

## Influences of *Pleurotus Sajor-caju* Diets on Performance and Biochemical Parameters in Experimental Rats

Molehin Olorunfemi Raphael and Oyetayo Folake Lucy\*

Department Of Biochemistry, Faculty of Science University of Ado Ekiti P.M.B.5363, Ado-Ekiti, Nigeria

### Abstract

The cap and stalk of *Pleurotus sajor-caju* mushroom were analysed for proximate composition and used for a 28 day feeding trial in experimental rats. Protein composition of cap (26.33%) and stalk (22.59 %) were relatively high while the crude fat cap (3.67%), stalk (2.6%) was low. Plasma urea of rats fed mushroom cap (25.6±0.3) and stalk (21.7±0.4) were lower than that of the control (31.0±0.3) diet fed rats. The plasma creatinine of rats fed mushroom cap (4.2 ±0.3) and stalk (3.1±0.1) were lower than that of the control (6.9±0.3). Performances of rats fed mushroom showed that daily feed intakes (g) were found to be lower in rats fed mushroom stalk (9.8±0.0) than those fed control diet. Also, daily feed intake was found to be higher in rats fed mushroom cap (10.2±0.0) than the control (9.8±0.0) diet. There were no significant differences in the feed gain ratio of rats fed cap, stalk, and casein control diet. The implication of these results is that mushroom diet is an important source of nutrients which supported growth without eliciting toxicological effects.

**Keywords:** *Pleurotus sajor-caju*; Proximate compositions; Urea; Creatinine; Crude protein; Feed gain ratio

### Introduction

Mushrooms are saprophytic fungi which produce a wide range of extracellular enzymes that enable them to degrade complex organic matter into soluble substances for the purpose of nutrition [1]. Edible mushrooms such as *Pleurotus sajor-caju*, *Agaricus bisporus*, *Pleurotus abolorus*, *Pleurotus florida* have been recognized as delicacies in human diet. They have been known to be cultivated by the Chinese for centuries [2].

Edible mushrooms are important sources of nutrients valuable as health foods because they are low in calories but high in vegetable proteins. These offer advantages over animal proteins in human nutrition in countries where animal proteins are expensive and inadequate and over plant proteins because it contains some essential amino acids especially lysine and leucine in high concentrations [3].

Edible mushrooms such as *Pleurotus* species contain substances that aid the body's immune systems e.g. *Pleurotus folrida* possess antioxidant and anti-tumor activities [4,5], while *Tremella fuciformis* (wood ear) is an immune stimulant. *Pleurotus sajor-caju* contains ingredients that modulate hypertension by affecting the rennin-angiotensin system [6,7]. Edible mushrooms represent a good source of high value of nutrients. They are good sources of proteins and vitamins such as vitamins A, B<sub>1</sub>, B<sub>2</sub>, C, and D<sub>2</sub>, niacin and minerals like phosphorus, iron and calcium [8,9].

*Pleurotus sajor-caju* (polyporales: polyporaceae) is an edible cosmopolitan mushroom [10]. Originating from India, it grows naturally on a succulent plant in the foothills of the Himalayas. It is believed to be indigenous to South East Asia where it is commercially being cultivated on farm waste products such as banana pseudo stems, rice or wheat, straw, pine needles and dust [3,11]. *Pleurotus sajor-caju* is an important commercially produced mushroom because it can produce a broad spectrum of lignocellulotic enzymes and this is reflected in its ability to grow on waste residue of widely varying composition [1]. Earlier report on *Pleurotus sajor-caju* have been on the chemical composition [12]. The foregoing hopes to draw attention to

influences of *Pleurotus sajor-caju* diet on some biochemical parameters in experimental rats.

### Materials and Methods

#### Collection of mushrooms

*Pleurotus sajor-caju* fruit bodies were obtained from local markets around Ado-Ekiti, Nigeria. Samples were cleaned, separated into cap and stalk, oven dried at 60°C and powdered in a Philips blender.

#### Experimental animals

Wistar strain male albino rats (n=15) of the same litter weighing 40-50g were obtained from the rat colony of the Department of Biochemistry of University of Ilorin, Nigeria. Rats were divided into 3 groups. Group 1 rats were fed mushroom cap diet; group 2 rats were fed mushroom stalk diet while group 3 rats were fed the casein control diet. The rats were acclimatized for seven days, thereafter housed in individual steel cages. Feeds and water were given to them ad libitum.

#### Diet formulation

The percentage composition of experimental diets is shown on Table 2. Powdered mushroom was incorporated into the diet group at 10% protein level. Vitamin/mineral mixture and vegetable oil were added at 5% level and corn starch was used to make up the diet to 100. Casein was the protein source for the control.

