

In Vitro Potency of Combinatorial Antifungal Drugs towards Candida Isolates from Patients

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ABSTRACT

Candidiasis is a fungal infection (mycosis) caused by an opportunistic yeast with only 8-10 pathogenic species. In spite of the multiple classes of antifungal drugs available to combat these infections, the treatment is hampered by drug toxicity, tolerability, emergence of drug resistant isolates and many more. Combination therapy has been suggested as a possible approach to improve these treatment outcomes. This study is, therefore, aimed at the analysis of the sensitivity of *Candida* isolates obtained from patients towards different classes of drugs affecting multiple pathways in the pathogen life cycle. Four different individual drugs with different mechanisms of action and six different combinations of these drugs at three different proportions (1:1, 1:3, and 3:1) were used for the study. All the drug sensitivity assays were performed by the agar well diffusion method. The results were statistically analyzed using Prism software. Results have shown synergistic effects of drug combinations on the sensitivity of the isolates without much variation in the stoichiometric ratios, thereby providing an alternative to monotherapy to combat the emergence of drug resistant isolates and drying existing drug pipelines.

Keywords: Candidiasis; Isolates; Drug combinations; Statistical analysis

INTRODUCTION

The last few years have witnessed progressive increase in mycotic infections which are one of the primary causes of morbidity and mortality especially in immunocompromised and severely ill patients. Candida is a fungal pathogen which is usually a member of normal flora but transits to pathogenic form by several virulence factors like adherence to host tissues and medical devices, secretion of extracellular hydrolytic enzymes etc. [1]. The species is responsible for various clinical manifestations including overgrowth as well as life threatening disseminated infections. Candida species have been reported as the fourth most common nosocomial bloodstream pathogen by National Nosocomial Infections Surveillance System (NNISS) with a mortality rate of about 45% [1]. Till date, more than 150 species of Candida are known with only 10 species responsible for the disease with C. albicans being responsible for majority of the infections. However, recent studies have shown the increase in incidences of infection due to non-albicans Candida species like Candida auris, Candida tropicalis, Candida glabrata, Candida parapsilosis, Candida krusei, and Candida lusitaniae [2,3]. The fatality of fungal infections in immune-compromised conditions has been recently observed with mucormycosis (black fungus) and *Candida* (white fungus) infections [4]. These infections spread worldwide in both COVID-19 positive patients as well as patients recovered from COVID-19. Studies have shown the presence of oropharyngeal Candidiasis in patients suffering from COVID-19 and *C. auris* infections in critically ill COVID-19 patients [5,6].

Several non-albicans species are inherently resistant or acquire resistance to commonly used antifungal drugs. Factors like severe immunosuppression, prematurity, use of broad-spectrum antibiotics, and usage of antimycotic drugs are associated with this change. Six different classes of antifungal drugs with different modes of action are available which selectively eliminate fungal pathogens from a host with minimal toxicity to the host. These include Azoles, Allylamines, Polyenes, Echinocandins, Nucleotide Analogue and Penicillin derivatives. Amphotericin and Azoles have been the choice of drugs for patients suffering from mucormycosis and *Candida* infections during the COVID-19 outbreak [7]. However, numerous recent studies have shown the emergence of drug resistant *Candida* strains

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and isolates. This along with increase in disease causing species has led to a dire need for development of newer drug strategies. One of these novel drug strategies is the combinatorial therapy consisting of multiple drugs affecting different pathways which have proven to be much more efficacious than a single counterpart. It offers numerous advantages such as no or vey less toxicity analysis, lower treatment failure rate, lower case-fatality ratios, fewer side-effects than monotherapy, slower development of resistance, and thus less money needed for the development of new drugs [8]. Thus, in view of this, the present study is focused on determining the drug sensitivity profiles of Candida isolates obtained from patients in Rajasthan, India. Further, we have also analyzed the efficacy of drug combinations of multiple drugs in three different stoichiometric ratios towards these isolates. All the results were appropriately statistically analyzed. Our results have clearly shown that all drug combinations are much more efficacious than a single drug towards these isolates without much variation observed at different stoichiometric ratios.

