petrushina I, Ghochikyan A, Mktrichyan M, Mamikonyan G, Movsesyan N, et al. (2007) Alzheimer's disease peptide epitope vaccine reduces insoluble but not soluble/oligomeric Abeta species in amyloid precursor protein transgenic mice. *J Neurosci* 27: 12721-12731.
71. Wilcock DM, Jantzen PT, Li Q, Morgan D, Gordon MN (2007) Amyloid-beta vaccination, but not nitro-nonsteroidal anti-inflammatory drug treatment, increases vascular amyloid and microhemorrhage while both reduce parenchymal amyloid. *Neuroscience* 144: 950-960.
72. Congdon EE, Necula M, Blackstone RD, Kuret J (2007) Potency of a tau fibrillization inhibitor is influenced by its aggregation state. *Arch Biochem Biophys* 465: 127-135.
73. Lovestone S, Reynolds CH (1997) The phosphorylation of tau: a critical stage in neurodevelopment and neurodegenerative processes. *Neuroscience* 78: 309-324.
74. Hung KS, Hwang SL, Liang CL, Chen YJ, Lee TH, et al. (2005) Calpain inhibitor inhibits p35-p25-Cdk5 activation, decreases tau hyperphosphorylation, and improves neurological function after spinal cord hemisection in rats. *J Neuropathol Exp Neurol* 64: 15-26.
75. Uno Y, Iwashita H, Tsukamoto T, Uchiyama N, Kawamoto T, et al. (2009) Efficacy of a novel, orally active GSK-3 inhibitor 6-Methyl-N-[3-[[3-(1-methylethoxy)propyl]carbonyl]-1H-pyrazol-4-yl]pyridine-3-carboxamide in tau transgenic mice. *Brain research* 1296: 148-163.
76. Cedazo-Minguez A, Popescu BO, Blanco-Millan JM, Akterin S, Pei JJ, et al. (2003) Apolipoprotein E and beta-amyloid (1-42) regulation of glycogen synthase kinase-3beta. *J Neurochem* 87: 1152-1164.
77. Ohkubo N, Lee YD, Morishima A, Terashima T, Kikkawa S, et al. (2003) Apolipoprotein E and Reelin ligands modulate tau phosphorylation through an apolipoprotein E receptor/disabled-1/glycogen synthase kinase-3beta cascade. *FASEB J* 17: 295-297.
78. Keeney JT, Swomley AM, Harris JL, Fiorini A, Mitov MI, et al. (2011) Cell Cycle Proteins in Brain in Mild Cognitive Impairment: Insights into Progression to Alzheimer Disease. *Neurotox Res*.
79. Davies P (1985) A critical review of the role of the cholinergic system in human memory and cognition. *Ann NY Acad Sci* 444: 212-217.
80. Zhu X, Lee HG, Perry G, Smith MA (2007) Alzheimer disease, the two-hit hypothesis: an update. *Biochim Biophys Acta* 1772: 494-502.
81. Zhu X, Raina AK, Perry G, Smith MA (2004) Alzheimer's disease: the two-hit hypothesis. *Lancet neurol* 3: 219-226.
82. Chen H, Chomyn A, Chan DC (2005) Disruption of fusion results in mitochondrial heterogeneity and dysfunction. *J Biol Chem* 280: 26185-26192.
83. Jendrach M, Mai S, Pohl S, Voith M, Bereiter-Hahn J (2008) Short- and long-term alterations of mitochondrial morphology, dynamics and mtDNA after transient oxidative stress. *Mitochondrion* 8: 293-304.
84. Small GW, Mazziotta JC, Collins MT, Baxter LR, Phelps ME, et al. (1995) Apolipoprotein E type 4 allele and cerebral glucose metabolism in relatives at risk for familial Alzheimer disease. *JAMA* 273: 942-947.
85. Yu T, Robotham JL, Yoon Y (2006) Increased production of reactive oxygen species in hyperglycemic conditions requires dynamic change of mitochondrial morphology. *Proc Natl Acad Sci U S A* 103: 2653-2658.
86. Blass JP (2000) The mitochondrial spiral. An adequate cause of dementia in the Alzheimer's syndrome. *Ann NY Acad Sci* 924: 170-183.
87. Aliev G, Liu J, Shenk JC, Fischbach K, Pacheco GJ, et al. (2009) Neuronal mitochondrial amelioration by feeding acetyl-L-carnitine and lipoic acid to aged rats. *J Cell Mol Med* 13: 320-333.