\*Corresponding author: Oyetayo Folake Lucy, Department of Biochemistry, Faculty Of Science, University of Ado Ekiti P.M.B.5363, Ado-Ekiti, Nigeria, Tel: +234-806-016-3025; E-mail: [ovounad@yahoo.com](mailto:ovounad@yahoo.com)

Received August 27, 2011; Accepted November 17, 2011; Published November 19, 2011

Citation: Raphael MO, Lucy OF (2011) Influences of *Pleurotus Sajor-caju* Diets on Performance and Biochemical Parameters in Experimental Rats. J Nutr Food Sci 1:112. doi:10.4172/2155-9600.1000112

Copyright: © 2011 Raphael MO, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## Blood sample collection

After 28 days of feeding experimental diets, rats were sacrificed by neck decapitation and blood were obtained from the rats via cardiac puncture and collected into EDTA bottles. The blood samples collected were analysed for plasma urea and creatinine concentrations.

## Estimations

Plasma urea concentration of metabolites was determined by the method of Searcy et al. [13] while a plasma creatinine concentration was determined by the method of Slot, [14]. Crude protein was determined by the method of AOAC, [15]. Proximate chemical composition of the samples was determined by the method of AOAC, [16]. Organs (liver, kidney, heart, and small intestines) were excised and weighed.

## Statistical analysis

Data obtained were expressed as means of triplicate determinations. The Level of significance was set at  $p \leq 0.05$  (17)

## Results

The bolden figures are corrected to 1 decimal places are directed by the reviewers.

## Discussion

The proximate composition of the cap and stalk of the *Pleurotus sajor-caju* are presented in Table 1. The crude protein concentration of cap *Pleurotus sajor-caju*, (26.3±0.01) is slightly higher than the stalk *Pleurotus sajor-caju* (22.5±0.3). The protein values of both cap and stalk is higher than the average proteins present in mushrooms (17.5%) [18]. Both cap and stalk is higher when compared with other mushrooms like *Agaricus bisporus* (16.40±0.01) and *Agaricus bisporus* (19.5±0.01) but lower when compared to *Termitomyces mammiformis* (36.8%). The moisture content of cap *Pleurotus sajor-caju* is comparable to those obtained from *Termitomyces mammiformis*. However, the cap *Pleurotus sajor-caju* moisture content is higher when compared with *Agaricus bisporus* (5.2±0.1) but lower to *Agaricus bitorquis* (12.1±0.13). The carbohydrate concentrations is comparable to previously reported for *Pleurotus sajor-caju* (39.8) by [7]. The ash content of (10.4±0.02) for cap is comparable to those obtained for *Agaricus bisporus* 11.01±0.02

Parameters	Cap	Stalk
Moisture	10.2±0.0	10.1±0.0
Crude Protein	26.3±0.0	22.5±0.3
Crude fibre	8.9±0.1	16.2±0.3
Crude fat	3.7±0.2	2.6±0.1
Ash	10.4±0.2	8.5 ±1.0
Total Carbohydrate	38.2±0.0	40.0± 0.1

Table 1: Proximate composition (%) of cap and stalk of *Pleurotus sajor-caju*.

Sample	Cap Diet	Stalk Diet	Control Diet
Casein	—	—	11.5
Cornstarch	41.6	21.3	78.5
Mushroom	48.4	68.7	—
Vegetable oil	5.0	5.0	5.0
Vit/Mineral	5.0	5.0	5.0
% crude protein	20.7	14.6	87.0

Table 2: Percentage composition of experimental diets.

Sample	Urea(mg/dL)	Creatinine (mg/dL)
Cap	25.6±0.3	4.20±0.3
Stalk	21.7±0.4	3.1±0.1
Control	31.0±0.3	6.9±0.3

Data are mean ± S.D values for triplicates of determinations.

Cap group = Basal diet +10% cap of *Pleurotus sajor-caju*

Stalk group = Basal diet +10% stalk of *Pleurotus sajor-caju*

Control group = Basal diet + casein

Table 3: Plasma urea and creatinine concentration (mg/dL) in blood of rats fed *Pleurotus sajor-caju* diets.

Sample	Daily weight gain (g)	Daily feed intake (g)	Feed gain ratio
Cap	3.3±0.0	10.2±0.0	3.1±0.0
Stalk	3.3±0.0	9.8±0.0	2.9±0.0
Control	3.3±0.0	9.8±0.02	2.9±0.0

Data are mean± S.D values for triplicates of determinations.