MATERIALS AND METHODS

Candida isolates and culture conditions

Candida isolates used in the study were kindly provided by MCRD-CIRD (Microbial Culture Repository Division-Centre for Innovation, Research and Department), Dr. B. Lal Clinical Laboratory Pvt. Ltd., Jaipur. These isolates were extracted from various infectious human samples such as urine, blood, pus and vaginal swabs obtained from Candidiasis patients in Rajasthan. Patient's demographic details such as sex, age and clinical information were recorded and used for analysis. The study consists of a total of 17 isolates which were cultivated and maintained on YEPD medium (Hi Media Laboratories, Mumbai, India) (unless specified) at 30°C. The isolates were confirmed macroscopically by visualizing the colony morphology (creamy colony in color) on YEPD agar plates as well as microscopically using Indian ink (Nigrosin) staining.

Preparation of antifungal drugs and drug combinations

A total of four drugs, Itraconazole, Amphotericin B, Micafungin and Terbinafine, belonging to different classes were used in the study. All the drugs were prepared in Di Methyl Sulfoxide (DMSO) as per Clinical and Laboratory Standards Institute (CLSI) guidelines. Itraconazole and Amphotericin B were used at a concentration of 13.5 mg/ml and 0.33 mg/ml while Micafungin and Terbinafine were used at a concentration of 50 mg/ml and 0.435 mg/ml. DMSO was used as a negative control for all the assays.

Depending on the results obtained from drug sensitivity assays using individual drugs, a total of six drug combinations were prepared at three different stoichiometric assays. The drug combinations used for the study are shown in Figure 1.

Antifungal drug sensitivity

All the drug sensitivity assays were performed as per standard procedures in accordance with National Committee for Clinical Laboratory (NCCLS) guidelines by agar well diffusion assay. Briefly, the isolates were primarily inoculated in Sabouraud Dextrose Broth (SDB) and incubated at 30°C for 24 hours. These grown cultures were then diluted to an O.D. of 0.2 and then swabbed on SDA plates. Wells were punctured in these plates and 50 μ L of the prepared drug/drug combination was added into wells. Wells containing DMSO were used as negative control. Plates were incubated for 24 hours

at 30°C and the clear zone obtained around the well, known as the zone of inhibition (ZOI), was measured by zone measuring scale.



Statistical analysis

The mean ZOI \pm standard deviation of all isolates for a drug or a drug combination was calculated and was used for the comparison of the significance of drug combinations to individual drugs as well as combinations at various proportions. All the statistical analysis was done by unpaired t test using PRISM version 5.0.

RESULTS

Demographic analysis of the isolates

Demographic analysis of the isolates was done on the basis of gender, age group, sample type as well as species causing the infections are shown in Figure 2a. The details of the isolates obtained are summarized in the supporting data (Table S1). As it is evident from the results, Candida infections are not gender biased and affect both males and females are shown in Figure 2a. However, previous studies have shown that females are slightly more prone to Candida infections owing to their higher susceptibility towards UTIs (Urinary Tract Infections) [9,10]. The deviation in our results from the previous studies could be attributed to the smaller number of isolates used in the study [11,12]. The analysis of these infections in different age groups suggested almost similar infectivity rate in all age groups with slight increased rates in older age groups owing to weaker immune systems are shown in Figure 2b. This is well in agreement with previous studies suggesting that Candida infections mostly occur in immunocompromised patients, so it can occur in any age group who have weaker immune response [13-16]. Our results showed that majority of samples were obtained from the patient's sputum and urine, indicating that Candida majorly affects respiratory and urinary systems in the human body are shown in Figure 2c. It is also correlated from previous studies that the organism majorly affects lungs causing diseases like pneumonitis [17,18]. Besides, Candida based UTI infections are also very prevalent mostly observed in females as vaginal candidiasis, vulvovaginal candidiasis caused by Candida albicans [19-21]. Isolates obtained from blood demonstrate the severity of the disease is shown in Figure 2c. Evidences have also suggested the occurrence of Candida pneumonia and Candiduria in severely immunocompromised individuals with disseminated disease [22,23]. Although the emergence of infections caused by non-albican species is increasing, the majority of the infections in this study were

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caused by albican species are shown in Figure 2d. This albican species bias has been observed previously as well from a number of studies [24-26]. However, infections caused by non-albican species such as auris, *glabrata*, etc. are rising continuously which are also causing an increase in emergence of drug resistance [27-29].



