

Insights into Glycobiology and Infection Mechanisms Using CT-GM1 Interaction

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DESCRIPTION

Cholera intoxication of human enteroids is a topic that explores the interaction between *Vibrio cholerae*, the bacterium responsible for causing cholera, and human enteroids, which are three-dimensional cell culture models of the human intestinal epithelium. This interaction is of significant interest in the fields of microbiology, infectious disease, and glycobiology. Glycobiology plays an important role in cholera intoxication as the interaction between *Vibrio cholerae* and host intestinal cells involves specific glycan-receptor interactions. A prominent example of this is the binding of Cholera Toxin (CT) to GM1 ganglioside, a specific glycan receptor on the surface of intestinal epithelial cells.

Cholera intoxication of human enteroids

***Vibrio cholerae* and cholera toxin:** Cholera is a severe diarrheal disease caused by *Vibrio cholerae*, a bacterium that infects the human intestine. Cholera toxin is the main virulence factor produced by *V. cholerae*, and it plays a crucial role in the pathogenesis of the disease. The toxin is responsible for the massive secretion of fluids and electrolytes in the intestine, leading to the characteristic watery diarrhea.

Human enteroids: Human enteroids are *in vitro* models of the human intestinal epithelium. They are derived from stem cells or biopsy samples and can replicate the structure and function of the human intestine, including the presence of various cell types found in the gut, such as enterocytes, goblet cells, and paneth cells. Enteroids are used to study various aspects of intestinal physiology and pathophysiology.

Glycobiology in cholera intoxication: Glycobiology is the study of complex carbohydrates (glycans) and their roles in biological processes. In the context of cholera, glycobiology is crucial because cholera toxin binds to specific glycans on the surface of intestinal epithelial cells. These glycans, including GM1 ganglioside, serve as receptors for the toxin. Understanding the glycan structures and their interactions with cholera toxin is essential for comprehending the initial steps of infection.

Cholera intoxication mechanism: Cholera toxin is taken up by enterocytes in the human intestine. Once inside the cell, it modifies the G protein, leading to the activation of adenylate cyclase and an increase in intracellular cAMP levels. This, in turn, triggers chloride ion secretion and inhibits sodium ion absorption, resulting in the massive outpouring of water and electrolytes into the intestinal lumen.

Use of enteroids in research: Human enteroids are valuable tools for studying cholera intoxication. Researchers can use these models to investigate the interaction between cholera toxin and intestinal epithelial cells, the specific glycan-receptor interactions, and the cellular responses to the toxin. This provides insights into the pathogenesis of cholera and can help in the development of therapeutic strategies.

Clinical implications: Understanding the molecular details of cholera intoxication of human enteroids can lead to the development of novel therapies and preventive strategies for cholera, which remains a significant global health concern in areas with poor sanitation and clean water access.

CT-GM1 interaction

GM1 ganglioside is a type of glycosphingolipid found on the surface of intestinal epithelial cells. It has a complex glycan structure consisting of sialic acid, galactose, glucose, and other sugar residues. GM1 serves as the primary receptor for cholera toxin. Cholera toxin is secreted by *Vibrio cholerae* and consists of two subunits: The A subunit (CTA) and the B subunit (CTB). The CTB subunit is responsible for binding to GM1 ganglioside. CTB has a high affinity for the glycan structure of GM1. Once CTB binds to GM1 on the surface of intestinal epithelial cells, the toxin-receptor complex is internalized through endocytosis. The glycan-receptor interaction is critical for this internalization process.

Inside the host cell, the cholera toxin undergoes a series of events that lead to its toxic effects. Cholera toxin modifies a host G protein by ADP-ribosylation, leading to the activation of adenylate cyclase. This results in increased levels of cyclic AMP

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(cAMP) in the cell, leading to altered ion transport and excessive secretion of fluids into the intestinal lumen. This is the primary cause of the profuse watery diarrhea characteristic of cholera. While GM1 is the best-characterized receptor for cholera toxin, variations in glycan structures exist among individuals. Some people may express GM1 with slightly different sugar compositions. Understanding the diversity in glycan structures and their interaction with cholera toxin is important for both research and potential therapeutic strategies. The knowledge of the CT-GM1 interaction has been utilized in the development of cholera vaccines. Some vaccines use an inactivated or genetically modified version of cholera toxin as an antigen to stimulate an immune response, thus generating protective antibodies against the toxin.

CONCLUSION

In summary, glycobiology is integral to cholera intoxication due to the specific interactions between cholera toxin and host cell receptors, such as GM1 ganglioside. The binding of cholera toxin to glycan receptors on intestinal cells is the initial step in the pathogenesis of cholera, leading to the toxin's internalization and subsequent toxic effects on the host, which result in severe diarrhea. The study of cholera intoxication of human enteroids provides a valuable platform for investigating the molecular and cellular interactions that underlie cholera pathogenesis, with a particular focus on the important role of glycobiology in the infection process. This research has clinical implications for addressing cholera and other enteric infections.