Cap group = Basal diet +10% cap of *Pleurotus sajor-caju*

Stalk group = Basal diet +10% stalk of *Pleurotus sajor-caju*

Control group = Basal diet + casein

Table 4: Nutrient Utilization and performance of rats fed *Pleurotus sajor-caju* diets.

Organ	Cap diet	Stalk diet	Control diet
Liver	3.4±0.2	4.6±0.0	5.9±0.2
Kidney	0.01±0.01	0.01±0.00	0.5±0.00
Heart	0.4±0.0	0.5±0.00	0.5±0.3
Small intestine	3.9±0.1	4.96±0.3	5.0±0.3

Data are mean± S.D values for triplicates of determinations

Cap group = Basal diet +10% cap of *Pleurotus sajor-caju*

Stalk group = Basal diet +10% stalk of *Pleurotus sajor-caju*

Control group = Basal diet + casein

Table 5: Relative Organ weight (g/100g body wt) of rats fed *Pleurotus sajor-caju* diets.

and *Agaricus bitorquis* 10.11±0.10 [2]. The crude fibre composition of the cap was relatively higher than that of the cap.

Table 3 shows the plasma urea and creatinine concentrations (mg/dl) of rats fed cap, stalk and control diet. The mean plasma urea concentrations (mg/dl) were found to be lower in rats fed mushroom cap (25.6±2.04) and stalk (21.7±2.45) diets compared with the control diet fed (31.0±2.02). Plasma urea is derived from dietary protein and endogenous protein catabolism [19]. A decrease in dietary proteins results in reduction of plasma urea concentration which is influenced by protein quality and quantity [20]. Hence plasma urea concentrations can be used to predict protein quality [21]. Reduction of dietary protein results in decreased faecal urinary and total nitrogen excretion [22]. The higher plasma mean urea concentration of the control rats (31.0±0.3) might be due to the higher quality of the control diet. Animal proteins are usually of higher quality than those of plant origin [23-25] stated that dietary proteins reduce plasma urea concentration. This might explain the lowered plasma urea concentration in cap and stalk diets fed rats. Plasma creatinine concentration distribution showed no significance differences ( $p \leq 0.05$ ) between the treatment groups and the control. This may suggest the low rate of conversion of muscle creatine of rats to creatinine [26].

Nutritional Performance of rats fed mushroom diets is shown on Table 4. There were no significant changes ( $p \leq 0.05$ ) in the daily feed intake in all the rats fed mushroom cap ( $10.2 \pm 0.03$ ) and stalk ( $9.8 \pm 0.01$ ) diets compared with the control fed rats ( $9.8 \pm 0.02$ ). Daily weight gain for the groups fed mushroom diets were lower than that of the control. There was an appreciable weight gain in all the rats fed mushroom cap and stalk diets [3]. Feed gain ratio in all the groups fed mushroom diets suggest adequate utilization of diets.

Table 5 shows the relative organ weights of rats fed mushroom and control diets. There was no significant difference ( $p \leq 0.05$ ) between the liver weights of rats fed wild cap diet and the control diet-fed rats. Organs from stalk diets fed rats significantly different from one another and the control. This suggests that the *Pleurotus sajor-caju* diets were not toxic as liver hypertrophy has been related to toxicity in rats. The kidney weights of rats fed cap and stalk s diets showed no significant differences ( $p \leq 0.05$ ) from one another and also when compared with the control. Muscle and liver tissues are more susceptible to effects of dietary proteins than kidneys [27]. Hence, an increase in kidney weight could be an indication of a toxic environment. The mushroom diet treatment groups had significant lower ( $p \leq 0.05$ ) relative heart weight than the control group. The above results showed that the cap and stalk wild-obtained *Pleurotus sajor-caju* diets are not likely to cause any toxicological insult to vital organs and hence, safe for consumption.

In conclusion, it can be seen from the results gotten in this research that *Pleurotus sajor-caju* is a good source of quality protein has been shown to enhance growth without eliciting toxicological insults on major organs in experimental rats. Since this species of mushroom is already widely accepted as an important source of inexpensive nutrients, we further encourage its husbandry.