Figure 2: The figure shows the proportions of isolates which are categorized on the basis of gender (a) Age groups; (b) Sample type; (c) Albicans vs non-albicans; (d) Species. **Note:** a) (**■**) Male; (**■**) Female; b) (**■**) 0-20; (**■**) 20-40; (**■**) 40-60; (**■**) 60-above, c) (**■**) Urine; (**■**) Blood; (**■**) Plus; (**■**) Sputum; (**■**) Vaginal Swab; d) (**■**) Albicans; (**■**) Non-Albicans.

Drug sensitivity of isolates towards monotherapeutic drugs

Four different antifungal drugs belonging to different classes were used for the study. Drug sensitivity analysis by standard well diffusion assay was done with DMSO as the negative control. Representative images of drug sensitivity assay are shown in the supporting data (Figure S1) and as per CLSI guidelines 17 mm ZOI was taken as the cut-off value. On the basis of results shown in Table 1, it is clear that all the isolates were sensitive to Amphotericin B and Micafungin while only 59% and 53% isolates were sensitive to Terbinafine and Itraconazole

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respectively. The sensitivity of the isolates towards Amphotericin B and Micafungin have also been previously reported to 99.5% and 100% respectively [30,31]. Similar results were observed with patients suffering from oral Candidiasis or autoimmune *Candida* dystrophy [32]. Similarly, emerging resistance was observed for *Candida* isolates towards Itraconazole [31-33]. Also,emergence of resistant has been observed with isolates towards Terbinafine [34], in patients suffering from Balanoposthitis caused by *Candida albicans* [35]. These results along with previous studies clearly indicate the emergence of drug resistant isolates in the area under the study and elsewhere towards Azoles and Terbinafine. Thus, alternate options need to be evaluated to combat the infections caused by the pathogen.

Drug sensitivity of isolates towards combinatorial therapy

Studies so far have indicated that individual drugs may not be the best choice in the current scenario. Multiple studies have illustrated drug combinations to be more efficacious preventing the emergence of resistant strains, lowering the concentration of drugs and treating critically ill patients rapidly [36]. Not just towards Candida infections, combinatorial drugs have proven to be effective in other infections as well such as Chaetomium species, invasive aspergillosis, invasive mold diseases in hematologic patients [37-39]. We, therefore, have analyzed the drug sensitivity of isolates towards drug combinations at three different stoichiometric ratios. The drugs to which more than and less than 60% isolates were sensitive were considered as S and R respectively. Representative images of drug sensitivity assay are shown in the supporting data (Figure S2). The drug combinations were used to evaluate the synergistic effects of drugs in combinations. On the basis of results shown in Tables 2-4, Itraconazole and Terbinafine were individually not effective to all the isolates but their combination was especially effective when used at 1:1 combination. Similarly, all the other five combinations showed statistically significant synergistic effects towards the growth of the isolates. These results are well in agreement with numerous previous studies which have been discussed here.

Table 1: Protein content of different strains of Pleurotuscystidiosus mushrooms (g/100 g of dried sample). Note: Resistant isolates are highlighted in red.

S.No.	Sample	Amphotericin	Terbinafine	ltraconazole	Micafungin
1	RPC 804	21 mm	14 mm	14 mm	17 mm
2	C1654	20 mm	12 mm	10 mm	19 mm
3	37/1561	21 mm	13 mm	14 mm	21 mm
4	RPC 937	20 mm	16 mm	16 mm	23 mm
5	64/316	22 mm	0 mm	10 mm	19 mm
6	20/519	19 mm	14 mm	17 mm	21 mm
7	MNC 2175	26 mm	26 mm	20 mm	21 mm
8	KNC 32	21 mm	23 mm	0 mm	29 mm
9	C1805	20 mm	16 mm	10 mm	17 mm
10	330/1994	23 mm	24 mm	31 mm	28 mm
11	C2325	26 mm	22 mm	18 mm	27 mm
12	C2262	27 mm	25 mm	18 mm	26 mm
13	C2264	26 mm	25 mm	19 mm	29 mm
14	C2321	25 mm	21 mm	20 mm	30 mm
15	C386	24 mm	25 mm	2 mm	29 mm
16	C128	21 mm	26 mm	21 mm	30 mm
17	C2554	25 mm	30 mm	18 mm	31 mm
	Percent Sensitivity	100 (17/17)	59 (10/17)	53 (10/17)	100 (17/17)

Note: Resistant isolates are highlighted in red.