88. Liu J, Head E, Gharib AM, Yuan W, Ingersoll RT, et al. (2002) Memory loss in old rats is associated with brain mitochondrial decay and RNA/DNA oxidation:

- partial reversal by feeding acetyl-L-carnitine and/or R-alpha -lipoic acid. *Proc Natl A Sci U S A* 99: 2356-2361.
89. Liu J, Killilea DW, Ames BN (2002) Age-associated mitochondrial oxidative decay: improvement of carnitine acetyltransferase substrate-binding affinity and activity in brain by feeding old rats acetyl-L- carnitine and/or R-alpha -lipoic acid. *Proc Natl Acad Sci U S A* 99: 1876-1881.
90. Long J, Gao F, Tong L, Cotman CW, Ames BN, et al. (2009) Mitochondrial decay in the brains of old rats: ameliorating effect of alpha-lipoic acid and acetyl-L-carnitine. *Neurochem Res* 34: 755-763.
91. Shenk JC, Liu J, Fischbach K, Xu K, Puchowicz M, et al. (2009) The effect of acetyl-L-carnitine and R-alpha-lipoic acid treatment in ApoE4 mouse as a model of human Alzheimer's disease. *J Neurol sci* 283: 199-206.
92. Cao Y, Qu HJ, Li P, Wang CB, Wang LX, et al. (2011) Single dose administration of L-carnitine improves antioxidant activities in healthy subjects. *Tohoku J Exp Med* 224: 209-213.
93. Heneka MT, Kummer MP, Weggen S, Bulic B, Multhaup G, et al. (2011) Molecular mechanisms and therapeutic application of NSAIDs and derived compounds in Alzheimer's disease. *Curr Alzheimer Res* 8: 115-131.
94. Lublin AL, Gandy S (2010) Amyloid-beta oligomers: possible roles as key neurotoxins in Alzheimer's Disease. *Mt Sinai J Med* 77: 43-49.
95. Zago W, Buttini M, Comery TA, Nishioka C, Gardai SJ, et al. (2012) Neutralization of Soluble, Synaptotoxic Amyloid beta Species by Antibodies Is Epitope Specific. *J Neurosci* 32: 2696-2702.
96. Lustbader JW, Cirilli M, Lin C, Xu HW, Takuma K, et al. (2004) ABAD directly links Abeta to mitochondrial toxicity in Alzheimer's disease. *Science* 304: 448-452.
97. Grueninger F, Bohrmann B, Czech C, Ballard TM, Frey JR, et al. (2010) Phosphorylation of Tau at S422 is enhanced by Abeta in TauPS2APP triple transgenic mice. *Neurobiol Dis* 37: 294-306.
98. Schuessel K, Schafer S, Bayer TA, Czech C, Pradier L, et al. (2005) Impaired Cu/Zn-SOD activity contributes to increased oxidative damage in APP transgenic mice. *Neurobiol Dis* 18: 89-99.
99. David DC, Hauptmann S, Scherping I, Schuessel K, Keil U, et al. (2005) Proteomic and functional analyses reveal a mitochondrial dysfunction in P301L tau transgenic mice. *J Biological Chem* 280: 23802-23814.
100. Markesbery WR (1997) Oxidative stress hypothesis in Alzheimer's disease. *Free Radic Biol Med* 23: 134-147.
101. Leibovitz BE, Siegel BV (1980) Aspects of free radical reactions in biological systems: aging. *J Gerontol* 35: 45-56.
102. Barouki R (2006) Ageing free radicals and cellular stress. *Med Sci (Paris)* 22: 266-272.
103. Butterfield DA, Reed TT, Perluigi M, De Marco C, Coccia R, et al. (2007) Elevated levels of 3-nitrotyrosine in brain from subjects with amnesic mild cognitive impairment: Implications for the role of nitration in the progression of Alzheimer's disease. *Brain Res* 1148: 243-248.
104. Carter MD, Simms GA, Weaver DF (2010) The development of new therapeutics for Alzheimer's disease. *Clini Pharmacol Ther* 88: 475-486.
105. Aksenov M, Aksenova M, Butterfield DA, Markesbery WR (2000) Oxidative modification of creatine kinase BB in Alzheimer's disease brain. *J Neurochem* 74: 2520-2527.
106. Castegna A, Aksenov M, Thongboonkerd V, Klein JB, Pierce WM, et al. (2002) Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part II: dihydropyrimidinase-related protein 2, alpha-enolase and heat shock cognate 71. *J Neurochem* 82: 1524-1532.
107. Aluise CD, Robinson RA, Beckett TL, Murphy MP, Cai J, et al. (2010) Preclinical Alzheimer disease: brain oxidative stress, Abeta peptide and proteomics. *Neurobiol Dis* 39: 221-228.

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