## References

1. Buswell I, Chang SI (1992) The production of the three enzymes of edible mushrooms. Mushroom science 9: 745 -760.
2. Sadiq Saiqa H, Nawal B, Muhammed AI (2008) Studies on chemical composition and Nutritive Evaluation of wild Edible mushrooms. Iran J chem chem Eng 27: 3.
3. Oyetayo FL, Oyetayo VO (2009) Assessment of nutritional quality of wild and cultivated *Pleurotus sajor-caju*. J Med Food 12: 1149-1153.
4. Nayana J, Janardhanan KK (2000) Antioxidant and anti tumor activity of *Pleurotus florida*. Current Science 79: 941-943.
5. Manpreet K, Giridhar S, Khanna, PK (2004) In vitro and in vivo antioxidant potentials of *Pleurotus florida* in experimental animals. Mushroom research 12: 21-26.
6. Chang R (1996) Functional Properties of edible mushroom. Nutrition Reviews 54: 91-93.
7. Alam N, Hossain S, Khair A, Rubul Amin SM, Asaduzzaman K (2007) Comparative effects of Oyster mushrooms on Plasma lipid profile of hypercholesterolaemic rats. Bangladesh J Mushroom 1:15-22.
8. Rayman M (2000) The importance of Selenium to human health. The lancet 356: 233-241.
9. Alam N, Hossain S, Khair A, Rubul Amin SM, Asaduzzaman K (2007) Nutritional analysis of Dietary mushroom-*Pleurotus florida* Eger and *Pleurotus sajor-caju* (Fr)Singer. Bangladesh J Mushroom 1: 1-7.
10. Zadrazil F, Kurtzman RH (1982) The biology of *Pleurotus* cultivation in the tropics In Tropical mushrooms (Chang ST, Quimio TH (Eds.), Hong Kong: The Chinese Press, 493: 277-298.
11. Jandaik CL, Kapur JN (1976) Effect of carbon and nitrogen nutrition of *Pleurotus sajor-caju*. Indian Phytopathology 29: 326-327.
12. Oyetayo FL, Akindahunsi AA (2006) Nutrient and antinutrient distribution of edible mushroom, *Pleurotus tuber-regium* (fries) singer. Swiss Society of Food Science and Technology LWT 39: 548-553.
13. Searcy RL, Reardon JE, Foreman JA (1967) A new photometric method for serum urea nitrogen determination. Am J Med Technol 33: 15-20
14. Slot C (1965) Plasma creatinine determination: A new and specific Jaffe Reaction method. Scand J Clin Lab Invest 17: 381.
15. Association of Official Analytical Chemist, AOAC (1984) Official methods of analysis of the association of official analytical chemists. 14 ed. Arlington VA 1141.
16. Association of Official Analytical Chemist, AOAC (1980): Official methods of analysis of the association of official analytical chemists. 14 ed. Arlington VA 1141.
17. Zar JH (1984) Biostatistical Analysis (2<sup>nd</sup> ed.) Prentice Hall. Inc.Upper Saddle River. NJ.
18. Sivrikaya, H, Bacak L, Teroglu I, Eroglu H, (2002) Truce Element in *Pleurotus sajor-caju* cultivated on chemi thermomechanical pulp for bioleaching. Food chem 79: 173.
19. Baron DN, Wicher JT, lee KE (1994) A new short textbook of Chemical pathology.5<sup>th</sup> ed. ELBS, London.pp 151-156.
20. Zervas S, Zijlstra RT (2002) Effect of dietary protein and oathhull fiber on nitrogen excretion patterns and postrandial plasma urea profiles in grower pigs. J Anim Sci 80: 3238-3246.
21. Coma J, Carrion, Zimmerman DR (1995) Use of plasma nitrogen as a rapid response criterion to determine the lysine requirement of pigs. J AM Science 73: 472 481.
22. Dourmal JY, Henry Y, Bourdon D, Quiniou N, Guillou D (1993) Effect of growth performance, carcass characteristics and nitrogen output in growing finishing pigs. Pages 206 -211 In Proc. 1st Int. Symposium on Nitrogen Floro in Pig production and Environmental consequence. EAAP publ. No. 69. Pudoc, Wageningen, the Netherlands.
23. Carpenter KJ (1994) Protein and energy: A study of changing ideas in nutrition. New York 1: 2 -4.
24. Lopez JRD, Goodband GL, Alle GW, Jesse JL, Nelssen MD, et al. (1994) The effect of diets formulated on an ideal protein basis on growth performance, carcass characteristics and thermal balance of finishing gilts housed in a hot, diurnal environment. J Anim Sci 72: 367-379.
25. Lenis NJ, Van Diepen HTM, Bukker P, Jongblood AW, Van der Meupen J (1999) Effect of the ratio between essential and non-essential amino acids in the diets on utilization of nitrogen and amino acids by growing pigs. J Animal Sci 77: 1777-1787.
26. Wyss M, Kaddurah-Daouk R (2000) Creatine and creatinine metabolism. Physiological Reviews 80: 1107.
27. Allison JB (1955) Biological evaluation of Protein. Physiol Rev 35: 664-670.