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Table 2: The table shows drug sensitivity profiles (ZOI in mm) of isolates towards combination of drugs at 1:1 ratio. Note: Resistant isolates are highlighted in red.

Zone of inhibition								
S.No.	Sample	l+T	M+T	A+T	l+M	A+I	A+M	
1	RPC 804	25 mm	30 mm	28 mm	28 mm	25 mm	29 mm	
2	C1654	23 mm	29 mm	18 mm	27 mm	20 mm	28 mm	
3	37/1 561	25 mm	28 mm	22 mm	29 mm	21 mm	24 mm	
4	RPC 937	27 mm	31 mm	21 mm	27 mm	24 mm	25 mm	
5	64/316	26 mm	28 mm	19 mm	26 mm	23 mm	26 mm	
6	20/519	40 mm	40 mm	35 mm	0 mm	31 mm	24 mm	
7	MNC 2175	31 mm	27 mm	23 mm	28 mm	20 mm	28 mm	
8	KNC 32	26 mm	32 mm	18 mm	26 mm	30 mm	25 mm	
9	C1805	28 mm	28 mm	24 mm	27 mm	26 mm	25 mm	
10	330/1994	25 mm	30 mm	23 mm	27 mm	28 mm	30 mm	
11	C2325	32 mm	32 mm	20 mm	31 mm	23 mm	29 mm	
12	C2262	27 mm	30 mm	21 mm	28 mm	24 mm	26 mm	
13	C2264	27 mm	33 mm	20 mm	32 mm	24 mm	28 mm	
14	C2321	28 mm	31 mm	24 mm	27 mm	22 mm	27 mm	
15	C386	34 mm	32 mm	19 mm	33 mm	23 mm	27 mm	
16	C128	30 mm	32 mm	20 mm	27 mm	25 mm	26 mm	
17	C2554	32 mm	30 mm	22 mm	32 mm	29 mm	33 mm	

Note: Resistant isolates are highlighted in red.

Table 3: The table shows drug sensitivity profiles (ZOI in mm) of isolates towards combination of drugs at 1:3 ratio. Note: Resistant isolates are highlighted in red.

Zone of inhibition								
S.No.	Sample	l+T	M+T	A+T	l+M	A+I	A+M	
1	RPC 804	24 mm	32 mm	25 mm	24 mm	24 mm	29 mm	
2	C1654	24 mm	30 mm	17 mm	27 mm	24 mm	28 mm	
3	3 7/1 561	25 mm	34 mm	17 mm	26 mm	24 mm	30 mm	
4	RPC 937	27 mm	32 mm	23 mm	27 mm	16 mm	30 mm	
5	64/316	26 mm	30 mm	23 mm	28 mm	17 mm	32 mm	
6	20/519	40 mm	40 mm	39 mm	31 mm	27 mm	31 mm	
7	MNC 2175	30 mm	36 mm	26 mm	29 mm	20 mm	31 mm	
8	KNC 32	29 mm	36 mm	23 mm	30 mm	24 mm	31 mm	
9	C1805	30 mm	32 mm	22 mm	28 mm	23 mm	31 mm	
10	3 3 0/1994	16 mm	18 mm	30 mm	11 mm	28 mm	31 mm	
11	C2325	40 mm	36 mm	30 mm	30 mm	27 mm	27 mm	
12	C2262	28 mm	30 mm	24 mm	24 mm	25 mm	31 mm	
13	C2264	27 mm	29 mm	25 mm	24 mm	26 mm	28 mm	
14	C2321	29 mm	32 mm	23 mm	28 mm	26 mm	28 mm	
15	C386	33 mm	30 mm	26 mm	24 mm	27 mm	28 mm	
16	C128	32 mm	32 mm	23 mm	24 mm	18 mm	28 mm	
17	C2554	34 mm	31 mm	27 mm	26 mm	27 mm	28 mm	

Note: Resistant isolates are highlighted in red.

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Table 4: The table shows drug sensitivity profiles (ZOI in mm) of isolates towards combination of drugs at 3:1 ratio. Note: Resistant isolates are highlighted in red.

S.No. Sample I+T M+T A+T I+M A+I A+M 1 RPC 804 22 mm 32 mm 24 mm 22 mm 23 mm 25 mm 2 C1654 21 mm 34 mm 16 mm 23 mm 20 mm 19 mm 3 37/1 561 24 mm 34 mm 18 mm 24 mm 20 mm 22 mm 4 RPC 937 27 mm 34 mm 19 mm 22 mm 18 mm 20 mm 20 mm 5 64/316 27 mm 34 mm 19 mm 22 mm 19 mm 19 mm 6 20/519 40 mm 40 mm 32 mm 32 mm 29 mm 29 mm	Zone of inhibition								
1 RPC 804 22 mm 32 mm 24 mm 22 mm 23 mm 25 mm 2 C1654 21 mm 34 mm 16 mm 23 mm 20 mm 19 mm 3 37/1 561 24 mm 34 mm 18 mm 24 mm 20 mm 22 mm 4 RPC 937 27 mm 34 mm 20 mm 22 mm 18 mm 20 mm 20 mm 5 64/316 27 mm 34 mm 19 mm 22 mm 19 mm 19 mm 6 20/519 40 mm 40 mm 32 mm 32 mm 29 mm 29 mm	S.No.	Sample	l+T	M+T	A+T	l+M	A+I	A+M	
2 C1654 21 mm 34 mm 16 mm 23 mm 20 mm 19 mm 3 37/1 561 24 mm 34 mm 18 mm 24 mm 20 mm 22 mm 4 RPC 937 27 mm 34 mm 20 mm 22 mm 18 mm 20 mm 20 mm 5 64/316 27 mm 34 mm 19 mm 22 mm 19 mm 19 mm 6 20/519 40 mm 40 mm 32 mm 32 mm 29 mm 29 mm	1	RPC 804	22 mm	32 mm	24 mm	22 mm	23 mm	25 mm	
3 37/1 561 24 mm 34 mm 18 mm 24 mm 20 mm 22 mm 4 RPC 937 27 mm 34 mm 20 mm 22 mm 18 mm 20 mm 5 64/316 27 mm 34 mm 19 mm 22 mm 19 mm 19 mm 6 20/519 40 mm 40 mm 32 mm 32 mm 29 mm 29 mm	2	C1654	21 mm	34 mm	16 mm	23 mm	20 mm	19 mm	
4 RPC 937 27 mm 34 mm 20 mm 22 mm 18 mm 20 mm 5 64/316 27 mm 34 mm 19 mm 22 mm 19 mm 19 mm 19 mm 6 20/519 40 mm 40 mm 32 mm 32 mm 29 mm 29 mm	3	37/1 561	24 mm	34 mm	18 mm	24 mm	20 mm	22 mm	
5 64/316 27 mm 34 mm 19 mm 22 mm 19 mm 19 mm 6 20/519 40 mm 40 mm 32 mm 32 mm 29 mm 29 mm	4	RPC 937	27 mm	34 mm	20 mm	22 mm	18 mm	20 mm	
6 20/519 40 mm 40 mm 32 mm 32 mm 29 mm 29 mm	5	64/316	27 mm	34 mm	19 mm	22 mm	19 mm	19 mm	
	6	20/519	40 mm	40 mm	32 mm	32 mm	29 mm	29 mm	
7 MNC 2175 29 mm 33 mm 25 mm 26 mm 20 mm 27 mm	7	MNC 2175	29 mm	33 mm	25 mm	26 mm	20 mm	27 mm	
8 KNC 32 27 mm 35 mm 22 mm 27 mm 22 mm 24 mm	8	KNC 32	27 mm	35 mm	22 mm	27 mm	22 mm	24 mm	
9 C1805 28 mm 31 mm 20 mm 26 mm 25 mm 21 mm	9	C1805	28 mm	31 mm	20 mm	26 mm	25 mm	21 mm	
10 330/1994 14 mm 16 mm 30 mm 12 mm 26 mm 34 mm	10	330/1994	14 mm	16 mm	30 mm	12 mm	26 mm	34 mm	
11 C2325 30 mm 35 mm 29 mm 28 mm 29 mm 28 mm	11	C2325	30 mm	35 mm	29 mm	28 mm	29 mm	28 mm	
12 C2262 28 mm 32 mm 22 mm 27 mm 22 mm 24 mm	12	C2262	28 mm	32 mm	22 mm	27 mm	22 mm	24 mm	
13 C2264 30 mm 31 mm 21 mm 28 mm 23 mm 25 mm	13	C2264	30 mm	31 mm	21 mm	28 mm	23 mm	25 mm	
14 C2321 30 mm 32 mm 27 mm 30 mm 25 mm 24 mm	14	C2321	30 mm	32 mm	27 mm	30 mm	25 mm	24 mm	
15 C386 30 mm 32 mm 23 mm 29 mm 24 mm 24 mm	15	C386	30 mm	32 mm	23 mm	29 mm	24 mm	24 mm	
16 C128 31 mm 34 mm 23 mm 27 mm 19 mm 22 mm	16	C128	31 mm	34 mm	23 mm	27 mm	19 mm	22 mm	
17 C2554 32 mm 34 mm 23 mm 27 mm 21 mm 26 mm	17	C2554	32 mm	34 mm	23 mm	27 mm	21 mm	26 mm	

Note: Resistant isolates are highlighted in red.

DISCUSSION

Studies have shown the active and synergistic interaction of azoles with Terbinafine, Amphotericin, and Echinocandins (micafungin) against Candida isolates [40-43]. Another study has reported that a combination of Terbinafine and azole resulted in a fungicidal action against C. albicans, although the drugs were fungistatic when they were used individually [44]. Studies have also shown simultaneous inhibition of different fungal cell targets, giving synergism between Echinocandins (cell wall active) and Amphotericin. Antifungal drug combinations have also been used against planktonic and sessile cells of Candida albicans [45]. The combinations showed a higher cure rate, fewer relapses, and less nephrotoxicity than monotherapy with Amphotericin [46]. Similarly, Terbinafine has also shown synergistic effects due to their mechanism of actions, when used in combination with different drugs like azoles against Candida albicans and Candida glabrata infections [47,48]. Thus, it can be concluded that combination therapy could provide an effective alternative to monotherapy for patients with invasive infections that are difficult to treat due to multi-resistant species.

Statistical analysis of combinatorial drugs at different proportions vs. monotherapy

All the results obtained from drug sensitivity analysis were statistically analyzed as described in materials and methods using Prism. The drug combinations were compared to the efficacy by individual drugs along with the three stoichiometric ratios of a combination. Mean ± Std Dev of all the combinations and individual drugs are shown in Figures 3 and 4. For all the combinations, it was observed that the combinations were much better than monotherapy; however, no significant difference was observed between stoichiometric ratios except in the case of A+T and A+M combinations. In the A+T combination, the mean was significantly different in 1:3 proportions while insignificant in other proportions as well as to individual drugs. In the A+M combination, the mean for 1:1 and 1:3 was significantly different from the individual drug, and when used in the combination of 3:1, no difference was observed. All the p values obtained for different drug and drug combinations are summarized in Figure 5. A complete analysis of sensitivity of isolates towards different drug combinations is shown in Figure 4.



Figure 4: The figure shows complete analysis of sensitivity of isolates towards different drug combinations.



Figure 5: The figure shows the graphs showing significant difference between p values of different drugs and drug combinations.

CONCLUSION

The study is focused on the drug sensitivity analysis of *Candida* isolates towards various drugs and their combinations. Four different classes of drugs used for the study belonged to different classes with a different mechanism of action (Itraconazole, Amphotericin B, Micafungin and Terbinafine). Results have shown that the monotherapy drugs were not as efficacious. Thus, combinatorial drugs were prepared and analyzed at three different stoichiometric ratios. The results from this study showed that different isolates showed maximum sensitivities to the combinatorial drugs at all proportions. Most of the combinations were statistically significant from drug monotherapy except for Amphotericin B. No statistical significance was observed between the three different proportions of drug combinations. Combination therapy could provide an alternative to monotherapy for patients with invasive infections that are difficult to treat due to the emergence of resistance.

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